Supplementary Figures:

Supplementary Figure 1 Study design for X chromosome analyses



STAGE 1 (genome-wide association studies)

The discovery stage (stage 1) included 14 studies and 33,009 individuals. One variant was followed up in stage 2 in 3 studies and 52,359 individuals.

Supplementary Figure 2 Association test statistics for FEV1, FEV1/FVC and FVC



a) Quantile-quantile plots. QQplots show –log₁₀(*P*) of observed genome-wide association results against expected results in case of no association for autosomal chromosomes for FEV₁, FEV₁/FVC and FVC. Only variants with effective sample size (product of sample size and imputation quality summed up across studies) ≥70% are included.

 FEV_1





FEV₁/FVC





FVC



b) Regional association plots for 16 new lung function signals. –log10(P) in stage 1 metaanalysis for the trait with strongest association are plotted against chromosomal position (NCBI 37) for 1Mb regions. Sentinel variants in each plot and represented in purple. Other variants are coloured according to their correlation with the sentinel variant (see legend in each plot). Annotation for each variant is presented as default in locuszoom (http://csg.sph.umich.edu/locuszoom/): framestop and splice, triangle; non synonymous, inverted triangle; synonymous and UTR, square; TFBScons , star; MCS44 Placental, square with diagonal lines and None-of-the-above, filled circle.



rs10850377 (FEV1)

rs7155279 (FEV1)











Cohorts





PIVUS

7

rs117068593 (FEV1)

rs134041 (FEV1)







Cohorts



Cohorts

Cohorts

9

rs148274477 (FEV1/FVC)

rs34886460 (FEV1/FVC)

rs12149828 (FEV1/FVC)







rs148274477 (FEV1/FVC)



Cohorts





ECRHS

PIVUS

Cohorts

rs113473882 (FEV1/FVC)

rs7050036 (FEV1/FVC)



rs113473882 (FEV1/FVC) rs7050036 (FEV1/FVC) - 1 1 - 1 ECRHS PIVUS 1 PIVUS TwinsUK 1 TwinsUK Cohorts UK BILEVE UK BILEVE Summary Summary Г Г -0.8 -0.4 0.0 -0.08 -0.04 0.00 0.08 -0.6 -0.2 0.2 0.04 Effects Effects

Cohorts







Cohorts

Cohorts

0.00 0.05 0.10 0.15

Effects



12

c) Forest plots for the 16 loci associated with lung function for stage 1 and stage 2 separately. Each of the SNPs included in the figure showed genome-wide significant association (*P*<5×10⁻⁸) with either FEV₁, FEV₁/FVC or FVC after meta-analysing stages 1 and 2. The contributing effect (transformed beta) from each study is shown by a square, with confidence intervals indicated by horizontal lines. The contributing weight of each study to the meta-analysis is indicated by the size of the square. The combined meta-analysis estimate is shown at the bottom of each graph.

Supplementary Figure 3 Transcriptomic profiling of candidate lung function genes in primary human

bronchial epithelial cells.











Transcript ID	Percent of total pool
TCONS_00071386	Between 1 and 5%
TCONS_00071385	Between 1 and 5%
TCONS_00071384	Less than 1%
TCONS_00071383	Between 1 and 5%
TCONS_00071382	Between 1 and 5%
TCONS_00071381	Less than 1%
TCONS_00071380	Less than 1%
TCONS_00071379	Less than 1%
TCONS_00071378	Between 1 and 5%
TCONS_00071377	Between 5 and 10%
TCONS_00071376	Between 60 and 70%
TCONS_00071375	Less than 1%
TCONS_00071374	Between 5 and 10%
TCONS_00071373	Between 1 and 5%
TCONS_00071372	Less than 1%



Transcript ID	Percent of total pool
TCONS_00095005	Less than 1%
TCONS_00095004	Between 20 and 30%
TCONS_00095003	Less than 1%
TCONS_00095002	Between 70 and 80%
TCONS_00095001	Less than 1%
TCONS_00095000	Less than 1%

		Transcript ID	Percent of total pool
	B	TCONS_00125914	Less than 1%
		TCONS_00125913	Less than 1%
CONF 0013	5914	TCONS_00125912	Between 1 and 5%
TCONS_00125	\$913	TCONS_00125911	Between 1 and 5%
CON5_00125912	- 6	TCONS_00125910	Between 1 and 5%
CON5_00125911	- 6	TCON5_00125909	Between 1 and 5%
ICONS 00125910		TCONS 00125908	Between 5 and 10%
TCONS_00125909	1	TCONS 00125907	Less than 1%
ICONE 00123908	1	TCONS 00125906	Between 1 and 5%
ECON5 00125907		TCONS 00125905	Between 1 and 5%
ICONS COLIZSION	1	TCONS 00125904	Between 1 and 5%
1000 00123903	1	TCONS 00125903	Less than 1%
1005 0013904	1	TCONS 00125902	Less than 1%
1CON3 00123903		TCONS 00125901	Between 10 and 20
10N5 00125902	- ř	TCONS 00125900	Less than 1%
BONS 0013901 + 1 + 1 + 1	1	TCONS 00125899	Less than 1%
KONE ON STATE		TCONS 00125898	Between 1 and 5%
TOTA OUT OF THE OWNER	i i	TCONS 00125897	Between 5 and 10%
	1	TCONS 00125896	Less than 1%
		TCONS 00125895	Less than 1%
Prove courses	1	TCONS 00125894	Less than 1%
	Ē	TCONS 00125893	Less than 1%
	-	TCONS 00125892	Retween 1 and 5%
TONE OUT OF		TCONS_00125891	Loss than 1%
	-	TCONS_00125891	Less than 1%
TCOM5 00125890		TCONE 00125890	Less than 1%
TONS OUTSERS	-	TCONS_00125889	Less than 176
TCONS 00125888		TCONS_00125888	Between 5 and 107
ICONS ODI25887	-	TCONS_00125887	Less than 1%
#60N5-00125886	- 1	TCONS_00125886	Between 1 and 5%
260N5-00125885	3 -	TCONS 00125885	Between 20 and 50
ICONS 00125884	-	TCONS_00125884	Between 1 and 5%
ACONS 00125883	3 -	TCONS_00125883	Less than 1%
TCONS 00125882	-	TCONS_00125882	Less than 1%
7G045-00125881	-	TCONS_00125881	Between 1 and 5%
ACONS,09125880	-	TCONS_00125880	Less than 1%
ECON5_09125829	1-	TCONS_00125879	Between 1 and 5%
ECOM5_091,358;78	2 -	TCONS_00125878	Less than 1%
ECONS_00125677	5	TCONS_00125877	Between 5 and 10%
ECONS.00125826 A	4	TCONS_00125876	Less than 1%
TCONS 00125875	- 2	TCONS_00125875	Less than 1%
TCONS 00125874	5 -	TCONS_00125874	Between 1 and 5%
TCONS_00125873	4	TCONS_00125873	Less than 1%
TCONS_00125822		TCONS_00125872	Less than 1%
TCONS 00125871 B118 8 83 88 8 8 8 8 8 8 8 8 8 8 8 8 8 8	1	TCONS_00125871	Less than 1%
TCONS_00125870 8 81 18 8 8 8 8 8 8 8 8 8 8 8 8 8 8 8	_	TCONS_00125870	Less than 1%
TCONS 00125869 8 81 18 8 8 8 8 8 8 8 8 8 8 8 8 8 8 8	3	TCONS_00125869	Less than 1%
ECONS 00125868 8 8118 8 88 88 8 8 8 8 8 8 8 8 8 8 8	~	TCONS_00125868	Between 1 and 5%
TCONS 00125867	2	TCONS_00125867	Less than 1%
TCONS 00125846 B B118 B1 B B B B B B B B B B B B B B B		TCONS_00125866	Less than 1%
TCONS 00125845 0 0110 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0		TCONS_00125865	Less than 1%
TCONS 00125864 8 81 18 8 8 8 8 8 8 8 8 8 8 8 8 8 8 8		TCONS_00125864	Less than 1%
ACONS 00125863	12	TCONS_00125863	Less than 1%
ECONS_00125862		TCONS_00125862	Less than 1%
#CONS_00125861	3	TCONS_00125861	Less than 1%
	-		20

Anovel isoforms



Transcript ID Percent of total pool

COM2_00108911	Less than 1%
CONS_00168916	Less than 1%
CONS_00168915	Between 40 and 50%
CONS_00168914	Less than 1%
CONS_00168913	Between 50 and 60%
CONS_00168912	Less than 1%



Figures show novel and previously described (Ensembl) mRNA isoforms and their percent abundance. A: Individual predicted gene's isoforms with indicated splice variation identified using RNA-seq reference annotation based transcript assembly. Different splice events are described in the box beneath the main graph and novel transcripts are indicated by a star. X-axis contains two outmost genomic coordinates. B: Percent abundance of individual transcripts in primary human bronchial epithelial cells (passage 3) grown under basal conditions. Supplementary Figure 4 Comparison of results before and after excluding individuals with asthma



Association results are presented for the 16 novel sentinel variants with the trait that showed the strongest association, for all individuals ("All", N=48,943) and after removing individuals with asthma ("Asthma free", N=41,455) using UK BiLEVE data.

Supplementary Figure 5 Comparison of results obtained with 1000 Genomes variants and with

HapMap for autosomal signals

a) Region plots for the new signals using HapMap and 1000 Genomes variants



i. rs6681426

ii. rs201204531 (chr1:219963090:I)



RDH1

←NT5C1B

18.6

iii. rs61067109



iv. rs6441207



v. rs6856422



vi. rs148274477



22



viii. rs2274116



ix. rs11383346 (chr12:28283187:I)



x. rs10850377



xi. rs7155279



xii. rs117068593



xiii. rs12149828



xiv. rs113473882



xv. rs134041



Region plots are presented with the 1000 Genomes results including all 1000 Genomes variants with *N* effective \geq 70% (plot on the left) and including only HapMap variants with *N* effective \geq 70% extracted from the previous HapMap meta-analysis ⁵ (plot on the right) for all the new signals.

b) Scatter plot of minor allele frequencies for HapMap sentinel variants vs. 1000





Shown are MAF for sentinel variants defined using 1000 Genomes stage 1 results vs. MAF for sentinel variants defined using 1000 Genomes stage 1 results restricted to HapMap variants only, in previously reported regions¹⁻⁵, defined as the previously reported sentinel variant^{3,4} +/- 500kb either side (*MFAP2, TGFB2, EFEMP1, TNS1, HDAC4, RARB, MECOM, FAM13A, GSTCD, HHIP, SPATA9, HTR4, ADAM19, BMP6, ZKSCAN3, NCR3, AGER, ARMC2, GPR126, PTCH1, CDC123, C10orf11, HSD17B12, PRDM11, LRP1, CCDC38, THSD4, MMP15, CFDP1, WWOX, KCNJ2, KCNE2*).

Supplementary Tables:

Supplementary Table 1 Sample population characteristics and genotyping platform details for each study in stage1 and stage2

a) Sample population characteristics for each study

Characteristics are shown for studies analysed in stage 1 (GWAS meta-analysis) and stage 2 (followup). Stage 1 studies: B58C (B58C-T1DGC, British 1958 Birth Cohort–Type 1 Diabetes Genetics Consortium and B58C-WTCCC, British 1958 Birth Cohort–Wellcome Trust Case Control Consortium); BHS1&2, Busselton Health Study 1 and 2; the CROATIA- Korcula study; the CROATIA-Split study; the CROATIA-Vis study; EPIC population based, European Prospective Investigation into Cancer and Nutrition Cohort; GS:SFHS, Generation Scotland: Scottish Family Health Study; H2000, Finnish Health 2000 survey; KORA F4, Cooperative Health Research in the Region of Augsburg; KORA S3, Cooperative Health Research in the Region of Augsburg; LBC1936, Lothian Birth Cohort 1936; NFBC1966, Northern Finland Birth Cohort of 1966; NSPHS, Northern Sweden Population Health Study; ORCADES, Orkney Complex Disease Study; SHIP, Study of Health in Pomerania; YFS, the Young Finish Study. Stage 2 studies: ECRHS, the European Community Respiratory Health Survey; PIVUS, Prospective Investigation of the Vasculature in Uppsala Seniors; the TwinsUK study; UK BiLEVE, UK Biobank Lung Exome Variant Evaluation study.

Study name	N Total	N male	N female	Age range (y) at lung function measur ement	Mean age, y (s.d.)	Mean height, cm (s.d.)	Mean FEV ₁ , L (s.d.)	Mean FVC, L (s.d.)	Mean FEV₁/FV C (s.d.)	N never smoke rs	N ever smok ers	Lambda FEV ₁	Lambda FVC	Lambda factor FEV1/FVC
Stage 1														
B58C	5934	2955	2979	44-45	45.12 (0.38)	169.43 (9.29)	3.30 (0.76)	4.19 (0.98)	0.79 (0.08)	1709	4225	1.022	1.011	1.024
BHS1&2	4355	1922	2433	17-97	51.21 (17.00)	168.00 (9.39)	3.01 (0.96)	3.88 (1.16)	0.77 (0.07)	2301	2054	1.015	1.017	1.015
CROATIA- Korcula	826	302	524	18-90	55.63 (13.50)	168.10 (9.20)	2.72 (0.83)	3.29 (0.96)	0.83 (0.1)	403	423	1.003	1.018	1.017
CROATIA-Split	493	210	283	18-85	49.08 (14.63)	172.60 (9.49)	3.19 (0.91)	3.80 (1.06)	0.84 (0.08)	239	254	1.003	1.011	0.997
CROATIA-Vis	925	390	535	18-88	55.90 (15.51)	167.80 (9.88)	3.42 (1.21)	4.41 (1.42)	0.77 (0.09)	388	537	1.002	0.982	0.969
EPIC population based	2339	1102	1237	39-77	59.20 (9.00)	167.10 (8.80)	2.50 (0.72)	3.04 (0.90)	0.83 (0.11)	1062	1277	1.005	1.008	1.014
GS:SFHS	8093	3385	4708	18-98	51.60 (13.34)	167.90 (9.55)	2.78 (0.87)	3.91 (1.01)	0.71 (0.01)	4319	3774	1.003	1.014	1.013
H2000	821	394	427	30-75	50.47 (10.91)	169.10 (9.14)	3.29 (0.9)	4.16 (1.07)	0.79 (0.07)	249	572	0.999	1.002	1
KORA F4	1474	717	757	41-84	55.08 (9.90)	169.15 (9.42)	3.23 (0.85)	4.19 (1.05)	0.77 (0.07)	556	918	0.998	1.003	1.008
KORA S3	1147	551	596	28-89	50.82 (15.23)	169.22 (9.32)	3.34 (0.90)	4.10 (1.06)	0.81 (0.08)	520	627	1.009	1.012	1.018
LBC1936	991	501	490	68-71	69.55 (0.84)	166.44 (8.93)	2.38 (0.67)	3.04 (0.87)	0.79 (0.10)	437	554	1.011	1.05	1.002
NFBC1966	4563	2180	2383	31-31	31.00 (0)	171.00 (9.54)	3.93 (0.80)	4.71 (0.99)	0.84 (0.07)	1648	2915	1.005	0.99	0.996
NSPHS	871	400	471	14-91	49.20 (20.00)	164.00 (10.10)	2.92 (0.90)	3.53 (1.06)	0.83 (0.09)	750	121	0.996	1.011	0.977
ORCADES	1802	719	1083	16-91	54.00 (15.00)	166.00 (9.20)	2.89 (0.83)	3.61 (0.99)	0.79 (0.08)	1022	780	1.016	1.013	1.009
SAPALDIA	1378	665	713	18-61	41.30 (11.20)	169.47 (9.12)	3.53 (0.86)	4.50 (1.04)	0.78 (0.08)	631	747	0.995	1.001	1.012
SHIP	1768	863	905	25-85	52.43 (13.58)	169.66 (9.12)	3.28 (0.89)	3.87 (1.03)	0.85	770	998	1.019	1.007	1.005
YFS	419	198	221	30-47	38.88 (5.07)	172.25 (8.90)	3.73 (0.75)	4.68 (0.99)	0.8 (0.06)	233	186	1.012	1.02	0.997
Total stage 1 sample size	38199									17237	20962			

Study name	N Total	N male	N female	Age range (y) at lung function measur ement	Mean age, y (s.d.)	Mean height, cm (s.d.)	Mean FEV ₁ , L (s.d.)	Mean FVC, L (s.d.)	Mean FEV₁/FV C (s.d.)	N never smoke rs	N ever smok ers	Lambda FEV ₁	Lambda FVC	Lambda factor FEV ₁ /FVC
Stage 2														
ECRHS	1747	823	924	20-48	33.97 (7.16)	170.32 (9.47)	3.73 (0.82)	4.54 (1.03)	0.82 (0.07)	763	984	NA	NA	NA
PIVUS	837	406	431	70-72	70.20 (0.17)	168.93 (9.21)	2.44 (0.68)	3.21 (0.87)	0.76 (0.10)	409	428	NA	NA	NA
TwinsUK	3023	444	2579	19-82	53.08 (13.15)	161.68 (6.21)	2.63 (0.61)	3.28 (0.65)	0.80 (0.08)	1865	1158	NA	NA	NA
UK BILEVE	48943	2448 9	24454	40-70	56.93 (7.89)	168.7 (9.13)	2.65 (0.87)	3.59 (1.05)	0.73 (0.08)	24483	24460	NA	NA	NA
Total stage 2 sample size	54550									27520	27030			

b) Study, genotyping, imputation and genotype-phenotype data

Imp'n, imputation.

Study name	GWAS platform	Calling algorithm	Individu al call rate filter (applied before imp'n)	SNP call rate filter (applied before imp'n)	SNP HWE <i>P</i> filter (applied before imp'n)	SNP MAF filter (applied before imp'n)	SNP MAF filter (applied before imp'n)		Imputation software and version	NCBI; 1000 Genomes version for imp'n	Genotype- phenotype association software
Stage 1		1	T	I	T	I		I	I	1	1
B58C	Illumina 550k/610k	GenCall	None	>=95%	≥0.0001 (tested on females only for chromos ome X)	≥1%	Consistent allele frequencies across data deposits ($P \ge 0.0001$ for pairwise comparisons) and for chrX SNPs, consistent allele frequencies between males and females ($P \ge 0.0001$).	500,521 (including 11,696 chrX)	MACH 1.0.18 & Minimac 2012-11-16	Phase 1 March 2012	probABEL 0.1-9e
BHS1&2	Illumina 610-Quad (N=1,168) & Illumina 660W- Quad (N=3,428)	BeadStudi o (N=1,168) & GenomeS tudio (N=3,428)	0.95	0.95	1.00E-06	0.01	Individuals were removed if they had sex inconsistencies, had heterozygosity > 5 s.d. from the mean, were PCA outliers, were 1 individual from a pair of duplicates or had IBD inconsistencies.	521,307	Minimac and MACH1 v1.0.18	b37; Phase 1 March 2012	ProbABEL
CROATIA- Korcula	Illumina HumanHa p370CNV duo chip	BeadStudi o	97%	98%	1.00E-06	0.01		316,879	SHAPEIT2, IMPUTE2	b37; ALL (Phase 1 integrated release v3, April 2012)	ProbABEL
CROATIA- Split	Illumina HumanHa p370CNV quad chip	BeadStudi o	97%	98%	1.00E-06	0.01		321,727	SHAPEIT2, IMPUTE2	b37; ALL (Phase 1 integrated release v3, April 2012)	ProbABEL

Study name	GWAS platform	Calling algorithm	Individu al call rate filter (applied before imp'n)	SNP call rate filter (applied before imp'n)	SNP HWE <i>P</i> filter (applied before imp'n)	SNP MAF filter (applied before imp'n)	Other filters	No of SNPs after filtering (before imp'n)	Imputation software and version	NCBI; 1000 Genomes version for imp'n	Genotype- phenotype association software
CROATIA- Vis	Illumina Infinium HumanHa p300 BeadChip	BeadStudi o	97%	98%	1.00E-06	0.01		273,671	SHAPEIT2, IMPUTE2	b37; ALL (Phase 1 integrated release v3, April 2012)	ProbABEL
EPIC population based	Affymetrix 500K	BRLMM	94%	90%	1.00E-06	0.01	Heterozygosity, ethnic outlier	392,573	IMPUTE 2.2.2	Mar-12	SNPTEST 2.4.0
GS:SFHS	Illumina OmniExpr ess+Exom e	GenomeS tudio	97% (Omni); 99% (Exome)	98%	1.00E-06	0.01 (Omni); 0.0001 (Exome)		690,759	SHAPEIT2, IMPUTE2	b37; ALL (Phase 1 integrated release v3, April 2012)	ProbABEL
H2000	Illumina HumanHa p 610K	Illuminus	0.95	0.95 (0.99 for SNPs with MAF < 0.05)	1.00E-06	0.01		553,722	IMPUTE version 2.2.2	1,000 Genomes haplotypes Phase I integrated variant set release (v3) in NCBI build 37 (hg19) coordinates	SNPTest
KORA F4	Affymetrix Axiom	Affymetrix Software	0.97	0.98	5x10-6	0.01	-missmatch of phenotypic and genetic gender - 5s.d. from mean heterozygosity rate - check for European ancestry - check for population outlier	523,260 (chr 1-26) 508,532 (chr 1-22) 14,096(chrX- nonPAR) 444(chrX- PAR1) 58(chrX- PAR2)	SHAPEIT v2, IMPUTE v2.3.0	1000g phase1 all (ALL_1000G_ph ase1integrated_ v3_impute_mac 1)	SNPTEST v2.4.1

Study name	GWAS platform	Calling algorithm	Individu al call rate filter (applied before imp'n)	SNP call rate filter (applied before imp'n)	SNP HWE <i>P</i> filter (applied before imp'n)	SNP MAF filter (applied before imp'n)	Other filters	No of SNPs after filtering (before imp'n)	Imputation software and version	NCBI; 1000 Genomes version for imp'n	Genotype- phenotype association software
KORA S3	Illumina Omni 2.5/ Illumina Omni Express	Genome Studio	0.97	0.98	5x10-6	0.01	person wise: - missmatch of phenotypic and genetic gender - 5s.d. from mean heterozygosity rate - check for European ancestry - check for population outlier SNP wise: only SNPs that were genotyped with good quality on both chips	600641 (chr 1-26) 588307 (chr 1-22) 14625 (chrX- nonPAR)	SHAPEIT v2, IMPUTE v2.3.0	1000g phase1 all (ALL_1000G_ph ase1integrated_ v3_impute_mac 1)	SNPTEST v2.4.1
LBC1936	Illumina 610- Quadv1	GenomeS tudio	0.95	0.98	≥0.001	0.01		549,692	minimac 2012-11-16	version 3, cosmopolitan	mach2qtl
NFBC1966	Illumina HumanCN V- 370DUO Analysis BeadChip	Beadstudi o	95%	95 % (99 % if MAF < 5 %)	1.00E-04	0.01	Duplicates concordance < 99%; IBS pairwise sharing>0.20; withdrew consent; gender mismatch; heterozygosity outliers>0.29; MDS outliers.	309,948	Impute version 2.3.0	1000G March 2012 pphase1 integrated v3	SNPTEST
NSPHS	Illumina Infinum HapMap 300 v2 & Illumina Human OmniExpr ess	BeadStudi o (Infinum) & GenoneSt udio (OmniExp ress)	0.9	0.95	3.2E-08 (Infinum) & 1.4E- 08 (OmniEx press)	0.01	FDR level of heterozygosity 0.01	306,086 (Infinum) & 631503 (OmniExpres s)	Impute2 (v 2.2.2)	hg19	ProbABEL

Study name	GWAS platform	Calling algorithm	Individu al call rate filter (applied before imp'n)	SNP call rate filter (applied before imp'n)	SNP HWE <i>P</i> filter (applied before imp'n)	SNP MAF filter (applied before imp'n)	Other filters	No of SNPs after filtering (before imp'n)	Imputation software and version	NCBI; 1000 Genomes version for imp'n	Genotype- phenotype association software
ORCADES	Illumina Hap300, Illumina Omni1 & Illumina OmniX	Beadstudi o using Hap300v2 cluster file (Hap300) & Genome Studio, using Illumina cluster files (Omni1 & OmniX)	98%	97%	1.00E-06	1% (Hap300) & monomor phic (Omni & OmniX)	Subject Heterozygosity FDR<1%	287,208 (Hap300), 843723 (Omni) & 654651 (OmniX)	shapeit.v2.r6 44.+impute_ v2.2.2_x86_ 64_static/im pute2	1000G Phase I Integrated Release Version 3 Haplotypes (2010-11 data freeze, 2012-03- 14 haplotypes).	probABEL v. 0.4.3
SAPALDIA	Illumina 610k quad	Gencall	95%	95%	1.00E-06	0.01	none	545,131	Mach 1.0.16.a, minimac- omp RELEASE STAMP 2012-05-29 (autosomes) & MiniMac RELEASE STAMP 2012-11-16 (chr X)	build37	probABEL
SHIP	Affymetrix SNP 6.0	Birdseed2	>92%	>80%	>0.0001	none	duplicate SNP positions, sex- mismatch or duplicate individuals (by IBD)	905,910	IMPUTE v2.2.2	b37, 1000Gv3 (Mar 2012)	QUICKTEST v0.95
YFS	Illumina 670k custom	Illuminus	0.95	0.95	1.00E-06	0.01	heterozygosity, relatedness	546,674	SHAPEIT v1 and IMPUTE v2.2.2	Phase 1, release v3, March 2012 haplotypes	SNPTEST v.2.4.1

Study name	GWAS platform	Calling algorithm	Individu al call rate filter (applied before imp'n)	SNP call rate filter (applied before imp'n)	SNP HWE <i>P</i> filter (applied before imp'n)	SNP MAF filter (applied before imp'n)	Other filters	No of SNPs after filtering (before imp'n)	Imputation software and version	NCBI; 1000 Genomes version for imp'n	Genotype- phenotype association software
Stage 2											
ECRHS	Human 610 Quad Chip (ILLUMIN A)	BeadStudi o	exclusion of males with high X heterozy gosity	95%	0.0001	1%	sex discrepancies, IBS analysis for relatedness, ancestry analysis using PCA	567,512	IMPUTE2	version 3	SNPTEST v2
PIVUS	Illumina OmniExpr ess and Metabocgi p	Bead Studio	≥0.95	≥0.95 (≥0.99 for MAF<0.0 5)	10-6	≥0.01		738,606	SHAPEIT and IMPUTE2	Mar-12	SNPTEST
TwinsUK	Illumina 300K Duo for and HumanHa p610- Quad		Illumina GenCall	if MAF>5% then 0.95, else 0.99	E-06	1%		unavailable	Impute 2	Mar-12	Gemma
UK BILEVE	Affymetrix Axiom UK BiLEVE array	Axiom® GT1 algorithm (Affymetri x Power Tools v1.15.1)	95%	None	1.00E-06	1%	Het. outliers, PCA outliers, sex mismatch, relateds (IBD)	unavailable (regional imputation undertaken)	SHAPEIT and IMPUTE2	Mar-12	SNPTEST

Supplementary Table 2 Follow-up results

a) Association results for 56 variants followed up in stage 2

Results for stage 1, stage 2 and the meta-analysis of both are presented for the 56 variants (55 from autosomal chromosomes and 1 from chromosome X) followed up in stage 2. P<5x10⁻⁸ after meta-analysing stage 1 and stage 2 are highlighted in bold. rs189059189 and rs117068593 appear twice on the table because they were selected for follow-up for two different traits. Chr., chromosome; dist., distance.

