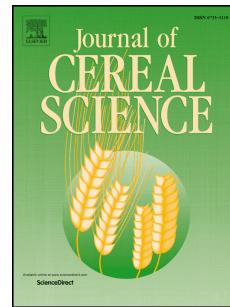


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1 **Characterization of Volatile Aroma Compounds after in-vial
2 Cooking of Foxtail Millet Porridge with Gas Chromatography
3 -Mass Spectrometry**

4
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14 **Short version of title:** Foxtail Millet Aroma Compounds

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22 **ABSTRACT:** Foxtail millet has become popular over recent years for its nutritional
23 value and ecological functions. The aroma of foxtail millet is not well characterized,

24 which is critical for its eating quality and understanding the biochemistry and
25 genetics of aroma is important for molecular breeding of millets rich in aroma. In this
26 study, the volatile aroma compounds of the elite millet variety Jingu 21 were
27 investigated at different cooking times, pH, processing methods, and compared with
28 3 other varieties. An in-vial cooking method was developed which combined solid
29 phase micro-extraction and gas chromatography-mass spectrometry for the
30 detection and identification of volatile compounds. The main findings were: a)
31 Twelve aroma compounds were identified during cooking, which were hexanal,
32 heptanal, octanal, (E)-2-heptenal, nonanal, trans-2-octenal, trans-2-nonenal,
33 2,4-nonadienal, (E,E)-2,4-decadienal, 1-octen-3-ol, 2-pentylfuran and 6-methyl-5-
34 hepten-2-one. b) Longer cooking times produced higher concentrations of aroma
35 compounds. c) Variations in cooking pH (from 6 to 8) had no obvious impact on the
36 aroma of the millet porridge. d) More volatile compounds were released from millet
37 flour compared to millet grain. e) There were significant differences among varieties
38 and Jingu 21 millet showed the highest abundance of most aroma compounds,
39 explaining partly why it is strongly favored by consumers for decades.

40 **Keywords:** Foxtail Millet; Volatile Aroma Compounds; In-vial Cooking; Gas
41 Chromatography-Mass Spectrometry

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46 **1. Introduction**

47 Foxtail millet (*Setaria italica*), one of the oldest cereal crops, has been grown in

48 China for thousands of years, and is now planted in India, North Korea, North Africa,
49 the United States and elsewhere (Lata et al., 2013). It is a drought-tolerant crop
50 adapted to arid or semi-arid areas and has a wide distribution in the northern region
51 of China. Foxtail millet has high utilization value, as seeds can be used for food, bran
52 for stock feed, and stalks for fuels or materials (Pantet al., 2016).

53 Millet food products such as millet wine, vinegar, crisps, porridge and infant
54 formula powder, are high in protein, vitamins, fatty acids, amino acids and some vital
55 nutrients which also have potential function to restrain type 2 diabetes (Song and
56 Gao, 2005; Ren et al., 2016). Millet flour and bran are also rich in antioxidant
57 components that help to prevent oxidative stress and may reduce free radical
58 damage in humans (Suma and Urooj, 2012). For these reasons, it can be expected
59 that consumer demand for millet will increase, and cultivation of high quality
60 varieties of foxtail millet will likely become a new and growing trend in agriculture.

61 The quality of foxtail millet is measured in three ways: nutritional quality,
62 physical appearance and eating quality (He et al., 2015). In Chinese traditional
63 cooking, millet is mainly used to make porridge. The flavour, colour, stickiness and
64 viscosity of porridge are the key features for evaluating the quality of foxtail millet.
65 Among these, aroma is critical for consumers and volatile compounds are vital for the
66 flavour characteristics of millet porridge. However, the millet aroma is quite subtle,
67 and not easy to measure and quantify during cooking, and there have been only a
68 few published studies addressing the flavour of millet porridge.

69 With gas chromatography-mass spectrometry (GC-MS) now in common use,
70 detection and analysis of volatiles produced upon cooking foxtail millet is possible.
71 An alternative way to detect volatile compounds in foxtail millet porridge is to use

72 simultaneous distillation extraction (SDE) combined with GC-MS (Liu et al., 2012), but
73 aroma compounds generated during cooking are very volatile and difficult to capture
74 for accurate GC-MS analysis, and solvent extraction methods used for sampling
75 flavour might not represent the volatile aroma compounds generated during cooking.
76 Furthermore, it is challenging to prepare homogeneous replicates in small batch
77 sizes.

78 In this study, foxtail millet from Jingu 21 was selected as the test material for its
79 high quality and popularity in China (Shi et al., 2001). The aim of this study was 1) to
80 develop an in-vial cooking method as a novel approach for multiple small volume
81 sampling of large batches to minimize variation in sample processing; 2) to compare
82 volatile aroma profile in the elite foxtail millet variety, Jingu 21 millet, with other
83 three varieties, Jingu 36, Daqinggu and Zhishenggu (Zhang et al., 2016); 3) to
84 investigate the impact of cooking times, pH of cooking water and processing method
85 (whole millet grain vs. millet flour) on twelve important volatile aroma compounds
86 contributing to aroma in plant foods (Table 1). This work will provide a foundation for
87 future research to develop foxtail millet varieties with improved cooking quality.

