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Early postnatal exposure to a cafeteria diet interferes with recency and spatial memory, but not open field habituation in adolescent rats

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For Peer Review

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3 Abstract
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8 The cafeteria diet (CD), an experimental diet that mimics the obesogenic Western diet,
9 can impair memory in adult rats. However, the suckling period is also particularly
10 susceptible to diet-induced behavioural modification. Here, following exposure to CD
11 feeding during lactation, 24-26 day old offspring were tested to determine maternal
12 dietary effects on either open field habituation, object location (OL) learning or on
13 recency learning. Whereas no impact on habituation learning could be demonstrated,
14 both OL and recency memory were impaired. In controls (C), OL memory was shown
15 both after a 5 min ($P<0.05$) or 60 min ($P<0.001$) inter-trial interval (ITI). After the 60
16 min ITI, the difference between C and CD was significant ($P<0.05$). Learning did not
17 occur in the CD group at any time point and was not observed after the 24hr ITI in in
18 either group. Whereas control rats demonstrated intact recency memory ($P<0.00001$),
19 no learning occurred in the CD group. Both groups differed significantly in their
20 exploration ratios ($P<0.01$). This study suggests a detrimental effect of exposure to an
21 unhealthy Western diet during lactation, on cognitive functions in adolescent rats. These
22 results could have implications for human cognition in the context of obesity epidemic.
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50 KEYWORDS
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52 Western diet, rodents, memory, lactation,
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1 INTRODUCTION

The cafeteria diet is an experimental diet that had been developed to model Western nutritional habits and their pathophysiological consequences in rodents (Rothwell & Stock, 1979; Sclafani & Springer, 1976). Although the composition of this diet is not strictly defined, it usually comprises high-calorie, highly palatable human food items. In rodents, this diet leads to hyperphagia and subsequently to increased energy intake based on overconsumption of fat and sugar. In comparison with high fat diets, cafeteria diet better mimics pathophysiological aspects of human obesity, which is the product of excessive energy intake due to unrestricted dietary choices and inadequate physical activity (Sampey et al., 2011).

Human obesity is also being linked to cognitive dysfunctions (Contu & Hawkes, 2017; Francis & Stevenson, 2013; Morris, Beilharz, Maniam, Reichelt, & Westbrook, 2015; Pedditzi, Peters, & Beckett, 2016; Reichelt, Stoeckel, Reagan, Winstanley, & Page, 2018; Stevenson et al., 2020; Yeomans, 2017), and even a short four day exposure to a high fat-high sugar breakfast can interfere with human memory (Attuquayefio, Stevenson, Oaten, & Francis, 2017). In this context, experimental studies demonstrated that cafeteria feeding has detrimental effects in a variety of rodent memory tests (Andre, Dinel, Ferreira, Laye, & Castanon, 2014; Beilharz, Kaakoush, Maniam, & Morris, 2018; Beilharz, Maniam, & Morris, 2014; Bondan, Cardoso, Martins, & Otton, 2019; Darling, Ross, Bartness, & Parent, 2013; Ferreira, Castro, Andrade, Dulce Madeira, & Cardoso, 2018; Kendig, Westbrook, & Morris, 2019; Kosari, Badoer, Nguyen, Killcross, & Jenkins, 2012; Lewis, Singh, & Youssef, 2019; Reichelt, Gibson, Abbott, & Hare, 2019; Reichelt, Loughman, et al., 2018; Tran & Westbrook, 2018, Abbott, Arnott, Westbrook, & Tran 2019). Whereas many of these studies have been conducted in adult rats, fewer studies specifically addressed memory effects of a cafeteria diet in adolescent rats (Reichelt et al., 2019; Reichelt, Loughman, et al., 2018, Noble & Kanoski, 2016). Of particular interest is a literature which demonstrates that exposure to a hypercaloric diet during early postnatal development impacts upon later behaviour and memory (DeCapo, Thompson, Dunn, & Sullivan, 2019; Moreton et al., 2019; Wright, Langley-Evans, & Voigt, 2011; Wright, Fone, Langley-Evans, & Voigt, 2011; Wright, King, Davey, Langley-Evans, & Voigt, 2014).

In rodents, exposure to cafeteria diet during lactation leads to changes in object discrimination memory in adult age (Alamy & Bengelloun, 2012). This suggests that postnatally developing structures and functions of the offspring's central nervous system are susceptible to challenges associated with hyper-energetic diets. One brain region that undergoes a protracted postnatal development, both in humans and rodents, is the

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3 prefrontal cortex (PFC) (Callaghan & Tottenham, 2016; Kolb et al., 2012; Petanjek et al.,
4 2011) which renders this brain region susceptible to environmental challenges including
5 malnutrition (Reichelt, 2016). In a previous study, we demonstrated that exposure to
6 cafeteria diet during suckling changes PFC dopamine and serotonin neurotransmitter
7 mechanism while also interfering with novel object recognition during adolescence
8 (Moreton et al., 2019). The PFC is also involved in in recency memory in rats (Barker,
9 Bird, Alexander, & Warburton, 2007; Hannesson, Vacca, Howland, & Phillips, 2004;
10 Nelson, Cooper, Thur, Marsden, & Cassaday, 2011), and a cafeteria diet interferes with
11 recency memory in adult rats (Tran & Westbrook, 2018).

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No study has yet considered the impact of early postnatal dietary CD effects on recency
memory. Considering the importance of the lactational period for behavioural and
memory development (Moreton et al., 2019; Speight, Davey, McKenna, & Voigt, 2017;
Wright et al., 2011a,b; Wright et al., 2014), we aimed to investigate the impact of CD
exposure during this period on recency memory. This was to address the wider
hypothesis that early postnatal overnutrition impacts on a range of memories. Thus we
compared the effects of lactational exposure to a cafeteria diet on prefrontal cortex-
dependent recency memory, hippocampus dependent object location memory (Ainge &
Langston, 2012; Dix & Aggleton, 1999) and habituation (Rankin et al., 2009) learning in
the open field (Leussis & Bolivar, 2006). We hypothesised a detrimental effect of the diet
in all three models of memory.

2 METHODS

2.1 Experimental animals

Virgin female Wistar rats (n = 24; Charles River UK) were mated with male rats at 9
weeks of age. Dams were housed two per cage until they had reached the last four days
of their gestation period when they were placed in individual cages. Standard laboratory
chow (Teklad Global 18% Protein Rodent Diet Harlan, UK) and water (filtered tap water)
were available ad libitum. The rats were maintained under a 12 h light dark cycle (with
1 h dusk and 1 h dawn, lights on at 07:30 h), between 20 and 22 degrees C° and at
55 ± 10% relative humidity. Light intensity was 370 lx. At birth, litter size was adjusted to
eight, 4 females and 4 males.

All experiments were performed with approval from the University of Nottingham Animal
Welfare and Ethical Review Body (AWERB) and in accordance with the Animals (Scientific
Procedures) Act, 1986 and ARRIVE guidelines.

2.2 Experimental diet

Following parturition, dams were randomly allocated to either standard laboratory chow diet (control) or the same chow diet in conjunction with a variety of highly palatable, energy-dense human foods (experimental cafeteria diet, CD). Food items consisted of shortbread, golden syrup cake, plain chocolate, pork pie, pâté, cocktail sausages, cheddar cheese, crisps, peanuts and strawberry jam. Of these items, four were provided in excess each day and placed in a bowl on the cage floor. At least one item was exchanged daily in order to maintain novelty and interest (Akyol, Langley-Evans, & McMullen, 2009). Food consumption of the dams was measured every other day during lactation. Energy intake (kJ) and macronutrient consumption (carbohydrates including sugar, fat and protein) were calculated from the manufacturers' data. The average daily percentage change in the weight of foods ranged from 0.0 to 6.2 % and corresponded to an average overestimation of energy intake by 2.51 % (7.5 kJ/d), which can be considered within an acceptable error of measurement (Akyol, Langley-Evans, & McMullen, 2009). Dam body weight was determined alongside food intake measures. On postnatal day 21, offspring were weaned from their dams and then housed in groups of four with littermates of the same sex. Weanlings were fed standard chow for the remainder of the study.