			Coded allele/ Non	Stage 1	•	•	Stage 2	•	•	Stage 1 + stage 2		
rs number (chr: position)	Gene (function)	Measure	coded allele	Beta	SE	Р	Beta	SE	Р	Beta	SE	Р
rs10798992 (chr1:34268324)	CSMD2 (intronic)	FEV ₁ /FVC	C/A	0.041	0.008	1.25E-07	0.002	0.006	7.14E-01	0.017	0.005	3.02E-04
	<i>CDC7</i> (dist=50181), <i>TGFBR3</i>						0.002	0.000			0.000	0.022 01
rs4658231 (chr1:92041502)	(dist=104398)	FVC	A/G	0.051	0.009	5.33E-09	0.011	0.007	1.20E-01	0.027	0.006	8.36E-07
rs2764553 (chr1:111732303)	DENND2D (intronic)	FEV ₁ /FVC	C/A	0.044	0.010	3.73E-06	0.020	0.008	7.27E-03	0.030	0.006	6.37E-07
rs4926386 (chr1:149886135)	SV2A (intronic)	FEV ₁	C/A	-0.062	0.012	1.99E-07	-0.030	0.010	3.58E-03	-0.044	0.008	2.09E-08
rs6681426 (chr1:150586971)	MCL1 (dist=34757), ENSA (dist=7628)	FEV ₁	G/A	0.042	0.008	1.07E-07	0.021	0.006	1.13E-03	0.029	0.005	4.35E-09
rs201204531 (chr1:219963090)	LYPLAL1 (dist=576883), RNU5F-1 (dist=83529)	FEV ₁ /FVC	A/ATG	-0.038	0.008	1.98E-06	-0.027	0.006	1.77E-05	-0.031	0.005	2.68E-10
rs61067109 (chr2·18292452)	<i>KCNS3</i> (dist=178227), <i>NT5C1B-RDH14</i> (dist=443537)	FEV./EV/C	G/A	-0.050	0.009	3 09E-08	-0.042	0.007	6 56E-09	-0.045	0.006	1 40E-15
rs1080871 (chr2:/685/708)				0.035	0.003	4 70E 06	0.042	0.006	2 28E 01	0.040	0.000	1.565.04
rs13386197 (chr2:47798588)	<i>KCNK12</i> (dist=1118), <i>MSH6</i> (dist=211633)	FVC	C/G	0.035	0.000	1.65E-06	0.011	0.000	4.44E-01	0.039	0.005	7.48E-04
	<i>INSIG2</i> (dist=343750), <i>LOC101927709</i>		07	0.400	0.007	4.045.00	0.000	0.000	0.005.04	0.055	0.004	1.005.00
rs185216854 (cnr2:119211347)	(dist=146012) CWC22 (dist=643603)	FEV ₁ /FVC	G/T	0.180	0.037	1.31E-06	-0.029	0.030	3.38E-01	0.055	0.024	1.99E-02
rs74445319 (chr2:181515383)	SCHLAP1 (dist=41448)	FVC	C/T	0.102	0.022	4.68E-06	0.021	0.014	1.49E-01	0.044	0.012	2.23E-04
rs11432697 (chr3:89210365)	EPHA3 (intronic)	FVC	T/TA	0.042	0.009	4.05E-06	0.018	0.007	1.75E-02	0.027	0.006	1.97E-06
rs10935473 (chr3:98416900)	CPOX (dist=104445), ST3GAL6-AS1 (dist=16277)	FVC	G/T	-0.038	0.008	4.87E-06	-0.007	0.006	2.76E-01	-0.018	0.005	3.37E-04
rs7356112 (chr3:143970905)	C3orf58 (dist=259695), PLOD2 (dist=1816323)	FVC	C/T	0.104	0.021	6.05E-07	0.011	0.015	4.83E-01	0.044	0.012	4.24E-04
rs6441207 (chr3:158282459)	RP11-538P18.2 (ncRNA_intronic)	FVC	C/T	0.036	0.008	4.54E-06	0.036	0.006	5.85E-09	0.036	0.005	1.27E-13
rs73223499 (chr4:14300501)	<i>LINC01085</i> (dist=158825), <i>CPEB2-AS1</i> (dist=611084)	FEV ₁ /FVC	C/T	-0.080	0.017	3.99E-06	-0.037	0.015	1.42E-02	-0.055	0.011	1.15E-06
rs35288667 (chr4:16544514)	LDB2 (intronic)	FVC	A/G	0.049	0.010	1.55E-06	-0.005	0.008	5.01E-01	0.015	0.006	1.81E-02
rs6856422 (chr4:106841962)	NPNT (intronic)	FEV ₁ /FVC	G/T	-0.044	0.008	1.30E-07	-0.055	0.006	1.15E-17	-0.051	0.005	1.51E-23
rs11269742 (chr4:177758035)	VEGFC (dist=44136), NEIL3 (dist=472956)	FVC	T/TGGGA AAAAAAT AC	0.043	0.009	9.91E-07	0.000	0.007	9.78E-01	0.017	0.005	2.19E-03
rs112226333 (chr5:31525207)	DROSHA (intronic)	FEV ₁	G/T	0.051	0.010	6.12E-07	0.016	0.008	4.42E-02	0.029	0.006	3.32E-06
rs11960365 (chr5:79650724)	CRSP8P (dist=2939), ZFYVE16 (dist=53108)	FVC	G/T	0.050	0.010	2.15E-06	-0.008	0.008	3.30E-01	0.014	0.006	3.25E-02
rc140178028 (chr5·106100235)	RAB9BP1 (dist=1673436), LOC102467213 (dist=41663)	FEV.	C/A	0 132	0.027	1 145-06	0.018	0.021	3 82E-01	0.060	0.016	2 57E-04
(0110.10010320)	LOC100294145 (dist=14952), HLA-DMB	ı ∟ v 1		0.132	0.021	1.142-00	0.010	0.021	5.02L-01	0.000	0.010	2.57 2-04
rs114372586 (chr6:32886487)	(dist=15919)	FVC	T/C	0.081	0.017	2.83E-06	0.019	0.012	1.20E-01	0.040	0.010	7.30E-05
			Coded allele/ Non	Stage 1			Stage 2			Stage 1 +	stage 2	
------------------------------	--	-----------------------	----------------------	---------	-------	----------	---------	-------	----------	-----------	---------	----------
rs number (chr: position)	Gene (function)	Measure	coded allele	Beta	SE	Р	Beta	SE	Р	Beta	SE	Р
rs79787160 (chr6:72212515)	<i>LINC00472</i> (dist=82067), <i>RIMS1</i> (dist=383891)	FEV ₁ /FVC	A/G	0.241	0.050	1.09E-06	0.020	0.038	5.93E-01	0.103	0.030	6.71E-04
rs9352253 (chr6:76687328)	IMPG1 (intronic)	FFV ₁	A/C	0.041	0.009	1.51E-06	-0.012	0.007	7.25E-02	0.008	0.005	1.25E-01
rs148274477 (chr6:142838173)	GPR126(dist=70770), <i>RP11- 440G9.1</i> (dist=9419)	FEV ₁ /FVC	С/Т	-0.161	0.027	2.68E-09	-0.162	0.019	5.63E-18	-0.162	0.015	9.58E-26
rs6570551 (chr6:143556810)	AIG1 (intronic)	FEV ₁ /FVC	A/G	-0.042	0.008	7.67E-08	-0.001	0.006	9.11E-01	-0.017	0.005	4.62E-04
rs4728570 (chr7:84728349)	SEMA3D (intronic)	FEV ₁	G/A	-0.037	0.008	3.06E-06	-0.006	0.006	3.45E-01	-0.018	0.005	2.80E-04
rs141779646 (chr7:90805723)	CDK14 (intronic)	FVC	G/C	0.087	0.019	3.80E-06	-0.016	0.016	3.23E-01	0.028	0.012	2.40E-02
rs112225874 (chr8:69228553)	LOC286189 (ncRNA_intronic)	FVC	C/T	0.114	0.025	3.84E-06	0.032	0.024	1.72E-01	0.071	0.017	2.82E-05
rs76163270 (chr9:24678364)	<i>IZUMO3</i> (dist=132690), <i>TUSC1</i> (dist=998023)	FEV ₁ /FVC	A/C	-0.079	0.017	4.65E-06	-0.018	0.014	1.80E-01	-0.042	0.011	9.64E-05
rs34886460 (chr9:119359372)	ASTN2 (intronic)	FEV ₁ /FVC	T/TA	0.041	0.008	6.75E-08	0.025	0.006	4.07E-05	0.031	0.005	4.72E-11
rs2274116 (chr9:139094805)	LHX3 (exonic)	FVC	C/T	0.041	0.009	2.68E-06	0.038	0.007	4.20E-09	0.039	0.005	5.55E-14
rs10735064 (chr12:888848)	WNK1 (intronic)	FVC	G/A	0.042	0.009	3.49E-06	0.014	0.007	5.41E-02	0.025	0.006	1.03E-05
rs12314259 (chr12:12272466)	<i>LRP</i> 6 (UTR3)	FVC	C/G	0.048	0.010	4.67E-06	-0.002	0.009	8.29E-01	0.018	0.007	6.46E-03
rs11383346 (chr12:28283187)	<i>PTHLH</i> (dist=158271), <i>CCDC91</i> (dist=126946)	FVC	A/AT	-0.043	0.008	3.83E-08	-0.041	0.006	4.42E-11	-0.042	0.005	9.52E-18
rs10850377 (chr12:115201436)	<i>TBX3</i> (dist=79467), <i>MED13L</i> (dist=1194945)	FEV ₁	G/A	-0.047	0.008	6.68E-09	-0.028	0.006	1.55E-05	-0.035	0.005	2.50E-12
rs6563127 (chr13:80216147)	NDFIP2 (dist=85935), LINC01080 (dist=369065)	FVC	A/G	-0.059	0.013	2.91E-06	-0.011	0.011	3.38E-01	-0.031	0.008	1.46E-04
rs189059189 (chr14:34231304)	NPAS3 (intronic)	FEV ₁	C/T	-0.355	0.067	1.11E-07	-0.035	0.051	4.99E-01	-0.154	0.041	1.61E-04
rs189059189 (chr14:34231304)	NPAS3 (intronic)	FVC	C/T	-0.318	0.067	2.19E-06	-0.041	0.051	4.23E-01	-0.143	0.041	4.46E-04
rs61980882 (chr14:75021464)	LTBP2 (intronic)	FVC	C/T	-0.039	0.008	3.72E-07	-0.007	0.006	2.22E-01	-0.020	0.005	3.65E-05
rs7155279 (chr14:92485881)	TRIP11 (intronic)	FEV ₁	G/T	-0.041	0.008	1.39E-07	-0.022	0.006	4.17E-04	-0.030	0.005	1.41E-09
rs117068593 (chr14:93118229)	RIN3 (exonic)	FEV ₁	C/T	-0.048	0.010	2.72E-06	-0.027	0.008	5.68E-04	-0.035	0.006	2.25E-08
rs117068593 (chr14:93118229)	RIN3 (exonic)	FVC	C/T	-0.054	0.010	1.74E-07	-0.022	0.008	5.83E-03	-0.033	0.006	8.10E-08
rs4906032 (chr14:101518910)	MIR381HG, MIR487A (downstream)	FVC	A/G	0.037	0.008	4.78E-06	0.004	0.007	5.75E-01	0.018	0.005	7.31E-04
rs8027068 (chr15:100339709)	DNM1P46 (ncRNA_intronic)	FEV ₁ /FVC	G/A	0.043	0.009	3.79E-06	-0.005	0.007	5.05E-01	0.012	0.006	2.97E-02
rs113733630 (chr16:4343067)	<i>TFAP4</i> (dist=20066), <i>GLIS2</i> (dist=39158)	FEV ₁ /FVC	A/AGTTT	0.042	0.009	3.78E-06	0.015	0.007	4.03E-02	0.025	0.006	6.92E-06
rs12149828 (chr16:10706328)	<i>EMP</i> 2 (dist=31789), <i>TEKT5</i> (dist=15033)	FEV ₁ /FVC	G/A	0.049	0.010	3.16E-06	0.035	0.008	3.33E-05	0.040	0.007	7.65E-10
rs12929007 (chr16:30929696)	FBXL19-AS1 (downstream)	FEV ₁	C/G	-0.049	0.010	6.21E-07	0.001	0.007	8.47E-01	-0.016	0.006	5.95E-03
rs7187961 (chr16:53826034)	FTO (intronic)	FVC	T/C	0.048	0.010	3.43E-06	-0.008	0.007	2.80E-01	0.011	0.006	6.76E-02
rs16953104 (chr16:54165373)	<i>FTO</i> (dist=16994), <i>IRX3</i> (dist=151839)	FEV ₁	С/Т	0.276	0.057	1.09E-06	-0.007	0.064	9.10E-01	0.152	0.042	3.35E-04

			Coded allele/ Non	Stage 1			Stage 2			Stage 1 +	- stage 2	
rs number (chr: position)	Gene (function)	Measure	coded allele	Beta	SE	Р	Beta	SE	Р	Beta	SE	Р
rs4792721 (chr17:16006827)	NCOR1 (intronic)	FEV ₁	A/G	0.038	0.008	1.67E-06	0.017	0.006	8.06E-03	0.025	0.005	4.42E-07
rs58270905 (chr18:42844617)	SLC14A2 (intronic)	FEV ₁ /FVC	A/G	0.252	0.055	4.51E-06	0.013	0.049	7.92E-01	0.118	0.036	1.20E-03
rs34000248 (chr18:67464012)	DOK6 (intronic)	FEV ₁	ACT/A	-0.070	0.014	5.75E-07	0.012	0.012	3.18E-01	-0.023	0.009	1.11E-02
rs113473882 (chr19:41124155)	LTBP4 (intronic)	FEV ₁ /FVC	T/C	-0.174	0.033	1.48E-07	-0.138	0.028	1.00E-06	-0.153	0.021	9.95E-13
rs134041 (chr22:28056338)	<i>MIAT</i> (dist=983898), <i>MN1</i> (dist=87927)	FEV ₁	T/C	-0.045	0.008	4.19E-09	-0.018	0.006	4.19E-03	-0.028	0.005	3.03E-09
rs79017381 (chr22:31496200)	SMTN (intronic)	FEV ₁ /FVC	C/T	0.120	0.026	4.41E-06	0.026	0.019	1.71E-01	0.059	0.015	1.36E-04
rs7050036 (chrX:15964845)	AP1S2 (dist=91708), GRPR (dist=176579)	FEV ₁ /FVC	T/A	-0.041	0.007	4.37E-09	-0.013	0.005	1.60E-02	-0.023	0.004	4.14E-08

b) Association results for FEV1, FEV1/FVC and FVC for the 16 sentinel variants

Results for FEV₁, FEV₁/FVC and FVC in stage 1, stage 2 and the meta-analysis of both are presented for the 16 sentinel variants. $P < 5 \times 10^{-8}$ after meta-analysing stage 1 and stage 2 are highlighted in bold. Chr., chromosome; dist., distance.

		Measure	Coded	Stage 1			Stage 2			Stage 1	+ stage	2
rs number (chr: position)	Gene (function)	Measure	Non coded allele	Beta	SE	Р	Beta	SE	Р	Beta	SE	Р
		FEV ₁	G/A	0.042	0.008	1.07E-07	0.021	0.006	1.13E-03	0.029	0.005	4.35E-09
rs6681426 (chr1:150586971)	MCL1(dist=34757), ENSA(dist=7628)	FEV ₁ /FVC	G/A	0.014	0.008	7.47E-02	-0.003	0.006	6.17E-01	0.004	0.005	4.65E-01
``````````````````````````````````````		FVC	G/A	0.036	0.008	4.74E-06	0.024	0.006	1.42E-04	0.029	0.005	5.56E-09
		FEV ₁	A/ATG	-0.007	0.008	3.75E-01	-0.017	0.006	7.49E-03	-0.013	0.005	8.09E-03
rs201204531 (chr1:219963090)	LYPLAL1(dist=576883), RNU5F-1 (dist=83529)	FEV ₁ /FVC	A/ATG	-0.038	0.008	1.98E-06	-0.027	0.006	1.77E-05	-0.031	0.005	2.68E-10
,		FVC	A/ATG	0.011	0.008	1.53E-01	-0.008	0.006	2.18E-01	0.000	0.005	9.31E-01
		FEV ₁	G/A	-0.039	0.009	1.83E-05	-0.036	0.007	3.93E-07	-0.037	0.006	3.21E-11
rs61067109 (chr2:18292452)	<i>KCNS3</i> (dist=178227), <i>NT5C1B-RDH14</i> (dist=443537)	FEV ₁ /FVC	G/A	-0.050	0.009	3.09E-08	-0.042	0.007	6.56E-09	-0.045	0.006	1.40E-15
``````````````````````````````````````		FVC	G/A	-0.009	0.009	3.05E-01	-0.026	0.007	3.07E-04	-0.020	0.006	5.22E-04
		FEV ₁	C/T	0.029	0.008	2.07E-04	0.033	0.006	1.34E-07	0.031	0.005	1.23E-10
rs6441207 (chr3:158282459)	AK097794 (ncRNA_intronic)	FEV ₁ /FVC	C/T	-0.003	0.008	6.56E-01	0.001	0.006	8.13E-01	0.000	0.005	9.25E-01
(********		FVC	C/T	0.036	0.008	4.54E-06	0.036	0.006	5.85E-09	0.036	0.005	1.27E-13
		FEV ₁	G/T	-0.045	0.008	6.22E-08	-0.037	0.006	1.26E-08	-0.040	0.005	5.60E-15
rs6856422 (chr4:106841962)	NPNT (intronic)	FEV ₁ /FVC	G/T	-0.044	0.008	1.30E-07	-0.055	0.006	1.15E-17	-0.051	0.005	1.51E-23
,		FVC	G/T	-0.020	0.008	1.54E-02	-0.019	0.006	3.97E-03	-0.019	0.005	1.70E-04
		FEV ₁	C/T	0.010	0.027	7.07E-01	-0.057	0.019	2.42E-03	-0.035	0.015	2.26E-02
rs148274477 (chr6:142838173)	<i>GPR126</i> (dist=70770),	FEV ₁ /FVC	C/T	-0.161	0.027	2.68E-09	-0.162	0.019	5.63E-18	-0.162	0.015	9.58E-26
(********	LOC153910(dist=9419)	FVC	C/T	0.114	0.027	3.07E-05	0.017	0.019	3.56E-01	0.048	0.015	1.77E-03
		FEV ₁	T/TA	0.023	0.008	2.85E-03	0.015	0.006	1.13E-02	0.018	0.005	1.21E-04
rs34886460 (chr9:119359372)	ASTN2 (intronic)	FEV ₁ /FVC	T/TA	0.041	0.008	6.75E-08	0.025	0.006	4.07E-05	0.031	0.005	4.72E-11
(* * * * * * * * * * * * * * * * * * *		FVC	T/TA	0.000	0.008	9.72E-01	0.007	0.006	2.33E-01	0.004	0.005	3.62E-01
		FEV ₁	C/T	0.029	0.009	6.27E-04	0.029	0.007	1.09E-05	0.029	0.005	2.53E-08
rs2274116 (chr9:139094805)	LHX3 (exonic)	FEV ₁ /FVC	C/T	-0.013	0.009	1.29E-01	0.001	0.007	9.00E-01	-0.004	0.005	4.13E-01
(FVC	C/T	0.041	0.009	2.68E-06	0.038	0.007	4.20E-09	0.039	0.005	5.55E-14
		FEV ₁	A/AT	-0.039	0.008	3.41E-07	-0.034	0.006	5.37E-08	-0.036	0.005	1.07E-13
rs11383346 (chr12:28283187)	PTHLH(dist=158271), CCDC91(dist=126946)	FEV ₁ /FVC	A/AT	-0.002	0.008	8.38E-01	-0.004	0.006	5.31E-01	-0.003	0.005	5.37E-01
(*******		FVC	A/AT	-0.043	0.008	3.83E-08	-0.041	0.006	4.42E-11	-0.042	0.005	9.52E-18
		FEV ₁	G/A	-0.047	0.008	6.68E-09	-0.028	0.006	1.55E-05	-0.035	0.005	2.50E-12
rs10850377 (chr12:115201436)	<i>TBX3</i> (dist=79467), <i>MED13L</i> (dist=1194945)	FEV ₁ /FVC	G/A	-0.020	0.008	1.10E-02	-0.010	0.006	1.38E-01	-0.014	0.005	6.00E-03
		FVC	G/A	-0.033	0.008	3.60E-05	-0.027	0.006	2.67E-05	-0.030	0.005	4.59E-09

rs number (chr:			Coded	Stage 1			Stage 2			Stage 1	+ stage	2
rs number (chr: position)	Gene (function)	Measure	Non coded allele	Beta	SE	Р	Beta	SE	Ρ	Beta	SE	Р
		FEV ₁	G/T	-0.041	0.008	1.39E-07	-0.022	0.006	4.17E-04	-0.030	0.005	1.41E-09
rs7155279 (chr14:92485881)	TRIP11 (intronic)	FEV ₁ /FVC	G/T	-0.023	0.008	3.49E-03	-0.015	0.006	2.10E-02	-0.018	0.005	2.83E-04
``````````````````````````````````````		FVC	G/T	-0.031	0.008	8.62E-05	-0.020	0.006	1.50E-03	-0.024	0.005	8.06E-07
		FEV ₁	C/T	-0.048	0.010	2.72E-06	-0.027	0.008	5.68E-04	-0.035	0.006	2.25E-08
rs117068593 (chr14:93118229)	RIN3 (exonic)	FEV ₁ /FVC	C/T	0.008	0.010	4.23E-01	-0.018	0.008	2.10E-02	-0.008	0.006	1.79E-01
``````````````````````````````````````		FVC	C/T	-0.054	0.010	1.74E-07	-0.022	0.008	5.83E-03	-0.033	0.006	8.10E-08
		FEV ₁	G/A	0.037	0.010	3.38E-04	0.022	0.008	7.31E-03	0.028	0.007	1.45E-05
rs12149828 (chr16:10706328)	<i>EMP2</i> (dist=31789), <i>TEKT5</i> (dist=15033)	FEV ₁ /FVC	G/A	0.049	0.010	3.16E-06	0.035	0.008	3.33E-05	0.040	0.007	7.65E-10
(**********		FVC	G/A	0.005	0.010	6.12E-01	0.007	0.008	3.96E-01	0.006	0.007	3.27E-01
		FEV ₁	T/C	-0.029	0.033	3.78E-01	-0.019	0.028	5.00E-01	-0.023	0.021	2.78E-01
rs113473882 (chr19:41124155)	LTBP4 (intronic)	FEV ₁ /FVC	T/C	-0.174	0.033	1.48E-07	-0.138	0.028	1.00E-06	-0.153	0.021	9.95E-13
,		FVC	T/C	0.073	0.033	2.88E-02	0.046	0.028	1.05E-01	0.057	0.022	8.02E-03
		FEV ₁	T/C	-0.045	0.008	4.19E-09	-0.018	0.006	4.19E-03	-0.028	0.005	3.03E-09
rs134041 (chr22:28056338)	<i>MIAT</i> (dist=983898), <i>MN1</i> (dist=87927)	FEV ₁ /FVC	T/C	-0.025	0.008	1.06E-03	-0.016	0.006	1.11E-02	-0.019	0.005	5.45E-05
(********		FVC	T/C	-0.031	0.008	5.44E-05	-0.017	0.006	7.20E-03	-0.022	0.005	3.69E-06
		FEV ₁	T/A	-0.014	0.007	4.41E-02	-0.003	0.005	6.03E-01	-0.007	0.004	1.06E-01
rs7050036 (chrX:15964845)	AP1S2(dist=91708), GRPR(dist=176579)	FEV ₁ /FVC	T/A	-0.041	0.007	4.37E-09	-0.013	0.005	1.60E-02	-0.023	0.004	4.14E-08
,,		FVC	T/A	0.010	0.007	1.47E-01	0.005	0.005	3.36E-01	0.007	0.004	1.01E-01

Supplementary Table 3 Functional characterization of novel lung function loci

a) eQTL results

Results from eQTL analyses in blood⁶ and lung⁷⁻⁹ (details in **Supplementary Methods**) are presented here for variants in 2Mb regions (sentinel +/- 1Mb) in LD ($r^2 > 0.3$) with the sentinel variant for all the 16 new sentinel variants, which (i) met an FDR threshold of 10%, and (ii) had eQTL $P < 5 \times$ 10^{-5} . For each probeset we ranked variants according to their correlation with the sentinel SNP (as measured by r^2) first and to their eQTL P value second, and we only present results for the top ranked variant for each probeset. Datasets were queried both for *cis* and *trans* eQTLs, but no *trans* results were significant, therefore the table only presents *cis* results. "Effect on expression" indicates whether the "Effect allele" was associated with increased gene expression ("+") or with decreased gene expression ("-"). In the lung eQTL analyses for the autosomal chromosomes results from three different studies were meta-analysed, as indicated by "Dataset" on the table, however for X chromosome analyses were undertaken separately for males and females in each study (details are given in the "Dataset" column). No blood eQTL results were available for the X chromosome. This table presents results for an additional SNP (rs4926386) next to the *ENSA* region. Chr., chromosome.