88 **2. Materials and Methods**

89 **2.1 Samples preparation**

90 Four varieties of foxtail millet (Jingu 21, Jingu36, Daqinggu, Zhishenggu) were
91 harvested in 2014, and dehusked using a rice huller (JLGJ-45, Hangzhou HR, China).
92 Jingu 21 millet grains were milled using a grinder (DēLonghi, KG49, Germany). All
93 samples were stored at 4 °C.

94 Millet (1.5 g), ultra-pure water (Pur1te ‘Select’ DI Water System, UK) (7.5 ml)
95 and 20 µl of internal standard (5 mg) 3-heptanone (Sigma, Saint Louis, USA) in 100 ml

96 methanol (Laboratory reagent grade, Fisher Scientific, UK) were added to a sample
97 vial (20 ml, 75.5 x 22.5 mm, Fisherbrand). Prepared samples were heated in the GC
98 incubator at 100 °C. Standard cooking time was 20 min, but additional tests were
99 done at 0, 10, 20, 30, 40 min. The pH of the water was adjusted to pH6, pH7, pH8 by
100 0.1M HCl and 0.1M NaOH; Two sizes of processed millet (millet grain, millet flour)
101 were compared.

102 **2.2 Identification and quantification of volatile aroma compounds by GC -MS**

103 All the samples were extracted using solid phase micro extraction fiber
104 (50/30μm, DVB/ CAR/ PDMS). Before extracting, SPME was heated at 250 °C for 1
105 hour, then desorbed for 10 min for analysis. A GC-MS (Thermo Fisher TRACE 1300,
106 USA) with a ZB-WAX Capillary GC Column (length 30 m, inner diameter 0.25 mm, film
107 thickness 1 μm; Phenomenex Inc., Macclesfield, UK) was utilized for analysing and
108 identifying the extracted compounds. The oven program was set as follows: Initial
109 temperature was 40 °C for 2 min, then increased to 250 °C at a rate of 6 °C per min
110 and held for 3 min. The inlet temperature was set at 250 °C in splitless mode. The
111 mass spectrometer settings were: Ionization mode EI, 70 eV; Filament emission
112 current 200 μA; Temperature ion source was 200 °C; Interface temperature was
113 250°C. Full scan mode was used to detect the volatile compounds (mass range from
114 m/z 20 to 300).

115 Volatile compounds were identified and selected by matching the mass spectra
116 with the spectra of reference compounds in the NIST/EPA/NIH mass spectral library
117 version 2.0 (Faircom Corporation, U.S.) using retention time. 3-heptanone was used
118 for standard calibration for quantitative analysis.

119 **2.3 Statistical analysis**

120 Millet processing was conducted in triplicate, and the data were analysed by
121 ANOVA SPSS 17.0 package. Significant differences in measurements were determined
122 by Tukey's HSD test ($\alpha=0.05$). Principal component analysis (*The Unscrambler® X*)
123 was performed to understand factors affecting the change of volatile components
124 under different treatments.

125 **3. Results and Discussion**

126 **3.1 Identification and quantification of volatile aroma compounds**

127 Twelve volatile aroma compounds were identified in Jingu 21 millet after 20 min
128 in-vial cooking, using SPME-GC-MS. Compared to traditional in-pot cooking, which
129 involves high water loss, is time consuming and produces a large amount of waste
130 materials, the new in-vial method was fast and reproducible, involved minimal
131 human handling, reduced working time and allowed the automatic running of larger
132 batches of samples. Furthermore, the coefficient of variation of the proposed
133 method was less than 10% for most volatile aroma compounds in millet porridge
134 (Table 1).

135 The 12 volatile compounds generated a unique profile (Table 1) and were
136 predicted to play an important contribution to the flavour of the millet porridge.
137 Their production is known to be closely related to the enzymatic activity of the fatty
138 acid metabolism pathway, and lipoxygenase activities (Liavonchanka and Feussner,
139 2006; Xu and Barringer, 2009), but their relative abundance and release during
140 cooking has not previously been studied in these millet varieties. The fatty acid
141 composition of foxtail millet has previously been reported to include linoleic acid,
142 oleic acid, palmitic acid, stearic acid and arachidic acid. Among these, linoleic acid is
143 the major fatty acid. Hexanal, 2-pentylfuran, (E)-2-heptenal, (E,E)-2,4-decadienal and

144 1-octen-3-ol are commonly derived from linoleic acid; heptanal, octanal and nonanal
145 were formed by the hydroperoxide degradation of oleic acid. 6-methyl
146 -5-heptene-2-one (Table 1) was probably derived from the oxidative cleavage of
147 acyclic carotenoid, as shown to be important in cooked rice (Widjaja et al., 1996).
148 The result indicate that aldehydes may be important in the formation of the aroma of
149 millet porridge (Liu et al., 2012). This finding could help to study further on the
150 implications of fatty acid metabolism and eating quality in foxtail millet.

151 **3.2 Impact of different millet varieties**

152 Foxtail millet germplasm resources are rich and diverse in China with more than
153 26,670 landraces of foxtail millet collected to date (Wang et al., 2012). We selected
154 four varieties of foxtail millet that are commonly accepted as being of good eating
155 quality, Jinggu 21, Jinggu 36, Zhishenggu and Daqinggu. Jinggu 21 millet is one of the
156 most popular varieties in China, due to its high eating quality and good nutritional
157 value.