2.3 Behavioural testing

Behavioural testing occurred in 24 to 26 day old offspring. Two randomly selected pups, of each sex from each litter were used for testing; a total of 151 offspring were tested. The remaining offspring were not used in the current study. Testing was undertaken between 08:30 and 13:30h. Only one male/female were used from each litter for each trial to avoid within-litter effects. Experiments were independent, i.e. each individual was only tested in one of the experiments outlined below. Likewise, different offspring were allocated to the separate intervals in the object location test (2.3.2).

All testing took place in an arena measured 60 x 60 x 30 cm, with a grey floor insert and plastic walls. A camera was positioned over the centre of the arena and connected to a computer screen in an adjacent room. On three of the walls, visual cues were displayed. These visual cues were images of a black triangle, circle or square on a white background, placed in the centre of each wall. Behaviour was analysed using Ethovision XT 7 (Noldus, Netherlands). The room was dimly lit (8.0 lux) as dim lighting was used as a measure to reduce stress and anxiety (Voigt et al., 2005). The arena and the objects were cleaned with 70% ethanol solution between uses by different rats, to remove any olfactory cues.

2.3.1 Open field habituation

Each rat to be used was removed from its cage, and placed in the centre of the arena facing the wall with the circle image. Each animal was exposed twice for 10 minutes to the open field, 24 hours apart. Intersession habituation was determined using the behavioural parameters of locomotion (in cm) and rearing (frequency) (Wilson, Voigt, Bader, Marsden, & Fink, 1996). Locomotion was tracked automatically using Ethovision. Rearing was defined as raising both forepaws simultaneously and was tracked manually using the same software.

2.3.2 Object location memory

Three proximal visual cues, black shapes (square, triangle and circle) on a white background were fixed to the centre point of three of the four walls. This was to facilitate spatial orientation in the arena, as the rats were able to see and use the cues, without physically touching them (Ainge & Langston, 2012; Dix & Aggleton, 1999). Clear plastic bottles, with three horizontal black strips, were used as objects for testing. They were filled with water and secured to the arena floor using blue tack. Two identical objects were placed 20 cm away from the corners of the arena. Placing the objects here gave the rat the ability to walk all the way around the object.

Each rat was briefly habituated to the empty arena for 3 minutes and then returned to the home cage. After 2 minutes, the rat was returned again to the arena for a 5 minute trial phase, in which two identical objects had been placed into the arena. All rats were placed into the centre of the arena facing the circle visual cue, regardless of object placement to ensure all rats started with the same spatial orientation in the arena. Exploration of the objects was defined by active pawing at, or sniffing of the objects, as well as rats having their nose directed towards the object, within a 1.5 cm distance. Climbing or rearing on the object were not recorded as object exploration. Contact time with the objects (s) was manually scored using Ethovision. After an inter-testing-interval (ITI) of either 5 minutes, 60 minutes or 24 hours, the rat re-entered the arena for a 5 minute test phase. During the ITI, the rat was returned to their home cage. In the test phase, one of the two objects was moved to another corner of the arena, whereas the other object which remained in the same familiar location. The distance between objects was kept the same throughout all trials. Between experiments the objects were placed in different corners, so that all corner combinations were used in order to remove any bias and counterbalance any corner preferences that the rats might develop. To quantify object preference, exploration times for each object from both trials were converted to an exploration ratio. This ratio represents the proportion of time spent exploring the object in the novel location divided by the total object exploration time during the test trial (t

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3 novel/ (t novel + t familiar) (Dix & Aggleton, 1999; Jablonski, Schreiber, Westbrook,
4 Brennan, & Stanton, 2013). If the rat showed a distinct preference for the object in the
5 novel location (exploration ratio > 0.5), then a learning effect would be considered
6 present.
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10 11 12 2.3.3 Recency memory

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14 Recency memory testing (Mitchell & Laiacona, 1998) took place in the same arena as
15 under 2.3.1 and 2.3.2., and the objects the rats were exposed to were plastic bottles
16 with either three black or white horizontal tape stripes around them (2 black striped
17 bottles, 2 white striped bottles). The bottles were placed roughly 20 x 20cm from the
18 walls of the arena. The order and position of these bottles were counterbalanced to
19 control for location and colour bias.
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24 Each rat was placed into the arena and exposed to the first pair of identical objects (e.g.
25 black stripes) for 5 minutes while the incidence and length of object exploration was
26 recorded using EthoVision. Contact with the objects was defined as active sniffing of the
27 object within a radius of 1.5 cm and/or touching with the front paws and nose. Climbing
28 or rearing on the object were not counted as exploration. After the first exposure, the rat
29 was returned to its cage for 60 minutes, after which it was placed back in the arena and
30 exposed to the second pair of identical objects (white stripes) for 5 minutes and the
31 same behaviours were recorded. After this exposure, the rat was placed back in its cage
32 for 15 minutes, before being placed in the arena for the test exposure. In this exposure,
33 the rat is exposed to one object from each pair (one black, one white) and contact with
34 the remote ('novel', black stripes in this example) and recent ('familiar', white stripes in
35 this example) objects is recorded, as per the previous trials.
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43 To quantify object preference during the exploration, times for each object from both
44 trials were converted to an exploration ratio. This ratio represents the proportion of time
45 spent exploring the remote ('novel') object divided by the total object exploration time
46 during the test trial (t novel/(t novel + t familiar) (Dix & Aggleton, 1999; Jablonski et al.,
47 2013; Mitchell & Laiacona, 1998). If the rat showed a distinct preference for the remote
48 object (exploration ratio > 0.5), then a learning effect would be considered present.
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54 55 2.4 Statistical analyses

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57 The statistical unit for macronutrient intake was the dam. The statistical unit for the
58 behavioural analyses was the litter.
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60 Student's t-test was used to analyse all nutritional data.

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3 For both object location and recency memory, object familiarisation was analysed by Two
4 Way ANOVA (sex, diet) to exclude that these two factors impact on the exploration of
5 object upon first exposure. Males and female offspring were then combined for
6 subsequent analyses since sex differences in memory do not manifest before puberty
7 (Cyrenne & Brown, 2011).
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11 Intersession habituation was assessed based on locomotion (distance travelled) and
12 frequency of rearing events. Both of these parameters were analysed independently
13 using a Two-Way RM ANOVA (diet, trial). The effect of diet on body weight of the dam
14 during lactation was analysed by a Two-Way RM ANOVA (diet, time).
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18 For both object location and recency memory, one sample t-test was used to compare
19 the discrimination ratio of each diet group with values above 0.5 indicating a preference
20 for the novel object and hence a learning effect (Dix & Aggleton, 1999). The exploration
21 ratios between the two diet groups were compared using a Student's t-test. In addition,
22 it is thought that different memory mechanisms underlie the separate time intervals
23 (Rosenzweig, Bennett, Colombo, Lee, & Serrano, 1993) and for these reasons object
24 location data at all ITIs were analysed independently. Consequently, no object location
25 data was analysed using an ANOVA and no between ITI comparisons were made. The use
26 of exploration ratio as a parameter for measuring the learning effect, parallels with other
27 spatial learning and recency memory studies (Nelson et al., 2011).
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34 All figures were created and statistical analysis was performed using GraphPad Prism
35 version 7 (GraphPad Software, USA). All values are shown as means + SEM. Differences
36 between groups were considered significant if the P value < 0.05.
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44 3 RESULTS

45 3.1 Macronutrient and energy intake in dams during lactation

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47 Fat ($t=22.71$, $P<0.0001$) and sugar ($t=9.93$, $P<0.0001$) intakes were significantly
48 increased in lactating CD-fed dams (Tab. 1) which led to a higher total energy intake in
49 these dams ($t=7.54$, $P<0.0001$). The overall protein intake was reduced by 13%
50 ($t=2.43$, $P<0.05$). Despite the increased sugar intake, CD-fed dams consumed less
51 carbohydrates in total ($t=3.58$, $P<0.01$) (Tab. 1). Mean daily chow consumption as
52 measured during lactation was reduced by 59 % in the CD-fed group (C: 59.52 ± 2.18 g;
53 CD: 21.45 ± 2.84 g; $t=10.61$, $P<0.0001$). Although body weight increased in both
54 groups during the lactational period ($F(10, 220) = 88.34$, $P<0.0001$), differences in
55 macronutrient intake did not affect body weight ($F(1, 22) = 0.28$, $P=0.59$).
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3.2 Open field habituation

When exposed to the open field arena, offspring from both cafeteria fed and chow fed dams explored the arena in a similar way. Both groups travelled a similar distance and showed similar numbers of rearing responses. Upon a second exposure, Two-WAY RM ANOVA revealed a decrease in the total distance travelled ($F(1,26)= 7.65, P<0.01$), and in the number of rearing responses ($F(1, 26) = 17.02, P<0.001$). As there was no effect of diet on either parameter, the observed decreases in locomotor activity and rearing suggest that habituation occurred similarly in both groups (Fig. 1).