Sentinel rs number (chr: position)	Proxy rs number (chr: position)	R ² with sentinel	eQTL <i>P</i>	Probeset id	Gene Symbol	Effect allele	Effect on expression	Tissue	Dataset
	rs4926386 (chr1:149886135)	1.00	7.72E-06	100138305_TGI_at	ARNT	С	-	lung	meta-analysed
rs4926386	rs3754047 (chr1:150039207)	0.60	1.51E-13	3890500	MRPS21	А	-	blood	-
(cnr1:149886135)	rs3738320 (chr1:149982626)	0.38	7.57E-06	100309612_TGI_at	SV2A	А	+	lung	meta-analysed
	rs4926399 (chr1:149924299)	0.35	1.82E-05	100123438_TGI_at	MTMR11	С	-	lung	meta-analysed
	rs6681426 (chr1:150586971)	1.00	2.10E-68	100138305_TGI_at	ARNT	А	-	lung	meta-analysed
	rs6681426 (chr1:150586971)	1.00	5.21E-15	100302368_TGI_at	LASS2	А	-	lung	meta-analysed
	rs6681426 (chr1:150586971)	1.00	1.44E-13	100135055_TGI_at	HORMAD1	А	-	lung	meta-analysed
	rs6681426 (chr1:150586971)	1.00	1.66E-12	100151271_TGI_at	LASS2	А	-	lung	meta-analysed
	rs6681426 (chr1:150586971)	1.00	1.85E-12	100309723_TGI_at	ARNT	А	+	lung	meta-analysed
	rs6681426 (chr1:150586971)	1.00	8.55E-11	100308717_TGI_at	GOLPH3L	А	-	lung	meta-analysed
	rs6681426 (chr1:150586971)	1.00	6.09E-10	100144443_TGI_at	cDNA clone that overlaps ADAMTSL4	A	+	lung	meta-analysed
	rs6681426 (chr1:150586971)	1.00	1.09E-07	100306322_TGI_at	ARNT	А	+	lung	meta-analysed
rs6681426	rs6681426 (chr1:150586971)	1.00	8.75E-06	100307575_TGI_at	MRPS21	А	+	lung	meta-analysed
(CHIT. 150580971)	rs6681426 (chr1:150586971)	1.00	1.49E-05	100313509_TGI_at	FAM63A	А	+	lung	meta-analysed
	rs2280078 (chr1:150600181)	0.99	1.08E-120	6330484	CTSS	С	+	blood	-
	rs2280078 (chr1:150600181)	0.99	1.13E-20	1430278	CTSK	С	+	blood	-
	rs2280078 (chr1:150600181)	0.99	1.94E-05	2190040	ARNT	С	-	blood	-
	rs4970969 (chr1:150612361)	0.91	1.58E-05	100310890_TGI_at	ARNT	Т	-	lung	meta-analysed
	rs11204718 (chr1:150709785)	0.84	4.12E-08	100159164_TGI_at	GOLPH3L	С	-	lung	meta-analysed
	rs11204718 (chr1:150709785)	0.84	3.00E-07	100143466_TGI_at	GOLPH3L	С	-	lung	meta-analysed
	rs7521898 (chr1:150707596)	0.80	2.22E-19	3370121	LASS2	А	+	blood	-
	rs7521898 (chr1:150707596)	0.80	2.52E-17	6380154	LASS2	А	+	blood	-

Sentinel rs number (chr: position)	Proxy rs number (chr: position)	R ² with sentinel	eQTL <i>P</i>	Probeset id	Gene Symbol	Effect allele	Effect on expression	Tissue	Dataset
• • •	rs11204691 (chr1:150644339)	0.76	4.27E-05	2640338	ARNT	Т	+	blood	-
	rs878471 (chr1:150547747)	0.73	4.55E-08	100144721_TGI_at	RPRD2	G	-	lung	meta-analysed
	rs878471 (chr1:150547747)	0.73	6.87E-06	100161206_TGI_at	ENSA	G	-	lung	meta-analysed
	rs897815 (chr1:150879101)	0.64	4.89E-11	4480192	CDC42SE1, C1orf56	G	-	blood	-
	rs897815 (chr1:150879101)	0.64	3.99E-07	4390372	BNIPL	G	-	blood	-
	rs897815 (chr1:150879101)	0.64	5.60E-07	4540288	CDC42SE1, C1orf56	G	-	blood	-
	rs3768013 (chr1:150815411)	0.63	3.14E-05	6450722	SETDB1	А	-	blood	-
	rs11204693 (chr1:150647987)	0.55	1.20E-05	7380100	MCL1	Т	-	blood	-
	rs11204664 (chr1:150531380)	0.48	2.47E-08	100156777_TGI_at	MRPS21	С	+	lung	meta-analysed
	rs11204664 (chr1:150531380)	0.48	1.50E-06	100130530_TGI_at	ECM1	С	+	lung	meta-analysed
	rs2305814 (chr1:150958977)	0.45	2.99E-05	4280162	SCNM1	С	-	blood	-
	rs486836 (chr1:150875967)	0.40	1.78E-05	100125530_TGI_at	CTSS	А	+	lung	meta-analysed
rs201204531 (chr1:219963090)	rs1338230 (chr1:219963090)	0.31	1.27E-06	100159938_TGI_at	SLC30A10	G	+	lung	meta-analysed
	rs6441207 (chr3:158282459)	1.00	2.30E-08	100301782_TGI_at	MLF1	С	-	lung	meta-analysed
	rs6441207 (chr3:158282459)	1.00	3.21E-06	100304858_TGI_at	MLF1	С	-	lung	meta-analysed
	rs112353818 (chr3:158239479)	0.82	2.63E-09	100150755_TGI_at	GFM1	G	+	lung	meta-analysed
	rs112353818 (chr3:158239479)	0.82	2.90E-06	100123606_TGI_at	MLF1	G	-	lung	meta-analysed
rs6441207 (chr3:158282459)	rs7620927 (chr3:158245883)	0.81	1.22E-05	3440195	GFM1	G	+	blood	-
	rs12492884 (chr3:158252566)	0.53	2.62E-06	60670	GFM1,LXN	G	-	blood	-
	rs1656377 (chr3:158285280)	0.50	2.32E-06	100312821_TGI_at	MLF1	С	+	lung	meta-analysed
	rs2731122 (chr3:158295280)	0.37	2.62E-06	100141104_TGI_at	GFM1	G	-	lung	meta-analysed
	rs63740961 (chr3:158295171)	0.31	8.55E-06	100301664_TGI_at	GFM1	С	-	lung	meta-analysed

Sentinel rs number (chr: position)	Proxy rs number (chr: position)	R ² with sentinel	eQTL <i>P</i>	Probeset id	Gene Symbol	Effect allele	Effect on expression	Tissue	Dataset
rs6856422	rs4600917 (chr4:106829645)	0.77	2.89E-06	4860553	INTS12	С	-	blood	-
(chr4:106841962)	rs2553449 (chr4:106698892)	0.42	3.17E-05	5360487	GSTCD	Т	+	blood	-
rs34886460 (chr9:119359372)	rs1372335 (chr9:119403743)	0.62	1.10E-06	5670088	TRIM32	Т	+	blood	-
	rs2274116 (chr9:139094805)	1.00	1.81E-06	100124395_TGI_at	QSOX2	С	+	lung	meta-analysed
rs2274116 (chr9:139094805)	rs10858246 (chr9:139102831)	0.88	4.78E-08	3830605	C9orf151	С	+	blood	-
	rs3780190 (chr9:139099073)	0.43	1.32E-05	3060494	CARD9	G	-	blood	-
	rs11383346 (chr12:28283187)	1.00	1.73E-13	100137632_TGI_at	cDNA clone that maps to CCDC91	NA	-	lung	meta-analysed
(chr12:28283187)	rs11049391 (chr12:28322273)	0.90	1.55E-05	100124630_TGI_at	CCDC91	G	+	lung	meta-analysed
	rs10843114 (chr12:28303296)	0.35	6.29E-06	100133373_TGI_at	CCDC91	С	-	lung	meta-analysed
	rs7155279 (chr14:92485881)	1.00	1.01E-10	100132576_TGI_at	ATXN3	G	-	lung	meta-analysed
	rs7155279 (chr14:92485881)	1.00	3.38E-06	100160661_TGI_at	TRIP11	G	-	lung	meta-analysed
	rs76497846 (chr14:92531888)	0.99	1.80E-05	100127017_TGI_at	CPSF2	G	-	lung	meta-analysed
	rs7157056 (chr14:92498033)	0.94	3.00E-34	4670441	FBLN5	G	+	blood	-
	rs7157056 (chr14:92498033)	0.94	1.23E-29	5690639	FBLN5	G	+	blood	-
rs7155279	rs7157056 (chr14:92498033)	0.94	1.12E-19	2760136	TRIP11	G	+	blood	-
(CNr14:92485881)	rs10151561 (chr14:92430184)	0.75	2.05E-06	100300526_TGI_at	ATXN3	А	-	lung	meta-analysed
	rs10129429 (chr14:92430831)	0.75	1.72E-05	100143127_TGI_at	NDUFB1	G	+	lung	meta-analysed
	rs7145052 (chr14:92461192)	0.69	3.35E-05	5090037	NDUFB1	Т	-	blood	-
	rs111269625 (chr14:92514370)	0.56	2.50E-13	100148617_TGI_at	ATXN3	G	+	lung	meta-analysed
	rs111269625 (chr14:92514370)	0.56	1.13E-08	100306579_TGI_at	ATXN3	G	+	lung	meta-analysed
	rs111269625 (chr14:92514370)	0.56	1.60E-05	100137755_TGI_at	ATXN3	G	-	lung	meta-analysed

Sentinel rs number (chr: position)	Proxy rs number (chr: position)	R ² with sentinel	eQTL <i>P</i>	Probeset id	Gene Symbol	Effect allele	Effect on expression	Tissue	Dataset
rs12149828 (chr16:10706328)	rs8053116 (chr16:10719601)	0.55	5.51E-06	100123005_TGI_at	EMP2	А	+	lung	meta-analysed
rs113473882 (chr19:41124155)	rs113434779 (chr19:41236459)	0.49	1.15E-05	100302701_TGI_at	NUMBL	С	-	lung	meta-analysed
	rs798183 (chrX:15840426)	0.62	1.96E-08	100302651_TGI_at	ZRSR2	Т	+	lung	UBC_male
	rs798183 (chrX:15840426)	0.62	3.18E-07	100302651_TGI_at	ZRSR2	Т	+	lung	Laval_male
rs7050036	rs798183 (chrX:15840426)	0.62	1.34E-06	100302651_TGI_at	ZRSR2	Т	+	lung	Groningen_ma le
(chrX:15964845)	rs798183 (chrX:15840426)	0.62	7.79E-06	100302651_TGI_at	ZRSR2	Т	+	lung	UBC_female
	rs798157 (chrX:15810140)	0.53	2.99E-09	100311509_TGI_at	AP1S2	Т	+	lung	Laval_male
	rs798157 (chrX:15810140)	0.53	5.49E-08	100122762_TGI_at	AP1S2	Т	+	lung	Laval_male

b) DNase hypersensitivity sites

DNase hypersensitivity sites as defined by ENCODE¹⁰, which contain at least one variant strongly correlated (r² > 0.8) with any of the new 16 sentinel variants are presented here. The "Signal Value" is a measurement of average enrichment in the region and the "Score value" is a normalized value that ranges from 0 to 1,000 based on the signal value^{10,11}. Only DNase hypersensitivity sites with "Score value" > 100 are presented. Cell type abbreviations are: A549, epithelial cell line derived from a lung carcinoma tissue; AG04450, fetal lung fibroblast; HMVEC-LBI, blood microvascular endothelial cells, lung-derived; HMVEC-LLy, lymphatic microvascular endothelial cells, lung-derived; HPF, pulmonary fibroblasts isolated from lung tissue; HPAEC, pulmonary artery endothelial cells; HPAF, pulmonary artery fibroblasts; NHLF, lung fibroblasts; WI-38, embryonic lung fibroblast cells, hTERT immortalized, includes Raf1 construct. Chr., chromosome.

Sentinel rs number (chr: position)	Proxy rs number (chr: position)	R ² with sentinel	Cell type	Chr.	Start	End	Score value	Signal value
rs6681426 (chr1:150586971)	rs6655975 (chr1:150542128)	0.97	HPAEC	1	150542080	150542230	102	15
rs6681426 (chr1:150586971)	rs6655975 (chr1:150542128)	0.97	HPAEC	1	150542080	150542230	102	15
rs201204531 (chr1:219963090)	rs10863500 (chr1:219961690)	1.00	AG04450	1	219961560	219961710	101	24
rs2274116 (chr9:119359372)	rs4836757 (chr9:119326613)	0.98	A549	9	119326565	119326715	100	21
rs11383346 (chr12:28283187)	rs2881788 (chr12:28272574)	0.82	WI-38	12	28272520	28272670	100	10
rs11383346 (chr12:28283187)	rs7310453 (chr12:28283780)	0.99	AG04450	12	28283700	28283850	106	173
rs11383346 (chr12:28283187)	rs7310453 (chr12:28283780)	0.99	HPAF	12	28283700	28283850	129	1340
rs11383346 (chr12:28283187)	rs7310453 (chr12:28283780)	0.99	HPAEC	12	28283720	28283870	137	272
rs11383346 (chr12:28283187)	rs7310453 (chr12:28283780)	0.99	HMVEC-LLy	12	28283740	28283890	110	739
rs11383346 (chr12:28283187)	rs7310453 (chr12:28283780)	0.99	HPF	12	28283740	28283890	115	253
rs11383346 (chr12:28283187)	rs7310453 (chr12:28283780)	0.99	WI-38	12	28283740	28283890	116	297
rs11383346 (chr12:28283187)	rs7310453 (chr12:28283780)	0.99	NHLF	12	28283740	28283890	121	948
rs11383346 (chr12:28283187)	rs7310453 (chr12:28283780)	0.99	A549	12	28283760	28283910	100	21
rs11383346 (chr12:28283187)	rs7310453 (chr12:28283780)	0.99	HMVEC-LBI	12	28283760	28283910	182	1082
rs11383346 (chr12:28283187)	rs7310453 (chr12:28285003)	0.99	HMVEC-LBI	12	28285000	28285150	101	16
rs11383346 (chr12:28283187)	rs7310453 (chr12:28406157)	0.82	HPAF	12	28406080	28406230	100	19
rs11383346 (chr12:28283187)	rs7310453 (chr12:28406157)	0.82	NHLF	12	28406080	28406230	102	84
rs10850377 (chr12:115201436)	rs78649052 (chr12:115200597)	0.97	HPAF	12	115200500	115200650	102	77
rs10850377 (chr12:115201436)	rs78649052 (chr12:115200597)	0.97	AG04450	12	115200500	115200650	103	81
rs10850377 (chr12:115201436)	rs78649052 (chr12:115200597)	0.97	HPF	12	115200500	115200650	109	148
rs10850377 (chr12:115201436)	rs78649052 (chr12:115200597)	0.97	WI-38	12	115200500	115200650	109	172
rs117068593 (chr14:93118229)	rs78440108 (chr14:93070286)	0.90	AG04450	14	93070260	93070410	103	74
rs117068593 (chr14:93118229)	rs78440108 (chr14:93070286)	0.90	AG04450	14	93070260	93070410	103	74
rs117068593 (chr14:93118229)	rs78440108 (chr14:93070286)	0.90	WI-38	14	93070260	93070410	103	57
rs117068593 (chr14:93118229)	rs78440108 (chr14:93070286)	0.90	WI-38	14	93070260	93070410	103	57
rs117068593 (chr14:93118229)	rs78440108 (chr14:93070286)	0.90	AG04450	14	93070260	93070410	103	74
rs117068593 (chr14:93118229)	rs78440108 (chr14:93070286)	0.90	AG04450	14	93070260	93070410	103	74
rs117068593 (chr14:93118229)	rs78440108 (chr14:93070286)	0.90	WI-38	14	93070260	93070410	103	57
rs117068593 (chr14:93118229)	rs78440108 (chr14:93070286)	0.90	WI-38	14	93070260	93070410	103	57
rs117068593 (chr14:93118229)	rs78440108 (chr14:93070286)	0.90	HPAF	14	93070260	93070410	104	164
rs117068593 (chr14:93118229)	rs78440108 (chr14:93070286)	0.90	HPAF	14	93070260	93070410	104	164
rs117068593 (chr14:93118229)	rs78440108 (chr14:93070286)	0.90	HPF	14	93070260	93070410	104	66
rs117068593 (chr14:93118229)	rs78440108 (chr14:93070286)	0.90	HPF	14	93070260	93070410	104	66
rs117068593 (chr14:93118229)	rs78440108 (chr14:93070286)	0.90	NHLF	14	93070260	93070410	104	191

Sentinel rs number (chr: position)	Proxy rs number (chr: position)	R ² with sentinel	Cell type	Chr.	Start	End	Score value	Signal value
rs117068593 (chr14:93118229)	rs78440108 (chr14:93070286)	0.90	NHLF	14	93070260	93070410	104	191
rs117068593 (chr14:93118229)	rs78440108 (chr14:93070286)	0.90	HPAF	14	93070260	93070410	104	164
rs117068593 (chr14:93118229)	rs78440108 (chr14:93070286)	0.90	HPAF	14	93070260	93070410	104	164
rs117068593 (chr14:93118229)	rs78440108 (chr14:93070286)	0.90	HPF	14	93070260	93070410	104	66
rs117068593 (chr14:93118229)	rs78440108 (chr14:93070286)	0.90	HPF	14	93070260	93070410	104	66
rs117068593 (chr14:93118229)	rs78440108 (chr14:93070286)	0.90	NHLF	14	93070260	93070410	104	191
rs117068593 (chr14:93118229)	rs78440108 (chr14:93070286)	0.90	NHLF	14	93070260	93070410	104	191
rs117068593 (chr14:93118229)	rs72697299 (chr14:93070367)	0.90	AG04450	14	93070260	93070410	103	74
rs117068593 (chr14:93118229)	rs72697299 (chr14:93070367)	0.90	AG04450	14	93070260	93070410	103	74
rs117068593 (chr14:93118229)	rs72697299 (chr14:93070367)	0.90	WI-38	14	93070260	93070410	103	57
rs117068593 (chr14:93118229)	rs72697299 (chr14:93070367)	0.90	WI-38	14	93070260	93070410	103	57
rs117068593 (chr14:93118229)	rs72697299 (chr14:93070367)	0.90	AG04450	14	93070260	93070410	103	74
rs117068593 (chr14:93118229)	rs72697299 (chr14:93070367)	0.90	AG04450	14	93070260	93070410	103	74
rs117068593 (chr14:93118229)	rs72697299 (chr14:93070367)	0.90	WI-38	14	93070260	93070410	103	57
rs117068593 (chr14:93118229)	rs72697299 (chr14:93070367)	0.90	WI-38	14	93070260	93070410	103	57
rs117068593 (chr14:93118229)	rs72697299 (chr14:93070367)	0.90	HPAF	14	93070260	93070410	104	164
rs117068593 (chr14:93118229)	rs72697299 (chr14:93070367)	0.90	HPAF	14	93070260	93070410	104	164
rs117068593 (chr14:93118229)	rs72697299 (chr14:93070367)	0.90	HPF	14	93070260	93070410	104	66
rs117068593 (chr14:93118229)	rs72697299 (chr14:93070367)	0.90	HPF	14	93070260	93070410	104	66
rs117068593 (chr14:93118229)	rs72697299 (chr14:93070367)	0.90	NHLF	14	93070260	93070410	104	191
rs117068593 (chr14:93118229)	rs72697299 (chr14:93070367)	0.90	NHLF	14	93070260	93070410	104	191
rs117068593 (chr14:93118229)	rs72697299 (chr14:93070367)	0.90	HPAF	14	93070260	93070410	104	164
rs117068593 (chr14:93118229)	rs72697299 (chr14:93070367)	0.90	HPAF	14	93070260	93070410	104	164
rs117068593 (chr14:93118229)	rs72697299 (chr14:93070367)	0.90	HPF	14	93070260	93070410	104	66
rs117068593 (chr14:93118229)	rs72697299 (chr14:93070367)	0.90	HPF	14	93070260	93070410	104	66
rs117068593 (chr14:93118229)	rs72697299 (chr14:93070367)	0.90	NHLF	14	93070260	93070410	104	191
rs117068593 (chr14:93118229)	rs72697299 (chr14:93070367)	0.90	NHLF	14	93070260	93070410	104	191
rs117068593 (chr14:93118229)	rs78440108 (chr14:93070286)	0.90	HPAEC	14	93070280	93070430	103	21
rs117068593 (chr14:93118229)	rs78440108 (chr14:93070286)	0.90	HPAEC	14	93070280	93070430	103	21
rs117068593 (chr14:93118229)	rs78440108 (chr14:93070286)	0.90	HPAEC	14	93070280	93070430	103	21
rs117068593 (chr14:93118229)	rs78440108 (chr14:93070286)	0.90	HPAEC	14	93070280	93070430	103	21
rs117068593 (chr14:93118229)	rs72697299 (chr14:93070367)	0.90	HPAEC	14	93070280	93070430	103	21
rs117068593 (chr14:93118229)	rs72697299 (chr14:93070367)	0.90	HPAEC	14	93070280	93070430	103	21
rs117068593 (chr14:93118229)	rs72697299 (chr14:93070367)	0.90	HPAEC	14	93070280	93070430	103	21

Sentinel rs number (chr: position)	Proxy rs number (chr: position)	R ² with sentinel	Cell type	Chr.	Start	End	Score value	Signal value
rs117068593 (chr14:93118229)	rs72697299 (chr14:93070367)	0.90	HPAEC	14	93070280	93070430	103	21
rs117068593 (chr14:93118229)	rs72697299 (chr14:93070367)	0.90	HMVEC-LLy	14	93070300	93070450	100	19
rs117068593 (chr14:93118229)	rs72697299 (chr14:93070367)	0.90	HMVEC-LLy	14	93070300	93070450	100	19
rs117068593 (chr14:93118229)	rs72697299 (chr14:93070367)	0.90	HMVEC-LLy	14	93070300	93070450	100	19
rs117068593 (chr14:93118229)	rs72697299 (chr14:93070367)	0.90	HMVEC-LLy	14	93070300	93070450	100	19
rs117068593 (chr14:93118229)	rs7143806 (chr14:93103729)	0.94	WI-38	14	93103660	93103810	102	44
rs117068593 (chr14:93118229)	rs7143806 (chr14:93103729)	0.94	WI-38	14	93103660	93103810	102	44
rs117068593 (chr14:93118229)	rs7143806 (chr14:93103729)	0.94	WI-38	14	93103660	93103810	102	44
rs117068593 (chr14:93118229)	rs7143806 (chr14:93103729)	0.94	WI-38	14	93103660	93103810	102	44
rs117068593 (chr14:93118229)	rs7143806 (chr14:93103729)	0.94	HPF	14	93103660	93103810	103	48
rs117068593 (chr14:93118229)	rs7143806 (chr14:93103729)	0.94	HPF	14	93103660	93103810	103	48
rs117068593 (chr14:93118229)	rs7143806 (chr14:93103729)	0.94	HPF	14	93103660	93103810	103	48
rs117068593 (chr14:93118229)	rs7143806 (chr14:93103729)	0.94	HPF	14	93103660	93103810	103	48
rs117068593 (chr14:93118229)	rs7143806 (chr14:93103729)	0.94	AG04450	14	93103680	93103830	101	29
rs117068593 (chr14:93118229)	rs7143806 (chr14:93103729)	0.94	AG04450	14	93103680	93103830	101	29
rs117068593 (chr14:93118229)	rs7143806 (chr14:93103729)	0.94	AG04450	14	93103680	93103830	101	29
rs117068593 (chr14:93118229)	rs7143806 (chr14:93103729)	0.94	AG04450	14	93103680	93103830	101	29
rs117068593 (chr14:93118229)	rs7143806 (chr14:93103729)	0.94	HPAF	14	93103720	93103870	100	23
rs117068593 (chr14:93118229)	rs7143806 (chr14:93103729)	0.94	HPAF	14	93103720	93103870	100	23
rs117068593 (chr14:93118229)	rs7143806 (chr14:93103729)	0.94	HPAF	14	93103720	93103870	100	23
rs117068593 (chr14:93118229)	rs7143806 (chr14:93103729)	0.94	HPAF	14	93103720	93103870	100	23
rs117068593 (chr14:93118229)	rs201493181 (chr14:93112309)	0.99	HPAF	14	93112300	93112450	102	85
rs117068593 (chr14:93118229)	rs201493181 (chr14:93112309)	0.99	HPAF	14	93112300	93112450	102	85
rs117068593 (chr14:93118229)	rs201493181 (chr14:93112309)	0.99	HPAF	14	93112300	93112450	102	85
rs117068593 (chr14:93118229)	rs201493181 (chr14:93112309)	0.99	HPAF	14	93112300	93112450	102	85
rs12149828 (chr16:10706328)	rs12596425 (chr16:10713425)	0.95	NHLF	16	10713420	10713570	100	12
rs134041 (chr22:28056338)	rs134077 (chr22:28063227)	0.97	HPAF	22	28063120	28063270	101	64
rs134041 (chr22:28056338)	rs134117 (chr22:28082715)	0.86	HPAEC	22	28082620	28082770	106	47
rs134041 (chr22:28056338)	rs134117 (chr22:28082715)	0.86	HMVEC-LLy	22	28082640	28082790	100	21
rs134041 (chr22:28056338)	rs134117 (chr22:28082715)	0.86	AG04450	22	28082640	28082790	101	28
rs134041 (chr22:28056338)	rs134117 (chr22:28082715)	0.86	HPF	22	28082640	28082790	103	59
rs134041 (chr22:28056338)	rs134117 (chr22:28082715)	0.86	WI-38	22	28082640	28082790	103	57
rs134041 (chr22:28056338)	rs134117 (chr22:28082715)	0.86	NHLF	22	28082640	28082790	104	190
rs134041 (chr22:28056338)	rs134117 (chr22:28082715)	0.86	HPAF	22	28082640	28082790	109	411

Sentinel rs number (chr: position)	Proxy rs number (chr: position)	R ² with sentinel	Cell type	Chr.	Start	End	Score value	Signal value
rs134041 (chr22:28056338)	rs134117 (chr22:28082715)	0.86	HMVEC-LBI	22	28082640	28082790	117	228
rs134041 (chr22:28056338)	rs134117 (chr22:28082715)	0.86	A549	22	28082680	28082830	100	45

c) Human Protein Atlas results

Protein expression results in the respiratory system for epithelial cells, pneumocytes and macrophages for a subset of genes for each of the regions that contain the 16 sentinel variants (see gene selection criteria in **Supplementary Methods**) were assessed in the Human Protein Atlas¹² and are presented here. If no results were available in the Human Protein Atlas a hyphen ("-") is presented on the table. Chr., chromosome; H, high; M, medium; L, low; ND not detected.

