1	The placental renin-angiotensin system and oxidative stress in pre-eclampsia		
2	Hiten D. Mistry ^{1*} , Lesia O. Kurlak ^{2*} & Fiona Broughton Pipkin ²		
3	¹ Division of Women's Health, King's College London, Women's Health Academic Centre		
4	KHP, St. Thomas' Hospital, London, UK		
5	² Division of Obstetrics & Gynaecology, School of Clinical Sciences, University of		
6	Nottingham, City Hospital, Nottingham, UK		
7	* Authors made equal contribution		
8	Corresponding author's address:	Dr. Hiten D. Mistry	
9		Division of Women's Health	
10		Women's Health Academic Centre, KHP	
11		King's College London	
12		10th Floor, North Wing	
13		St Thomas' Hospital	
14		Westminster Bridge Road	
15		London, SE1 7EH	
16		Tel: +44 (0) 207 188 8151	
17		Fax: +44 (0) 207 620 1227	
18			
19			
20	Short Tital: Placental RAS in Pre-eclampsia		
21	Sources of Funding: Part of this work was funded by the Biotechnology and Biological		
22	Research Council (BBSRC) BBS/S/P/2003/10412A.		
23	Conflict of Interest: No conflict of interest for all authors.		
24			
25	Abstract		

There is an inverse correlation between human birthweight and umbilical venous angiotensin
II (AngII) concentrations. Oxidative stress and increased prorenin receptor (PRR) both
enhance the cleavage of angiotensin I from angiotensinogen (AGT). Pre-eclampsia, a
hypertensive disorder of pregnancy, manifests as high blood pressure and proteinuria, and is a
state of increased oxidative stress.

Objectives, study design and main outcome measures: Hypothesis: Pre-eclampsia will be associated with increased placental expression of components of the renin-angiotensin system, which could result in reduced infant birthweight. Biopsies were taken 1cm from the placental edge from 27 normotensive controls and 23 pre-eclamptic White European women. Immunohistochemistry was performed for AGT, PRR, glutathione peroxidase 3 (GPx3) and the AT1R and AT2R AngII receptors. Protein expression was semi-quantitatively assessed (H-score).

38 **Results:** AT1R expression was significantly increased in pre-eclamptic placentae, and negatively correlated with birthweight (r = -0.529, P = 0.009). AT1R expression was also 39 40 negatively correlated with GPx3 expression overall (r = -0.647; P = 0.005). AT2R expression positively correlated with AGT (r = 0.615, P = 0.002) in the pre-eclamptic placentae only. 41 42 Conclusions: The raised AT1R expression in pre-eclampsia, together with inadequate antioxidant protection, possibly through lower GPx activity, might enhance the 43 44 vasoconstrictor effect of locally-generated AngII, contributing to the restricted fetal growth 45 characteristic of pre-eclampsia. Conversely, the AT2R:AGT association within the pre-46 eclamptic placenta may provide a compensatory mechanism.

47

Keywords: Pre-eclampsia, Renin-angiotensin-system, angiotensin receptors, angiotensinogen,
pro-renin receptor, placenta.

50 Introduction

51 Pre-eclampsia is a hypertensive disorder of pregnancy that occurs in ~3% of all pregnancies. 52 This disease, which manifests as high blood pressure and proteinuria, is probably mediated 53 by endothelial damage and may affect multiple systems of the body and contribute to adverse 54 pregnancy outcomes including maternal death, preterm birth, intrauterine growth restriction (IUGR) and fetal death [1]. Together with other hypertensive disorders of pregnancy, pre-55 56 eclampsia is responsible for at least 60,000 maternal deaths each year [2] and increases 57 perinatal mortality 5-fold [3]. Placental and maternal systemic oxidative stress are 58 components of the syndrome [4, 5] and contribute to a generalised maternal systemic 59 inflammatory activation [6]. Placental ischemia-reperfusion injury has been implicated in excessive production of reactive oxygen species (ROS), which could cause release of 60 placental factors that mediate the inflammatory responses [7]. We have recently shown that 61 62 placental antioxidant glutathione peroxidise (GPx) activity and expression are reduced in pre-63 eclampsia [8].