158 The volatile aroma compounds produced in different varieties of millet were
159 compared using a principal component analysis (PCA). The PCA results showed that a
160 total of 80% of the variance could be explained in the first two dimensions (Fig. 1a).
161 Along PC1, the 4 varieties were separated into different groups. Jinggu 21 millet was
162 clustered on the right side with the highest level of most aldehydes: hexanal,
163 2-pentylfuran, (E)-2-heptenal, 6-methyl-5-heptene-2-one, trans-2-octenal, 1-octen-3
164 -ol, trans-2-nonenal, 2,4-nonadienal and (E,E)-2,4-decadienal, and the Daqinggu
165 millet was clustered on the left side and was associated with octanal, nonanal and
166 heptanal. Jinggu 36 and Zhishenggu millets were clustered together and located in
167 the middle. Along PC2, Daqinggu was clustered on the top left quadrant, whereas

168 Jingu 21 clustered in the bottom right.

169 The results showed that Jingu 21 millet had more abundant volatile aroma
170 compounds than the other varieties tested. Most of the aroma compounds in Jingu
171 21 were present at a higher level than the other three varieties with no significant
172 difference ($p < 0.05$) except for trans-2-octenal, 6-methyl-5-hepten-2-one (Figure. 1b).

173 Trans-2-octenal in Jingu 21 was significantly higher than Daqinggu and Jingu 36
174 ($p < 0.05$). 6-methyl-5-hepten-2-one in Jingu 21 millet was present at a higher
175 concentration compared to the other three varieties ($p < 0.05$) (Fig. 1b). Octenal,
176 nonanal, heptanal in Daqinggu were in a higher level than other varieties. ANOVA
177 showed that octenal in Daqinggu was significant difference with Jingu 36 ($p < 0.05$),
178 but not significant difference with Jingu 21 and Zhishenggu ($p < 0.05$). Nonanal in
179 Daqinggu was both higher and significant difference than other varieties ($p < 0.05$).
180 Heptanal in Daqinggu was higher and significant difference than Jingu 36 and
181 Zhishenggu, but not significant difference with Jingu 21 (Fig. 1b).

182 Jingu 21 produced more aroma compounds than the other three varieties,
183 whereas Daqinggu contained a higher level of heptenal, octanal and nonanal. The
184 difference in the profiles may arise from their different contents of protein, fatty acid
185 and amino acids, which under heat-treatment may undergo maillard reaction, stecker
186 degradation and lipid oxidation, thereby producing different levels of heterocyclic
187 compounds, aldehydes, pyrazine, hydrocarbons and other aroma compounds (Smith
188 and Barringer, 2014). Based on data for Jingu 21, Jingu 36 and Daqinggu (Zhishenggu
189 not available) from the Chinese Crop Germplasm Resources Information System
190 (CCGRIS), Jingu 21 does show higher protein (15.12%) and fat (5.76%) contents than
191 the other varieties (Jingu 36 13.38%, 4.29% and Daqinggu 10.74%, 4.92%). The study

192 showed the flavour of millet may not be determined by a single aroma compound
193 but is rather the result of a combination of several compounds, similar to the findings
194 with rice aroma by Widjaja et al. (1996). The flavour of millet can also be influenced
195 by other factors, such as the ecological environment, the weather and the soil quality
196 where the crop is grown (Minet al., 2005). In this study, we only chose four varieties
197 of foxtail millet for primary testing, but the in-vial method will make it possible to
198 carry out extensive further research by which the effect of different genetic
199 backgrounds and geographical conditions on aroma profiles could be assessed.

200 **3.3 Impact of cooking time**

201 Cooking quality is an important index for assessing foxtail millet. It is impacted
202 by the cooking time, the gelatinization temperature of millet starch, the content of
203 amylase and amylopectin (He et al., 2015). Usually, cooking time influences the
204 stickiness and viscosity of millet porridge, but there is no information on the
205 relationship between volatile aroma compounds and cooking time. A range of
206 cooking times (0, 10, 20, 30, 40 min) was tested for aroma profiles, with the
207 traditional cooking time (20min), which is the standard residential and commercial
208 productions and preparation in catering establishments.

209 In order to better understand the changes in aroma compounds under different
210 cooking time, a PCA chart was used, which showed 93% of the variance could be
211 explained in the first two dimensions (Fig. 2a). Along PC1, different cooking times
212 were separated into 5 groups. The longer cooking times, 20, 30 and 40 min, were
213 clustered on the right side, whereas the shorter times of 0 and 10 min were clearly
214 clustered on the left side.

215 The radar chart (Figure 2b) showed hexanal, heptenal, 2-pentylfuran, octanal,

216 (E)-2-heptenal, nonanal, trans-2-octenal, 1-octen-3-ol, trans-2-nonenal, 2,4-
217 nonadienal and (E,E)-2,4-decadienal increased significantly in concentration from 0 to
218 40 min. Concentrations of most of the volatile aroma compounds were significantly
219 different after different cooking times ($p<0.05$), with the exception of 6-methyl-5-
220 heptene-2-one (Fig. 2b).