3.2 Object location

No effects of diet or sex on the time exploring the objects could be observed in any of the three experiments during the 5 min sampling period (trial 1) (Tab. 2).

When exposed to the two objects 5 and 60 minutes later, but with one object in a novel location, offspring from the control group explored the object in the novel location for longer than the object that remained in the familiar location ($t=2.66$ $df=19, P<0.05$; $t=4.658$ $df=13, P<0.001$). No learning occurred in the cafeteria group though, and the cafeteria group had a lower exploration ratio compared to control at the 60 minutes test session ($t=2.686$ $df=24, P<0.05$). By contrast to open field habituation, no learning occurred in either of the two groups when tested 24hrs following the first exposure (Fig. 2).

3.3 Object recency

As shown before (3.2), there was no impact of sex or diet on object exploration during the first 5 minute exposure to the two objects (Tab. 3).

Fifteen minutes after exposure to the recent and the distant object, rats from chow-fed dams explored the distant object for longer than the more recent object ($t=6.99$ $df=13, P<0.0001$), whereas such a learning effect was not observed in the cafeteria group. Both groups differ significantly in their exploration ratios ($t=3.60$ $df=23, P<0.01$) (Fig. 3).

4 DISCUSSION

The macronutrient intake pattern as obtained in the present study is very similar to previous reports from our laboratory. Increased overall energy intake was due to increased consumption of fat and sugars and is accompanied by a light but significant reduction of protein intake (Akyol et al., 2009; Akyol, McMullen, & Langley-Evans, 2012; George et al., 2019; Speight et al., 2017; Wright et al., 2014). Hence it is safe to say that the macronutrient and energy intake as independent variables are similar across previous behavioural studies including the present one. As discussed previously (Moreton et al., 2019), any observed behavioural effects are most likely mediated via maternal milk consumption rather than active pre-weaning CD intake by the pups. Milk composition during lactation reflects dietary intake when animals are fed CD (Rolls et al., 1986). Milk consumption of pups peaks around postnatal day 15 whereas pups start feeding solid food around day 17 (Ostadalova & Babicky, 2012). Given that milk consumption remains high at least to postnatal day 19 (Ostadalova & Babicky, 2012), direct CD consumption by the pups would be minimal, as they were weaned on day 21. Hence the observed effects are most likely mediated via maternal milk ingestion rather than a direct CD consumption.

Cafeteria diet during lactation had no effect on body weight in the dam. This finding is in line with previous studies, both in lactating (Akyol et al., 2012) and non-lactating females of similar age (Warneke et al., 2014). Compensatory mechanisms, of which the induction of thermogenesis appears to be particularly important (Rothwell & Stock, 1979; Rothwell and Stock, 1980) can buffer some of the effects of cafeteria feeding in younger rats. Hyperenergetic diets can also reduce the expression of hypothalamic orexigenic signalling in young rats to counter the obesogenic effects of the diet (Archer et al., 2004). CD is more likely to increase body weight in adult rats (Sclafani & Gorman, 1977).

The manipulation of maternal diet during lactation, impaired novel object discrimination (recognition) learning in a study under similar experimental conditions (Moreton et al., 2019). In previous studies, demonstrating detrimental effects on spatial memory, rats have either been exposed to CD either after weaning (Ferreira et al., 2018) or in adult age (Beilharz et al., 2014; Kendig et al., 2019; Kosari et al., 2012; Pini, Ferreira do Vales, Braga Costa, & Almeida, 2017). Here we demonstrate for the first time a detrimental effect of lactational exposure to CD on object location memory. We also present the novel finding that exposure during suckling to a hyper-energetic highly palatable diet leads to impaired recency memory. Despite these effects on memory, there was no evidence that the diet had any effect on object exploration during the sample phase in either of the two tests, although object exploration in the sample phase is possibly not a decisive factor as shown for spatial memory (Ozawa, Yamada, & Ichitani, 2011).

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3 Although the observed effects of maternal cafeteria feeding on spatial learning were
4 generally in line with other studies, the lack of a learning effect following a 24 hour ITI,
5 observed in this study, was in contrast to previous results (Westbrook et al., 2014).
6 According to Westbrook, 26- and 31-day-old, but not 21-day-old rats retained the object
7 location memory for a 24h ITI. Age differences could explain the discrepancy between
8 the current and the aforementioned study. Our rats were between 24 and 26 days old,
9 possibly just below the point where 24 h memory is fully developed. Strain differences
10 are another factor to be considered, but the nature of the test can play a role as well
11 (Kumar et al. 2019, Ennaceur et al., 2005). On a mechanistic level, it has been
12 demonstrated that hippocampal mossy fibres are still growing and remodelling up to
13 postnatal day 24 in rats (Holahan et al., 2007). These fibres are involved in spatial
14 memory formation and their postnatal development and functional anatomy is strain
15 dependent in rodents (Holahan et al., 2007; Crusio & Schwegler, 2005). Another study
16 (Ainge & Langston, 2012), albeit using a model of associate spatial learning, revealed
17 that spatial recognition does occur in 30-day-old rats, but not in 24-day-old rats. Of note
18 that study used a short 1-2 minute ITI. One could speculate that our weak learning effect
19 after a 5 minute ITI was rather in line with their finding, suggesting a rather late
20 development of associative spatial learning. However, as this would contradict our finding
21 after the 60 minute ITI, one could further speculate that different memories could be
22 involved (Rosenzweig, Bennett, Colombo, Lee, & Serrano, 1993).
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34 In contrast to spatial and recency memory, long-term habituation learning as studied
35 during repeated exposure to the same open field arena was not affected by maternal
36 diet. Habituation is a decreased response upon repeated exposure to the same stimulus
37 or environment and occurs if the exposure has neither positive nor negative
38 consequences (Rankin et al., 2009). Fully established 24 hour habituation of exploration
39 has been demonstrated before in rats of this age (Parsons, Fagan, & Spear, 1973). The
40 current results demonstrating established habituation learning in both the control and
41 experimental group established are in line with this earlier finding. To explain the lack of
42 dietary effects in this test, one could speculate that rats were 'over-trained' as a shorter
43 than 10 minutes exposure could possibly reduce the strength of the memory formation,
44 making it more susceptible to dietary challenge. This warrants further investigation
45 including testing a range of intersession retention intervals, as adult rats have been
46 shown to retain habituation over a period of 1 week (Richardson & Campbell, 1991). This
47 latter finding suggests an alternative interpretation of our results in that habituation
48 learning in the present paradigm is a robust and non-complex task which makes it less
49 susceptible to the dietary manipulation.
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58 Rats exposed to an open field arena as in the present study, are also exposed to distant
59 spatial cues at the same time. Thus spatial long-term habituation requires a functioning
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3 hippocampus (Riedel et al., 1999; Vianna et al., 2000). Consequently, as the dietary
4 interference with hippocampal function is established (Kanoski & Davidson, 2011;
5 Kanoski, Zhang, Zheng, & Davidson, 2010), dietary effects could have been expected.
6 However, considering that brain regions involved in habituation learning are potentially
7 more widespread (Yamaguchi & Knight, 1991), compensatory mechanisms could
8 maintain functioning of long-term habituation in adolescent CD fed offspring.
9 Hippocampus-dependent spatial memory has been shown to be particularly sensitive to
10 unbalanced hyper-energetic diets (Noble & Kanoski, 2016). Of note, consumption of a
11 Western diet reduces hippocampal volume, both in adults (Jacka, Cherbuin, Anstey,
12 Sachdev, & Butterworth, 2015) and in obese children (Mestre et al., 2017). However,
13 there is currently no evidence that developmental exposure to a cafeteria diet leads to
14 impairment of spatial memory in humans.
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22 A majority of, but not all (Nguyen et al., 2017; Pini et al., 2017), rodents studies
23 suggests that CD feeding interferes with hippocampus mediated spatial learning (Beilharz
24 et al., 2014; Ferreira et al., 2018; Kendig et al., 2019; Kosari et al., 2012; Tran &
25 Westbrook 2015) but functioning of other brain structures can also be affected (Nguyen
26 et al., 2017). Two different pathways have been described for underlying either object
27 recognition memory or spatial and hippocampal memory although both require PFC,
28 albeit different substructures of the PFC, functioning (Barker et al., 2007; Steckler,
29 Drinkenburg, Sahgal, & Aggleton, 1998). Interestingly, recency memory has also been
30 attributed directly to hippocampal function (Albasser, Amin, Lin, Iordanova, & Aggleton,
31 2012), but also PFC functioning (Mitchell & Laiacona, 1998; Nelson et al., 2011). Of note,
32 hippocampal lesion at birth decreases PFC functioning with a behavioural readout of
33 reduced performance in spatial and temporal memory tests in adolescence (Kruger et al.,
34 2012) indicating functional connections between these two structures. As outlined above
35 many studies that have examined the impact of cafeteria diets on memory used adult
36 rats rather than younger animals in which the postnatal brain is still under development.
37 The interaction between diet and memory could be different in young compared with
38 adult rats. The prefrontal cortex is a brain structure that undergoes a protracted
39 postnatal development (Kolb et al., 2012; Petanjek et al., 2011). Therefore the PFC is
40 particularly susceptible to early-life environmental factors which can result in either
41 positive or detrimental consequences in the adult (Callaghan & Tottenham, 2016; Kieling,
42 Goncalves, Tannock, & Castellanos, 2008; McCrory, De Brito, & Viding, 2010; Selemon &
43 Zecevic, 2015). A protracted development has not only been shown for the PFC alone but
44 could also affect functional interactions with the hippocampus (Murty et al., 2016). Here
45 we show for the first time that PFC and hippocampus dependent (Mitchell & Laiacona,
46 1998; Nelson et al., 2011) temporal memory is impaired due to exposure to cafeteria
47 diet during suckling. However, other brain regions generally important in information
48 processing like the subcortical thalamus are less studied regarding the impact of hyper-
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3 energetic diets (Dumont & Aggleton, 2013; Li et al., 2019). Adding complexity to this, CD
4 feeding alters not only a range of brain neurotransmitters and of behaviours, but also gut
5 microbiota (Leigh, Kaakoush, Bertoldo, Westbrook, & Morris, 2020; Moreton et al., 2019;
6 Reichelt, Loughman, et al., 2018; Wright et al., 2011). The latter could impact on
7 neurodevelopmental processes and cognitive performance (Cowan, Dinan, & Cryan,
8 2020; Cryan et al., 2019).