rs number (chr: position)	Measure	Gene Symbol	Respiratory epithelial cells, Nasopharynx (antibody1 staining, antibody2 staining, annotated protein expression)	Respiratory epithelial cells, Bronchus(antibody1 staining, antibody2 staining, annotated protein expression)	Pneumocytes, lung (antibody1 staining, antibody2 staining, annotated protein expression)	Macrophages, lung(antibody1 staining, antibody2 staining, annotated protein expression)
rs6681426 (chr1:150586971)	FEV ₁	ARNT	M,M,M	M,L,L	M,M,M	M,M,M
rs6681426 (chr1:150586971)	FEV ₁	CTSS	M,L,M	M,M,M	ND,ND,ND	Н,Н,Н
rs6681426 (chr1:150586971)	FEV ₁	LASS2	L,-,-	M,-,-	Н,-,-	ND,-,-
rs201204531 (chr1:219963090)	FF	SLC30A10	ND,-,-	ND,-,-	L,-,-	M,-,-
rs61067109 (chr2:18292452)	FF	KCNS3	M,-,-	-,-,-	L,-,-	M,-,-
rs6441207 (chr3:158282459)	FVC	GFM1	M,-,-	Н,-,-	ND,-,-	Н,-,-
rs6441207 (chr3:158282459)	FVC	MLF1	Н,-,-	M,-,-	ND,-,-	Н,-,-
rs6856422 (chr4:106841962)	FF	NPNT	ND,-,-	ND,-,-	Н,-,-	ND,-,-
rs6856422 (chr4:106841962)	FF	INTS12	Н,-,-	M,-,-	M,-,-	M,-,-
rs6856422 (chr4:106841962)	FF	GSTCD	M,-,-	M,-,-	ND,-,-	M,-,-
rs148274477 (chr6:142838173)	FF	GPR126	ND,-,-	ND,-,-	ND,-,-	ND,-,-
rs34886460 (chr9:119359372)	FF	TRIM32	M,-,-	M,-,-	ND,-,-	ND,-,-
rs2274116 (chr9:139094805)	FVC	QSOX2	M,-,-	Н,-,-	ND,-,-	L,-,-
rs11383346 (chr12:28283187)	FVC	CCDC91	Н,-,-	Н,-,-	M,-,-	M,-,-
rs10850377 (chr12:115201436)	FEV ₁	ТВХ3	L,-,-	L,-,-	ND,-,-	Н,-,-
rs7155279 (chr14:92485881)	FEV ₁	TRIP11	Н,-,-	Н,-,-	Н,-,-	Н,-,-
rs7155279 (chr14:92485881)	FEV ₁	ATXN3	M,H,H	ND,H,H	ND,H,H	ND,H,H
rs117068593 (chr14:93118229)	FEV ₁	RIN3	-,-,-	ND,-,-	ND,-,-	M,-,-
rs12149828 (chr16:10706328)	FF	EMP2	ND,-,-	M,-,-	M,-,-	M,-,-
rs113473882 (chr19:41124155)	FF	LTBP4	M,-,-	-,-,-	M,-,-	M,-,-
rs134041 (chr22:28056338)	FEV ₁	MN1	Н,-,-	Н,-,-	Н,-,-	ND,-,-
rs7050036 (chrX:15964845)	FF	ZRSR2	L,-,-	M,-,-	M,-,-	M,-,-
rs7050036 (chrX:15964845)	FF	AP1S2	ND,-,-	L,-,-	ND,-,-	ND,-,-

d) Differential gene expression in fetal lung

Publicly available gene expression data from fetal lung samples spanning pseudoglandular and canalicular stages of lung development¹³ were queried for a subset of genes for each of the regions that contain the 16 sentinel variants (see criteria in the **Supplementary Methods**). Adj. P < 0.05 are highlighted in bold, and genes with all the probes showing an effect on the same direction and with at least 33% of the probes with adjusted P < 0.05 are also highlighted in bold. Chr., chromosome, Probe id, Affymetrix probe id; Average expression, average expression for probe during the entire time period (genes with Average expression values > 8 can be considered highly expressed); Adj. P, adjusted P (Benjamini & Hochberg method¹⁴) for differential expression over time; Beta, mean change in gene expression per day during the studied period (7-22 weeks of gestational age); chr., chromosome.

rs number (chr: position)	Probe id	Gene Symbol	Average expression	Adj. P	Beta
rs6681426 (chr1:150586971)	233724_at	ARNT	4.55	4.58E-03	0.009
rs6681426 (chr1:150586971)	230619_at	ARNT	7.55	6.32E-03	0.012
rs6681426 (chr1:150586971)	210828_s_at	ARNT	5.03	3.13E-01	0.003
rs6681426 (chr1:150586971)	218222_x_at	ARNT	6.82	3.52E-01	0.002
rs6681426 (chr1:150586971)	218221_at	ARNT	8.11	5.17E-01	0.002
rs6681426 (chr1:150586971)	231016_s_at	ARNT	7.36	9.51E-01	0.000
rs6681426 (chr1:150586971)	232617_at	CTSS	6.17	3.02E-04	0.015
rs6681426 (chr1:150586971)	202901_x_at	CTSS	3.59	5.90E-03	0.010
rs6681426 (chr1:150586971)	202902_s_at	CTSS	6.06	1.20E-01	0.007
rs6681426 (chr1:150586971)	222212_s_at	LASS2	9.61	1.97E-01	0.003
rs201204531 (chr1:219963090)	220435_at	SLC30A10	3.42	2.56E-01	0.002
rs61067109 (chr2:18292452)	205968_at	KCNS3	7.98	9.79E-01	0.000
rs6441207 (chr3:158282459)	232296_s_at	GFM1	7.02	1.10E-02	-0.008
rs6441207 (chr3:158282459)	225158_at	GFM1	7.73	2.39E-02	-0.008
rs6441207 (chr3:158282459)	225161_at	GFM1	7.76	4.34E-02	-0.006
rs6441207 (chr3:158282459)	232295_at	GFM1	3.57	1.94E-01	-0.002
rs6441207 (chr3:158282459)	225153_at	GFM1	9.50	6.64E-01	0.001
rs6441207 (chr3:158282459)	220903_at	GFM1	3.81	8.44E-01	0.000
rs6441207 (chr3:158282459)	204783_at	MLF1	7.23	1.42E-02	0.010
rs6441207 (chr3:158282459)	204784_s_at	MLF1	8.41	9.07E-01	0.001
rs6856422 (chr4:106841962)	235387_at	GSTCD	6.32	8.55E-04	-0.012
rs6856422 (chr4:106841962)	1554518_at	GSTCD	5.80	1.50E-02	-0.005
rs6856422 (chr4:106841962)	220063_at	GSTCD	5.61	8.62E-02	-0.004
rs6856422 (chr4:106841962)	241126_at	GSTCD	3.46	4.88E-01	0.001
rs6856422 (chr4:106841962)	218616_at	INTS12	8.22	2.10E-01	-0.004
rs6856422 (chr4:106841962)	225911_at	NPNT	9.01	6.65E-06	0.011
rs6856422 (chr4:106841962)	244747_at	NPNT	4.36	9.75E-01	0.000
rs148274477 (chr6:142838173)	1553025_at	GPR126	3.77	1.69E-02	0.004
rs148274477 (chr6:142838173)	213094_at	GPR126	9.73	1.98E-01	-0.003
rs148274477 (chr6:142838173)	233887_at	GPR126	7.66	9.87E-01	0.000
rs34886460 (chr9:119359372)	236233_at	TRIM32	4.75	4.48E-01	0.002
rs34886460 (chr9:119359372)	203846_at	TRIM32	8.21	5.55E-01	0.001
rs2274116 (chr9:139094805)	235239_at	QSOX2	5.35	6.01E-02	-0.003
rs2274116 (chr9:139094805)	227146_at	QSOX2	7.05	1.34E-01	-0.003
rs11383346 (chr12:28283187)	218545_at	CCDC91	7.08	6.76E-01	0.002
rs11383346 (chr12:28283187)	1570571_at	CCDC91	4.55	8.94E-01	0.001
rs10850377 (chr12:115201436)	219682_s_at	TBX3	8.67	9.91E-03	0.006
rs10850377 (chr12:115201436)	225544_at	TBX3	9.56	2.70E-01	0.002
rs10850377 (chr12:115201436)	222917_s_at	TBX3	5.83	4.56E-01	0.002
rs10850377 (chr12:115201436)	229576_s_at	TBX3	7.36	5.21E-01	0.002
rs10850377 (chr12:115201436)	229565_x_at	TBX3	4.41	6.14E-01	0.001
rs10850377 (chr12:115201436)	228344_s_at	TBX3	4.41	7.96E-01	-0.001
rs10850377 (chr12:115201436)	243234_at	TBX3	3.63	9.83E-01	0.000
rs7155279 (chr14:92485881)	216657_at	ATXN3	3.89	9.9/E-04	0.006
rs7155279 (chr14:92485881)	21/321_x_at	ATXN3	3.38	1.00E-01	0.002
rs7155279 (chr14:92485881)	238723_at	ATXN3	6.46	1.89E-01	0.007

rs number (chr: position)	Probe id	Gene Symbol	Average expression	Adj. P	Beta
rs7155279 (chr14:92485881)	205416_s_at	ATXN3	5.48	4.77E-01	-0.002
rs7155279 (chr14:92485881)	235240_at	ATXN3	6.87	6.96E-01	-0.001
rs7155279 (chr14:92485881)	205415_s_at	ATXN3	6.50	9.51E-01	0.000
rs7155279 (chr14:92485881)	233182_x_at	ATXN3	5.27	9.59E-01	0.000
rs7155279 (chr14:92485881)	230211_at	TRIP11	4.74	3.53E-01	0.003
rs7155279 (chr14:92485881)	210760_x_at	TRIP11	4.62	6.02E-01	0.001
rs7155279 (chr14:92485881)	236160_at	TRIP11	6.77	6.46E-01	-0.001
rs7155279 (chr14:92485881)	209778_at	TRIP11	5.61	6.75E-01	0.002
rs117068593 (chr14:93118229)	1562005_at	RIN3	4.13	5.05E-02	0.004
rs117068593 (chr14:93118229)	60471_at	RIN3	7.46	2.57E-01	0.002
rs117068593 (chr14:93118229)	220439_at	RIN3	4.30	4.91E-01	-0.001
rs117068593 (chr14:93118229)	219457_s_at	RIN3	5.94	7.43E-01	-0.001
rs117068593 (chr14:93118229)	219456_s_at	RIN3	5.00	7.92E-01	0.001
rs12149828 (chr16:10706328)	204975_at	EMP2	8.29	3.68E-06	0.014
rs12149828 (chr16:10706328)	225078_at	EMP2	9.08	1.42E-05	0.011
rs12149828 (chr16:10706328)	225079_at	EMP2	8.71	8.62E-05	0.010
rs12149828 (chr16:10706328)	238500_at	EMP2	4.73	9.78E-02	0.004
rs113473882 (chr19:41124155)	227989_at	LTBP4	4.68	1.30E-01	-0.003
rs113473882 (chr19:41124155)	204442_x_at	LTBP4	8.44	3.89E-01	0.004
rs113473882 (chr19:41124155)	213176_s_at	LTBP4	8.03	9.31E-01	0.001
rs113473882 (chr19:41124155)	210628_x_at	LTBP4	6.83	9.96E-01	0.000
rs134041 (chr22:28056338)	205330_at	MN1	5.44	1.60E-01	-0.005
rs7050036 (chrX:15964845)	228415_at	AP1S2	7.82	9.12E-05	-0.009
rs7050036 (chrX:15964845)	230264_s_at	AP1S2	9.67	3.97E-02	-0.004
rs7050036 (chrX:15964845)	203300_x_at	AP1S2	9.07	1.52E-01	-0.004
rs7050036 (chrX:15964845)	203299_s_at	AP1S2	7.55	6.26E-01	-0.001
rs7050036 (chrX:15964845)	208174_x_at	ZRSR2	6.99	4.29E-01	0.002
rs7050036 (chrX:15964845)	213876_x_at	ZRSR2	7.18	5.82E-01	0.001

e) Differential gene expression in COPD vs. non COPD ever smoker samples

Publicly available gene expression data from bronchial brushings of individuals with COPD and of ever smoking individuals without $COPD^{15}$ were queried for a subset of genes for each of the regions that contain the 16 sentinel variants (see criteria in **Supplementary Methods**) are presented. There was no probe available for *ZRSR2* in this analysis. Adj. P < 0.05 are highlighted in bold. "Probe id", affymetrix probe id; Adj.P, *P* value after adjustment for multiple testing using the Benjamini & Hochberg method¹⁴; FC, fold change between cases and controls (>1 up regulated in cases); chr., chromosome.

rs number (chr: position)	Probe id	Gene Symbol	Adj. P	FC
rs6681426 (chr1:150586971)	405_at	ARNT	8.53E-02	1.04
rs6681426 (chr1:150586971)	1520_at	CTSS	1.04E-01	1.06
rs6681426 (chr1:150586971)	29956_at	LASS2	5.78E-01	1.02
rs201204531 (chr1:219963090)	55532_at	SLC30A10	9.35E-01	1.00
rs61067109 (chr2:18292452)	3790_at	KCNS3	9.23E-01	0.99
rs6441207 (chr3:158282459)	85476_at	GFM1	7.28E-01	0.99
rs6441207 (chr3:158282459)	4291_at	MLF1	1.74E-03	1.12
rs6856422 (chr4:106841962)	79807_at	GSTCD	3.84E-01	1.03
rs6856422 (chr4:106841962)	57117_at	INTS12	2.57E-01	1.04
rs6856422 (chr4:106841962)	255743_at	NPNT	7.90E-02	1.10
rs148274477 (chr6:142838173)	57211_at	GPR126	2.86E-01	1.07
rs34886460 (chr9:119359372)	22954_at	TRIM32	8.29E-01	1.01
rs2274116 (chr9:139094805)	169714_at	QSOX2	3.22E-01	0.97
rs11383346 (chr12:28283187)	55297_at	CCDC91	1.26E-01	1.05
rs10850377 (chr12:115201436)	6926_at	ТВХ3	5.43E-01	0.97
rs7155279 (chr14:92485881)	4287_at	ATXN3	2.07E-02	1.07
rs7155279 (chr14:92485881)	9321_at	TRIP11	5.72E-01	1.03
rs117068593 (chr14:93118229)	79890_at	RIN3	1.65E-01	0.96
rs12149828 (chr16:10706328)	2013_at	EMP2	9.84E-01	1.00
rs113473882 (chr19:41124155)	8425_at	LTBP4	1.18E-01	0.95
rs134041 (chr22:28056338)	4330_at	MN1	8.36E-01	0.99
rs7050036 (chrX:15964845)	8905_at	AP1S2	6.61E-01	1.02
rs7050036 (chrX:15964845)	NA	ZRSR2	NA	NA

f) Sentinel and functional variants annotation and joint analysis results

Annotations were provided using ENSEMBL's Variant Effect Predictor¹⁶. Functional variants were defined as variants within 1 Mb of the sentinel variant, which were annotated as 'deleterious' by SIFT¹⁷, 'probably damaging' or 'potentially damaging' by PolyPhen-2¹⁸, had a CADD¹⁹ scaled score \geq 20, or had a GWAVA score >0.5, and were in linkage disequilibrium (LD) with the sentinel variant (r^2 >0.3) and/or had nominal evidence of association (P<5×10⁻⁴) (see **Supplementary Methods**). Only functional variants, which explained the association of the sentinel variant (sentinel variant joint *P*>0.01), are listed. *RIN3* locus, where the sentinel variant is a missense variant, is also presented. Stage 1 unconditional analysis results and joint analysis results are shown in alternating white and grey colours. Conditional analyses were undertaken using GCTA²⁰ and LD information was estimated using UK BiLEVE data (details in the **Supplementary Methods**). GCTA did not produce joint results for rs34093919 and rs113473882, since they are almost in perfect LD. Chr., chromosome; freq., frequency.

rs number (chr:	Variant	Function	Gene	SIFT	PolyPhen	GWAVA	CADD	Measure	R ² with	MAE	Unconditional results		ults	Joint analysis		
position)	type	Function	Symbol	3IF I	FolyFilen	score	score	MedSure	sentinel	WAF	Beta	SE	Ρ	Beta	SE	Р
rs6681426 (chr1:150586971)	sentinel	Intron variant	ENSA	-	-	0.21	0.93	FEV_1	1.00	0.36	0.042	0.008	1.07E-07	0.034	0.014	1.23E-02
rs9803935 (chr1:150552622)	functional	Upstream gene variant	MCL1	-	-	0.51	7.67	FEV_1	0.68	0.48	0.036	0.008	2.14E-06	0.009	0.013	4.83E-01
rs6681426 (chr1:150586971)	sentinel	Intron variant	ENSA	-	-	0.21	0.93	FEV_1	1.00	0.36	0.042	0.008	1.07E-07	0.032	0.014	2.02E-02
rs1053732 (chr1:150597890)	functional	Intron variant	ENSA	-	-	0.55	3.88	FEV_1	0.68	0.48	0.037	0.008	1.20E-06	0.011	0.013	3.93E-01
rs6441207 (chr3:158282459)	sentinel	Intron variant, nc transcript variant	AK097794	-	-	0.1	0.68	FVC	1.00	0.41	0.036	0.008	4.54E-06	0.030	0.014	3.76E-02
rs939117 (chr3:158200869)	functional	Intron variant	RSRC1	-	-	0.6	16.60	FVC	0.73	0.47	0.031	0.008	3.89E-05	0.007	0.014	6.29E-01
rs148274477 (chr6:142838173)	sentinel	Intergenic variant	-	-	-	0.04	12.33	FEV₁/ FVC	1.00	0.02	-0.161	0.027	2.68E-09	-0.095	0.056	8.98E-02
rs17280293 (chr6:142688969)	functional	Missense variant	GPR126	Deleterious (0)	Possibly damaging (0.602)	-	21.90	FEV ₁ / FVC	0.85	0.02	-0.154	0.026	4.72E-09	-0.073	0.055	1.82E-01
rs12149828 (chr16:10706328)	sentinel	Intergenic variant	-	-	-	0.21	1.06	FEV₁/ FVC	1.00	0.17	0.049	0.010	3.16E-06	0.048	0.021	1.88E-02
rs12448455 (chr16:10711678)	functional	Intergenic variant	-	-	-	0.6	3.84	FEV₁/ FVC	0.80	0.19	0.039	0.010	5.74E-05	0.000	0.019	9.86E-01
rs113473882 (chr19:41124155)	sentinel	Intron variant	LTBP4	-	-	0.11	2.46	FEV₁/ FVC	1.00	0.02	-0.174	0.033	1.48E-07	NA	NA	NA
rs34093919 (chr19:41117300)	functional	Missense variant, splice region variant	LTBP4	Deleterious (0.05)	Possibly damaging (0.841)	-	18.35	FEV₁/ FVC	0.99	0.02	-0.163	0.032	4.82E-07	NA	NA	NA
rs134041 (chr22:28056338)	sentinel	Intergenic variant	-	-	-	0.46	19.28	FEV_1	1.00	0.43	-0.045	0.008	4.19E-09	-0.036	0.016	2.63E-02
rs134117 (chr22:28082715)	functional	Intron variant, nc transcript variant	RP1- 213J1P_B .1	-	-	0.6	12.58	FEV ₁	0.78	0.41	-0.042	0.008	4.43E-08	-0.010	0.016	5.51E-01
rs117068593 (chr4:93118229)	sentinel	Missense variant	RIN3	Tolerated (0.07)	Unknown (0)	-	14.37	FEV_1	1	0.18	-0.048	0.010	2.72E-06	NA	NA	NA

Supplementary Table 4 Literature search results for genes overlapping or proximal to novel lung function sentinel SNP and genes which contain a SNP with $r^2>0.3$ with lead SNP

A systematic literature search was performed for each gene. Three main websites were used. Firstly, each 'gene name' was searched for in Gene Cards (www.genecards.org), as well as the International Mouse Phenotyping Consortium database (www.mousephenotype.org). Secondly, each 'gene name', 'gene name' and 'lung', 'gene name' and 'lung disease', and 'gene name' and 'development' were searched for in Pubmed (www.pubmed.com). Information regarding gene function, association to human disease and animal model phenotyping was identified. Genes included in the gene column include genes containing SNP with r^2 >0.3 with sentinel SNP or genes selected as priority genes for each region (see criteria in **Supplementary Methods**). Chr., chromosome..

rs number (chr: postion:variant type, function)	Genes containing SNPs with r ² >0.3 with sentinel variant or implicated through eQTL analyses	Description of genes in region and biological plausibility	Animal model with lung function or pulmonary phenotype
rs6681426 (1:150586971:SNP, intergenic)	ADAMTSL4;ADAMTSL 4- AS1;ANXA9;ARNT;CE RS2 (LASS2);CTSK;CTSS; CYCSP51;ENSA;GOL PH3L;HORMAD1;LINC 00568;MCL1;RN7SL47 3P;RP11- 235D19.2;RP11- 363I22.3;SETDB1;SN ORA40	ADAMTSL4: Member of the ADAMTS (a disintegrin and metalloproteinase with thrombospondin motifs)-like gene family. Associated diseases include eye disorders such as ectopia lentis ²¹ . ADAMTSL4-AS1: ADAMTSL4 antisense RNA 1. ANXA9: Annexins are calcium-dependent phospholipid-binding proteins. Has calcium-binding sites which are required for annexins to aggregate and cooperatively bind anionic phospholipids and extracellular matrix proteins. ARNT: Binds to ligand-bound aryl hydrocarbon receptor. Aids the movement of a complex to the nucleus, where it promotes the expression of genes involved in xenobiotic metabolism. Also a co-factor for transcriptional regulation by hypoxia- inducible factor 1 which is important in lung development ²² . Chromosomal translocation ARNT with ETV6 gene on chromosome 12 has been described in leukaemia ²³ . CERS2 (LASS2): Also known as LASS2. Involved in sphingolipid synthesis ²⁴ . Associated with Breast Cancer ²⁵ . CTSK: Lysosomal cysteine proteinase involved in bone remodeling and resorption ²⁶ . Mutations in this gene are the cause of pycnodysostosis ²⁷ . CTSS: Lysosomal cysteine proteinase which is involved in the degradation of antigenic proteins to peptides for presentation on MHC class II molecules ³⁶ . Associated diseases include Athersclerosis, Lung Fibrosis and defects in the immune system ³⁹ . CYCSP51: Cytochrome C, Somatic Pseudogene 51. ENSA: Inhibits sulfonylurea binding to beta-cell membranes, reduces cloned K(ATP) channel currents, and stimulates insulin secretion from beta-cells, therefore ia a candidate gene involved in Diabetes ³⁰ . GOLPH3L: Potential regulatory role in Golgi trafficking ³¹ . Associated with Rhabdomyosarcoma ³² . HORMAD1: May play a role in the progression of meiosis and is a potential marker for cancer ³³ . LINC00568: Long Intergenic Non-Protein Coding RNA 568. MCL11: Regulates apoptosis ³⁴ . Mutations found in Leukemia ³⁵ . SNORA40: Small nucleolar RNA40.	<i>CERS2 (LASS2):</i> null mice show lung inflammation and airway obstruction ³⁷ . <i>CTSK:</i> null mice develop more lung fibrosis than wild type mice ³⁸ .
rs201204531 (1:219963090:INDE L, intergenic)	AC096643.1;RNA5SP7 6;SLC30A10	RNA5SP76: RNA, 5S Ribosomal Pseudogene 76, no data available. SLC30A10: Protein critical in maintaining manganese levels with loss of function mutations resulting in dystonia, adult-onset parkinsonism and chronic liver disease ³⁹ .	
rs61067109 (2:18292452:SNP, intergenic)	KCNS3	<i>KCNS3:</i> Regualtes the resting membrane potential and SNP's in this gene are associated with airway hyper- responsiveness ⁴⁰ . Associated with migraine ⁴¹ .	