221 The higher concentrations of volatile compounds found after extended cooking
222 times maybe due to extend chemical reactions such as lipid oxidation and maillard
223 reactions. It should be noted that the millet samples were prepared in a sealed vial
224 from which the gas could not easily escape to the atmosphere, and with a longer
225 cooking time, the concentration of volatile compounds in the vial was an accurate
226 reflection of the total volatiles produced. These results were similar to those for rice,
227 with longer cooking time releasing more flavour and increasing stickyness
228 (Champagne et al., 2008).

229 **3.4 Impact of different pH**

230 The pH of tap water in different areas of China varies and might influence the
231 cooking quality of millet. Normally, alkalinizing agents are used to increase the pH and
232 retain the colour of certain foods (Andrés-Bello et al., 2013) and sodium bicarbonate,
233 is commonly added to the pot during the cooking process for millet porridge by local
234 people to shorten the cooking time and to promote a good texture of millet porridge.
235 However, it is not known whether or not the alkaline condition of water influences
236 the generation of flavour aroma compounds. Thus, in this study, we set a pH range as
237 6, 7 and 8.

238 The PCA chart showed that 88% of the variance could be explained in the first
239 two dimensions (Fig. 3a). However, the pH zone and the volatile compounds were

240 not clustered clearly in dimensions in the PCA chart. Also, the radar chart showed no
241 general trends and ANOVA showed no major significant difference for the 12 volatile
242 aroma compounds tested (Fig. 3b), which suggests that the pH of the cooking water
243 had little or no impact on the production of volatile aroma compounds.

244 It is known, however, that the pH value can influences the colour and texture of
245 cooked millet during cooking, and alkaline conditions ($\text{pH}>8$) could help to prevent
246 colour and texture loss due to its effect on the protein properties of food, enzymatic
247 activities, gelification, and possibly chemistry (Shen et al., 2015).

248 **3.5 Impact of whole grain and ground flour**

249 Millet flour is important for making and developing processed food products,
250 and flavour is critical for the quality assessment of millet because it closely affects
251 consumer preference. Thus, we studied the extractability of flavour components
252 from flour and grain of Jingu 21 millet after cooking for 20 min and observed the
253 volatile changes.

254 The radar chart (Fig. 4) clearly shows higher production of flavour components
255 from flour compared with grain. Almost every flavour components was significantly
256 different comparing grain and flour ($p<0.05$), and hexanal, heptanal, 2-pentylfuran,
257 octanal, (E)-2-heptenal, 6-methyl-5-heptene-2-one, trans-2-octenal, 1-octen-3-ol,
258 2,4-nonadienal, but not nonanal, trans-2-nonenal, (E,E)-2,4-decadienal, were higher
259 using flour. It is assumed that this is related to fact that the particle size of millet flour
260 is substantially smaller than millet grains and the increased surface area of millet
261 leads to enhanced production or release of volatile compounds from flour. Similar
262 results were found in a previous study by Bhumiratana et al. (2011) in coffee, where
263 the ground coffee released more aroma compounds than coffee beans due to the

264 processing of grinding, which helped to expose and release the volatile aroma
265 compounds.

266 **4. Conclusion**

267 In this study, we developed an in-vial cooking method for the automatic cooking
268 and detection of volatile aroma compounds in the foxtail millet, obtained a
269 preliminary understanding of the volatile aroma compounds in millet porridge by
270 observing the changes of volatiles with different treatment methods. Among the 12
271 volatile aroma compounds selected, aldehydes played an important role in the aroma
272 profile of the millet varieties analysed. During extended cooking (40min), a higher
273 concentration of aroma compounds was produced. The pH of cooking water had no
274 impact on the volatile compounds. When millet grain was milled to flour, higher
275 concentrations of aroma compounds were released than for the whole grain.
276 Comparisons among four varieties of millet showed most of the aroma compounds
277 were at a higher level in Jingu 21. Secondary products from lipid oxidation were
278 elevated in the Daqinggu variety. Additional research on sensory evaluation will be
279 helpful to improve the quality of foxtail millet in the future. Improved understanding
280 and facile measurement of the aroma components should prove useful in breeding
281 programs to improve these attributes.

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Table 1. Volatile aroma compounds identified from Jingu 21 millet porridge

RT	Compound and+Cas#	Odor Description	References	Coefficient of Variation in Present in-vial Experiments
Alkanals				
4.45	Hexanal (66-25-1)	Green, grass-like	Lei & Boatright, 2008	7.1%
6.5	Heptanal (111-71-1)	Fruity, fatty	Lee, Xiao, Zhang, Ebeler & Mitchell, 2014	5.9%
8.82	Octanal (124-13-0)	Green, citrus-like	Park & Drake, 2016	3.9%
9.56	(E)-2-Heptenal (18829-55-5)	Herbaceous	Globisch, Schindler, Kressler & Henle, 2014 Franklin, Chapman, King, Mau, Huang & Mitchell, 2017.	5.9%
11.2	Nonanal (124-19-6)	Soapy, citrus-like		4.0%