64

65 The renin–angiotensin system (RAS) is known to be an important regulator of blood pressure, sodium and fluid homeostasis. In non-pregnant models, enhanced RAS activity causes 66 hypertension [9], salt retention [10], and hyperaldosteronism [11]. Although the exact 67 mechanism of pre-eclampsia remains unclear, it appears that angiotensinogen (AGT) could 68 69 play critical roles in its development [12]. AGT is the only renin substrate, and is thus a 70 major molecule in the RAS. Prorenin (PR) is the biosynthetic precursor of renin, the structure of which includes a peptide which obstructs the access of AGT to the active site of renin, 71 which binds to its receptor, prorenin receptor (PRR or ATPase H⁺ transporting lysosomal 72 73 accessory protein 2; ATP6AP2) [13]. Oxidative stress and increased PRR expression both 74 enhance the cleavage of angiotensin I (Ang I) from AGT. Hypoxaemia in the chronicallycannulated fetal lamb was associated with a doubling of circulating angiotensin II (AngII) 75

concentrations [14], and umbilical venous Ang II concentrations are higher in smaller babies
[15, 16]. AngII exerts part of its vasoconstrictor effect through the generation of ROS [17].

79 Ang II exerts a majority of its effects through two major angiotensin receptors: AT1R and 80 AT2R. These highly conserved seven-transmembrane G-protein–coupled receptors share a 34% sequence identity and have comparable affinities for Ang II [18]. Most of the actions of 81 82 Ang II, including vasoconstriction and stimulation of aldosterone synthesis, angiogenesis and cell growth are mediated by the AT1R [17]. The AT2R is implicated in apoptosis, reduction 83 84 in endothelial cell growth and migration, and vasodilation in the adult, the reduction in neointima formation after vascular injury [19-23], although it is usually expressed at low 85 density. However, expression is much higher during fetal life, where it may counterbalance 86 87 the effects of the AT1Rs during fetal development. Endothelial cell growth, migration, angiogenesis and apoptosis allowing spiral artery remodelling are all central to placentation. 88 89

In the placenta, an intrinsic angiotensin-generating system has been well documented based 90 91 on the presence of major RAS components including renin, AGT, angiotensin-converting 92 enzyme (ACE), and Ang II as well as its receptor subtypes [24-26]. The conversion of Ang I to Ang II by ACE in the feto-placental bed leads to a potent vasoconstrictor activity in the 93 94 feto-placental circulation [27, 28]. Placental trophoblasts are particularly AT1R rich [29]. A 95 functioning placental RAS appears necessary for an uncomplicated pregnancy [30]. There is an inverse correlation between human birthweight and umbilical venous AngII concentrations 96 [31]. An elevated fetal plasma renin activity [32] and fetal Ang II concentration [16] have 97 98 also been reported in IUGR. However, there is controversy about the expression of placental 99 angiotensin receptors in pre-eclampsia.

100	These early studies did not examine the placental RAS in detail. A recent paper has reported
101	on placental expression of PRR and AGT in normal term pregnancy [33], but these have not
102	been described in relation to pre-eclampsia. We thus hypothesised that pre-eclampsia, a state
103	of increased oxidative stress, will be associated with increased placental expression of
104	components of the RAS, which could result in reduced infant birthweight.
105	
106	
107	
108	
109	
110	
111	
112	
113	
114	
115	
116	
117	
118	
119	
120	
121	
122	
123	
124	