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13 Experimental studies, including the current one, provide accumulating evidence that
14 obesogenic diets interfere with a range memory processes (Reichelt, Loughman, et al.,
15 2018; Tran & Westbrook, 2018). This goes along with an increasing awareness that
16 exposure to hyperenergetic and unbalanced diets relate to cognitive disturbances and
17 psychiatric diseases later in later live as reviewed in (Adan et al., 2019; Francis &
18 Stevenson, 2013; Reichelt, Stoeckel, et al., 2018). Notably, two randomised controlled
19 trials demonstrated improvements in depressive patients when their diet was changed
20 from a poor Western diet to a healthier diet. (Francis et al., 2019; Jacka et al., 2017).
21 The finding that the impact of a poor quality diet is pronounced even when the exposure
22 occurs during early development, is of particular significance.

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29 In conclusion, we demonstrate here that exposure to a palatable, but unbalanced, hyper-
30 energetic cafeteria diet during lactation impairs recency memory and object location
31 memory in early adolescence, whereas the impact on spatial habituation learning
32 warrants further investigation. These results further support the notion that dietary
33 challenges during early postnatal development impact on learning in early adolescence.
34 Experimental findings like this, in conjunction with an increasing number of human
35 studies, suggest a detrimental effect of unhealthy Western diets on cognitive functions.

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REFERENCES

- Abbott, K. N., Arnott, C. K., Westbrook, R. F., & Tran, D. M. D. (2019). The effect of high fat, high sugar, and combined high fat-high sugar diets on spatial learning and memory in rodents: A meta-analysis. *Neuroscience & Biobehavioral Reviews*, 107, 399-421. doi:10.1016/j.neubiorev.2019.08.010
- Adan, R. A. H., van der Beek, E. M., Buitelaar, J. K., Cryan, J. F., Hebebrand, J., Higgs, S., Schellekens, H., Dickson, S. L. (2019). Nutritional psychiatry: Towards improving mental health by what you eat. *European Neuropsychopharmacology*, 29(12), 1321-1332. doi:10.1016/j.euroneuro.2019.10.011
- Ainge, J. A., & Langston, R. F. (2012). Ontogeny of neural circuits underlying spatial memory in the rat. *Frontiers in Neural Circuits*, 6, 8. doi:10.3389/fncir.2012.00008
- Akyol, A., Langley-Evans, S. C., & McMullen, S. (2009). Obesity induced by cafeteria feeding and pregnancy outcome in the rat. *British Journal of Nutrition*, 102(11), 1601-1610. doi:10.1017/S0007114509990961
- Akyol, A., McMullen, S., & Langley-Evans, S. C. (2012). Glucose intolerance associated with early-life exposure to maternal cafeteria feeding is dependent upon post-weaning diet. *British Journal of Nutrition*, 107(7), 964-978. doi:10.1017/S0007114511003916
- Alamy, M., & Bengelloun, W. A. (2012). Malnutrition and brain development: an analysis of the effects of inadequate diet during different stages of life in rat. *Neuroscience & Biobehavioral Reviews*, 36(6), 1463-1480. doi:10.1016/j.neubiorev.2012.03.009
- Albasser, M. M., Amin, E., Lin, T. C., Iordanova, M. D., & Aggleton, J. P. (2012). Evidence that the rat hippocampus has contrasting roles in object recognition memory and object recency memory. *Behavioral Neuroscience*, 126(5), 659-669. doi:10.1037/a0029754
- Andre, C., Dinel, A. L., Ferreira, G., Laye, S., & Castanon, N. (2014). Diet-induced obesity progressively alters cognition, anxiety-like behavior and lipopolysaccharide-induced depressive-like behavior: focus on brain indoleamine 2,3-dioxygenase activation. *Brain Behavior, and Immunity*, 41, 10-21. doi:10.1016/j.bbi.2014.03.012
- Archer, Z. A., Moar, K. M., Logie, T. J., Reilly, L., Stevens, V., Morgan, P. J., & Mercer, J. G. (2007). Hypothalamic neuropeptide gene expression during recovery from food restriction superimposed on short-day photoperiod-induced weight loss in the Siberian hamster. *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology*, 293(3), R1094-1101. doi:10.1152/ajpregu.00345.2007

1
2
3 Attuquayefio, T., Stevenson, R. J., Oaten, M. J., & Francis, H. M. (2017). A four-day
4 Western-style dietary intervention causes reductions in hippocampal-dependent learning
5 and memory and interoceptive sensitivity. *PLoSOne*, 12(2), e0172645.
6 doi:10.1371/journal.pone.0172645
7

8
9
10 Barker, G. R., Bird, F., Alexander, V., & Warburton, E. C. (2007). Recognition memory for
11 objects, place, and temporal order: a disconnection analysis of the role of the medial
12 prefrontal cortex and perirhinal cortex. *Journal of Neuroscience*, 27(11), 2948-2957.
13 doi:10.1523/JNEUROSCI.5289-06.2007
14

15
16
17 Beilharz, J. E., Kaakoush, N. O., Maniam, J., & Morris, M. J. (2018). Cafeteria diet and
18 probiotic therapy: cross talk among memory, neuroplasticity, serotonin receptors and gut
19 microbiota in the rat. *Molecular Psychiatry*, 23(2), 351-361. doi:10.1038/mp.2017.38
20

21
22 Beilharz, J. E., Maniam, J., & Morris, M. J. (2014). Short exposure to a diet rich in both
23 fat and sugar or sugar alone impairs place, but not object recognition memory in rats.
24 *Brain Behavior, and Immunity*, 37, 134-141. doi:10.1016/j.bbi.2013.11.016
25

26
27
28 Bondan, E. F., Cardoso, C. V., Martins, M. F. M., & Otton, R. (2019). Memory
29 impairments and increased GFAP expression in hippocampal astrocytes following
30 hypercaloric diet in rats. *Arquivos de Neuro-Psiquiatria*, 77(9), 601-608.
31 doi:10.1590/0004-282X20190091
32