11.95	Trans-2-octenal (2548-87-0)	Green, fatty	Shi, Li, Wang, Zhang, Qiu, Han, Wang, Chang & Guo, 2015.	8.6%
14.29	Trans-2-nonenal (18829-56-6)	Fatty, tallowy	Kaneko, Kumazawa & Ni-shimura, 2001	10.3%
17.72	2,4-Nonadienal (6750-03-4)	Fatty, beany	Park & Drake, 2016	18.3%
19.84	(E,E)-2,4-Decadienal (25152-84-5)	Fatty, waxy	Pan, Huang, Hsu, Lee, Liu, Cheng, Tsai, Shen & Lin, 2014.	22.0%
Phenols and Alcohols				
12.55	1-Octen-3-ol (3391-86-4)	Green, beany	Sugawara, Ito, Odagiri, Kubota & Kobayashi, 2014	5.2%
Heterocycles				
7.54	2-Pentylfuran (3777-69-3)	Beany	Pripdeevech, Moonggoot, Popluechai & Chukeatirote, 2014	7.0%
9.93	6-Methyl-5-hepten-2-one (110-93-0)	Banana-like, floral	Christensen, Edelenbos & Kreutzmann, 2007	3.9%

Volatile compounds were identified after 20 min cooking by GC-MS retention time (RT) compared with internal standard compounds.