125 Methods

126 Subjects:

127 The study population consisted of White European women who had either a normotensive or 128 pre-eclamptic pregnancy as previously described [4]. The investigations were approved by the Nottingham Hospital Ethics Committee and written, informed consent was obtained from 129 130 each subject. Cases were defined on admission with a clinical diagnosis of pre-eclampsia, 131 defined using the International Study for the Study of Hypertension in Pregnancy guidelines 132 [34]. Medical and obstetric histories, including delivery data, were obtained for each woman. 133 The birthweight centile for each baby was computed, correcting for gestation age, sex, 134 maternal parity and body mass index (BMI) [35]. 135 136 Sample collection: 137 Full depth placental biopsies were taken 1cm from the placental edge from 27 normotensive 138 and 23 pre-eclamptic women, avoiding placental infarcts. Tissues were taken within 10 min 139 of delivery, membranes removed and tissue washed in ice cold $1 \times PBS$ to remove maternal blood contamination. Biopsies were formalin fixed and wax-embedded for 140 141 immunohistochemical analysis. 142 143 Immunohistochemistry: 144 Immunohistochemical staining was performed on serial sections as previously described [8] using the Dako Envision[™] visualization system (Dako, Ely, UK). Table 1 provides further 145 details on antibody dilutions used, which were optimised by performing a dilution series for 146 147 each antibody. The heat-induced epitope retrieval was achieved by heating in a citrate buffer

148 (pH 6.0) using a microwave oven for 15 min. A negative control was performed for each test

section by incubation with mouse or rabbit IgG as appropriate. Sections were dehydrated and

cleaned in ascending concentrations of alcohol and xylene before coverslips were mounted (DPX mountant, BDH). Protein expression was semi-quantitatively assessed (H-score) at x200 magnification (Nikon Eclipse II microscope) by two blinded observers as described previously [36]. Between-observer agreement for H-scoring was excellent (kappa 0.97). Maternal plasma thiobarbituric acid reactive substances (TBARS) concentrations were measured as a global measure of oxidative stress; plasma total GPx activity and placental GPx3 protein expression were measured as indices of specific antioxidant activity. These data have been previously reported [4]. Statistical analysis All tests were performed using SPSS for Windows version 19.0. Summary data are presented as median (interquartile range) as appropriate for their distribution after testing for normality using the Kolmogorov-Smirnov test. Mann-Whitney U-tests were used to test differences between normotensive and pre-eclampsia groups. Spearman's Rho correlation and Kendall's Tau ranking tests were also conducted as appropriate. The null hypothesis was rejected where *P* < 0.05.

175 **Results**

176 All women conceived naturally and carried singleton pregnancies. The demographic, 177 obstetric and pregnancy data of the 50 women (27 normotensive, 23 pre-eclampsia) who 178 participated in the study are shown in Table 2. The complete and detailed data have been previously published [4]. In summary, women in the normotensive group gave birth without 179 180 developing hypertension or proteinuria, to infants weighing >2500 g, delivered at 37 weeks 181 or later. Overall, the pre-eclamptic women all had moderate to severe disease and had lower gestational ages at delivery than the control group (P < 0.05; Table 2). No pre-eclamptic 182 183 woman had Haemolysis Elevated Liver enzymes, and Low Platelet count (HELLP). All 184 neonates from both pregnancy groups survived. 185 186 All components of the RAS that were measured in this study were found to be expressed in 187 placental tissue, mainly localised to the villous syncytiotrophoblast, with some stromal 188 staining (Figure 1). Table 3 summarises the staining intensity of AGT, PRR, AT1R, AT2R 189 and GPx3 in placentae from normotensive and pre-eclamptic women. AT1R expression was significantly increased in pre-eclamptic compared to normotensive placentae (P = 0.032; 190 191 Table 3); all other RAS components were similarly expressed in both groups. Placental GPx3 expression (Table 3) and maternal plasma total GPx activity [4, 8] were significantly reduced 192 in pre-eclampsia while TBARS were highly significantly increased (1.2 [0.6, 1.6] compared 193 to 0.45 [0.2, 0.8] μ mol/l (P < 0.001)[4]. 194 195

196 A negative association was observed between AT1R and birthweight in the pre-eclamptic

- 197 group (Figure 2; r = -0.529, *P*=0.009) but not in the normotensive group (*P* > 0.5). A
- 198 significant inverse correlation was also found overall between placental AT1R and GPx3
- 199 protein expression (Figure 3; r = 0.634; P = 0.006). AT2R expression was found to be

positively related to AGT in pre-eclampsia (r = 0.615, P = 0.002), but not control, placentae (r = 0.064, P > 0.05; Figure 4).Although staining density was diffuse throughout the villous syncytiotrophoblasts, there was also dense staining localised specifically around fetal vessels in 18 (PRR) and 8 (AGT) of the 50 placenta (Figure 1). The presence of perivascular staining for PRR and AGT was found to be associated with increased maternal TBARS (Chi-squared P = 0.001 and P = 0.033).