rs number (chr: postion:variant type, function)	Genes containing SNPs with r ² >0.3 with sentinel variant or implicated through eQTL analyses	Description of genes in region and biological plausibility	Animal model with lung function or pulmonary phenotype
rs6441207 (3:158282459:SNP, ncRNA_intronic)	GFM1; MLF1;MTAPP1;RARR ES1;AK097794(RP11- 538P18.2);RP11- 79M21.3;RSRC1	<i>GFM1</i> : Mitochondrial GTPase that catalyzes the GTP-dependent ribosomal translocation step during translation elongation ⁴² . Diseases associated with GFM1 include combined oxidative phosphorylation deficiency 1 ⁴³ . <i>MLF1</i> : Involved in lineage commitment of primary haemopoietic progenitors by restricting erythroid formation and enhancing myeloid formation ⁴⁴ . Mutations involved in myelodysplastic syndromes and myeloid leukemia ⁴⁵ . Binds DNA and affects the expression of a number of genes so may function as a transcription factor in the nucleus ⁴⁶ . <i>MTAPP1</i> : methylthioadenosine phosphorylase pseudogene 1. <i>RARRES1</i> : Retinoid acid (RA) receptor-responsive gene which inhibits the cytoplasmic carboxypeptidase AGBL2 and may regulate the alpha-tubulin tyrosination cycle ⁴⁷ . Associated with Ovarian Endometriosis and downregulated in prostate cancer ⁴⁸ . <i>RSRC1</i> : Encodes a serine and arginine rich-related protein involved in mRNA splicing. Associated with height ⁴⁹ and Schizophrenia ⁵⁰ .	
rs6856422 (4:106841962:SNP, intronic)	GSTCD;INTS12;NPNT	GSTCD : Has homology with the glutathione S-transferase (GST) super family of enzymes ⁵¹ . May be involved in the detoxification of products of oxidative stress and synthesis of steroid hormones ⁵² . However, GSTCD lacks key functional domains important for GST activity. <i>GSTCD</i> expression in human lung tissue is ubiquitous ⁹ . <i>INTS12</i> : Subunit of the Integrator complex, which associates with the C-terminal domain of RNA polymerase II large subunit and mediates 3` end processing of small nuclear RNAs ⁵³ . <i>INTS12</i> expression in human lung is found predominantly in epithelial cells and pneumocytes ⁹ <i>NPNT</i> : Potentially involved in the development of many tissues ⁵⁴ . Associated with include acute and chronic hepatitis ⁵⁵ .	
rs148274477 (6:142838173:SNP, intergenic)	GPR126;LOC153910(RP11-440G9.1)	<i>GPR126:</i> Encodes a G protein-coupled receptor, with variations affecting stature ^{49,56-58} , birth length ⁵⁹ , and adolescent idiopathic scoliosis ⁶⁰ . May be required for normal differentiation of promyelinating Schwann cells and for normal myelination of axons ⁶¹ . It is expressed in adult mice lung ⁶² . It is required for mice embryonic viability and cardiovascular development ⁶³	
rs34886460 (9:119359372:INDE L, intronic)	TRIM32	ASTN2: Expressed in the brain and may function in neuronal migration. Associated with migraine ⁶⁴ . TRIM32: Protein interacts with the activation domain of the HIV-1 Tat protein. Associated with Bardet-Biedl syndrome (a ciliopathic disorder) ⁶⁵ .	
rs2274116 (9:139094805:SNP, exonic)	LHX3;QSOX2	<i>LHX3:</i> Carries the LIM domain, a unique cysteine-rich zinc-binding domain. Transcription factor required for pituitary development and motor neuron specification. Mutations cause combined pituitary hormone deficiency 3 ⁶⁶ . <i>QSOX2:</i> May contribute to disulfide bond formation in a variety of secreted proteins. Regulates the sensitization of neuroblastoma cells for IFN-gamma-induced cell death ⁶⁷ .	
rs11383346 (12:28283187:INDE L, intergenic)	CCDC91;RP11- 874G11.1;RP11- 967K21.1;RP11- 967K21.2	CCDC91: Associated with brain white matter structure and development ⁶⁸ .	
rs10850377(12:115201436:SNP, intergenic)	RP4-601P9.1;RP4- 601P9.2; TBX3	TBX3: Encodes a transcription factor involved in the regulation of developmental processes in limbs and genitals ⁶⁹ . Mutations cause ulnar-mammary and Holt-Oram syndromes ⁷⁰ . It is involved in the TGFβ1 signaling pathway ^{71,72} .	
rs7155279 (14:92485881:SNP, intronic)	ATXN3;FBLN5;RP11- 529H20.6;TRIP11	 ATXN3: Deubiquitinating enzyme involved in protein homeostasis maintenance, transcription, cytoskeleton regulation, myogenesis and degradation of misfolded chaperone substrates. Associated with Machado-Joseph disease (spinocerebellar ataxia-3)⁷³. FBLN5: Encodes a secreted, extracellular matrix protein. Promotes adhesion of endothelial cells through interaction of integrins and the RGD motif⁷⁴. Defects in gene cause Cutis Laxa, autosomal recessive, type ia, and Hereditary Sensorimotor Neuropathy with Hyperelastic Skin⁷⁵. Involved in tissue repair in COPD⁷⁶ and elastogenesis and lung 	<i>FBLN5:</i> Homozygous mutant mice have expanded lungs containing dilated alveoli (most severe in peripheral regions) ⁸⁰ .

rs number (chr: postion:variant type, function)	Genes containing SNPs with r ² >0.3 with sentinel variant or implicated through eQTL analyses	Description of genes in region and biological plausibility	Animal model with lung function or pulmonary phenotype
		development ⁷⁷ . TRIP11: Protein is associated with the Golgi apparatus ⁷⁸ . Mutations in this gene cause achondrogenesis type IA (most severe form of dwarfism bone/cartilage malformations) ⁷⁹ .	Adult <i>fibulin-5 -/-</i> mice have disruptions in the alveolar structure and disruption to distal airways was found in P3 neonates ⁸¹ .
rs117068593 (14:93118229:SNP, exonic)	RIN3	<i>RIN3:</i> Member of the RIN family of Ras interaction-interference protein ⁸² . Associated with Paget's disease ⁸³ , COPD ⁸⁴ and Alzheimer's disease ⁸⁵ .	
rs12149828 (16:10706328:SNP, intergenic)	EMP2;TEKT5	EMP2: Involved in molecular interactions during implantation ⁸⁶ . Diseases associated include charcot-marie-tooth disease type 1 ⁸⁷ . TEKT5: Tektins are structural components of ciliary and flagellar microtubules. TEKT5 is implicated in sperm motility in mice ⁸⁸ .	
rs113473882 (19:41124155:SNP, intronic)	CTC- 490E21.12;CYP2B6;C YP2B7P;ITPKC;LTBP4 ;NUMBL;RAB4B	CYP2B6 and CYP2B7P: Cytochrome P450 proteins are monooxygenases which are involved in drug metabolism and synthesis of cholesterol, steroids and lipids. Enzymes are involved in an NADPH-dependent electron transport pathway. <i>ITPKC:</i> Encodes inositol 1,4,5-trisphosphate (Ins(1,4,5)P(3)) 3-kinase enzymes. Has nuclear import and export activity. Associated with Kawasaki disease ⁸⁹ . <i>LTBP4:</i> Binds transforming growth factor beta (TGFB) as it is secreted and targeted to the extracellular matrix. COPD risk associated gene with deregulated expression in the basal cells of smokers ⁹⁰ . <i>NUMBL:</i> Has a role in neurogenesis ⁹¹ . <i>RAB4B:</i> RAB proteins are members of the RAS superfamily of small GTPases that are involved in vesicular trafficking ⁹² .	LTBP4: Mice deficient in Latent TGFβ Binding Protein 4 (Ltbp4) display a defect in lung septation and elastogenesis ⁷⁷ . Mice with <i>LTBP4</i> disruption show a pulmonary emphysema- like condition at birth which increases in severity with age. The lungs of <i>LTBP4</i> -disrupted mice had less alveolar spaces and those present were enlarged and inflated. Mice also developed cardiomyopathy and colorectal cancers ³³ .
rs134041(22:28056338:SNP, intergenic)	MN1; RP1- 213J1PB.1;RP11- 375H17.1	<i>MN1:</i> Translocation of MN1 is associated with Leukemia ⁹⁴ .	
rs7050036(X:15964 845:SNP, intergenic)	AP1S2;GRPR;RNU5F- 7P;SETP15;ZRSR2	<i>AP1S2:</i> Adaptor protein complex 1 subunit 2 found at the cytoplasmic face of coated vesicles in the Golgi complex. Mediates the recruitment of clathrin and the recognition of sorting signals within transmembrane receptors. Associated diseases include dandy-walker malformation with mental retardation ^{95,96} , and basal ganglia disease ⁹⁷ . <i>GRPR:</i> Receptor for Gastrin-releasing peptide which regulates gastrointestinal and central nervous system functions including smooth muscle cell contraction and epithelial cell proliferation.Expressed by pulmonary neuroendocrine cells and known to stimulate growth and maturation of lung development ^{98,99} . <i>RNU5F-7P:</i> RNA, U5F Small Nuclear 7, Pseudogene. <i>SETP15:</i> SET Pseudogene 15. <i>ZRSR2: Zinc</i> Finger with RNA-Binding Motif which encodes an essential splicing factor which associates with U2 snRNA. Linked to myelodysplasia ¹⁰⁰ .	

Supplementary Table 5 Associations of top signals with other traits

Associations with lung function in children (a)), separately in ever and never smokers (b)), with smoking behaviour (c)) and with other traits as in the GWAS catalog¹⁰¹ (d)).

a) Effect of top hits on children

Stage 2 results, as well as results in 5,0625062 children (8-9 years of age) in ALSPAC¹⁰² are presented for the new 16 sentinel variants. *P* values that meet a Bonferroni correction for 16 tests (*P* < 3.13×10^{-3}) in ALSPAC are highlighted in bold. Chr., chromosome; dist., distance; Freq., frequency; N., effective sample size; Imp., imputation.

		Measure	Coded allele		Stage 2				ALSPAC				
rs number (chr:position)	Gene Symbol (function)			Other allele	Coded allele freq.	Beta	SE	Р	Coded allele freq.	lmp quality	Beta	SE	Р
rs6681426 (chr1:150586971)	MCL1 (dist=34757), ENSA (dist=7628)	FEV ₁	G	А	0.35	0.021	0.006	1.11E-03	0.35	0.99	0.046	0.030	1.25E-01
rs201204531 (chr1:219963090)	LYPLAL1 (dist=576883), RNU5F-1 (dist=83529)	FEV ₁ /FVC	А	ATG	0.38	-0.027	0.006	1.76E-05	0.38	0.97	-0.048	0.029	9.79E-02
rs61067109 (chr2:18292452)	<i>KCNS3</i> (dist=178227), <i>NT5C1B-RDH14</i> (dist=443537)	FEV ₁ /FVC	G	A	0.77	-0.042	0.007	6.61E-09	0.76	1.00	-0.043	0.033	1.93E-01
rs6441207 (chr3:158282459)	RP11-538P18.2 (ncRNA_intronic)	FVC	С	Т	0.59	0.036	0.006	5.70E-09	0.59	0.98	0.118	0.029	4.72E-05
rs6856422 (chr4:106841962)	NPNT (intronic)	FEV ₁ /FVC	G	т	0.56	-0.055	0.006	1.13E-17	0.55	0.86	-0.074	0.030	1.36E-02
rs148274477 (chr6:142838173)	<i>GPR126</i> (dist=70770), <i>RP11-440G9.1</i> (dist=9419)	FEV ₁ /FVC	С	т	0.97	-0.162	0.019	5.66E-18	0.97	0.76	-0.165	0.094	7.92E-02
rs34886460 (chr9:119359372)	ASTN2 (intronic)	FEV ₁ /FVC	т	ТА	0.54	0.025	0.006	4.09E-05	0.54	0.98	-0.019	0.028	4.97E-01
rs2274116 (chr9:139094805)	LHX3 (exonic)	FVC	С	т	0.66	0.038	0.007	4.16E-09	0.68	0.84	0.060	0.033	6.90E-02
rs11383346 (chr12:28283187)	<i>PTHLH</i> (dist=158271), <i>CCDC91</i> (dist=126946)	FVC	А	AT	0.41	-0.041	0.006	4.29E-11	0.42	0.97	-0.014	0.029	6.29E-01
rs10850377 (chr12:115201436)	<i>TBX3</i> (dist=79467), <i>MED13L</i> (dist=1194945)	FEV ₁	G	А	0.65	-0.028	0.006	1.50E-05	0.65	0.96	-0.031	0.031	3.17E-01
rs7155279 (chr14:92485881)	TRIP11 (intronic)	FEV ₁	G	Т	0.64	-0.022	0.006	4.02E-04	0.64	0.98	-0.013	0.031	6.75E-01
rs117068593 (chr14:93118229)	RIN3 (exonic)	FEV ₁	С	т	0.81	-0.027	0.008	5.68E-04	0.82	0.94	-0.021	0.039	5.90E-01
rs12149828 (chr16:10706328)	<i>EMP2</i> (dist=31789), <i>TEKT5</i> (dist=15033)	FEV ₁ /FVC	G	А	0.83	0.035	0.008	3.33E-05	0.83	0.91	-0.007	0.039	8.58E-01
rs113473882 (chr19:41124155)	LTBP4 (intronic)	FEV ₁ /FVC	Т	С	0.99	-0.138	0.028	9.94E-07	0.99	0.76	0.177	0.147	2.29E-01
rs134041 (chr22:28056338)	<i>MIAT</i> (dist=983898), <i>MN1</i> (dist=87927)	FEV ₁	т	С	0.44	-0.018	0.006	4.12E-03	0.44	0.99	0.003	0.029	9.18E-01
rs7050036 (chrX:15964845)	<i>AP1S2</i> (dist=91708), <i>GRPR</i> (dist=176579)	FEV ₁ /FVC	Т	А	0.39	-0.013	0.005	1.60E-02	0.40	0.98	0.017	0.023	4.70E-01

b) Effect of novel signals on ever smokers and never smokers in stage 1

Stage 1 results for genome-wide significant associations ($P < 5 \times 10^{-8}$) with either FEV₁, FEV₁/FVC or FVC separately in ever smokers and never smokers, and the interaction *P* value are shown. Chr., chromosome.

rs number	Coded	Measure	Ever smo	okers		Never sm	okers		Interaction
(chr:position)	allele	Measure	Beta	SE	Р	Beta	SE	Р	Р
rs6681426 (chr1:150586971)	G	FEV ₁	0.038	0.010	2.54E-04	0.049	0.012	2.07E-05	4.82E-01
rs201204531 (chr1:219963090)	А	FEV ₁ /FVC	-0.039	0.010	1.94E-04	-0.036	0.012	2.63E-03	8.32E-01
rs61067109 (chr2:18292452)	G	FEV ₁ /FVC	-0.043	0.012	3.61E-04	-0.054	0.013	4.04E-05	5.51E-01
rs6441207 (chr3:158282459)	С	FVC	0.029	0.010	5.38E-03	0.042	0.012	3.02E-04	4.09E-01
rs6856422 (chr4:106841962)	G	FEV ₁ /FVC	-0.043	0.011	1.02E-04	-0.044	0.012	4.27E-04	9.69E-01
rs148274477 (chr6:142838173)	С	FEV ₁ /FVC	-0.195	0.037	1.01E-07	-0.118	0.039	2.51E-03	1.51E-01
rs34886460 (chr9:119359372)	Т	FEV ₁ /FVC	0.027	0.010	6.96E-03	0.055	0.011	7.06E-07	6.27E-02
rs2274116 (chr9:139094805)	С	FVC	0.038	0.011	8.04E-04	0.041	0.013	1.19E-03	8.75E-01
rs11383346 (chr12:28283187)	А	FVC	-0.036	0.010	5.67E-04	-0.049	0.011	1.54E-05	3.91E-01
rs10850377 (chr12:115201436)	G	FEV ₁	-0.052	0.011	1.66E-06	-0.042	0.012	3.20E-04	5.35E-01
rs7155279 (chr14:92485881)	G	FEV ₁	-0.049	0.010	3.11E-06	-0.035	0.011	2.47E-03	3.66E-01
rs117068593 (chr14:93118229)	С	FEV ₁	-0.053	0.014	7.86E-05	-0.038	0.015	1.09E-02	4.45E-01
rs12149828 (chr16:10706328)	G	FEV ₁ /FVC	0.057	0.014	3.87E-05	0.038	0.015	1.41E-02	3.45E-01
rs113473882 (chr19:41124155)	Т	FEV ₁ /FVC	-0.206	0.044	3.32E-06	-0.135	0.048	5.42E-03	2.78E-01
rs134041 (chr22:28056338)	Т	FEV ₁	-0.043	0.010	2.25E-05	-0.045	0.011	4.93E-05	8.88E-01
rs7050036 (chrX:15964845)	Т	FEV ₁ /FVC	-0.048	0.011	1.72E-05	-0.033	0.015	3.16E-02	4.46E-01

c) Associations with smoking behaviour in UK BiLEVE for novel signals

Association of the 16 sentinel variants with heavy vs. never smoking status in UK BiLEVE (N heavy=24,457, N never=24,474). *P* that met a Bonferroni corrected threshold for 16 tests (P<3.13x10⁻ ³) are highlighted in bold. Chr., chromosome.

rs number (chr: position)	Coded allele	Other allele	MAF	Imputation quality	Beta	SE	Р
rs6681426 (chr1:150586971)	А	G	0.35	1.00	-0.003	0.013	8.07E-01
rs201204531 (chr1:219963090)	ATG	А	0.38	0.99	0.042	0.013	1.54E-03
rs61067109 (chr2:18292452)	А	G	0.23	1.00	-0.007	0.015	6.33E-01
rs6441207 (chr3:158282459)	Т	С	0.40	1.00	-0.008	0.013	5.30E-01
rs6856422 (chr4:106841962)	Т	G	0.45	0.95	-0.010	0.013	4.50E-01
rs148274477 (chr6:142838173)	Т	С	0.03	0.98	-0.067	0.040	9.45E-02
rs10512249 (chr9:98256309)	А	G	0.10	0.99	0.034	0.022	1.16E-01
rs34886460 (chr9:119359372)	ТА	Т	0.47	1.00	0.020	0.013	1.23E-01
rs2274116 (chr9:139094805)	Т	С	0.34	0.98	-0.018	0.014	1.75E-01
rs11383346 (chr12:28283187)	AT	А	0.41	1.00	-0.029	0.013	2.47E-02
rs10850377 (chr12:115201436)	А	G	0.34	0.98	-0.019	0.014	1.52E-01
rs7155279 (chr14:92485881)	Т	G	0.36	1.00	0.015	0.013	2.63E-01
rs117068593 (chr14:93118229)	Т	С	0.19	0.98	-0.003	0.016	8.50E-01
rs12149828 (chr16:10706328)	А	G	0.16	0.94	-0.017	0.018	3.53E-01
rs113473882 (chr19:41124155)	С	Т	0.01	0.95	-0.150	0.067	2.55E-02
rs134041 (chr22:28056338)	С	Т	0.44	0.98	-0.007	0.013	5.80E-01
rs7050036 (chrX:15964845)	A	Т	0.39	0.98	0.000	0.011	9.69E-01

d) Associations of top hits with other traits

The GWAS catalog¹⁰¹ was searched for variants in 2Mb regions centred on the sentinel variant for the 16 sentinel variants. Variants that were genome-wide significant (P<5x10⁻⁸) in the GWAS catalog¹⁰¹ and were in LD (r^2 >0.3) with the sentinel variants, or were in genes that contained at least one variant in LD (r^2 >0.3) with the sentinel variants were selected. Chr., chromosome.

Sentinel rs number (chr:position)	rs number (chr:position)	R ² with sentinel	PubMed id	Trait	Gene Symbol	Р
rs6681426 (chr1:150586971)	rs2230061 (chr1:150727539)	6.44E-01	24064335	Fat body mass	CTSS	4.00E-08
rs6681426 (chr1:150586971)	rs7412746 (chr1:150860471)	4.09E-01	21983785	Melanoma	ARNT, SETDB1, LASS2, ANXA9, MCL1, CTSK	9.00E-11
rs6681426 (chr1:150586971)	rs267734 (chr1:150951477)	1.30E-01	20383146	Chronic kidney disease	ANXA9,FAM63A,PRUNE,BNIPL,LAS S2,SETDB1	1.00E-12
rs6681426 (chr1:150586971)	rs267733 (chr1:150958836)	8.34E-02	24097068	LDL cholesterol	ANXA9, CERS2	5.00E-09
rs201204531 (chr1:219963090)	rs2820443 (chr1:219753509)	2.93E-02	23754948	Sexual dimorphism in anthropometric traits	LYPLAL1, SLC30A10, ZC3H11B	7.00E-16
rs6441207 (chr3:158282459)	rs2362965 (chr3:158109379)	6.33E-01	23563607	Height	RSRC1,SHOX2	2.00E-09
rs6856422 (chr4:106841962)	rs11727189 (chr4:106619140)	7.48E-02	20010835	Pulmonary function	INTS12, NPNT, FLJ20184, GSTCD	5.00E-17
rs6856422 (chr4:106841962)	rs10516526 (chr4:106688904)	7.50E-02	20010834	Pulmonary function	GSTCD	2.00E-23
rs6856422 (chr4:106841962)	rs17331332 (chr4:106808107)	7.89E-02	23284291	Pulmonary function (interaction)	INTS12, GSTCD, NPNT	1.00E-16
rs148274477 (chr6:142838173)	rs7741741 (chr6:142655801)	6.51E-02	23563607	Height	GPR126	1.00E-20
rs148274477 (chr6:142838173)	rs6570507 (chr6:142679572)	5.94E-02	23666238	Scoliosis	GPR126	4.00E-14
rs148274477 (chr6:142838173)	rs6570507 (chr6:142679572)	5.94E-02	19343178	Height	GPR126	4.00E-11
rs148274477 (chr6:142838173)	rs4896582 (chr6:142703877)	5.52E-02	18391950	Height	GPR126	2.00E-18
rs148274477 (chr6:142838173)	rs3817928 (chr6:142750516)	1.02E-01	23284291	Pulmonary function (interaction)	GPR126	3.00E-12
rs148274477 (chr6:142838173)	rs3817928 (chr6:142750516)	1.02E-01	20010835	Pulmonary function	GPR126	1.00E-09
rs148274477 (chr6:142838173)	rs3748069 (chr6:142767633)	6.58E-02	18391951	Height	GPR126	5.00E-14
rs148274477 (chr6:142838173)	rs7763064 (chr6:142797289)	6.47E-02	20881960	Height	GPR126	1.00E-33
rs34886460 (chr9:119359372)	rs6478241 (chr9:119252629)	5.27E-02	23793025	Migraine - clinic-based	ASTN2	1.00E-09
rs34886460 (chr9:119359372)	rs6478241 (chr9:119252629)	5.27E-02	22683712	Migraine	ASTN2	4.00E-08
rs2274116 (chr9:139094805)	rs7860634 (chr9:139089679)	3.74E-01	23408906	Thyroid hormone levels	LHX3	2.00E-14
rs2274116 (chr9:139094805)	rs7849585 (chr9:139111870)	1.51E-01	20881960	Height	QSOX2	5.00E-14
rs2274116 (chr9:139094805)	rs12338076 (chr9:139121740)	1.55E-01	20189936	Height	LHX3, QSOX2	2.00E-08
rs11383346 (chr12:28283187)	rs2638953 (chr12:28534415)	3.00E-01	20881960	Height	CCDC91	7.00E-17

Sentinel rs number (chr:position)	rs number (chr:position)	R ² with sentinel	PubMed id	Trait	Gene Symbol	Р
rs11383346 (chr12:28283187)	rs10843164 (chr12:28569714)	2.89E-01	23563607	Height	CCDC91	6.00E-12
rs7155279 (chr14:92485881)	rs7153027 (chr14:92427222)	7.41E-01	18391951	Height	TRIP11, FBLN5, ATXN3, CPSF2	1.00E-10
rs7155279 (chr14:92485881)	rs8007661 (chr14:92459958)	6.45E-01	18391950	Height	TRIP11,ATXN3	6.00E-10
rs7155279 (chr14:92485881)	rs7155279 (chr14:92485881)	1.00E+00	20881960	Height	TRIP11	1.00E-10
rs117068593 (chr14:93118229)	rs10498633 (chr14:92926952)	2.75E-05	24162737	Alzheimer's disease (late onset)	SLC24A4, RIN3	6.00E-09
rs117068593 (chr14:93118229)	rs10498635 (chr14:93103309)	9.39E-01	21623375	Paget's disease	RIN3	3.00E-11
rs117068593 (chr14:93118229)	rs754388 (chr14:93115410)	9.94E-01	24621683	Chronic obstructive pulmonary disease (moderate to severe)	RIN3	5.00E-09
rs117068593 (chr14:93118229)	rs754388 (chr14:93115410)	9.94E-01	24945404	Bone mineral density (paediatric, total body less head)	RIN3	3.00E-09
rs117068593 (chr14:93118229)	rs754388 (chr14:93115410)	9.94E-01	24945404	Bone mineral density (paediatric, lower limb)	RIN3	1.00E-10
rs113473882 (chr19:41124155)	rs28493229 (chr19:41224204)	6.63E-04	22081228	Kawasaki disease	ІТРКС	2.00E-12
rs113473882 (chr19:41124155)	rs2233152 (chr19:41281016)	5.98E-04	22446962	Kawasaki disease	NUMBL,MIA	4.00E-10
rs113473882 (chr19:41124155)	rs7937 (chr19:41302706)	5.19E-03	22080838	Chronic obstructive pulmonary disease	RAB4B, EGLN2, CYP2A6	3.00E-09

Supplementary Table 6 List of 49 lung function signals detected to date

Variants associated with FEV₁, FEV₁/FVC or FVC in lung function GWAS to date are presented. The study in which each variant was firstly identified is indicated ("Identified in"), and the results presented ("Results from") correspond to the study with largest sample size available for the previously discovered variants and to the current study for the novel variants. Effect sizes correspond to inverse normally transformed lung function traits in all instances except for those FVC results extracted from Loth 2014⁴, which correspond to ml. Previous studies referred to in the table are: Wilk_2009¹, Repapi_2010⁵, Hancock_2010², Soler_Artigas_2011³ and Loth_2014⁴.