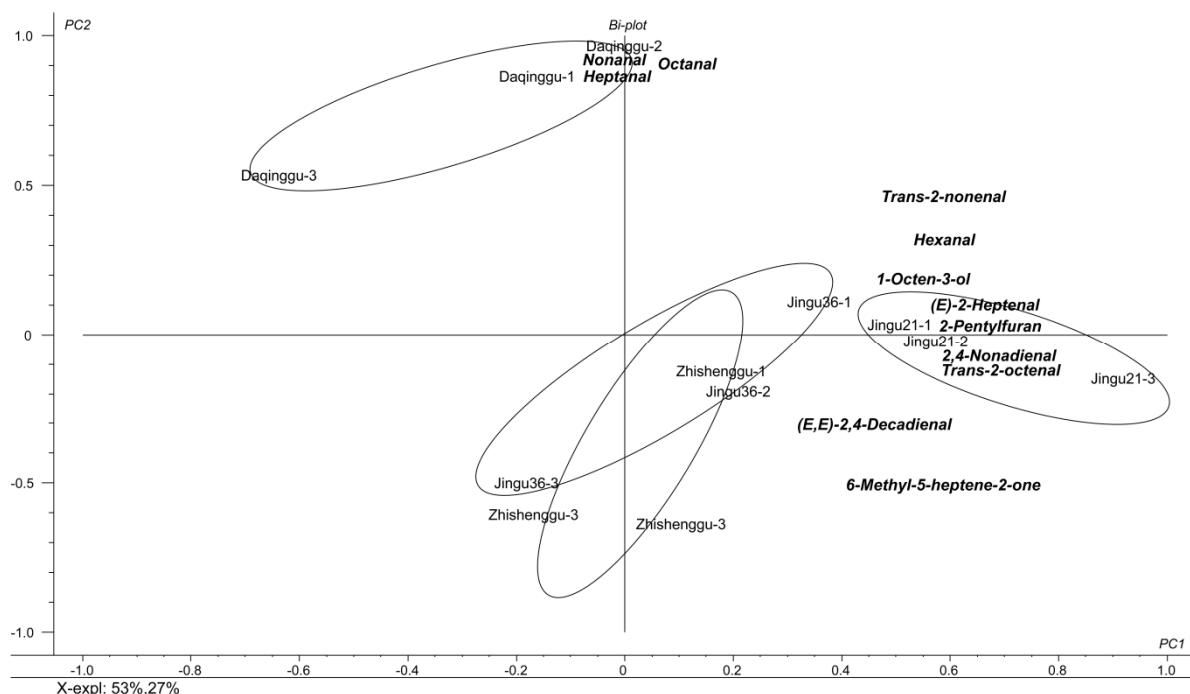


Fig. 1a. PCA analysis of aroma components for 4 varieties of millet

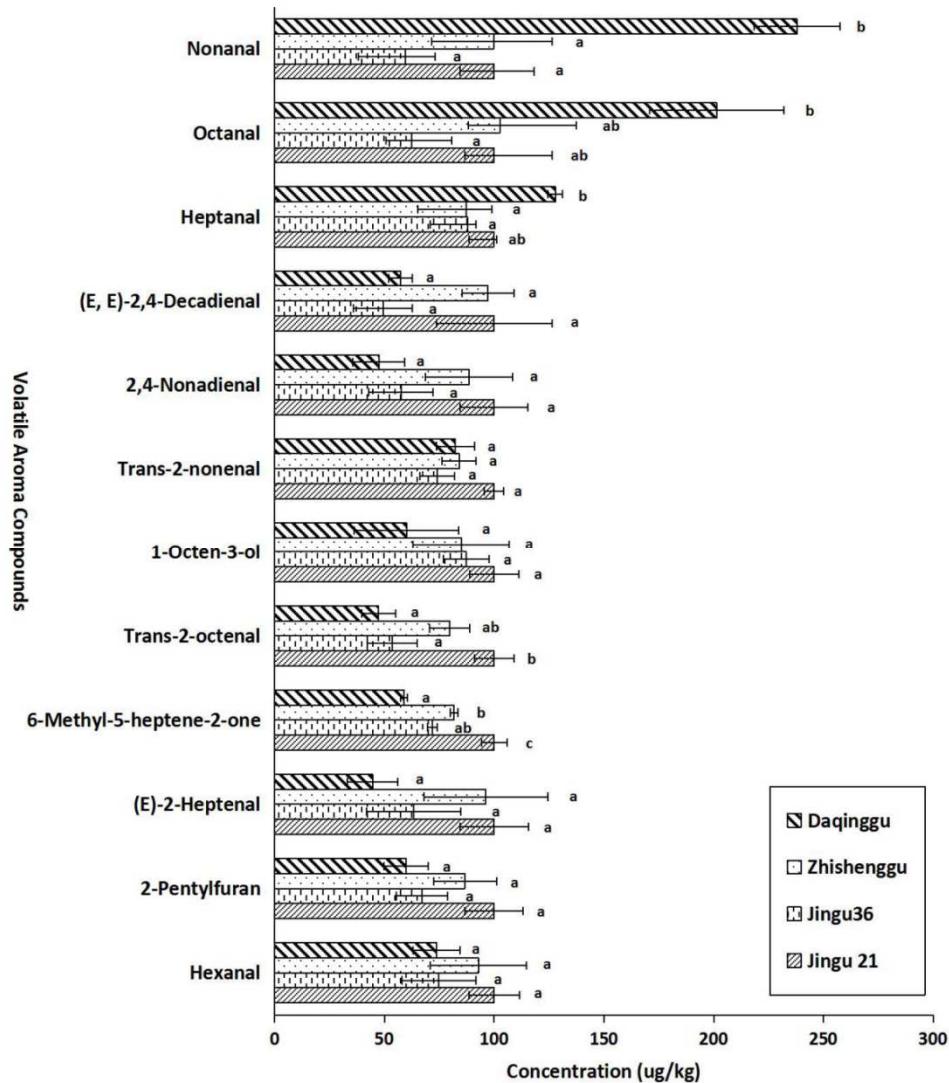


Fig. 1b. Comparison of volatile aroma compounds from four varieties of millet. Cooking time was 20 minutes. Different letters indicate significant differences between varieties ($p<0.05$). Jingu 21 millet values were normalized to 100%.

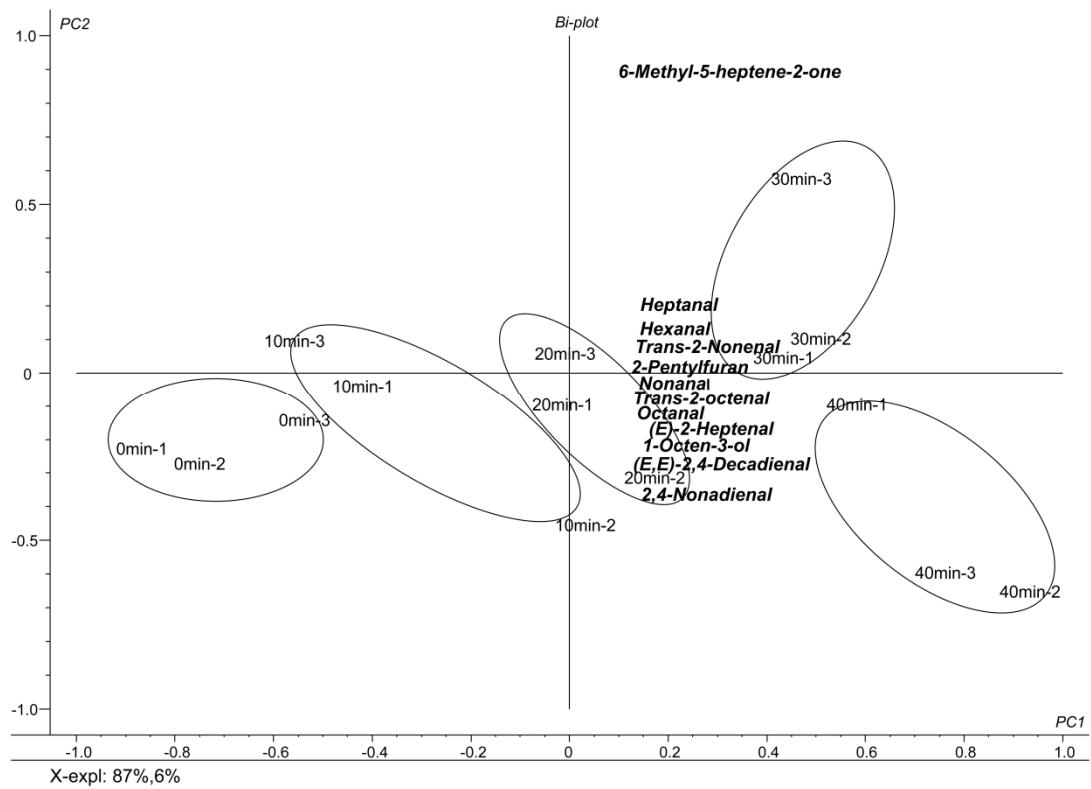


Fig. 2a. PCA analysis of aroma components of millet Jingu 21 produced after different cooking times