225 Discussion

It has been known for nearly 40 years [37] that there is a tissue-based RAS in the 226 227 uteroplacental unit, suggesting a locally acting RAS. Our localised staining of the RAS 228 components to the syncytiotrophoblast is consistent with other very recent results [38]. Our study demonstrates that the placental RAS may be involved in pre-eclampsia, with raised 229 230 AT1R expression, together with the raised AngII concentrations previously reported [15, 16], 231 contributing to the restricted fetal growth characteristically observed in this syndrome. The 232 pressor response to AngII is well known to be enhanced before pre-eclampsia is clinically 233 detectable [39] and is associated with an increased density of AT1Rs [40]. Agonist 234 autoantibodies to the AT1R are also increased in pre-eclampsia [41]. 235 236 The inverse correlation between placental AT1R and GPx3 expression (Figure 3) is particularly interesting in light of the stimulatory effect of AngII on ROS generation [17]. 237 238 The absence of adequate antioxidant protection in pre-eclampsia might enhance the 239 vasoconstrictor effect of locally-generated AngII. The specific clustering of both the PRR and AGT around the vessels, in conjunction with elevated measures of oxidative stress (TBARS), 240 241 suggests that the RAS may be contributing to the heightened state of oxidative stress in placental tissue of pre-eclamptic pregnancies. ROS superoxide anion (O_2) has been 242 243 implicated in Ang II-mediated hypertension [42] and experimental data in the non-pregnant 244 state support the concept that Ang II-mediated hypertension in pregnancy may also be due, in 245 part, to effects on the oxidative state in vascular-endothelial tissue [43]. 246 247 There is some controversy surrounding the regulation of placental AT1R in pre-eclampsia. The increase in placental AT1R expression in pre-eclampsia we observed is in agreement 248 249 with previous studies in placental [44] and decidual [17] tissue. However, two earlier studies

250 reported that the capacity and affinity of AT1R were significantly lower in placentae from 251 pregnancies complicated by pre-eclampsia and IUGR [45] and in a previous study we also 252 found no significant increase in placental AT1R expression [26]. It is now well established 253 from several studies that gradients in gene expression [46] or enzyme activity [8] exist across the placenta. In our original paper [26], results were reported from biopsies taken within 1cm 254 255 of the cord insertion. We have subsequently demonstrated an increase in placental ACE activity at the periphery [47], where sensitivity of chorionic plate arteries to Ang II is greatest 256 257 [48]. In the current investigation, we therefore studied samples collected from 1cm from the 258 periphery of the placenta. This adds further weight to the requirement for structured, rather 259 than random, sampling across the placental disc, with clear reporting of sampling site.

260

We have, for the first time, identified a clear relationship between placental AT2R and AGT expression in pre-eclamptic, but not normotensive pregnancy (Figure 4). The AT2R appears to function as an 'antagonist' of AT1R [49], and is predominantly expressed in fetal tissue [50]. Ang II binding to AT2Rs increases apoptosis, causes vasodilation and is thought to be involved in fetal tissue development [51, 52]. Thus, the strong positive associations observed in the pre-eclamptic placentae between AT2R and AGT may indicate a potential compensatory mechanism, not activated in normotensive pregnancy.

268

The human placenta possesses an autonomous RAS, allowing the local generation of Ang II and its fragments. Our data suggest that, at term, increased activity in this local system, particularly evident in the syncytiotrophoblast, may contribute to both pre-eclampsia itself and the IUGR which frequently accompanies it. We have not addressed the question of distinguishing between the effects of placentally-generated AngII and fetal systemic AngII.