33
34
35 Callaghan, B. L., & Tottenham, N. (2016). The neuro-environmental loop of plasticity: A
36 cross-species analysis of parental effects on emotion circuitry development following
37 typical and adverse caregiving. *Neuropsychopharmacology*, 41(1), 163-176.
38 doi:10.1038/npp.2015.204
39

40
41
42 Contu, L., & Hawkes, C. A. (2017). A review of the impact of maternal obesity on the
43 cognitive function and mental health of the offspring. *International Journal of Molecular
44 Sciences*, 18(5). doi:10.3390/ijms18051093
45

46
47
48 Cowan, C. S. M., Dinan, T. G., & Cryan, J. F. (2020). Annual Research Review: Critical
49 windows - the microbiota-gut-brain axis in neurocognitive development. *Journal of Child
50 Psychology and Psychiatry*, 61(3), 353-371. doi:10.1111/jcpp.13156
51

52
53
54 Crusio, W. E., & Schwegler, H. (2005). Learning spatial orientation tasks in the radial-
55 maze and structural variation in the hippocampus in inbred mice. *Behavioral and Brain
56 Functions*, 1(1), 3. doi:10.1186/1744-9081-1-3
57

58
59
60 Cryan, J. F., O'Riordan, K. J., Cowan, C. S. M., Sandhu, K. V., Bastiaanssen, T. F. S.,
Boehme, M., . . . Dinan, T. G. (2019). The microbiota-gut-brain axis. *Physiological
Reviews*, 99(4), 1877-2013. doi:10.1152/physrev.00018.2018

1
2
3
4
5 Cyrenne, D. L., & Brown, G. R. (2011). Ontogeny of sex differences in response to novel
6 objects from adolescence to adulthood in lister-hooded rats. *Developmental*
7 *Psychobiology*, 53(7), 670-676. doi:10.1002/dev.20542
8
9

10 Darling, J. N., Ross, A. P., Bartness, T. J., & Parent, M. B. (2013). Predicting the effects
11 of a high-energy diet on fatty liver and hippocampal-dependent memory in male rats.
12 *Obesity*, 21(5), 910-917. doi:10.1002/oby.20167
13
14

15
16 DeCapo, M., Thompson, J. R., Dunn, G., & Sullivan, E. L. (2019). Perinatal nutrition and
17 programmed risk for neuropsychiatric disorders: A focus on animal models. *Biological*
18 *Psychiatry*, 85(2), 122-134. doi:10.1016/j.biopsych.2018.08.006
19
20

21
22 Dix, S. L., & Aggleton, J. P. (1999). Extending the spontaneous preference test of
23 recognition: evidence of object-location and object-context recognition. *Behavioral Brain*
24 *Research*, 99(2), 191-200. doi.org/10.1016/S0166-4328(98)00079-5
25
26

27
28 Dumont, J. R., & Aggleton, J. P. (2013). Dissociation of recognition and recency memory
29 judgments after anterior thalamic nuclei lesions in rats. *Behavioral Neuroscience*, 127(3),
30 415-431. doi:10.1037/a0032750
31
32

33
34 Ennaceur, A., Michalikova, S., Bradford, A., & Ahmed, S. (2005). Detailed analysis of the
35 behavior of Lister and Wistar rats in anxiety, object recognition and object location tasks.
36 *Behavioral Brain Research*, 159(2), 247-266. doi:10.1016/j.bbr.2004.11.006
37
38

39
40 Ferreira, A., Castro, J. P., Andrade, J. P., Dulce Madeira, M., & Cardoso, A. (2018).
41 Cafeteria-diet effects on cognitive functions, anxiety, fear response and neurogenesis in
42 the juvenile rat. *Neurobiology of Learning and Memory*, 155, 197-207.
43 doi:10.1016/j.nlm.2018.07.014
44
45

46 Francis, H., & Stevenson, R. (2013). The longer-term impacts of Western diet on human
47 cognition and the brain. *Appetite*, 63, 119-128. doi:10.1016/j.appet.2012.12.018
48
49

50
51 Francis, H. M., Stevenson, R. J., Chambers, J. R., Gupta, D., Newey, B., & Lim, C. K.
52 (2019). A brief diet intervention can reduce symptoms of depression in young adults - A
53 randomised controlled trial. *PLoS One*, 14(10), e0222768.
54 doi:10.1371/journal.pone.0222768
55
56

57
58 George, G., Draycott, S. A. V., Muir, R., Clifford, B., Elmes, M. J., & Langley-Evans, S. C.
59 (2019). The impact of exposure to cafeteria diet during pregnancy or lactation on
60 offspring growth and adiposity before weaning. *Scientific Reports*, 9(1), 14173.
doi:10.1038/s41598-019-50448-x

1
2
3
4
5 Hannesson, D. K., Vacca, G., Howland, J. G., & Phillips, A. G. (2004). Medial prefrontal
6 cortex is involved in spatial temporal order memory but not spatial recognition memory
7 in tests relying on spontaneous exploration in rats. *Behavioral Brain Research*, 153(1),
8 273-285. doi:10.1016/j.bbr.2003.12.004
9

10
11 Holahan, M. R., Honegger, K. S., & Routtenberg, A. (2007). Expansion and retraction of
12 hippocampal mossy fibers during postweaning development: strain-specific effects of
13 NMDA receptor blockade. *Hippocampus*, 17(1), 58-67. doi:10.1002/hipo.20242
14
15

16
17 Jablonski, S. A., Schreiber, W. B., Westbrook, S. R., Brennan, L. E., & Stanton, M. E.
18 (2013). Determinants of novel object and location recognition during development.
19 *Behavioral Brain Research*, 256, 140-150. doi:10.1016/j.bbr.2013.07.055
20
21

22
23 Jacka, F. N., Cherbuin, N., Anstey, K. J., Sachdev, P., & Butterworth, P. (2015). Western
24 diet is associated with a smaller hippocampus: a longitudinal investigation. *BMC*
25 *Medicine*, 13, 215. doi:10.1186/s12916-015-0461-x
26
27

28
29 Jacka, F. N., O'Neil, A., Opie, R., Itsiopoulos, C., Cotton, S., Mohebbi, M., Berk, M.
30 (2017). A randomised controlled trial of dietary improvement for adults with major
31 depression (the 'SMILES' trial). *BMC Medicine*, 15(1), 23. doi:10.1186/s12916-017-0791-
32 y
33

34
35 Kanoski, S. E., & Davidson, T. L. (2011). Western diet consumption and cognitive
36 impairment: links to hippocampal dysfunction and obesity. *Physiology & Behavior*,
37 103(1), 59-68. doi:10.1016/j.physbeh.2010.12.003
38
39

40
41 Kanoski, S. E., Zhang, Y., Zheng, W., & Davidson, T. L. (2010). The effects of a high-
42 energy diet on hippocampal function and blood-brain barrier integrity in the rat. *Journal*
43 *of Alzheimer's Disease*, 21(1), 207-219. doi:10.3233/JAD-2010-091414
44
45

46
47 Kendig, M. D., Westbrook, R. F., & Morris, M. J. (2019). Pattern of access to cafeteria-
48 style diet determines fat mass and degree of spatial memory impairments in rats.
49 *Scientific Reports*, 9(1), 13516. doi:10.1038/s41598-019-50113-3
50
51

52
53 Kieling, C., Goncalves, R. R., Tannock, R., & Castellanos, F. X. (2008). Neurobiology of
54 attention deficit hyperactivity disorder. *Child and Adolescent Psychiatric Clinics of North*
55 *America*, 17(2), 285-307, viii. doi:10.1016/j.chc.2007.11.012
56
57

58
59 Kolb, B., Mychasiuk, R., Muhammad, A., Li, Y., Frost, D. O., & Gibb, R. (2012).
60 Experience and the developing prefrontal cortex. *Proceedings of the National Academy of*

1
2
3 *Sciences of the United States of America*, 109 Suppl 2, 17186-17193.
4 doi:10.1073/pnas.1121251109

5
6
7
8 Kosari, S., Badoer, E., Nguyen, J. C., Killcross, A. S., & Jenkins, T. A. (2012). Effect of
9 western and high fat diets on memory and cholinergic measures in the rat. *Behavioral*
10 *Brain Research*, 235(1), 98-103. doi:10.1016/j.bbr.2012.07.017