rs number (chr: position)	Gene (closest)	Measure	Identified in	Results from	Alleles	Coded allele	Coded allele freq	Beta	SE	Pval
rs2284746 (chr1:17306675)	MFAP2	FEV1/FVC	Soler_Artigas_2011	Soler_Artigas_2011	C/G	G	0.58	-0.040	0.005	7.50E-16
rs6681426 (chr1:150586971)	ENSA	FEV1	current	current	A/G	G	0.36	0.029	0.005	4.35E-09
rs993925 (chr1:218860068)	TGFB2	FEV1/FVC	Soler_Artigas_2011	Soler_Artigas_2011	C/T	Т	0.42	0.034	0.006	1.16E-08
rs201204531 (chr1:219963090)	RNU5F-1	FEV1/FVC	current	current	ATG/A	А	0.41	-0.031	0.005	2.68E-10
rs61067109 (chr2:18292452)	KCNS3	FEV1/FVC	current	current	A/G	G	0.77	-0.045	0.006	1.40E-15
rs1430193 (chr2:56120853)	EFEMP1	FVC	Loth_2014	Loth_2014	A/T	Т	0.37	- 21.125	2.999	1.86E-12
rs2571445 (chr2:218683154)	TNS1	FEV1	Repapi_2010	Soler_Artigas_2011	A/G	G	0.62	0.047	0.007	9.83E-11
rs12477314 (chr2:239877148)	HDAC4	FEV1/FVC	Soler_Artigas_2011	Soler_Artigas_2011	C/T	Т	0.13	0.041	0.006	1.68E-12
rs1529672 (chr3:25520582)	RARB	FEV1/FVC	Soler_Artigas_2011	Soler_Artigas_2011	A/C	С	0.86	-0.048	0.006	3.97E-14
rs6441207 (chr3:158282459)	RP11- 538P18.2	FVC	current	current	T/C	С	0.59	0.036	0.005	1.27E-13
rs1344555 (chr3:169300219)	MECOM	FEV1	Soler_Artigas_2011	Soler_Artigas_2011	C/T	Т	0.17	-0.034	0.006	2.65E-08
rs2045517 (chr4:89870964)	FAM13A	FEV1/FVC	Hancock_2010	Soler_Artigas_2011	C/T	Т	0.36	-0.047	0.007	2.00E-11
rs10516526 (chr4:106688904)	GSTCD	FEV1	Repapi_2010 & Hancock_2010	Soler_Artigas_2011	A/G	G	0.08	0.108	0.014	4.75E-14
rs6856422 (chr4:106841962)	NPNT	FEV1/FVC	current	current	T/G	G	0.53	-0.051	0.005	1.51E-23
rs11100860 (chr4:145479139)	HHIP	FEV1/FVC	Wilk_2009	Soler_Artigas_2011	A/G	G	0.43	0.064	0.007	6.81E-20
rs153916 (chr5:95036700)	SPATA9	FEV1/FVC	Soler_Artigas_2011	Soler_Artigas_2011	C/T	Т	0.52	-0.031	0.005	2.12E-08
rs1985524 (chr5:147847788)	HTR4	FEV1	Repapi_2010	Soler_Artigas_2011	C/G	G	0.61	-0.048	0.007	3.06E-11
rs11134779 (chr5:156936766)	ADAM19	FEV1/FVC	Hancock_2010	Soler_Artigas_2011	A/G	G	0.31	-0.042	0.007	6.01E-09
rs number (chr: position)	Gene (closest)	Measure	Identified in	Results from	Alleles	Coded allele	Coded allele	Beta	SE	Pval
------------------------------	-------------------	----------	-------------------------------	--------------------	---------	--------------	--------------	-------------	-------	----------
	(,						freq			
rs6923462 (chr6:7801112)	BMP6	FVC	Loth_2014	Loth_2014	C/T	Т	0.84	30.883	4.288	5.89E-13
rs6903823 (chr6:28322296)	ZKSCAN3	FEV1	Soler_Artigas_2011	Soler_Artigas_2011	A/G	G	0.19	-0.037	0.006	2.18E-10
rs2857595 (chr6:31568469)	NCR3	FEV1/FVC	Soler_Artigas_2011	Soler_Artigas_2011	A/G	G	0.79	0.037	0.006	2.28E-10
rs2070600 (chr6:32151443)	AGER	FEV1/FVC	Repapi_2010 & Hancock_2010	Soler_Artigas_2011	C/T	Т	0.05	0.126	0.016	9.07E-15
rs2798641 (chr6:109268050)	ARMC2	FEV1/FVC	Soler_Artigas_2011	Soler_Artigas_2011	C/T	Т	0.13	-0.041	0.007	8.35E-09
rs148274477 (chr6:142838173)	GPR126	FEV1/FVC	current	current	T/C	С	0.98	-0.162	0.015	9.58E-26
rs262129 (chr6:142853144)	LOC153910	FEV1/FVC	Hancock_2010	Soler_Artigas_2011	A/G	G	0.26	0.056	0.008	2.91E-13
rs16909859 (chr9:98204792)	PTCH1	FEV1/FVC	Soler_Artigas_2011	Soler_Artigas_2011	A/G	G	0.89	0.080	0.013	7.45E-10
rs16909898 (chr9:98231008)	PTCH1	FEV1/FVC	Hancock_2010	Soler_Artigas_2011	A/G	G	0.85	-0.072	0.012	3.94E-09
rs34886460 (chr9:119359372)	ASTN2	FEV1/FVC	current	current	TA/T	Т	0.53	0.031	0.005	4.72E-11
rs2274116 (chr9:139094805)	LHX3	FVC	current	current	T/C	С	0.67	0.039	0.005	5.55E-14
rs7068966 (chr10:12277992)	CDC123	FEV1/FVC	Soler_Artigas_2011	Soler_Artigas_2011	C/T	Т	0.43	0.033	0.005	6.13E-13
rs11001819 (chr10:78315224)	C10orf11	FEV1	Soler_Artigas_2011	Soler_Artigas_2011	A/G	G	0.50	-0.029	0.004	2.98E-12
rs4237643 (chr11:43648368)	HSD17B12	FVC	Loth_2014	Loth_2014	T/G	Т	0.31	- 16.666	3.023	3.53E-08
rs2863171 (chr11:45250732)	PRDM11	FVC	Loth_2014	Loth_2014	A/C	С	0.16	23.924	3.906	8.97E-10
rs11383346 (chr12:28283187)	CCDC91	FVC	current	current	AT/A	А	0.41	-0.042	0.005	9.52E-18
rs11172113 (chr12:57527283)	LRP1	FEV1/FVC	Soler_Artigas_2011	Soler_Artigas_2011	C/T	Т	0.66	-0.032	0.006	1.24E-08
rs1036429 (chr12:96271428)	CCDC38	FEV1/FVC	Soler_Artigas_2011	Soler_Artigas_2011	C/T	Т	0.25	0.038	0.006	2.30E-11
rs10850377 (chr12:115201436)	ТВХ3	FEV1	current	current	A/G	G	0.67	-0.035	0.005	2.50E-12
rs7155279 (chr14:92485881)	TRIP11	FEV1	current	current	T/G	G	0.64	-0.030	0.005	1.41E-09
rs117068593 (chr14:93118229)	RIN3	FEV1	current	current	T/C	С	0.82	-0.035	0.006	2.25E-08
rs8033889 (chr15:71680080)	THSD4	FEV1/FVC	Repapi_2010 & Hancock_2010	Soler_Artigas_2011	G/T	Т	0.14	-0.072	0.008	2.03E-17
rs12149828 (chr16:10706328)	EMP2	FEV1/FVC	current	current	A/G	G	0.83	0.040	0.007	7.65E-10
rs12447804 (chr16:58075282)	MMP15	FEV1/FVC	Soler_Artigas_2011	Soler_Artigas_2011	C/T	Т	0.24	-0.038	0.007	3.59E-08
rs2865531 (chr16:75390316)	CFDP1	FEV1/FVC	Soler_Artigas_2011	Soler_Artigas_2011	A/T	Т	0.47	0.031	0.005	1.77E-11
rs1079572 (chr16:78187138)	WWOX	FVC	Loth_2014	Loth_2014	G/A	G	0.42	16.258	2.837	9.95E-09
rs6501431 (chr17:68976415)	KCNJ2	FVC	Loth_2014	Loth_2014	C/T	Т	0.80	23.053	3.884	2.94E-09
rs113473882 (chr19:41124155)	LTBP4	FEV1/FVC	current	current	C/T	Т	0.98	-0.153	0.021	9.95E-13
rs9978142 (chr21:35652239)	KCNE2	FEV1/FVC	Soler_Artigas_2011	Soler_Artigas_2011	A/T	Т	0.22	-0.043	0.008	2.65E-08

rs number (chr: position)	Gene (closest)	Measure	Identified in	Results from	Alleles	Coded allele	Coded allele freq	Beta	SE	Pval
rs134041 (chr22:28056338)	MN1	FEV1	current	current	C/T	Т	0.43	-0.028	0.005	3.03E-09
rs7050036 (chrX:15964845)	AP1S2	FEV1/FVC	current	current	A/T	Т	0.38	-0.023	0.004	4.14E-08

Supplementary Table 7 Proportion of variance explained

Stage 1 results for the 33 previously discovered signals¹⁻⁵ and the 16 novel signals are shown with the proportion of the variance of the lung function traits that they explain (r^2), separately for FEV₁, FEV₁/FVC and FVC. Additive polygenic variance is estimated assuming a heritability for lung function of 40%¹⁰³⁻¹⁰⁵. Chr., chromosome; freq., frequency.

rs number (chr: position)	Closest gene	Coded allele	Non coded allele	Coded allele freq.	Beta (SE)	Р	N	r² (%)
Known				•				
rs2284746 (chr1:17306675)	MFAP2	С	G	0.50	-0.011 (0.008)	1.40E-01	36224	0.006
rs993925 (chr1:218860068)	TGFB2	С	Т	0.66	-0.016 (0.008)	5.32E-02	35599	0.010
rs1430193 (chr2:56120853)	EFEMP1	A	Т	0.61	0.02 (0.008)	1.42E-02	35824	0.017
rs2571445 (chr2:218683154)	TNS1	А	G	0.40	-0.034 (0.008)	8.56E-06	37553	0.053
rs12477314 (chr2:239877148)	HDAC4	С	Т	0.80	-0.032 (0.009)	5.28E-04	37932	0.032
rs1529672 (chr3:25520582)	RARB	С	А	0.83	-0.025 (0.01)	1.17E-02	36309	0.018
rs1344555 (chr3:169300219)	МЕСОМ	С	Т	0.79	0.036 (0.01)	1.62E-04	35867	0.040
rs2045517 (chr4:89870964)	FAM13A	С	Т	0.59	0 (0.008)	9.52E-01	38095	0.000
rs10516526 (chr4:106688904)	GSTCD	А	G	0.94	-0.100 (0.016)	1.52E-10	37284	0.110
rs11100860 (chr4:145479139)	HHIP	А	G	0.58	-0.028 (0.008)	1.96E-04	38063	0.036
rs153916 (chr5:95036700)	SPATA9	С	Т	0.44	-0.004 (0.008)	5.90E-01	37448	0.001
rs1985524 (chr5:147847788)	HTR4	G	С	0.54	-0.045 (0.008)	3.11E-09	37265	0.094
rs11134779 (chr5:156936766)	ADAM19	А	G	0.66	0.029 (0.008)	1.85E-04	38090	0.037
rs6923462 (chr6:7801112)	BMP6	Т	С	0.84	0.037 (0.01)	3.00E-04	37005	0.035
rs6903823 (chr6:28322296)	ZKSCAN3	А	G	0.78	0.035 (0.01)	3.71E-04	37973	0.033
rs114327456 (chr6:31568469)	NCR3	G	А	0.78	0.036 (0.01)	3.22E-04	34997	0.037
rs114177847 (chr6:32151443)	AGER	С	Т	0.95	-0.03 (0.021)	1.63E-01	25796	0.008
rs2798641 (chr6:109268050)	ARMC2	С	Т	0.82	0.026 (0.01)	8.57E-03	37706	0.018
rs262129 (chr6:142853144)	LOC153910	Α	G	0.70	-0.01 (0.008)	2.24E-01	37585	0.004
rs16909859 (chr9:98204792)	PTCH1	G	Α	0.92	-0.007 (0.014)	6.06E-01	33748	0.001
rs16909898 (chr9:98231008)	PTCH1	А	G	0.90	-0.031 (0.013)	1.53E-02	36168	0.016
rs7068966 (chr10:12277992)	CDC123	С	Т	0.48	-0.037 (0.008)	9.53E-07	37832	0.063
rs11001819 (chr10:78315224)	C10orf11	G	A	0.52	-0.027 (0.008)	3.44E-04	38018	0.034
rs4237643 (chr11:43648368)	HSD17B12	Т	G	0.30	-0.026 (0.008)	1.27E-03	38018	0.027
rs2863171 (chr11:45250732)	PRDM11	A	С	0.84	-0.032 (0.01)	2.11E-03	38138	0.025
rs11172113 (chr12:57527283)	LRP1	Т	С	0.60	-0.011 (0.008)	1.46E-01	37724	0.006
rs1036429 (chr12:96271428)	CCDC38	Т	С	0.19	0.011 (0.01)	2.29E-01	38123	0.004
rs8033889 (chr15:71680080)	THSD4	G	Т	0.78	0.032 (0.009)	3.29E-04	38080	0.034
rs12447804 (chr16:58075282)	MMP15	С	Т	0.77	0.008 (0.009)	3.89E-01	37782	0.002
rs2865531 (chr16:75390316)	CFDP1	Т	Α	0.41	0.025 (0.008)	1.24E-03	37942	0.027
rs1079572 (chr16:78187138)	WWOX	G	A	0.43	0.016 (0.008)	3.41E-02	37809	0.012
rs6501431 (chr17:68976415)	KCNJ2	С	Т	0.21	-0.021 (0.009)	2.35E-02	37869	0.014
rs9978142 (chr21:35652239)	KCNE2	A	Т	0.85	0.010 (0.011)	3.46E-01	37633	0.002
Total r ² for known variants								0.855
Total additive polygenic variance	e for known v	ariants						2.138
New								
rs6681426 (chr1:150586971)	ENSA	G	А	0.36	0.042 (0.008)	1.07E-07	37944	0.074
rs201204531 (chr1:219963090)	RNU5F-1	А	ATG	0.41	-0.007 (0.008)	3.75E-01	34887	0.002

rs number (chr: position)	Closest gene	Coded allele	Non coded allele	Coded allele freq.	Beta (SE)	Р	N	r ² (%)	
rs61067109 (chr2:18292452)	KCNS3	G	А	0.77	-0.039 (0.009)	1.83E-05	37433	0.049	
rs6441207 (chr3:158282459)	AK097794	С	Т	0.59	0.029 (0.008)	2.07E-04	36426	0.038	
rs6856422 (chr4:106841962)	NPNT	G	Т	0.53	-0.045 (0.008)	6.22E-08	31460	0.093	
rs148274477 (chr6:142838173)	LOC153910	С	Т	0.98	0.010 (0.027)	7.07E-01	30318	0.000	
rs34886460 (chr9:119359372)	ASTN2	Т	ТА	0.53	0.023 (0.008)	2.85E-03	37583	0.024	
rs2274116 (chr9:139094805)	LHX3	С	Т	0.67	0.029 (0.009)	6.27E-04	32226	0.036	
rs11383346 (chr12:28283187)	CCDC91	А	AT	0.41	-0.039 (0.008)	3.41E-07	37510	0.069	
rs10850377 (chr12:115201436)	TBX3	G	А	0.67	-0.047 (0.008)	6.68E-09	37268	0.090	
rs7155279 (chr14:92485881)	TRIP11	G	Т	0.64	-0.041 (0.008)	1.39E-07	37691	0.074	
rs117068593 (chr14:93118229)	RIN3	С	Т	0.82	-0.048 (0.01)	2.72E-06	34496	0.064	
rs12149828 (chr16:10706328)	TEKT5	G	А	0.83	0.037 (0.01)	3.38E-04	34022	0.038	
rs113473882 (chr19:41124155)	LTBP4	Т	С	0.98	-0.029 (0.033)	3.78E-01	32220	0.002	
rs134041 (chr22:28056338)	MN1	Т	С	0.43	-0.045 (0.008)	4.19E-09	37669	0.092	
rs7050036 (chrX:15964845)	AP1S2	Т	А	0.38	-0.014 (0.007)	4.41E-02	32292	0.013	
Total r2 for new variants								0.758	
Total additive polygenic variand	ce for new var	iants						1.896	
Total r ² for known and new vari	ants							1.614	
Total additive polygenic variance for known and new variants									

rs number (chr: position)	Closest gene	Coded allele	Non coded allele	Coded allele freq.	Beta (SE)	Р	N	r² (%)
Known				-				
rs2284746 (chr1:17306675)	MFAP2	С	G	0.50	0.043 (0.008)	2.19E-08	36208	0.086
rs993925 (chr1:218860068)	TGFB2	С	Т	0.66	-0.021 (0.008)	1.04E-02	35582	0.018
rs1430193 (chr2:56120853)	EFEMP1	А	Т	0.61	-0.012 (0.008)	1.25E-01	35808	0.007
rs2571445 (chr2:218683154)	TNS1	А	G	0.40	-0.012 (0.008)	1.11E-01	37537	0.007
rs12477314 (chr2:239877148)	HDAC4	С	Т	0.80	-0.052 (0.009)	3.59E-08	37915	0.080
rs1529672 (chr3:25520582)	RARB	С	А	0.83	-0.046 (0.01)	6.43E-06	36292	0.056
rs1344555 (chr3:169300219)	MECOM	С	Т	0.79	0.021 (0.01)	2.81E-02	35856	0.013
rs2045517 (chr4:89870964)	FAM13A	С	Т	0.59	0.035 (0.008)	4.78E-06	38078	0.055
rs10516526 (chr4:106688904)	GSTCD	А	G	0.94	-0.051 (0.016)	1.08E-03	37267	0.029
rs11100860 (chr4:145479139)	HHIP	А	G	0.58	-0.054 (0.008)	8.60E-13	38046	0.134
rs153916 (chr5:95036700)	SPATA9	С	Т	0.44	0.032 (0.008)	3.55E-05	37429	0.046
rs1985524 (chr5:147847788)	HTR4	G	С	0.54	-0.039 (0.008)	2.50E-07	37246	0.071
rs11134779 (chr5:156936766)	ADAM19	А	G	0.66	0.036 (0.008)	4.06E-06	38073	0.056
rs6923462 (chr6:7801112)	BMP6	Т	С	0.84	0 (0.01)	9.92E-01	36987	0.000
rs6903823 (chr6:28322296)	ZKSCAN3	А	G	0.78	0.013 (0.01)	1.78E-01	37958	0.005
rs114327456 (chr6:31568469)	NCR3	G	А	0.78	0.023 (0.01)	2.08E-02	34980	0.015
rs114177847 (chr6:32151443)	AGER	С	Т	0.95	-0.118 (0.021)	2.08E-08	25785	0.122
rs2798641 (chr6:109268050)	ARMC2	С	Т	0.82	0.033 (0.01)	7.86E-04	37689	0.030
rs262129 (chr6:142853144)	LOC153910	А	G	0.70	-0.033 (0.008)	5.92E-05	37568	0.043
rs16909859 (chr9:98204792)	PTCH1	G	А	0.92	0.055 (0.014)	1.43E-04	33738	0.043
rs16909898 (chr9:98231008)	PTCH1	А	G	0.90	0.052 (0.013)	5.48E-05	36153	0.045
rs7068966 (chr10:12277992)	CDC123	С	Т	0.48	-0.034 (0.008)	6.00E-06	37816	0.054
rs11001819 (chr10:78315224)	C10orf11	G	А	0.52	-0.018 (0.008)	2.00E-02	38002	0.014
rs4237643 (chr11:43648368)	HSD17B12	Т	G	0.30	-0.004 (0.008)	6.64E-01	38001	0.000
rs2863171 (chr11:45250732)	PRDM11	А	С	0.84	-0.009 (0.01)	3.77E-01	38121	0.002
rs11172113 (chr12:57527283)	LRP1	Т	С	0.60	-0.032 (0.008)	3.41E-05	37707	0.046
rs1036429 (chr12:96271428)	CCDC38	Т	С	0.19	0.03 (0.01)	1.84E-03	38106	0.025
rs8033889 (chr15:71680080)	THSD4	G	Т	0.78	0.071 (0.009)	4.31E-15	38063	0.162
rs12447804 (chr16:58075282)	MMP15	С	Т	0.77	0.024 (0.009)	6.36E-03	37766	0.020
rs2865531 (chr16:75390316)	CFDP1	Т	А	0.41	0.037 (0.008)	1.29E-06	37925	0.062
rs1079572 (chr16:78187138)	WWOX	G	А	0.43	-0.002 (0.008)	7.84E-01	37792	0.000
rs6501431 (chr17:68976415)	KCNJ2	С	Т	0.21	0.014 (0.009)	1.24E-01	37852	0.006
rs9978142 (chr21:35652239)	KCNE2	А	Т	0.85	0.028 (0.011)	7.58E-03	37623	0.019
Total r ² for known variants								1.372
Total additive polygenic varianc	e for known va	ariants						3.430
New					1	I		
rs6681426 (chr1:150586971)	ENSA	G	А	0.36	0.014 (0.008)	7.47E-02	37927	0.008
rs201204531 (chr1:219963090)	RNU5F-1	А	ATG	0.41	-0.038 (0.008)	1.98E-06	34866	0.065

b. FEV_1/FVC

rs number (chr: position)	Closest gene	Coded allele	Non coded allele	Coded allele freq.	Beta (SE)	Р	N	r² (%)		
rs61067109 (chr2:18292452)	KCNS3	G	А	0.77	-0.05 (0.009)	3.09E-08	37416	0.082		
rs6441207 (chr3:158282459)	AK097794	С	Т	0.59	-0.003 (0.008)	6.56E-01	36408	0.001		
rs6856422 (chr4:106841962)	NPNT	G	Т	0.53	-0.044 (0.008)	1.30E-07	31446	0.089		
rs148274477 (chr6:142838173)	LOC153910	С	Т	0.98	-0.161 (0.027)	2.68E-09	30398	0.116		
rs34886460 (chr9:119359372)	ASTN2	Т	ТА	0.53	0.041 (0.008)	6.75E-08	37567	0.078		
rs2274116 (chr9:139094805)	LHX3	С	Т	0.67	-0.013 (0.009)	1.29E-01	32205	0.007		
rs11383346 (chr12:28283187)	CCDC91	А	AT	0.41	-0.002 (0.008)	8.38E-01	37497	0.000		
rs10850377 (chr12:115201436)	TBX3	G	А	0.67	-0.02 (0.008)	1.10E-02	37245	0.017		
rs7155279 (chr14:92485881)	TRIP11	G	Т	0.64	-0.023 (0.008)	3.49E-03	37675	0.023		
rs117068593 (chr14:93118229)	RIN3	С	Т	0.82	0.008 (0.01)	4.23E-01	34476	0.002		
rs12149828 (chr16:10706328)	TEKT5	G	А	0.83	0.049 (0.01)	3.16E-06	33999	0.064		
rs113473882 (chr19:41124155)	LTBP4	Т	С	0.98	-0.174 (0.033)	1.48E-07	32207	0.086		
rs134041 (chr22:28056338)	MN1	Т	С	0.43	-0.025 (0.008)	1.06E-03	37653	0.028		
rs7050036 (chrX:15964845)	AP1S2	Т	А	0.38	-0.041 (0.007)	4.37E-09	32285	0.107		
Total r ² for new variants								0.772		
Total additive polygenic varianc	e for new vari	ants						1.931		
Total r ² for known and new varia	ants							2.144		
Total additive polygenic variance for known and new variants 5										

rs number (chr: position)	Closest gene	Coded allele	Non coded allele	Coded allele freq.	Beta (SE)	Р	N	r² (%)
Known								
rs2284746 (chr1:17306675)	MFAP2	С	G	0.50	-0.039 (0.008)	3.61E-07	35967	0.072
rs993925 (chr1:218860068)	TGFB2	С	Т	0.66	-0.006 (0.008)	4.86E-01	35348	0.001
rs1430193 (chr2:56120853)	EFEMP1	А	Т	0.61	0.032 (0.008)	9.67E-05	35571	0.043
rs2571445 (chr2:218683154)	TNS1	А	G	0.40	-0.031 (0.008)	6.97E-05	37297	0.042
rs12477314 (chr2:239877148)	HDAC4	С	Т	0.80	0.002 (0.009)	8.15E-01	37678	0.000
rs1529672 (chr3:25520582)	RARB	С	А	0.83	0.003 (0.01)	7.58E-01	36053	0.000
rs1344555 (chr3:169300219)	MECOM	С	Т	0.79	0.023 (0.01)	1.46E-02	35627	0.017
rs2045517 (chr4:89870964)	FAM13A	С	Т	0.59	-0.023 (0.008)	2.66E-03	37839	0.024
rs10516526 (chr4:106688904)	GSTCD	А	G	0.94	-0.076 (0.016)	1.54E-06	37028	0.062
rs11100860 (chr4:145479139)	HHIP	А	G	0.58	0.009 (0.008)	2.30E-01	37807	0.004
rs153916 (chr5:95036700)	SPATA9	С	Т	0.44	-0.029 (0.008)	1.62E-04	37195	0.038
rs1985524 (chr5:147847788)	HTR4	G	С	0.54	-0.02 (0.008)	1.13E-02	37010	0.017
rs11134779 (chr5:156936766)	ADAM19	А	G	0.66	0.007 (0.008)	3.60E-01	37834	0.002
rs6923462 (chr6:7801112)	BMP6	Т	С	0.84	0.048 (0.01)	3.52E-06	36750	0.059
rs6903823 (chr6:28322296)	ZKSCAN3	А	G	0.78	0.03 (0.01)	2.55E-03	37719	0.024
rs114327456 (chr6:31568469)	NCR3	G	А	0.78	0.025 (0.01)	1.47E-02	34741	0.017
rs114177847 (chr6:32151443)	AGER	С	Т	0.95	0.048 (0.022)	2.79E-02	25540	0.019
rs2798641 (chr6:109268050)	ARMC2	С	Т	0.82	0.007 (0.01)	4.81E-01	37450	0.001
rs262129 (chr6:142853144)	LOC153910	А	G	0.70	0.005 (0.008)	5.48E-01	37330	0.001
rs16909859 (chr9:98204792)	PTCH1	G	А	0.92	-0.048 (0.014)	9.58E-04	33507	0.033
rs16909898 (chr9:98231008)	PTCH1	А	G	0.90	-0.072 (0.013)	2.87E-08	35924	0.086
rs7068966 (chr10:12277992)	CDC123	С	Т	0.48	-0.015 (0.008)	5.33E-02	37577	0.010
rs11001819 (chr10:78315224)	C10orf11	G	А	0.52	-0.016 (0.008)	3.26E-02	37763	0.012
rs4237643 (chr11:43648368)	HSD17B12	Т	G	0.30	-0.028 (0.008)	7.14E-04	37763	0.030
rs2863171 (chr11:45250732)	PRDM11	А	С	0.84	-0.031 (0.01)	3.23E-03	37882	0.023
rs11172113 (chr12:57527283)	LRP1	Т	С	0.60	0.013 (0.008)	8.95E-02	37468	0.008
rs1036429 (chr12:96271428)	CCDC38	Т	С	0.19	-0.006 (0.01)	5.48E-01	37867	0.001
rs8033889 (chr15:71680080)	THSD4	G	Т	0.78	-0.011 (0.009)	2.25E-01	37824	0.004
rs12447804 (chr16:58075282)	MMP15	С	Т	0.77	-0.005 (0.009)	6.06E-01	37525	0.001
rs2865531 (chr16:75390316)	CFDP1	Т	А	0.41	0.004 (0.008)	6.11E-01	37685	0.001
rs1079572 (chr16:78187138)	WWOX	G	А	0.43	0.017 (0.008)	2.21E-02	37553	0.014
rs6501431 (chr17:68976415)	KCNJ2	С	Т	0.21	-0.033 (0.009)	3.74E-04	37612	0.034
rs9978142 (chr21:35652239)	KCNE2	А	Т	0.85	-0.011 (0.011)	2.81E-01	37382	0.003
Total r ² for known variants								0.703
Total additive polygenic varianc	e for known va	ariants						1.757
New								
rs6681426 (chr1:150586971)	ENSA	G	А	0.36	0.036 (0.008)	4.74E-06	37688	0.056
rs201204531 (chr1:219963090)	RNU5F-1	А	ATG	0.41	0.011 (0.008)	1.53E-01	34632	0.006

rs number (chr: position)	Closest gene	Coded allele	Non coded allele	Coded allele freq.	Beta (SE)	Р	N	r ² (%)		
rs61067109 (chr2:18292452)	KCNS3	G	А	0.77	-0.009 (0.009)	3.05E-01	37177	0.003		
rs6441207 (chr3:158282459)	AK097794	С	Т	0.59	0.036 (0.008)	4.54E-06	36173	0.058		
rs6856422 (chr4:106841962)	NPNT	G	Т	0.53	-0.02 (0.008)	1.54E-02	31226	0.019		
rs148274477 (chr6:142838173)	LOC153910	С	Т	0.98	0.114 (0.027)	3.07E-05	30110	0.058		
rs34886460 (chr9:119359372)	ASTN2	Т	ТА	0.53	0 (0.008)	9.72E-01	37330	0.000		
rs2274116 (chr9:139094805)	LHX3	С	Т	0.67	0.041 (0.009)	2.68E-06	32004	0.069		
rs11383346 (chr12:28283187)	CCDC91	А	AT	0.41	-0.043 (0.008)	3.83E-08	37254	0.081		
rs10850377 (chr12:115201436)	TBX3	G	А	0.66	-0.033 (0.008)	3.60E-05	37015	0.046		
rs7155279 (chr14:92485881)	TRIP11	G	Т	0.64	-0.031 (0.008)	8.62E-05	37439	0.041		
rs117068593 (chr14:93118229)	RIN3	С	Т	0.82	-0.054 (0.01)	1.74E-07	34254	0.080		
rs12149828 (chr16:10706328)	TEKT5	G	А	0.83	0.005 (0.01)	6.12E-01	33774	0.001		
rs113473882 (chr19:41124155)	LTBP4	Т	С	0.98	0.073 (0.033)	2.88E-02	31982	0.015		
rs134041 (chr22:28056338)	MN1	Т	С	0.43	-0.031 (0.008)	5.44E-05	37415	0.044		
rs7050036 (chrX:15964845)	AP1S2	Т	А	0.38	0.01 (0.007)	1.47E-01	32283	0.007		
Total r ² for new variants								0.582		
Total additive polygenic varianc	e for new varia	ants						1.454		
Total r ² for known and new variants 1.										
Total additive polygenic variance for known and new variants 3										

Supplementary Table 8 Results for 1000 Genomes sentinel variants with lower allele frequency than HapMap sentinel variants in two known loci

Shown are stage 1 and UK BiLEVE results for sentinel variants in previously reported lung function regions ²⁻⁵ which had minor allele frequency (<10%) more than two-fold lower than the HapMap sentinel variants in these regions, with the exception of a signal in *GPR126* (rs148274477) whose results are presented in **Table 1**. "GWAS sentinel" and "GWAS gene" present the sentinel SNP reported in a previous GWAS ³ and the closest gene to sentinel SNP respectively. Chr., chromosome.