274

275	Acknowledgements
276	We thank all the women who participated in the study and the midwives and doctors whose
277	support made this study possible. We are also grateful to Dr. Paula Williams (University of
278	Nottingham) for help and advice regarding immunohistochemical quantification.
279	
280	
281	
282	
283	
284	
285	
286	
287	
288	
289	
290	
291	
292	
293	
294	
295	
296	
297	
298	
299	

300 **References**

- 301 [1] Steegers EA, von Dadelszen P, Duvekot JJ and Pijnenborg R. Pre-eclampsia. Lancet.
 302 2010;376(9741):631-44.
- 303 [2] Broughton Pipkin F. Risk factors for preeclampsia. N Engl J Med. 2001;344(12):925304 6.
- 305 [3] Roberts JM and Lain KY. Recent Insights into the pathogenesis of pre-eclampsia.
- 306 Placenta. 2002;23(5):359-72.
- 307 [4] Mistry HD, Wilson V, Ramsay MM, Symonds ME and Broughton Pipkin F. Reduced
- 308 selenium concentrations and glutathione peroxidase activity in preeclamptic pregnancies.
- 309 Hypertension. 2008;52(5):881-8.
- 310 [5] Poston L. The Role of Oxidative Stress. 2004; (Ed.) Critchley H, MacLean A, Poston L
- and Walker J. London: RCOG Press.
- [6] Redman CW and Sargent IL. Pre-eclampsia, the placenta and the maternal systemic
 inflammatory response--a review. Placenta. 2003;24 Suppl A:S21-7.
- 314 [7] Hung TH and Burton GJ. Hypoxia and reoxygenation: a possible mechanism for
- 315 placental oxidative stress in preeclampsia. Taiwan J Obstet Gynecol. 2006;45(3):189-
- 316 200.
- [8] Mistry HD, Kurlak LO, Williams PJ, Ramsay MM, Symonds ME and Broughton Pipkin F.
- Differential expression and distribution of placental glutathione peroxidases 1, 3 and 4 in
- normal and preeclamptic pregnancy. Placenta. 2010;31(5):401-8.
- 320 [9] MacGregor GA, Markandu ND, Roulston JE, Jones JC and Morton JJ. Maintenance of
- 321 blood pressure by the renin-angiotensin system in normal man. Nature.
- 322 1981;291(5813):329-31.
- 323 [10] Hall JE, Mizelle HL, Brands MW and Hildebrandt DA. Pressure natriuresis and
- 324 angiotensin II in reduced kidney mass, salt-induced hypertension. Am J Physiol.
- 325 1992;262(1 Pt 2):R61-71.
- 326 [11] Gordon RD, Klemm SA, Tunny TJ and Stowasser M. Primary aldosteronism:
- hypertension with a genetic basis. Lancet. 1992;340(8812):159-61.

- 328 [12] Zhou A, Carrell RW, Murphy MP, Wei Z, Yan Y, Stanley PL, Stein PE, Broughton
- 329 Pipkin F and Read RJ. A redox switch in angiotensinogen modulates angiotensin release.