11
12
13 Kruger, H. S., Brockmann, M. D., Salamon, J., Ittrich, H., & Hanganu-Opatz, I. L. (2012).
14 Neonatal hippocampal lesion alters the functional maturation of the prefrontal cortex and
15 the early cognitive development in pre-juvenile rats. *Neurobiology of Learning and*
16 *Memory*, 97(4), 470-481. doi:10.1016/j.nlm.2012.04.001

17
18
19
20 Kumar, G., Talpos, J., & Steckler, T. (2015). Strain-dependent effects on acquisition and
21 reversal of visual and spatial tasks in a rat touchscreen battery of cognition. *Physiology &*
22 *Behavior*, 144, 26-36. doi:10.1016/j.physbeh.2015.03.001

23
24
25
26 Leigh, S. J., Kaakoush, N. O., Bertoldo, M. J., Westbrook, R. F., & Morris, M. J. (2020).
27 Intermittent cafeteria diet identifies fecal microbiome changes as a predictor of spatial
28 recognition memory impairment in female rats. *Translational Psychiatry*, 10(1), 36.
29 doi:10.1038/s41398-020-0734-9

30
31
32
33 Leussis, M. P., & Bolivar, V. J. (2006). Habituation in rodents: a review of behavior,
34 neurobiology, and genetics. *Neuroscience & Biobehavioral Reviews*, 30(7), 1045-1064.
35 doi:10.1016/j.neubiorev.2006.03.006

36
37
38
39 Lewis, A. R., Singh, S., & Youssef, F. F. (2019). Cafeteria-diet induced obesity results in
40 impaired cognitive functioning in a rodent model. *Heliyon*, 5(3), e01412.
41 doi:10.1016/j.heliyon.2019.e01412

42
43
44
45 Li, J., Guo, Y., Li, Q., Miao, K., Wang, C., Zhang, D., . . . Zhang, S. (2019). Presence of
46 white matter lesions associated with diabetes-associated cognitive decline in male rat
47 models of pre-type 2 diabetes. *Medical Science Monitor*, 25, 9679-9689.
48 doi:10.12659/MSM.918557

49
50
51
52 McCrory, E., De Brito, S. A., & Viding, E. (2010). Research review: the neurobiology and
53 genetics of maltreatment and adversity. *Journal of Child Psychology and Psychiatry*,
54 51(10), 1079-1095. doi:10.1111/j.1469-7610.2010.02271.x

55
56
57
58
59
60 Mestre, Z. L., Bischoff-Grethe, A., Eichen, D. M., Wierenga, C. E., Strong, D., & Boutelle,
K. N. (2017). Hippocampal atrophy and altered brain responses to pleasant tastes among
obese compared with healthy weight children. *International Journal of Obesity (Lond)*,
41(10), 1496-1502. doi:10.1038/ijo.2017.130

1
2
3
4
5 Mitchell, J. B., & Laiacona, J. (1998). The medial frontal cortex and temporal memory:
6 tests using spontaneous exploratory behaviour in the rat. *Behavioral Brain Research*,
7 97(1-2), 107-113. doi:10.1016/s0166-4328(98)00032-1
8
9

10 Moreton, E., Baron, P., Tiplady, S., McCall, S., Clifford, B., Langley-Evans, S. C., . . .
11 Voigt, J. P. (2019). Impact of early exposure to a cafeteria diet on prefrontal cortex
12 monoamines and novel object recognition in adolescent rats. *Behavioral Brain Research*,
13 363, 191-198. doi:10.1016/j.bbr.2019.02.003
14
15

16
17 Morris, M. J., Beilharz, J. E., Maniam, J., Reichelt, A. C., & Westbrook, R. F. (2015). Why
18 is obesity such a problem in the 21st century? The intersection of palatable food, cues
19 and reward pathways, stress, and cognition. *Neuroscience & Biobehavioral Reviews*, 58,
20 36-45. doi:10.1016/j.neubiorev.2014.12.002
21
22

23
24 Murty, V. P., Calabro, F., & Luna, B. (2016). The role of experience in adolescent
25 cognitive development: Integration of executive, memory, and mesolimbic systems.
26 *Neuroscience & Biobehavioral Reviews*, 70, 46-58. doi:10.1016/j.neubiorev.2016.07.034
27
28

29
30 Nelson, A. J., Cooper, M. T., Thur, K. E., Marsden, C. A., & Cassaday, H. J. (2011). The
31 effect of catecholaminergic depletion within the prelimbic and infralimbic medial
32 prefrontal cortex on recognition memory for recency, location, and objects. *Behavioral*
33 *Neuroscience*, 125(3), 396-403. doi:10.1037/a0023337
34
35

36
37 Nguyen, J. C., Ali, S. F., Kosari, S., Woodman, O. L., Spencer, S. J., Killcross, A. S., &
38 Jenkins, T. A. (2017). Western diet chow consumption in rats induces striatal neuronal
39 activation while reducing dopamine levels without affecting spatial memory in the radial
40 arm maze. *Frontiers in Behavioral Neuroscience*, 11, 22. doi:10.3389/fnbeh.2017.00022
41
42

43 Noble, E. E., & Kanoski, S. E. (2016). Early life exposure to obesogenic diets and learning
44 and memory dysfunction. *Current Opinion in Behavioral Sciences*, 9, 7-14.
45 doi:10.1016/j.cobeha.2015.11.014
46
47

48
49 Ostadalova, I., & Babicky, A. (2012). Periodization of the early postnatal development in
50 the rat with particular attention to the weaning period. *Physiological Research*, 61, S1-
51 S7.
52
53

54
55 Ozawa, T., Yamada, K., & Ichitani, Y. (2011). Long-term object location memory in rats:
56 effects of sample phase and delay length in spontaneous place recognition test.
57 *Neuroscience Letters*, 497(1), 37-41. doi:10.1016/j.neulet.2011.04.022
58
59
60

1
2
3 Parsons, P. J., Fagan, T., & Spear, N. E. (1973). Short-term retention of habituation in
4 the rat: a developmental study from infancy to old age. *Journal of Comparative and*
5 *Physiological Psychology*, 84(3), 545-553. doi:10.1037/h0034889
6
7

8
9 Pedditz, E., Peters, R., & Beckett, N. (2016). The risk of overweight/obesity in mid-life
10 and late life for the development of dementia: a systematic review and meta-analysis of
11 longitudinal studies. *Age and Ageing*, 45(1), 14-21. doi:10.1093/ageing/afv151
12
13

14
15 Petanjek, Z., Judas, M., Simic, G., Rasin, M. R., Uylings, H. B., Rakic, P., & Kostovic, I.
16 (2011). Extraordinary neoteny of synaptic spines in the human prefrontal cortex.
17 *Proceedings of the National Academy of Sciences of the U S A*, 108(32), 13281-13286.
18 doi:10.1073/pnas.1105108108
19

20
21 Pini, R. T. B., Ferreira do Vales, L. D. M., Braga Costa, T. M., & Almeida, S. S. (2017).
22 Effects of cafeteria diet and high fat diet intake on anxiety, learning and memory in adult
23 male rats. *Nutritional Neuroscience*, 20(7), 396-408.
24 doi:10.1080/1028415X.2016.1149294
25
26

27
28 Rankin, C. H., Abrams, T., Barry, R. J., Bhatnagar, S., Clayton, D. F., Colombo, J., . . .
29 Thompson, R. F. (2009). Habituation revisited: an updated and revised description of the
30 behavioral characteristics of habituation. *Neurobiology of Learning and Memory*, 92(2),
31 135-138. doi:10.1016/j.nlm.2008.09.012
32
33

34
35 Reichelt, A. C. (2016). Adolescent maturational transitions in the prefrontal cortex and
36 dopamine signaling as a risk factor for the development of obesity and high fat/high
37 sugar diet induced cognitive deficits. *Frontiers in Behavioral Neuroscience*, 10, 189.
38 doi:10.3389/fnbeh.2016.00189
39
40

41
42 Reichelt, A. C., Gibson, G. D., Abbott, K. N., & Hare, D. J. (2019). A high-fat high-sugar
43 diet in adolescent rats impairs social memory and alters chemical markers characteristic
44 of atypical neuroplasticity and parvalbumin interneuron depletion in the medial prefrontal
45 cortex. *Food & Function*, 10(4), 1985-1998. doi:10.1039/c8fo02118j
46
47