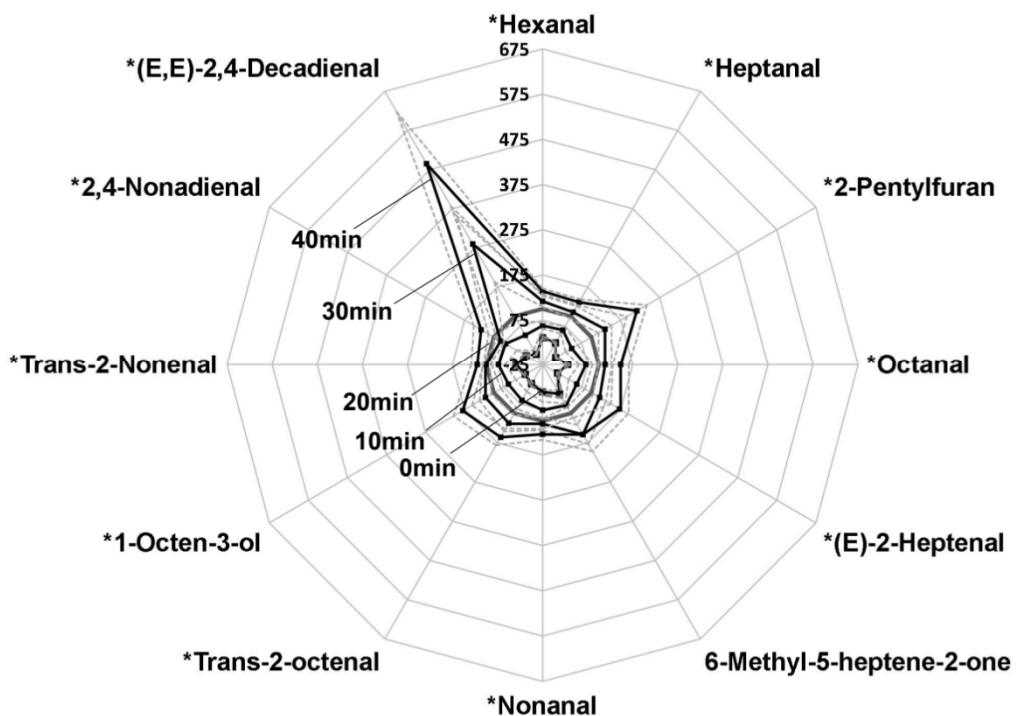


Fig. 2b. Effect of cooking times on volatile production of millet Jingu 21. Aroma profiles generated after 20 minutes (standard cooking time) compared with 0, 10, 20, 30, 40min. with +/- standard errors showed by grey dotted lines. * indicates significant differences for each compound ($p=0.05$).

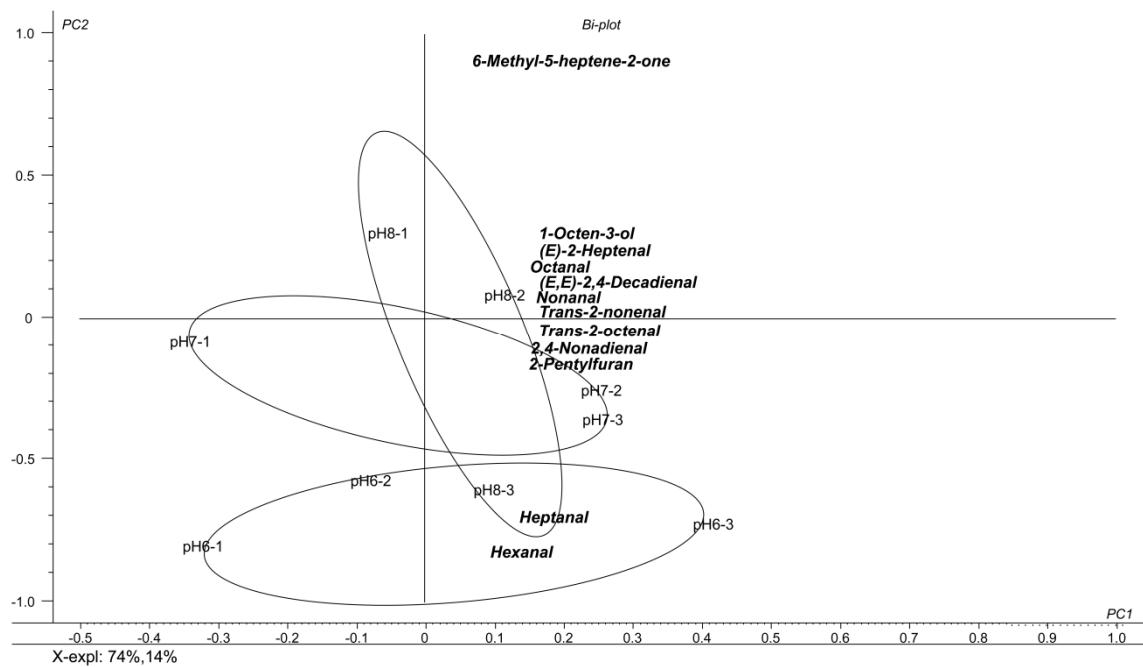


Fig. 3a. PCA analysis showing poor correlation between different cooking pH and aroma compound production of millet Jingu 21.