330 Nature. 2010;468(7320):108-11.

- 331 [13] Danser AH and Deinum J. Renin, prorenin and the putative (pro)renin receptor. J
- Renin Angiotensin Aldosterone Syst. 2005;6(3):163-5.
- 333 [14] Broughton Pipkin F, Lumbers ER and Mott JC. Factors influencing plasma renin and
- angiotensin II in the conscious pregnant ewe and its foetus. J Physiol. 1974;243(3):619-
- 335 36.
- 336 [15] Broughton Pipkin F and Symonds EM. Factors affecting angiotensin II
- concentrations in the human infant at birth. Clin Sci Mol Med. 1977;52(5):449-56.
- 338 [16] Kingdom JC, McQueen J, Connell JM and Whittle MJ. Fetal angiotensin II levels and
- 339 vascular (type I) angiotensin receptors in pregnancies complicated by intrauterine
- growth retardation. Br J Obstet Gynaecol. 1993;100(5):476-82.
- 341 [17] Higuchi S, Ohtsu H, Suzuki H, Shirai H, Frank GD and Eguchi S. Angiotensin II
- 342 signal transduction through the AT1 receptor: novel insights into mechanisms and
- 343 pathophysiology. Clin Sci (Lond). 2007;112(8):417-28.
- 344 [18] Chung O, Kuhl H, Stoll M and Unger T. Physiological and pharmacological
- implications of AT1 versus AT2 receptors. Kidney Int Suppl. 1998;67:S95-9.
- 346 [19] Nguyen Dinh Cat A and Touyz RM. A new look at the renin-angiotensin system--
- focusing on the vascular system. Peptides. 2011;32(10):2141-50.
- 348 [20] Csikos T, Chung O and Unger T. Receptors and their classification: focus on
- angiotensin II and the AT2 receptor. J Hum Hypertens. 1998;12(5):311-8.
- 350 [21] Arima S, Endo Y, Yaoita H, Omata K, Ogawa S, Tsunoda K, Abe M, Takeuchi K, Abe
- 351 K and Ito S. Possible role of P-450 metabolite of arachidonic acid in vasodilator
- 352 mechanism of angiotensin II type 2 receptor in the isolated microperfused rabbit afferent
- 353 arteriole. J Clin Invest. 1997;100(11):2816-23.
- [22] Janiak P, Pillon A, Prost JF and Vilaine JP. Role of angiotensin subtype 2 receptor in
- neointima formation after vascular injury. Hypertension. 1992;20(6):737-45.

- 356 [23] Haberl RL, Anneser F, Villringer A and Einhaupl KM. Angiotensin II induces
- endothelium-dependent vasodilation of rat cerebral arterioles. Am J Physiol. 1990;258(6
 Pt 2):H1840-6.
- 359 [24] Cooper AC, Robinson G, Vinson GP, Cheung WT and Broughton Pipkin F. The
- 360 localization and expression of the renin-angiotensin system in the human placenta
- 361 throughout pregnancy. Placenta. 1999;20(5-6):467-74.
- 362 [25] Poisner AM. The human placental renin-angiotensin system. Front Neuroendocrinol.
 363 1998;19(3):232-52.
- 364 [26] Williams PJ, Mistry HD, Innes BA, Bulmer JN and Broughton Pipkin F. Expression of
- 365 AT1R, AT2R and AT4R and their roles in extravillous trophoblast invasion in the human.
- 366 Placenta. 2010;31(5):448-55.
- 367 [27] Hosokawa T, Howard RB and Maguire MH. Conversion of angiotensin I to
- angiotensin II in the human foetoplacental vascular bed. Br J Pharmacol.
- 369 1985;84(1):237-41.
- 370 [28] Adamson SL, Morrow RJ, Bull SB and Langille BL. Vasomotor responses of the
- umbilical circulation in fetal sheep. Am J Physiol. 1989;256(5 Pt 2):R1056-62.
- 372 [29] Shibata E, Powers RW, Rajakumar A, von Versen-Hoynck F, Gallaher MJ, Lykins DL,
- 373 Roberts JM and Hubel CA. Angiotensin II decreases system A amino acid transporter
- 374 activity in human placental villous fragments through AT1 receptor activation. Am J
- 375 Physiol Endocrinol Metab. 2006;291(5):E1009-16.
- 376 [30] Irani RA and Xia Y. Renin angiotensin signaling in normal pregnancy and
- 377 preeclampsia. Semin Nephrol. 2011;31(1):47-58.
- 378 [31] Broughton Pipkin F and Symonds EM. Factors affecting angiotensin II
- 379 concentrations in the human infant at birth. Clin Sci Mol Med. 1977;52(5):449-56.
- 380 [32] Weiner CP and Robillard JE. Atrial natriuretic factor, digoxin-like immunoreactive
- 381 substance, norepinephrine, epinephrine, and plasma renin activity in human fetuses and
- their alteration by fetal disease. Am J Obstet Gynecol. 1988;159(6):1353-60.