01/140	01/140		D0 with	Coded/	Stage 1					UK BILEVE				
sentinel	gene	(chr:position)	sentinel	allele	MAF	Neff	Beta	SE	Р	MAF	Neff	Beta	SE	Р
rs993925	TGFB2	rs147187942 (chr1:218773216)	1.73E-02	T/A	0.090	31881	-0.047	0.014	8.72E-04	0.085	46263	-0.032	0.012	5.72E-03
rs12447804	MMP15	rs150232756 (chr16:57691218)	9.78E-06	C/T	0.005	28940	0.203	0.057	3.65E-04	0.005	42760	-0.057	0.051	2.67E-01

Supplementary Table 9 MAGENTA results

MAGENTA results both for the main analysis ("main") and the sensitivity analysis after removing the HLA region ("sensitivity (no HLA)") for pathways that met a Bonferroni threshold corrected for the number of pathways tested per database and had an FDR<5% in the main analysis are presented.

Measure	Analysis	Database	Bonferroni corrected threhold	Pathway	Ρ	FDR
FVC	main	Ingenuity	6.17E-04	PDGF.Signaling	2.00E-04	0.0028
FVC	main	PANTHER BIOLOGICAL PROCESS	2.30E-04	Chromatin packaging and remodeling	1.82E-04	0.0357
FVC	sensitivity (no HLA)	Ingenuity	6.17E-04	PDGF Signaling	6.60E-05	0.0023
FVC	sensitivity (no HLA)	PANTHER BIOLOGICAL PROCESS	2.30E-04	Chromatin packaging and remodeling	4.00E-04	0.0781

Supplementary Notes:

Supplementary Note 1: Heterogeneity test

A Chi-square test for heterogeneity was undertaken and I^2 calculated for the stage 1 results across studies for the 16 novel sentinel variants. Results are presented below; none of the results meets a Bonferroni correted threshold for 16 tests (*P*<3.13x10⁻³).

rs number (chr:position)	Gene (function)	Measure	X ²	Р	Degrees of freedom	l ²
rs6681426 (chr1:150586971)	MCL1 (dist=34757), ENSA (dist=7628)	FEV ₁	12.37	7.18E-01	16	0.00
rs201204531 (chr1:219963090)	LYPLAL1 (dist=576883), RNU5F-1 (dist=83529)	FEV ₁ /FVC	12.51	5.65E-01	14	0.00
rs61067109 (chr2:18292452)	<i>KCNS3</i> (dist=178227), <i>NT5C1B</i> (dist=443537)	FEV ₁ /FVC	17.30	3.66E-01	16	7.54
rs6441207 (chr3:158282459)	AK097794 (ncRNA_intronic)	FVC	11.69	7.02E-01	15	0.00
rs6856422 (chr4:106841962)	NPNT (intronic)	FEV ₁ /FVC	9.62	8.43E-01	15	0.00
rs148274477 (chr6:142838173)	<i>GPR126</i> (dist=70770), <i>LOC153910</i> (dist=9419)	FEV ₁ /FVC	25.12	6.77E-02	16	36.31
rs34886460 (chr9:119359372)	ASTN2 (intronic)	FEV ₁ /FVC	9.20	9.05E-01	16	0.00
rs2274116 (chr9:139094805)	LHX3 (exonic)	FVC	25.52	4.34E-02	15	41.22
rs11383346 (chr12:28283187)	<i>PTHLH</i> (dist=158271), <i>CCDC91</i> (dist=126946)	FVC	18.35	3.04E-01	16	12.82
rs10850377 (chr12:115201436)	<i>TBX3</i> (dist=79467), <i>MED13L</i> (dist=1194945)	FEV ₁	25.10	6.81E-02	16	36.26
rs7155279 (chr14:92485881)	TRIP11 (intronic)	FEV ₁	12.43	7.14E-01	16	0.00
rs117068593 (chr14:93118229)	RIN3 (exonic)	FEV ₁	16.86	3.27E-01	15	11.02
rs12149828 (chr16:10706328)	<i>EMP</i> 2 (dist=31789), <i>TEKT5</i> (dist=15033)	FEV ₁ /FVC	19.17	2.06E-01	15	21.75
rs113473882 (chr19:41124155)	LTBP4 (intronic)	FEV ₁ /FVC	15.91	4.59E-01	16	0.00
rs134041 (chr22:28056338)	<i>MIAT</i> (dist=983898), <i>MN1</i> (dist=87927)	FEV ₁	13.89	6.07E-01	16	0.00
rs7050036 (chrX:15964845)	AP1S2 (dist=91708), GRPR (dist=176579)	FF	9.49	7.35E-01	13	0.00

Supplementary Note 2: Stepwise conditional analysis to identify multiple signals within each region

In order to identify independent signals within regions already associated with lung function, we undertook a stepwise conditional analysis using GCTA²⁰ for each of the previously discovered¹⁻⁵ and new regions. This method uses a stepwise procedure to select a subset of independent variants that when fitted together in the model each has a joint *P* value below a pre specified threshold. In order to be comprehensive we defined the threshold for inclusion in the model as joint *P* value < 10⁻³ and we defined regions as 1Mb either side of the sentinel variant; when two regions overlapped we merged them into one larger region.

We identified one additional independent ($r^2 < 0.02$ with sentinel variant) variant with joint *P* value < 10^{-5} in two regions, near the previously discovered signals in *ZKSCAN3* and *CCDC38* (see bold *P*-values in the table below). Models including the variants selected by the stepwise conditional analysis in these two regions were fitted using UK BiLEVE data and none of these two variants was significant.

	Known		Code			Uncondition	s		Joint ar	Correlation			
Known sentinel rs number (chr:position)	GWAS gene symbol	rs number (chr:position)	allele / other allele	R ² with sentinel	Dataset	Coded allele frequency	Beta	SE	Р	Beta	SE	Р	(r) with the next variant
		rs73395396	СЛ	0.02	stage 1	0.99	-0.046	0.045	3.13E-01	0.389	0.107	2.76E-04	0.92
		(chr6:27479793)	0/1	0.02	UK BILEVE	0.99	0.055	0.032	8.78E-02	0.090	0.089	3.10E-01	0.93
rs6903823 (cbr6:28322206)	ZKSCAN3	rs200955	T/C	0.02	stage 1	0.99	-0.123	0.042	3.68E-03	-0.477	0.100	2.00E-06	0.14
(CIIIO.20022290)		(chr6:27839312)	1/0	0.02	UK BILEVE	0.99	-0.044	0.032	1.75E-01	-0.065	0.090	4.67E-01	0.14
		rs1778483	T/G	0.66	stage 1	0.73	0.034	0.009	2.25E-04	0.036	0.009	8.64E-05	
(chr6:28240991)	(chr6:28240991)	1/6	0.00	UK BILEVE	0.68	0.037	0.007	6.71E-08	0.036	0.007	1.84E-07		
		rs200233711	тло	0.00	stage 1	0.60	0.033	0.009	1.23E-04	0.038	0.009	9.20E-06	-0.05
	(chr12:95306163)	1/10	0.00	UK BILEVE	0.59	0.013	0.007	5.37E-02	0.013	0.007	5.63E-02	-0.04	
		rs66498225	СЛ	0.00	stage 1	0.94	-0.052	0.016	9.66E-04	-0.056	0.016	4.11E-04	-0.06
		(chr12:95330172)	0/1	0.00	UK BILEVE	0.93	-0.036	0.013	4.54E-03	-0.036	0.013	5.20E-03	-0.07
		rs2060304		0.00	stage 1	0.94	-0.049	0.016	1.58E-03	-0.065	0.016	4.12E-05	0.01
rs1036429	CCDC38	(chr12:95486073)	AG	0.00	UK BILEVE	0.93	0.000	0.013	9.69E-01	-0.007	0.013	6.08E-01	0.00
(CHI12.90271420)		rs150198783	C/A	0.00	stage 1	0.99	0.154	0.037	3.11E-05	0.155	0.037	2.57E-05	0.02
		(chr12:96141778)	G/A	0.00	UK BILEVE	0.99	0.020	0.043	6.38E-01	0.015	0.043	7.19E-01	0.05
		rs7134025	СЛ	0.25	stage 1	0.15	0.045	0.011	2.43E-05	0.046	0.011	1.38E-05	0.01
		(chr12:96153846)	0/1	0.25	UK BILEVE	0.14	0.023	0.009	1.60E-02	0.023	0.009	1.47E-02	0.03
		rs7959665	G/A	0.00	stage 1	0.02	-0.096	0.028	6.89E-04	-0.103	0.028	2.71E-04	
		(chr12:97069873)	G/A	0.00	UK BILEVE	0.02	-0.026	0.023	2.45E-01	-0.028	0.023	2.13E-01	

Supplementary Note 3: Individual study descriptions

GWAS samples and phenotype measurement

This section describes study-specific characteristics that are not presented in the tables. All participants provided written informed consent and studies were approved by local Research Ethics Committees and/or Institutional Review boards.

Details of the **British 1958 Birth Cohort** biomedical follow-up have been previously reported ¹¹⁷. Spirometry at age 44–45 years was done in the standing position without nose clips, using a Vitalograph handheld spirometer as previously described ¹¹⁸. In the analysis, all readings with a besttest variation greater than 10% were excluded.

The **Busselton Health Study** (BHS) is a longitudinal survey of the town of Busselton in the southwestern region of Western Australia that began in 1966. In 1994/1995 a cross-sectional community follow-up study was undertaken where blood was taken for DNA extraction. A sample of 1,168 European-ancestry individuals were genotyped using the Illumina 610-Quad BeadChip (BHS1), and subsequent genotyping was carried out on an independent group of 3,428 European-ancestry individuals using Illumina 660W-Quad (BHS2). Spirometric measures of forced expired volume in one second (FEV₁) and forced vital capacity (FVC) were assessed.

The CROATIA study was initiated to investigate the use of isolated rather than urban populations for the identification of genes associated with medically-relevant quantitative traits. Three cohorts have been recruited as part of the CROATIA study: **CROATIA-Vis**¹¹⁹, **CROATIA-Korcula**¹²⁰ and

CROATIA-Split¹²¹. CROATIA-Vis was the first to be collected when 1,008 Croatians aged 18-93 recruited from the villages of Komiza and Vis on the Dalmatian island of Vis. Recruitment occurred from 2003 to 2004 with participants donating blood for DNA extraction and biochemical measurements as well as undergoing some anthropometric measurements and physiological tests to measure traits such as height, weight and blood pressure, and finally completing several questionnaires relating to general health, medical history, diet and lifestyle. CROATIA-Korcula was recruited from 2007 to 2008 from the town of Korcula and the villages of Lumbarda, Zrnovo and Racisce on the island of Korcula, Croatia with 969 adults aged 18-98 agreeing to participate. This study followed the same recruitment procedures as CROATIA-Vis and the same samples and tests were collected with a few additions to reflect the research interests and expertise in Edinburgh. Volunteers were recruited to be part of the CROATIA-Split cohort in 2009-2010 from the Dalmatian mainland city of Split. This is the main ferry port to the islands and is the second largest city in Croatia and the largest along the Dalmatian coast. 1,012 adults aged 18-85 were recruited using the same methodology and with the same samples collected as in CROATIA-Korcula. Ethical approval was obtained from appropriate regulatory bodies in both Scotland and Croatia and participants gave informed consent prior to joining the study.

The **EPIC Norfolk** GWA cohort includes 2,566 participants randomly selected from the EPIC-Norfolk Study, a population-based cohort study of 25,663 men and women of European descent aged 39-79 years recruited in Norfolk, UK between 1993 and 1997¹²². Respiratory function was assessed by spirometry^{123,124}. Forced expiratory volume in 1 second (FEV₁) and forced vital capacity (FVC) were measured twice using a portable spirometer (Micro Medical, Rochester, United Kingdom). The higher values of the two readings for FEV₁ and FVC were used for the analyses. The **Generation Scotland: Scottish Family Health Study** is a collaboration between the Scottish Universities and the NHS, funded by the Chief Scientist Office of the Scottish Government. GS:SFHS is a family-based genetic epidemiology cohort with DNA, other biological samples (serum, urine and cryopreserved whole blood) and socio-demographic and clinical data from ~24,000 volunteers, aged 18-98 years, in ~7,000 family groups. Participants were recruited across Scotland, with some family members from further afield, from 2006-2011. Most (87%) participants were born in Scotland and 96% in the UK or Ireland. The cohort profile has been published¹²⁵. GS:SFHS operates under appropriate ethical approvals, and all participants gave written informed consent. Generation Scotland is a collaboration between the University Medical Schools and National Health Service in Aberdeen, Dundee, Edinburgh and Glasgow (UK).

The DNA archive established from the **Health 2000** Survey Cohort was used. Details of this study population and phenotyping procedures have been previously reported¹²⁶. Genome-wide genotyping was available for 2124 individuals selected from the Health 2000 cohort as metabolic syndrome cases and their matched controls¹²⁷. Spirometry was done in the standing position without nose clips, using a Vitalograph 2150 spirometer. In the analysis, the maximum permissible difference between the two highest FEV₁ and FVC values was 10%.

The KORA studies (Cooperative Health Research in the Region of Augsburg) are a series of independent population based studies from the general population living in the region of Augsburg, Southern Germany^{128,129}. **KORA F4** including 3,080 individuals was conducted from 2006-2008 as a follow-up study to KORA S4 (1999-2001). Lung function tests were performed in a random subsample of subjects born between 1946 and 1965 (age range 41–63 years). Spirometry was performed in line with the ATS/ERS recommendations¹³⁰ using a pneumotachograph-type spirometer 89

(Masterscreen PC, CardinalHealth, Würzburg, Germany) before and after inhalation of 200µg salbutamol. The present study is based on maximum values of FEV₁ and FVC measured before bronchodilation. The spirometer was calibrated daily using a calibration pump (CardinalHealth, Würzburg, Germany), and additionally, an internal control was used to ensure constant instrumental conditions. For KORA F4 participants without spirometry measurements in 2006-2008, we used measurements from the KORA-Age time point conducted in 2008/09. KORA Age contains subjects from all KORA studies born until 1943 (aged 65-90 years)¹³¹. Spirometry was measured in 935 randomly selected participants. Conditions including the examiner were the same as in 2008/09 except that inhalation of salbutamol was not performed due to the high number of contraindications anticipated in this aged population.

The KORA studies (Cooperative Health Research in the Region of Augsburg) are a series of independent population based studies from the general population living in the region of Augsburg, Southern Germany^{128,129}. The **KORA S3** study including 4,856 individuals was conducted in 1994/95. Spirometry was measured during a follow up in 1997/98 for all participants younger than 60 years who did not smoke or use inhalers one hour before the test. All spirometric tests were performed strictly adhering to the ECRHS protocol^{132,133} using Biomedin Spirometers (Biomedin srl, Padova, Italy). Tests were accounted valid if at least two technically satisfactory manoeuvres could be obtained throughout a maximum of nine trials. FEV₁ and FVC were defined as the maximum value within all valid manoeuvers. For KORA S3 participants without spirometry measurements in 1997/98 we used measurements from the KORA-Age time point conducted in 2008/09. KORA Age contains subjects from all KORA studies born until 1943 (aged 65–90 years) ¹³¹. Spirometry was measured in 935 randomly selected participants. Conditions including the examiner were the same as in KORA F4

(see below) except that inhalation of salbutamol was not performed due to the high number of contraindications anticipated in this aged population.

The **Lothian Birth Cohort 1936** consists of 1,091 relatively healthy individuals assessed on cognitive and medical traits at about 70 years of age. They were all born in 1936 and most took part in the Scottish Mental Survey of 1947. At baseline the sample of 548 men and 543 women had a mean age 69.6 years (s.d. = 0.8). They were all Caucasian, community-dwelling, and almost all lived in the Lothian region (Edinburgh city and surrounding area) of Scotland. A full description of participant recruitment and testing can be found elsewhere¹³⁴. Genotyping was performed at the Wellcome Trust Clinical Research Facility, Edinburgh. Quality control measures were applied and 1,005 participants remained. Lung function assessing peak expiratory flow rate, forced expiratory volume in 1 second, and forced vital capacity (each the best of three), using a Micro Medical Spirometer was assessed, sitting down without nose clips, at age 70 years. The accuracy of the spirometer is ±3% (to ATS recommendations Standardisation of Spirometry 1994 update for flows and volumes).

The **Northern Finland Birth Cohort** (NFBC) study programme was initiated in the 1960s. The cohort of women and their offspring was established in the provinces of Oulu and Lapland and had an expected date of birth in 1966 comprising 12,231 children (NFBC1966, 15). The NFBC1966 had spirometry and other measurements done at the age of 31 years. In NFBC1966, we used a Vitalograph P-model spirometer (Vitalograph Ltd., Buckingham, UK), with a volumetric accuracy of ±2% or ±50 mL whichever was greater. The spirometer was calibrated regularly using a 1-Litre precision syringe. The spirometric manoeuvre was performed three times but was repeated if the coefficient of variation between two maximal readings was >4%.

91

The **Northern Sweden Population Health Study** (NSPHS) represents a cross-sectional study conducted in the communities of Karesuando (samples gathered in 2006) and Soppero (2009) in the subarctic region of the County of Norrbotten, Sweden. Spirometry was performed in sitting position without noseclips using a MicroMedicalSpida 5 spirometer (http://www. medisave.co.uk). Three consecutive 28 lung function measurements per participant were done and the maximum value per measured lung function parameter was used for further analysis. Relatedness was taken into account by applying the "polygenic" linear mixed effects model. Genome-wide association analysis was performed using a score test, a family-based association test¹³⁵ which uses the residuals and the variance-covariance matrix from the polygenic model and the SNP fixed effect coded under an additive model.

The **Orkney Complex Disease Study** (ORCADES) is an ongoing family-based, cross-sectional study in the isolated Scottish archipelago of Orkney. Spirometry was performed in the sitting position without nose clips, using a Spida handheld spirometer. Measurements were repeated once and the better reading was used for analysis.

The **SAPALDIA** cohort is a population-based multi-center study in eight geographic areas representing the range of environmental, meteorological and socio-demographic conditions in Switzerland^{136,137}. It was initiated in 1991 (SAPALDIA 1) with a follow-up assessment in 2002 (SAPALDIA 2) and 2010 (SAPALDIA3). This study has specifically been designed to investigate longitudinally lung function, respiratory and cardiovascular health; to study and identify the

associations of these health indicators with individual long term exposure to air pollution, other toxic inhalants, life style and molecular factors.

The Study of Health in West Pomerania (SHIP) is a cross-sectional, population based survey in a region in the Northeast of Germany. Study details are given elsewhere^{138,139}. The examinations were conducted using a bodyplethysmograph equipped with a pneumotachograph (VIASYS Healthcare, JAEGER, Hoechberg, Germany) which meets the American Thoracic Society (ATS) criteria¹⁴⁰. The volume signal of the equipment was calibrated with a 3.0 litre syringe connected to the pneumotachograph in accordance with the manufacturer's recommendations and at least once on each day's testing. Barometric pressure, temperature and relative humidity were registered every morning. Calibration of reference gas and volume was examined under ATS-conditions (Ambient Temperature Pressure) and the integrated volumes were BTPS (Body Temperature Pressure Saturated) corrected^{140,141}. Lung function variables were measured continuously throughout the baseline breathing and the forced manoeuvres using a VIASYS HEALTHCARE system (MasterScreen Body/Diff.). Spirometry flow volume loops were conducted in accordance with ATS recommendations 40 in a sitting position and with wearing noseclips. The participants performed at least three forced expiratory lung function manoeuvres in order to obtain a minimum of two acceptable and reproducible values 41. Immediate on-screen error codes indicating the major acceptability (includingstart, duration and end of test) and reproducibility criteria supported the attempt for standardised procedures. The procedure was continuously monitored by a physician. The best results for FVC, FEV₁, peak expiratory flow (PEF) and expiratory flow at 75%, 50%, 25% of FVC (MEF 75, MEF 50, MEF 25) were taken. The ratio of FEV₁ to FVC was calculated from the largest FEV₁ and FVC.

The **YFS** is a population-based follow up-study started in 1980¹⁴². The main aim of the YFS is to determine the contribution made by childhood lifestyle, biological and psychological measures to the risk of cardiovascular diseases in adulthood. In 1980, over 3,500 children and adolescents all around Finland participated in the baseline study. The follow-up studies have been conducted mainly with 3-year intervals. The latest 30-year follow-up study was conducted in 2010-2011 (ages 33-49 years) with 2,063 participants. The study was approved by the local ethics committees (University Hospitals of Helsinki, Turku, Tampere, Kuopio and Oulu) and was conducted following the guidelines of the Declaration of Helsinki. All participants gave their written informed consent.

Follow-up samples and phenotype measurement

The **Avon Longitudinal Study of Parents and their Children** (ALSPAC) is a population-based birth cohort study consisting initially of over 13,000 women and their children recruited in the county of Avon, UK in the early 1990s¹⁴³. At 8–9 years of age, children's lung function was measured by spirometry (Vitalograph 2120, Maids Moreton, UK) according to American Thoracic Society criteria. Flow-volume curves were reviewed by a respiratory physician to ensure adherence to standards. GWA data and lung function measurements were available on 5,062 of the children from this study. Ethical approval for the study was obtained from the ALSPAC Ethics and Law Committee and the Local Research Ethics Committees. Please note that the study website contains details of all the data that is available through a fully searchable data dictionary

(http://www.bris.ac.uk/alspac/researchers/data-access/data-dictionary).

Details of the methods of **ECRHS I** and **ECRHS II**, a multicentre international cohort study, have been published elsewhere^{132,144}. Participants within the ECRHS were eligible for inclusion in this analysis if

they were identified by random sampling of those who fulfilled the following criteria 1) lived in centres that took part in genome-wide genotyping initiative under the auspices of GABRIEL¹⁴⁵ AND 2) were initially selected to take part in the ECRHS clinical measurements as part of the random sample (ie not specifically selected for inclusion because of any pre-existing disease). Participants were included in this analysis if they also provided a technically satisfactory forced expiratory manoeuvre, compliant with ATS spirometry criteria, at the time of the first survey (aged 20-48). Most centers used the BIOMEDIN water-sealed spirometer for lung function measures and all centers conducted manoeuvres in the sitting position with nose clips on. Further details are available in¹⁴⁶.

The **Prospective Investigation of the Vasculature in Uppsala Seniors** (PIVUS)¹⁴⁷ is a populationbased study of the cardiovascular health in the elderly. The main purpose of PIVUS was to investigate the role The Prospective Investigation of the Vasculature in Uppsala Seniors (PIVUS) of endothelial function in cardiovascular risk. Mailed invitations were sent to subjects who lived in Uppsala, Sweden, within 2 months after their 70th birthday. The subjects were randomly selected from the community register. A total of 1,016 men and women participated in the baseline investigation (participation rate, 50.1%). Spirometry was performed in 901 subjects at baseline in accordance with American Thoracic Society recommendations (α spirometer; Vitalograph Ltd; Buckingham, UK). The best value from three recordings was used. The Ethics Committee of the University of Uppsala approved the study, and the participants gave their informed consent. Genotyping of all samples was undertaken using the Illumina Omni Express and CardioMetabochip. Genotypes were called using GENCALL. A total of 738,879 SNPs passed quality control (thresholds: call rate < 0.95, and call rate < 0.99 for MAF<5%; HWE $P < 10^{-6}$). SNPs with MAF<1% were removed from the imputation scaffold. Imputation was performed using IMPUTE up to CEU haplotypes from Phase II HapMap reference panel.