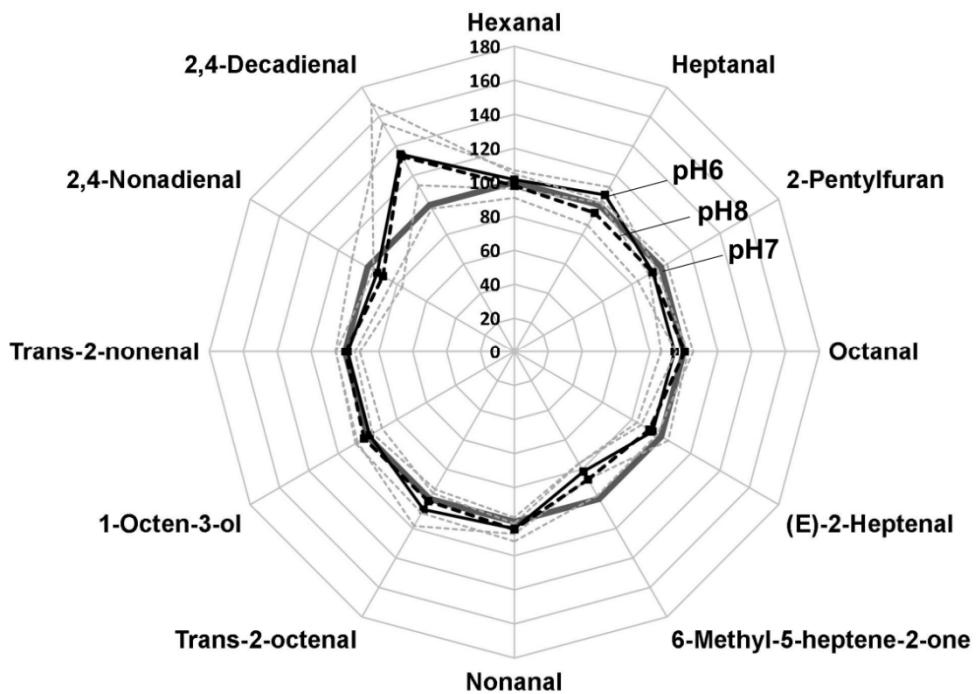


Fig. 3b. Aroma volatiles production of millet Jingu 21 at different pH. Aroma profile produced at the standard pH 7 (gray smooth circle, set at 100%). Average values of volatiles produced at pH 6 (black line), 7 (gray line) and 8 (dashed line) with +/- standard errors showed by grey dotted lines.

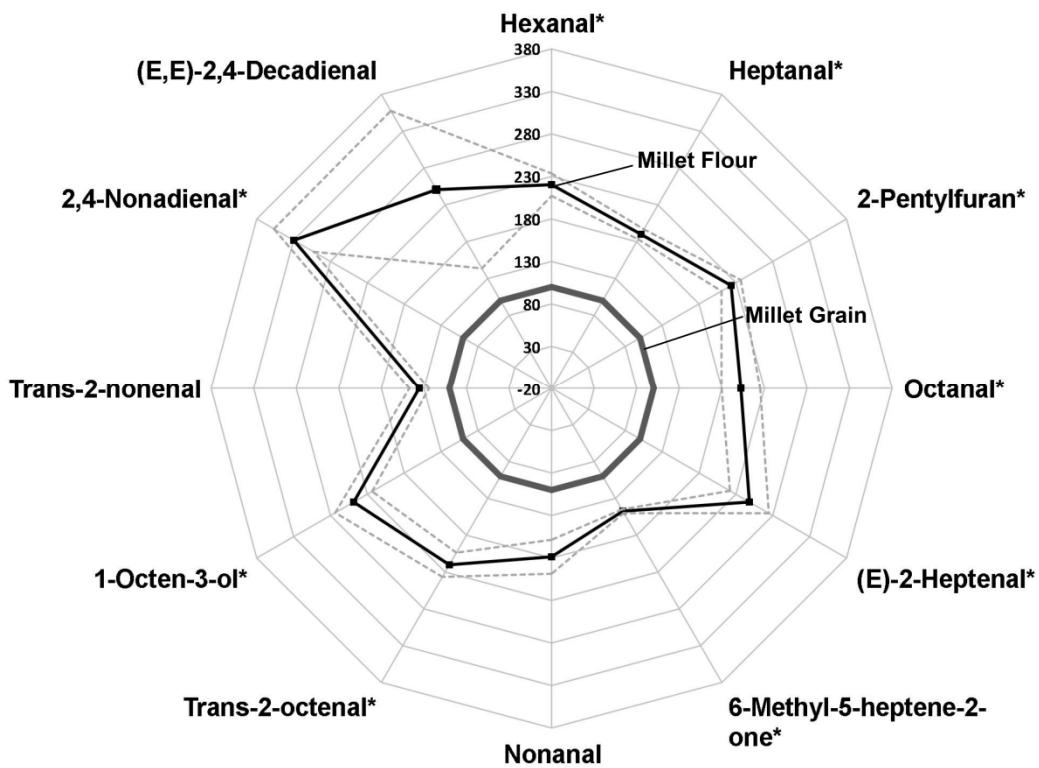


Fig. 4. Comparison of aroma profiles between flour and grain of millet Jingu 21. Aroma profile of the millet grain (smooth circle at 100%), millet flour (average value showed in black solid with square markers, with +/- standard errors showed by grey dotted lines). * indicate significant difference ($p=0.05$).

Research Highlight

- An in-vial cooking was developed to analyse aroma compounds in foxtail millet
- Within 40 min, longer cooking times result in higher levels of volatiles
- More aroma compounds were released from millet flour than grain during cooking
- Popular variety Jingu21 released higher levels of aroma compounds than the others