- [33] Pringle KG, Tadros MA, Callister RJ and Lumbers ER. The expression and localization
 of the human placental prorenin/renin-angiotensin system throughout pregnancy: roles
 in trophoblast invasion and angiogenesis? Placenta. 2011;32(12):956-62.
- 386 [34] Brown MA, Lindheimer MD, de Swiet M, Van Assche A and Moutquin JM. The
- 387 classification and diagnosis of the hypertensive disorders of pregnancy: statement from
- the International Society for the Study of Hypertension in Pregnancy (ISSHP). Hypertens
 Pregnancy. 2001;20(1):IX-XIV.
- [35] Gardosi J and Francis A. Customised Centile Calculator. 2006. Gestational network GROW centile v 5.1.
- 392 [36] McCarthy KSJ, Miller LS, Cox EB, Konrath J and McCarthy KSS. Estrogen receptor
- 393 analyses. Correlation of biochemical and immunohistochemical methods using
- 394 monoclonal antireceptor antibodies. Archives of Pathology and Laboratory Medicine
- 395 1985;109:716-21.
- 396 [37] Skinner SL, Lumbers ER and Symonds EM. Renin concentration in human fetal and
 397 maternal tissues. Am J Obstet Gynecol. 1968;101(4):529-33.
- 398 [38] Marques FZ, Pringle KG, Conquest A, Hirst JJ, Markus MA, Sarris M, Zakar T, Morris
 399 BJ and Lumbers ER. Molecular characterization of renin-angiotensin system components
 400 in human intrauterine tissues and fetal membranes from vaginal delivery and cesarean
- 401 section. Placenta. 2011;32(3):214-21.
- 402 [39] Gant NF, Daley GL, Chand S, Whalley PJ and MacDonald PC. A study of angiotensin
 403 II pressor response throughout primigravid pregnancy. J Clin Invest. 1973;52(11):2682-
- 404 9.
- 405 [40] Morgan L, Crawshaw S, Baker PN, Edwards R, Broughton Pipkin F and Kalsheker N.
- 406 Functional and genetic studies of the angiotensin II type 1 receptor in pre-eclamptic and
- 407 normotensive pregnant women. J Hypertens. 1997;15(12 Pt 1):1389-96.
- 408 [41] Herse F, Staff AC, Hering L, Muller DN, Luft FC and Dechend R. AT1-receptor
- 409 autoantibodies and uteroplacental RAS in pregnancy and pre-eclampsia. J Mol Med
- 410 (Berl). 2008;86(6):697-703.

411 [42] McIntyre M, Bohr DF and Dominiczak AF. Endothelial function in hypertension: the

role of superoxide anion. Hypertension. 1999;34(4 Pt 1):539-45.

413 [43] Shah DM. Role of the renin-angiotensin system in the pathogenesis of

414 preeclampsia. Am J Physiol Renal Physiol. 2005;288(4):F614-25.

- 415 [44] Leung PS, Tsai SJ, Wallukat G, Leung TN and Lau TK. The upregulation of
- 416 angiotensin II receptor AT(1) in human preeclamptic placenta. Mol Cell Endocrinol.

417 2001;184(1-2):95-102.

418 [45] Li X, Shams M, Zhu J, Khalig A, Wilkes M, Whittle M, Barnes N and Ahmed A.

419 Cellular localization of AT1 receptor mRNA and protein in normal placenta and its

420 reduced expression in intrauterine growth restriction. Angiotensin II stimulates the

421 release of vasorelaxants. J Clin Invest. 1998;101(2):442-54.

- 422 [46] Tzschoppe AA, Struwe E, Dorr HG, Goecke TW, Beckmann MW, Schild RL and
- 423 Dotsch J. Differences in gene expression dependent on sampling site in placental tissue
- 424 of fetuses with intrauterine growth restriction. Placenta. 2010;31(3):178-85.

425 [47] Brameld JM, Hold R and Broughton Pipkin F. Regional variation in angiotensin

426 converting enzyme activity in the human placenta. Placenta. 2011;32(11):906-8.