48
49 Reichelt, A. C., Loughman, A., Bernard, A., Raipuria, M., Abbott, K. N., Dachtler, J., . . .
50 Moore, R. J. (2018). An intermittent hypercaloric diet alters gut microbiota, prefrontal
51 cortical gene expression and social behaviours in rats. *Nutritional Neuroscience*, 1-15.
52 doi:10.1080/1028415X.2018.1537169
53
54

55
56 Reichelt, A. C., Stoeckel, L. E., Reagan, L. P., Winstanley, C. A., & Page, K. A. (2018).
57 Dietary influences on cognition. *Physiology & Behavior*, 192, 118-126.
58 doi:10.1016/j.physbeh.2018.02.052
59
60

1
2
3 Richardson, R., & Campbell, B. A. (1991). Ontogeny of long-term nonassociative memory
4 in the rat. *Animal Learning & Behavior*, 19(1), 1-10. doi:10.3758/bf03197854
5

6
7
8 Riedel, G., Micheau, J., Lam, A. G., Roloff, E. L., Martin, S. J., Bridge, H., . . . Morris, R.
9 G. (1999). Reversible neural inactivation reveals hippocampal participation in several
10 memory processes. *Nature Neuroscience*, 2(10), 898-905. doi:10.1038/13202
11

12
13
14 Rolls, B. A., Gurr, M. I., van Duijvenvoorde, P. M., Rolls, B. J., & Rowe, E. A. (1986).
15 Lactation in lean and obese rats: effect of cafeteria feeding and of dietary obesity on milk
16 composition. *Physiology & Behavior*, 38(2), 185-190.
17

18
19
20 Rosenzweig, M. R., Bennett, E. L., Colombo, P. J., Lee, D. W., & Serrano, P. A. (1993).
21 Short-term, intermediate-term, and long-term memories. *Behavioral Brain Research*,
22 57(2), 193-198. doi 10.1016/0166-4328(93)90135-d
23

24
25 Rothwell, N. J., & Stock, M. J. (1979). Regulation of energy balance in two models of
26 reversible obesity in the rat. *Journal of Comparative and Physiological Psychology*, 93(6),
27 1024-1034. doi: 10.1037/h0077631
28

29
30
31 Rothwell, N. J., & Stock, M. J. (1980). Thermogenesis induced by cafeteria feeding in
32 young growing rats. *Proceedings of the Nutrition Society*, 39(2), 45A.
33

34
35
36 Sampey, B. P., Vanhoose, A. M., Winfield, H. M., Freerman, A. J., Muehlbauer, M. J.,
37 Fueger, P. T., . . . Makowski, L. (2011). Cafeteria diet is a robust model of human
38 metabolic syndrome with liver and adipose inflammation: comparison to high-fat diet.
39 *Obesity (Silver Spring)*, 19(6), 1109-1117. doi:10.1038/oby.2011.18
40

41
42
43 Sclafani, A., & Springer, D. (1976). Dietary obesity in adult rats: similarities to
44 hypothalamic and human obesity syndromes. *Physiology & Behavior*, 17(3), 461-471.
45 doi: 10.1016/0031-9384(76)90109-8
46

47
48
49 Sclafani, A., & Gorman, A. N. (1977). Effects of age, sex, and prior body weight on the
50 development of dietary obesity in adult rats. *Physiology & Behavior*, 18(6), 1021-1026.
51 doi:10.1016/0031-9384(77)90006-3
52

53
54
55 Selemon, L. D., & Zecevic, N. (2015). Schizophrenia: a tale of two critical periods for
56 prefrontal cortical development. *Translational Psychiatry*, 5, e623.
57 doi:10.1038/tp.2015.115
58

59
60
Speight, A., Davey, W. G., McKenna, E., & Voigt, J. W. (2017). Exposure to a maternal
cafeteria diet changes open-field behaviour in the developing offspring. *International
Journal of Developmental Neuroscience*, 57, 34-40. doi:10.1016/j.ijdevneu.2016.12.005

1
2
3
4
5 Steckler, T., Drinkenburg, W. H., Sahgal, A., & Aggleton, J. P. (1998). Recognition
6 memory in rats--II. Neuroanatomical substrates. *Progress in Neurobiology*, 54(3), 313-
7 332. doi:10.1016/s0301-0082(97)00061-0
8
9

10
11 Stevenson, R. J., Francis, H. M., Attuquayefio, T., Gupta, D., Yeomans, M. R., Oaten, M.
12 J., & Davidson, T. (2020). Hippocampal-dependent appetitive control is impaired by
13 experimental exposure to a Western-style diet. *Royal Society Open Science*, 7(2),
14 191338. doi:10.1098/rsos.191338
15
16

17
18 Tran, D. M., & Westbrook, R. F. (2015). Rats Fed a Diet Rich in Fats and Sugars Are
19 Impaired in the Use of Spatial Geometry. *Psychological Science*, 26(12), 1947-1957.
20 doi:10.1177/0956797615608240
21
22

23
24 Tran, D. M. D., & Westbrook, R. F. (2018). Dietary effects on object recognition: The
25 impact of high-fat high-sugar diets on recollection and familiarity-based memory. *Journal*
26 *of Experimental Psychology: Animal Learning and Cognition*, 44(3), 217-228.
27 doi:10.1037/xan0000170
28
29

30
31 Vianna, M. R., Alonso, M., Viola, H., Quevedo, J., de Paris, F., Furman, M., . . . Izquierdo,
32 I. (2000). Role of hippocampal signaling pathways in long-term memory formation of a
33 nonassociative learning task in the rat. *Learning & Memory*, 7(5), 333-340.
34 doi:10.1101/lm.34600
35
36

37
38 Voigt, J., Hörtnagl, H., Rex, A., van Hove, L., Bader, M., & Fink, H. (2005). Brain
39 angiotensin and anxiety-related behavior: the transgenic rat TGR(ASrAOPEN)680. *Brain*
40 *Research*, 1046(1-2), 145-156. doi: 10.1016/j.brainres.2005.03.048
41
42

43
44 Warneke, W., Klaus, S., Fink, H., Langley-Evans, S. C., & Voigt, J. P. (2014). The impact
45 of cafeteria diet feeding on physiology and anxiety-related behaviour in male and female
46 Sprague-Dawley rats of different ages. *Pharmacology Biochemistry & Behavior*, 116, 45-
47 54. doi:10.1016/j.pbb.2013.11.016
48
49

50
51 Westbrook, S. R., Brennan, L. E., & Stanton, M. E. (2014). Ontogeny of object versus
52 location recognition in the rat: acquisition and retention effects. *Developmental*
53 *Psychobiology*, 56(7), 1492-1506. doi:10.1002/dev.21232
54
55

56
57 Wilson, W., Voigt, P., Bader, M., Marsden, C. A., & Fink, H. (1996). Behaviour of the
58 transgenic (mREN2)27 rat. *Brain Reserach*, 729(1), 1-9. doi.org/10.1016/0006-
59 8993(96)00114-X
60

1
2
3 Wright, T., Langley-Evans, S. C., & Voigt, J. P. (2011a). The impact of maternal cafeteria
4 diet on anxiety-related behaviour and exploration in the offspring. *Physiology & Behavior*
5 103(2), 164-172. doi:10.1016/j.physbeh.2011.01.008
6
7

8
9 Wright, T. M., Fone, K. C., Langley-Evans, S. C., & Voigt, J. P. (2011b). Exposure to
10 maternal consumption of cafeteria diet during the lactation period programmes feeding
11 behaviour in the rat *International Journal of Developmental Neuroscience*, 29(8), 785-
12 793. doi:10.1016/j.ijdevneu.2011.09.007
13
14

15
16 Wright, T. M., King, M. V., Davey, W. G., Langley-Evans, S. C., & Voigt, J. P. (2014).
17 Impact of cafeteria feeding during lactation in the rat on novel object discrimination in
18 the offspring. *British Journal of Nutrition*, 112(12), 1933-1937.
19 doi:10.1017/S0007114514003134
20
21

22
23 Yamaguchi, S., & Knight, R. T. (1991). P300 generation by novel somatosensory stimuli.
24 *Electroencephalography and Clinical Neurophysiology*, 78(1), 50-55. doi:10.1016/0013-
25 4694(91)90018-y
26
27

28
29 Yeomans, M. R. (2017). Adverse effects of consuming high fat-sugar diets on cognition:
30 implications for understanding obesity. *Proceedings of the Nutrition Society*, 76(4), 455-
31 465. doi:10.1017/S0029665117000805
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34 35 Data sharing statement 36

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39 The data that support the findings of this study are available from the corresponding
40 author upon reasonable request.
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TABLE 1 Average daily energy and macronutrient intake in lactating dams

Diet	Energy (kJ/d)		Carbohydrate total (g/d)		Sucrose (g/d)		Fat (g/d)		Protein (g/d)	
	Mean	SEM	Mean	SEM	Mean	SEM	Mean	SEM	Mean	SEM
Chow	786.20	20.49	26.79 (64.06)	0.70	2.12	0.05	3.76 (8.99)	0.10	11.27 (26.95)	0.29
Cafeteria	1001.00 ****	19.77	22.24 ** (50.29)	1.06	4.91 ****	0.27	11.99 **** (27.11)	0.35	9.99 * (22.59)	0.43

Data represent mean values from 12 dams/group as collected over 21 days of lactation.