- 427 [48] Tulenko TN. Regional sensitivity to vasoactive polypeptides in the human
- 428 umbilicoplacental vasculature. Am J Obstet Gynecol. 1979;135(5):629-36.
- 429 [49] AbdAlla S, Lother H, Abdel-tawab AM and Quitterer U. The angiotensin II AT2
- 430 receptor is an AT1 receptor antagonist. J Biol Chem. 2001;276(43):39721-6.

431 [50] Ozono R, Wang ZQ, Moore AF, Inagami T, Siragy HM and Carey RM. Expression of

432 the subtype 2 angiotensin (AT2) receptor protein in rat kidney. Hypertension.

- 433 1997;30(5):1238-46.
- 434 [51] Tebbs C, Pratten MK and Broughton Pipkin F. Angiotensin II is a growth factor in the
 435 peri-implantation rat embryo. J Anat. 1999;195 (Pt 1):75-86.
- 436 [52] Grishko V, Pastukh V, Solodushko V, Gillespie M, Azuma J and Schaffer S. Apoptotic
 437 cascade initiated by angiotensin II in neonatal cardiomyocytes: role of DNA damage. Am
 438 Delaying the second page (2000) 1000 (2000) 1000 (2000)
- 438 J Physiol Heart Circ Physiol. 2003;285(6):H2364-72.

439 Figure Legends

440 Figure 1: Immunohistochemical staining of AGT (A1 to A3), PRR (B1 to B3), AT1R (C1 to

441 C3) and AT2R (D1 to D3) at x200 magnification. Positive staining shown in brown, black

- 442 arrows indicate syncytiotrophoblast cells and red arrows show fetal vessels. ATIR expression
- 443 was increased in the pre-eclamptic compared to the normotensive control placentae (P <
- 444 0.05).
- **Figure 2:** Scatter plot illustrating the inverse association between AT1R expression in the
- placenta, and the birthweight in the pre-eclamptic group (r = -0.529; P = 0.009); not observed

447 in the controls (r = -0.117; P > 0.05).

448 Figure 3: Inverse association between the expression of GPx3 and AT1R in the placenta (r =

-0.634; *P* = 0.006). Placental GPx3 activity was only measured in 10 pre-eclampsia and 10
normotensive control samples.

Figure 4: Scatter plots showing the positive association between the placental AGT and AT2R expression in pre-eclamptic (r = 0.615; P = 0.002) but not normotensive controls (r = 0.064; P > 0.05).

454

455

456

457

458

Table 1. Details of antibody sources and dilutions.

Antigen	Suppler information	Concentration
		(mg/mL)
PRR	Sigma Prestige, rabbit polyclonal: HPA003156	1.8
AGT	Sigma Prestige, rabbit polyclonal: HPA0031557	0.26
AT1R	Abcam, mouse momoclonal: ab9391	80
AT2R	Abcam, rabbit polyclonal: ab19134	5.2

- **Table 2:** Obstetric data of subject groups. Data are presented as mean ±SD or median [IQR]
- 472 as appropriate. *P < 0.05, **P < 0.001; more detailed data has been previously published
- 473 (Mistry et al 2008).

Parameter	Normotensive control	Pre-eclampsia
	(n = 27)	(n = 23)
Gestational age at delivery (weeks)	40 ± 1.1	$36.4 \pm 3.8*$
Birthweight (kg)	3.55 [3.25, 3.86]	2.92 [1.92, 3.51]*
Birthweight centiles (%)	45 [23, 67]	35 [2, 87]

- **Table 3:** Placental protein expression (H-Score) of AGT, PRR, AT1R, ATT2R and GPx3. **P*
- 489 < 0.05; #previously published (Mistry et al 2008) and placental GPx3 activity was only
- 490 measured in 10 pre-eclampsia and 10 normotensive control samples.

Protein expression	H-Score (Median [interquartile range])		
	Normotensive control	Pre-eclampsia	
AGT	34 [10,95]	30 [9,60]	
PRR	117 [16,144]	139 [72,187]	
AT1R	40 [10,70]	68 [30,112]*	
AT2R	152 [120,180]	184 [74,220]	
Placental GPx3 expression (AU) #	197 [124, 254]	110 [82, 124]*	

492