Percent of total macronutrient intake are given in brackets. Student's t-test.

*P<0.05, **P<0.01, ***P < 0.001. ****P < 0.0001 vs. chow fed controls.

TABLE 2 Exploration time of the two objects during training (sampling period)

Intertrial interval	Diet	Exploration time (s) during familiarisation Male offspring		n	Exploration time (s) during familiarisation Female offspring		n
		Mean	SEM		Mean	SEM	
5 min	C	52	8.1	9	50	3.2	11
	CD	52	3.6	10	61	4.2	9
60 min	C	38	6.0	7	40	3.3	7
	CD	40	3.0	6	38	2.5	6
24 h	C	52	7.3	6	46	7.0	8
	CD	55	7.0	9	53	3.4	10

Familiarisation data for each of the three time points have been analysed separately. No significant effects of sex and diet have been observed. Sex: 5 min: $F(1, 35) = 1.18, P=0.28$; 60 min: $F(1, 22) = 0.01, P=0.92$; 24 h: $F(1, 29) = 0.36, P=0.55$. Diet: 5 min: $F(1, 35) = 0.67, P=0.42$; 60 min: $F(1, 22) = 0.01, P=0.92$; 24 h: $F(1, 29) = 0.75, P=0.39$. Two-Way ANOVA (diet, sex).

TABLE 3 Exploration times of the two objects during first exposure in the recency test

Diet	Exploration time (s) first exposure Male offspring		n	Exploration time (s) first exposure Female offspring		n
	Mean	SEM		Mean	SEM	
C	36.2	2.9	7	44.6	2.9	7
CD	46.4	6.4	6	44.4	7.9	5

No significant effects of sex ($F(1, 21) = 0.41, P=0.5299$) and diet ($F(1, 21) = 1.009, P=0.33$) have been observed. Two-Way ANOVA (diet, sex).

LEGENDS

FIGURE 1 Impact of lactational cafeteria diet (CD) on open field habituation in 24-26 day old weaner rats. Control (C) dams were fed on show. Both groups travelled a shorter distance (A) (** $P < 0.001$) and showed significantly reduced rearing (B) (** $P < 0.01$) upon a second exposure (effect of time) to the open field arena. Left columns of each pair represent the first exposure to the open field, right columns the second exposure 24 hrs later. No effects of diet on either rearing ($F(1, 26) = 1.41, P = 0.24$) or locomotion ($F(1, 26) = 1.07, P = 0.31$) were observed. Mean \pm SEM. Two-Way RM ANOVA (time, diet). $N = 16$ (C) and 12 (CD).

FIGURE 2 Impact of lactational cafeteria diet (CD) on object location memory in 24-26 day old weaner rats. Values over 0.5 (dotted line) represent memory. In controls (C), memory occurred both after a 5 min (* $P < 0.05$) or 60 min (** $P < 0.001$) inter-trial interval (ITI). After the 60 min interval, the difference between C and CD was significant (+ $P < 0.05$). Memory did not occur in the CD group at any time point 5 min ($t = 1.10, df = 18, P = 0.28$), 60 min ($t = 1.34, df = 11, P = 0.21$) and 24 h ($t = 1.27, df = 13, P = 0.22$) and was not observed after 24hrs in the CD group either ($t = 0.94, df = 18, P = 0.36$). *One-sample t-test. + Student's t-test. N : 5 min C=20 CD=19; 60 min: C=14, CD=12; 24 h: C=14, CD=19.

FIGURE 3 Impact of lactational cafeteria diet (CD) on recency memory in 24-26 day old weaner rats. Values over 0.5 represent learning. Controls (C; $n = 14$) explore the distant object longer than the more recent object, whereas such a learning effect is not observed in the cafeteria group (CD; $n = 11$; $t = 1.55, df = 10, P = 0.15$). Both groups differ significantly in their exploration ratios. **** $P < 0.0001$. One-sample t-test. + $P < 0.05$. Student's t-test.

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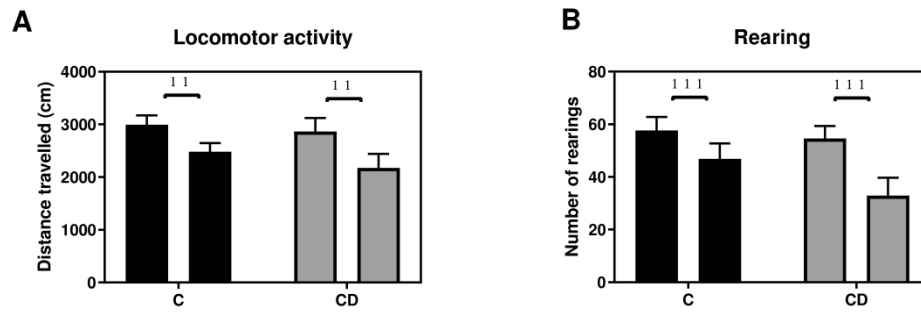


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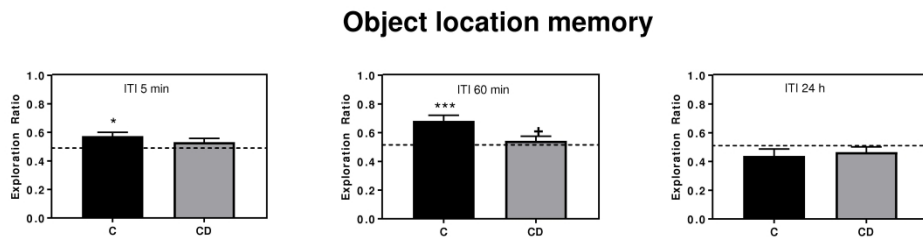
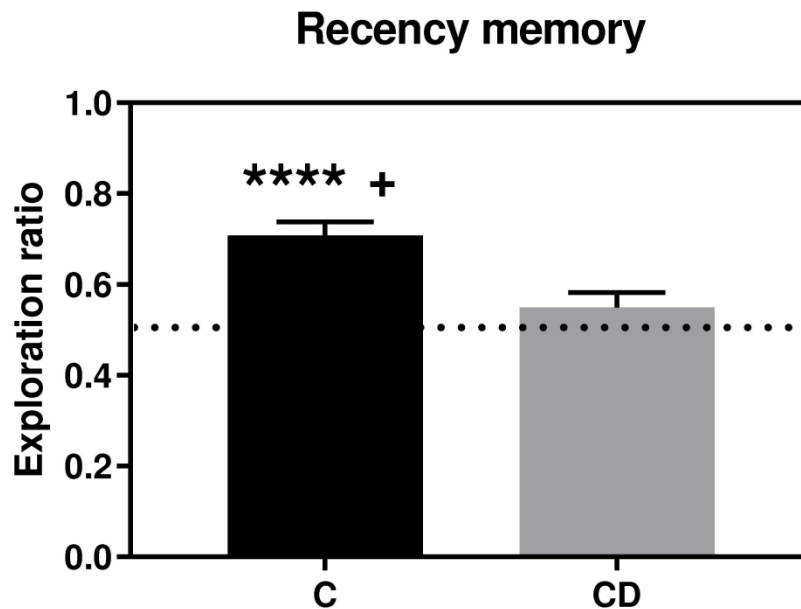


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