For this analysis, 3,023 participants (85% female and all of Caucasian ancestry) within the **TwinsUK** adult twin registry based at St. Thomas' Hospital in London were analyzed for whom both genotype and IOP information was available. Twins largely volunteered unaware of the eye studies interests at the time of enrolment and gave fully informed consent under a protocol reviewed by the St. Thomas' Hospital Local Research Ethics Committee. Subjects were genotyped in two different batches of approximately the same size, using two genotyping platforms from Illumina: 300K Duo for and HumanHap610-Quad arrays. Whole genome imputation of the genotypes was performed using HapMap2 (www.hapmap.org) haplotypes. Stringent quality control (QC) measures were implemented, including minimum genotyping success rate (>95%), Hardy-Weinberg equilibrium (P>10⁻⁶), minimum MAF (>1%) and imputation quality score (>0.7). Subjects of non Caucasian ancestry were excluded from the analysis. Association was calculated for all SNPs using a score test as implemented in the software GEMMA¹⁴⁸ (http://www.xzlab.org/software/gemma-0.94.tar.gz), accounting for possible covariates as explained in the sections explaining the general methodology.

The **UK Biobank Lung Exome Variant Evaluation** (UK BiLEVE) project was the first genetic study in UK Biobank and is described in¹⁴⁹. UK Biobank (http://www.ukbiobank.ac.uk/) contains data from 502,682 individuals (94% of self-reported European ancestry) with extensive health and life-style questionnaire data, physical measures (including spirometry) and DNA. Spirometry was undertaken using a Vitalograph Pneumotrac 6800. The participant was asked to record two to three blows (lasting for at least 6 seconds) within a period of about 6 minutes. The computer compared the reproducibility of the first two blows and, if acceptable (defined as a <5% difference in forced volume vital capacity (FVC) and Forced Expiratory Volume in 1 second (FEV₁), a third blow was not required. For the UK BiLEVE project, a sampling frame of 275,939 individuals was defined as those who were of European ancestry and had spirometry measures which met ERS/ATS guidelines¹³⁰. A total of 50,008 samples were selected from the extremes and middle of the distributions of percent predicted FEV₁, separately in never smokers and heavy smokers (20,005 individuals with low FEV₁, 19,997 with average FEV₁ and 10,006 with high FEV₁). DNA was extracted and genotyped with the custom-designed Affymetrix Axiom UK BiLEVE array. Following thorough variant and sample QC, 48,943 unrelated individuals remained for further analysis.

Supplementary Note 4: Investigators, contributions, funding and acknowledgments per study

SpiroMeta consortium

Analysis group: Ian P Hall (co-chair), Suzanne Miller, Nick Shrine, María Soler Artigas, David P Strachan, Martin D Tobin (co-chair), Louise V Wain.

Bioinformatics and functional assessment group: Ian P Hall, Abdul K Kheirallah, Erik Melén, Suzanne Miller, Ioanna Ntalla, Ian Sayers, María Soler Artigas, Martin D Tobin, Louise V Wain.

Writing group: Ian P Hall (co-chair), Suzanne Miller, María Soler Artigas, David P Strachan, Martin D Tobin (co-chair), Louise V Wain.

Funding & Acknowledgements:

The research undertaken by MDT, MSA and LVW was part-funded funded by the National Institute for Health Research (NIHR). The views expressed are those of the author(s) and not necessarily those of the NHS, the NIHR or the Department of Health. MDT holds a Medical Research Council Senior Clinical Fellowship (G0902313). This research used the ALICE High Performance Computing Facility at the University of Leicester. The Universities of Leicester and Nottingham acknowledge receipt of a Collaborative Research and Development grant from the Healthcare and Bioscience iNet, a project funded by the East Midlands Development Agency (EMDA), part-financed by the European Regional Development Fund and delivered by Medilink East Midlands. IPH holds a Medical Research Council programme grant (G1000861).

GWAS studies

B58C

Investigators: Wendy L McArdle, David P Strachan.

Contributions: project conception, design and management: D.P.S., phenotype collection and data management: W.L.M., D.P.S., data analysis: D.P.S. **Funding & Acknowledgements**: We acknowledge use of phenotype and genotype data from the British 1958 Birth Cohort DNA collection, funded by the Medical Research Council grant G0000934 and the Wellcome Trust grant 068545/Z/02. (http://www.b58cgene.sgul.ac.uk/). Genotyping for the B58C-WTCCC subset was funded by the Wellcome Trust grant 076113/B/04/Z. The B58C-T1DGC genotyping utilized resources provided by the Type 1 Diabetes Genetics Consortium, a collaborative clinical study sponsored by the National Institute of Diabetes and Digestive and Kidney Diseases (NIDDK), National Institute of Allergy and Infectious Diseases (NIAID), National Human Genome Research Institute (NHGRI), National Institute of Child Health and Human Development (NICHD), and Juvenile Diabetes Research Foundation International (JDRF) and supported by U01 DK062418. B58C-T1DGC GWAS data were deposited by the Diabetes and Inflammation Laboratory, Cambridge Institute for Medical Research (CIMR), University of Cambridge, which is funded by Juvenile Diabetes Research Foundation International, the Wellcome Trust and the National Institute for Health Research Cambridge Biomedical Research Centre; the CIMR is in receipt of a Wellcome Trust Strategic Award (079895).

The B58C-GABRIEL genotyping was supported by a contract from the European Commission Framework Programme 6 (018996) and grants from the French Ministry of Research.

BHS1&2

Investigators: John Beilby, Jennie Hui, Alan L James, AW Musk.

Contributions: phenotype collection and data management: J.B., J. Hui, A.L.J, A.W.M. **Funding & Acknowledgements**: The Busselton Health Study (BHS) acknowledges the generous support for the 1994/5 follow-up study from Healthway, Western Australia and the numerous Busselton community volunteers who assisted with data collection and the study participants from the Shire of Busselton. The Busselton Health Study is supported by The Great Wine Estates of the Margaret River region of Western Australia. GWAS genotyping was supported by a research collaboration with Pfizer.

CROATIA-Korcula

Investigators: Caroline Hayward, Jennifer E Huffman, Pau Navarro.

Contributions: project conception, design and management: C.H., phenotype collection and data management: C.H., J.E.H., P.N., data analysis: J.E.H., P.N. **Funding & Acknowledgements**: The CROATIA study was supported through grants from the Medical Research Council UK, the Ministry of Science, Education and Sport in the Republic of Croatia (number 216-1080315-0302), Croatian Science Foundation (grant 8875) and the European Union framework program 6 EUROSPAN project (contract no. LSHG-CT-2006-018947). SNP genotyping for CROATIA-Vis was performed by the Wellcome Trust Clinical Research Facility (WTCRF) at the Western General Hospital, Edinburgh, UK. CROATIA-Korcula was genotyped by Helmholz Zentrum München, GmbH, Neuherberg, Germany and CROATIA-Split by AROS Applied Biotechnology, Aarhus, Denmark. We would like to acknowledge the invaluable contributions of the recruitment teams in Croatia (including those from the Institute of Anthropological Research in Zagreb and the Croatian Centre for Global Health at the University of Split), the administrative teams in Croatia and Edinburgh and the people of Korcula, Vis and Split.

CROATIA-Split

Investigators: Ivana Kolcic, Ozren Polasek, Veronique Vitart, Tatijana Zemunik. **Contributions**: project conception, design and management: I.K., O.P., V.V., T.Z., phenotype collection and data management: I.K., O.P., V.V., T.Z., **Funding & Acknowledgements**: The CROATIA study was supported through grants from the Medical Research Council UK, the Ministry of Science, Education and Sport in the Republic of Croatia (number 216-1080315-0302), Croatian Science Foundation (grant 8875) and the European Union framework program 6 EUROSPAN project (contract no. LSHG-CT-2006-018947). SNP genotyping for CROATIA-Vis was performed by the Wellcome Trust Clinical Research Facility (WTCRF) at the Western General Hospital, Edinburgh, UK. CROATIA-Korcula was genotyped by Helmholz Zentrum München, GmbH, Neuherberg, Germany and CROATIA-Split by AROS Applied Biotechnology, Aarhus, Denmark. We would like to acknowledge the invaluable contributions of the recruitment teams in Croatia (including those from the Institute of Anthropological Research in Zagreb and the Croatian Centre for Global Health at the University of Split), the administrative teams in Croatia and Edinburgh and the people of Korcula, Vis and Split.

CROATIA-Vis

Investigators: Jonathan Marten, Igor Rudan, Alan F Wright.

Contributions: project conception, design and management: I.R., A.F.W., phenotype collection and data management: J.M., data analysis: J.M. **Funding & Acknowledgements**: The CROATIA study was supported through grants from the Medical Research Council UK, the Ministry of Science, Education and Sport in the Republic of Croatia (number 216-1080315-0302), Croatian Science Foundation (grant 8875) and the European Union framework program 6 EUROSPAN project (contract no. LSHG-CT-2006-018947). SNP genotyping for CROATIA-Vis was performed by the Wellcome Trust Clinical Research Facility (WTCRF) at the Western General Hospital, Edinburgh, UK. CROATIA-Korcula was genotyped by Helmholz Zentrum München, GmbH, Neuherberg, Germany and CROATIA-Split by AROS Applied Biotechnology, Aarhus, Denmark. We would like to acknowledge the invaluable contributions of the recruitment teams in Croatia (including those from the Institute of Anthropological Research in Zagreb and the Croatian Centre for Global Health at the University of Split), the administrative teams in Croatia and Edinburgh and the people of Korcula, Vis and Split.

EPIC

Investigators: Jing Hua Zhao, Robert A Scott, Nicholas J Wareham.

Contributions: project conception, design and management: J.H.Z., R.A.S., N.J.W., phenotype collection and data management: J.H.Z., data analysis: J.H.Z., **Funding & Acknowledgements**: The EPIC Norfolk Study is funded by program grants from the Medical Research Council UK and Cancer Research UK, and by additional support from the European Union, Stroke Association, British Heart Foundation, Department of Health, Food Standards Agency, and the Wellcome Trust.

GS:SFHS

Investigators: Lynne Hocking, Sandosh Padmanabhan, Generation Scotland, Holly Trochet. **Contributions**: project conception, design and management: L.H., S.P., G.S., phenotype collection and data management: L.H., S.P., G.S., H.T. **Funding & Acknowledgements**: GS:SFHS is funded by the Scottish Executive Health Department, Chief Scientist Office, grant number CZD/16/6. Exome array genotyping for GS:SFHS was funded by the Medical Research Council UK and performed at the Wellcome Trust Clinical Research Facility Genetics Core at Western General Hospital, Edinburgh, UK. We would like to acknowledge the invaluable contributions of the families who took part in the 100 Generation Scotland: Scottish Family Health Study, the general practitioners and Scottish School of Primary Care for their help in recruiting them, and the whole Generation Scotland team, which includes academic researchers, IT staff, laboratory technicians, statisticians and research managers.

H2000

Investigators: Markku Heliövaara, Mika Kähönen, Samuli Ripatti, Ida Surakka. **Contributions**: project conception, design and management: M. Heliövaara, M.K., phenotype collection and data management: M. Heliövaara, M.K., S.R, I. Surakka, data analysis: M.K., S.R, I. Surakka **Funding & Acknowledgements**: This study was financially supported by the Medical Research Fund of the Tampere University Hospital. S.R. was supported by the Academy of Finland (251217 and 255847), Center of Excellence in Complex Disease Genetics, EU FP7 projects ENGAGE (201413) and BioSHaRE (261433), the Finnish Foundation for Cardiovascular Research, Biocentrum Helsinki, and the Sigrid Juselius Foundation.

KORA F4

Investigators: Eva Albrecht, Harald Grallert, Joachim Heinrich, Janina S Ried. **Contributions**: project conception, design and management: J. Heinrich, phenotype collection and data management: H.G., J.S.R., data analysis: E.A., J.S.R. **Funding & Acknowledgements**: The KORA authors acknowledge all members of field staffs who were involved in the planning and conduction of the KORA Augsburg studies, as well as all KORA study participants. The KORA research platform (KORA, Cooperative Health Research in the Region of Augsburg) was initiated and financed by the Helmholtz Zentrum München - German Research Center for Environmental Health which is funded by the German Federal Ministry of Education and Research and by the State of Bavaria. The KORA-Age project was financed by the German Federal Ministry of Education and Research (BMBF FKZ 01ET0713 and 01ET1003A) as part of the 'Health in old age' program. Furthermore, KORA research was supported within the Munich Center of Health Sciences (MC Health), Ludwig-Maximilians-Universität, as part of LMUinnovativ. Further support was provided by the Competence Network ASCONET, subnetwork COSYCONET (FKZ 01GI0882).

KORA S3

Investigators: Christian Gieger, Stefan Karrasch, Rajesh Rawal, Holger Schulz. **Contributions**: project conception, design and management: C.G., S.K., H.S., phenotype collection and data management: C.G., S.K., R.R., H.S., data analysis: R.R., **Funding & Acknowledgements**: The KORA authors acknowledge all members of field staffs who were involved in the planning and conduction of the KORA Augsburg studies, as well as all KORA study participants. The KORA research platform (KORA, Cooperative Health Research in the Region of Augsburg) was initiated and financed by the Helmholtz Zentrum München - German Research Center for Environmental Health which is funded by the German Federal Ministry of Education and Research and by the State of Bavaria. The KORA-Age project was financed by the German Federal Ministry of Education and Research (BMBF FKZ 01ET0713 and 01ET1003A) as part of the 'Health in old age' program. Furthermore, KORA research was supported within the Munich Center of Health Sciences (MC Health), Ludwig-Maximilians-Universität, as part of LMUinnovativ. Further support was provided by the Competence Network ASCONET, subnetwork COSYCONET (FKZ 01GI0882).

LBC1936

Investigators: Ian J Deary, Sarah E Harris, Lorna M Lopez, John M Starr **Contributions**: project conception, design and management: I.J.D., J.M.S., phenotype collection and data management: I.J.D., S.E.H., L.M.L., J.M.S., data analysis: L.M.L. **Funding & Acknowledgements**: We thank the cohort participants who contributed to this study. Genotyping was supported by the UK's Biotechnology and Biological Sciences Research Council (BBSRC) (Ref. BB/F019394/1). Phenotype collection was supported by Research Into Ageing (continues as part of Age UK's The Disconnected Mind project). The work was undertaken by The University of Edinburgh Centre for Cognitive Ageing and Cognitive Epidemiology, part of the cross council Lifelong Health and Wellbeing Initiative (MR/K026992/1). Funding from the BBSRC and Medical Research Council (MRC) is gratefully acknowledged.

NFBC1966

Investigators: Alexessander Couto Alves, Anna-Liisa Hartikainen, Momoko Horikoshi, Marjo-Riitta Jarvelin.

Contributions: project conception, design and management: M-R.J., phenotype collection and data management: A.C.A., A-L.H., M-R.J., data analysis: A.C.A., M. Horikoshi, M-R.J. **Funding & Acknowledgements**: We thank the late Professor Paula Rantakallio (launch of NFBC1966), and Ms Outi Tornwall and Ms MinttuJussila (DNA biobanking). NFBC1966 received financial support from the Academy of Finland (project grants 104781, 120315, 129269, 1114194, 24300796), University Hospital Oulu, Biocenter, University of Oulu, Finland (75617), NHLBI grant 5R01HL087679-02 through the STAMPEED program (1RL1MH083268-01), NIH/NIMH (5R01MH63706:02), EU FP7 (HEALTH-F4-2007-201413), EU FP8 (277849), Medical Research Council, UK (G0500539, G1002319, G0600705).

NSPHS

Investigators: Stefan Enroth, Ulf Gyllensten, Åsa Johansson.

Contributions: project conception, design and management: U.G., phenotype collection and data management: S.E., A.J., data analysis: S.E. **Funding & Acknowledgements**: U.G. acknowledges Swedish Medical Research Council (K2007-66X-20270-01-3, 2012-2884), Foundation for Strategic Research (SSF) and European Commission FP6 STRP (LSHG-CT-2006-01947). A.J. acknowledges Swedish Society for Medical Research (SSMF).

ORCADES

Investigators: Harry Campbell, Peter K Joshi, Sarah H Wild, James F Wilson. **Contributions**: project conception, design and management: H.C., S.H.W., J.F.W., phenotype collection and data management: H.C., S.H.W., J.F.W., data analysis: P.K.J. **Funding & Acknowledgements**: The ORCADES study was funded by the Chief Scientist Office of the Scottish Government, the Royal Society and the MRC Human Genetics Unit. DNA extraction was performed at the Wellcome Trust Clinical Research Facility in Edinburgh. Genotyping was funded by the European Union Framework Programme 6 EUROSPAN project.

SAPALDIA

Investigators: Medea Imboden, Ashish Kumar, Nicole M Probst-Hensch. **Contributions:** project conception, design and management: N.M.P-H., phenotype collection and data management: M.I., A.K., N.M.P-H., data analysis: M.I., A.K., N.M.P-H. Funding & Acknowledgements: Study directorate: NM Probst-Hensch (PI; e/g); T Rochat (p), C Schindler (s), N Künzli (e/exp), JM Gaspoz (c) Scientific team: JC Barthélémy (c), W Berger (g), R Bettschart (p), A Bircher (a), C Brombach (n), PO Bridevaux (p), L Burdet (p), Felber Dietrich D (e), M Frey (p), U Frey (pd), MW Gerbase (p), D Gold (e), E de Groot (c), W Karrer (p), F Kronenberg (g), B Martin (pa), A Mehta (e), D Miedinger (o), M Pons (p), F Roche (c), T Rothe (p), P Schmid-Grendelmeyer (a), D Stolz (p), A Schmidt-Trucksäss (pa), J Schwartz (e), A Turk (p), A von Eckardstein (cc), E Zemp Stutz (e). Scientific team at coordinating centers: M Adam (e), I Aguilera (exp), S Brunner (s), D Carballo (c), S Caviezel (pa), I Curjuric (e), A Di Pascale (s), J Dratva (e), R Ducret (s), E Dupuis Lozeron (s), M Eeftens (exp), I Eze (e), E Fischer (g), M Foraster (e), M Germond (s), L Grize (s), S Hansen (e), A Hensel (s), M Imboden (g), A Ineichen (exp), A Jeong (g), D Keidel (s), A Kumar (g), N Maire (s), A Mehta (e), R Meier (exp), E Schaffner (s), T Schikowski (e), M Tsai (exp) (a) allergology, (c) cardiology, (cc) clinical chemistry, (e) epidemiology, (exp) exposure, (g) genetic and molecular biology, (m) meteorology, (n) nutrition, (o) occupational health, (p) pneumology, (pa) physical activity, (pd) pediatrics, (s) statistic. The study could not have been done without the help of the study participants, technical and administrative support and the medical teams and field workers at the local study sites. Local fieldworkers: Aarau: S Brun, G Giger, M Sperisen, M Stahel, Basel: C Bürli, C Dahler, N Oertli, I Harreh, F Karrer, G Novicic, N Wyttenbacher, Davos: A Saner, P Senn, R Winzeler, Geneva: F Bonfils, B Blicharz, C Landolt, J Rochat, Lugano: S Boccia, E Gehrig, MT Mandia, G Solari, B Viscardi, Montana: AP Bieri, C Darioly, M Maire, Payerne: F Ding, P Danieli A Vonnez, Wald: D Bodmer, E Hochstrasser, R Kunz, C Meier, J Rakic, U Schafroth, A Walder. Administrative staff: N Bauer Ott, C Gabriel, R Gutknecht. Funding: The Swiss National Science Foundation (grants no 33CS30-148470/1, 33CSCO-134276/1, 33CSCO-108796, 3247BO-104283, 3247BO-104288, 3247BO-104284, 3247-065896, 3100-059302, 3200-052720, 3200-042532, 4026-028099, PMPDP3 129021/1, PMPDP3 141671/1), the Federal Office for the Environment, the Federal Office of Public Health, the Federal Office of Roads and Transport, the canton's government 103

of Aargau, Basel-Stadt, Basel-Land, Geneva, Luzern, Ticino, Valais, and Zürich, the Swiss Lung League, the canton's Lung League of Basel Stadt/ Basel Landschaft, Geneva, Ticino, Valais, Graubünden and Zurich, Stiftung ehemals Bündner Heilstätten, SUVA, Freiwillige Akademische Gesellschaft, UBS Wealth Foundation, Talecris Biotherapeutics GmbH, Abbott Diagnostics, European Commission 018996 (GABRIEL), Wellcome Trust WT 084703MA.

SHIP

Investigators: Sven Gläser, Beate Koch, Alexander Teumer, Henry Völzke. **Contributions**: project conception, design and management: S.G., B.K., H.V., phenotype collection and data management: S.G., B.K., A.T., H.V., data analysis: A.T. **Funding & Acknowledgements**: SHIP is part of the Community Medicine Research net of the University of Greifswald, Germany, which is funded by the Federal Ministry of Education and Research, the Ministry of Cultural Affairs as well as the Social Ministry of the Federal State of Mecklenburg-West Pomerania, and the network 'Greifswald Approach to Individualized Medicine (GANI_MED)' funded by the Federal Ministry of Education and Research, and the German Asthma and COPD Network (COSYCONET) (grant no.01ZZ9603, 01ZZ0103, 01ZZ0403, 03IS2061A, BMBF 01GI0883). Genome-wide data have been supported by the Federal Ministry of Education and Research and a joint grant from Siemens Healthcare, Erlangen, Germany and the Federal State of Mecklenburg- West Pomerania (grant no. 03ZIK012). The University of Greifswald is a member of the 'Center of Knowledge Interchange' program of the Siemens AG and the Caché Campus program of the InterSystems GmbH.

YFS

Investigators: Nina Hutri-Kähönen, Terho Lehtimäki, Leo-Pekka Lyytikäinen, Olli T Raitakari. **Contributions**: project conception, design and management: T.L., O.T.R., phenotype collection and data management: N.H-K., T.L., O.T.R., data analysis: L-P.L. **Funding & Acknowledgements**: The authors acknowledge Academy of Finland (126925, 121584, 124282), Academy of Finland (Eye) (134309), Academy of Finland (Salve) (129378), Academy of Finland (Gendi) (117787), Academy of Finland (Skidi) (41071), Social Insurance Institution of Finland, Tampere University Hospital Medical Funds (X51001 for T.L.), Kuopio University Hospital Medical Funds, Turku University Hospital Medical Funds, Juho Vainio Foundation, Paavo Nurmi Foundation, Finnish Foundation of Cardiovascular Research (T.L.), Finnish Cultural Foundation, Tuberculosis Foundation (T.L.), Emil Aaltonen Foundation (T.L.), and Yrjö Jahnsson Foundation (T.L.).

Follow-up studies

ALSPAC

Investigators: David M Evans, Raquel Granell, John Henderson, John P Kemp.

Contributions: phenotype collection and data management: R.G., J. Henderson, J.P.K., data analysis: D.M.E. **Funding & Acknowledgements**: We are extremely grateful to all the families who took part in the study, the midwives for their help in recruiting them, and the whole ALSPAC team, which includes interviewers, computer and laboratory technicians, clerical workers, research scientists, volunteers, managers, receptionists and nurses. The UK Medical Research Council and the Wellcome Trust (Grant ref: 092731) and the University of Bristol provide core support for ALSPAC. This publication is the work of the authors and D.M.E will serve as guarantor for the contents of this paper.

UK BILEVE

Funding & Acknowledgements: This work was funded by a Medical Research Council (MRC) strategic award to MDT, IPH, DS and LVW (MC_PC_12010). This research has been conducted using the UK Biobank Resource. MDT has been supported by MRC fellowships G0501942 and G0902313. IPH is supported by an MRC programme grant (G1000861). Jonathan Marchini is funded by an ERC Consolidator Grant (617306). This article presents independent research funded partially by the National Institute for Health Research (NIHR). The views expressed are those of the author(s) and not necessarily those of the NHS, the NIHR or the Department of Health. We would like to acknowledge all members of the UK Biobank Array Design Group: Peter Donnelly (chair) (University of Oxford), Jeff Barrett (Wellcome Trust Sanger Institute), Jose Bras (University College London), Adam Butterworth (University of Cambridge), Richard Durbin (Wellcome Trust Sanger Institute), Paul Elliott (Imperial College London), Ian Hall (University of Nottingham), John Hardy (University College London), Mark McCarthy (University of Oxford), Gil McVean (University of Oxford), Tim Peakman (UK Biobank), Nazneen Rahman (The Institute of Cancer Research), Nilesh Samani (University of Leicester), Martin Tobin (University of Leicester), Hugh Watkins (University of Oxford).

ECRHS

Investigators: Claudia Flexeder, Juan R González, Joachim Heinrich, Deborah L Jarvis. **Contributions**: project conception, design and management: J. Heinrich, D.L.J., phenotype collection and data management: J.R.G., J. Heinrich, D.L.J., data analysis: C.F. **Funding & Acknowledgements**: Authors acknowledge EU funding (GABRIEL GRANT Number: 018996, ECRHS II Coordination Number: QLK4-CT-1999-01237).

PIVUS

Investigators: Erik Ingelsson, Lars Lind, Anubha Mahajan, Andrew P Morris. **Contributions**: project conception, design and management: L.L., phenotype collection and data management: E.I., L.L., A.M., A.P.M., data analysis: A.M., A.P.M. **Funding & Acknowledgements**: A.P.M. acknowledges the Wellcome Trust (WT098017,WT064890, WT090532). The PIVUS study acknowledges The Swedish Foundation for Strategic Research (ICA08-0047), The Swedish 105 Research Council (2012-1397), The Swedish Heart-Lung Foundation (20120197), The Swedish Society of Medicine and Uppsala University. The computations were performed on resources provided by SNIC through Uppsala Multidisciplinary Center for Advanced Computational Science (UPPMAX) under Project p2013056. APM is a Wellcome Trust Senior Research Fellow in Basic Biomedical Science (grant number WT098017).

TwinsUK

Investigators: Chris J Hammond, Pirro G Hysi, Tim D Spector, Ana Viñuela. **Contributions**: project conception, design and management: T.D.S., phenotype collection and data management: C.J.H., P.G.H., T.D.S., A.V., data analysis: P.G.H. **Funding & Acknowledgements**: This cohort received funding from the Wellcome Trust; the European Community's Seventh Framework Programme (FP7 /2007-13); US National Institutes of Health/National Eye Institute (1RO1EY018246); NIH Center for Inherited Disease Research; the National Institute for Health Research (NIHR)- funded BioResource, Clinical Research Facility and Biomedical Research Centre based at Guy's and St Thomas' NHS Foundation Trust in partnership with King's College London.

Lung eQTL study

Investigators: Yohan Bossé, Corry-Anke Brandsma, David C Nickle, Ma'en Obeidat, Don D Sin. **Contributions**: phenotype collection and data management: Y.B., C-A.B., D.C.N., D.D.S., data analysis: M.O. **Funding & Acknowledgements**: The authors would like to thank the research staff at the Respiratory Health Network Tissue Bank of the FRQS for collecting lung specimens for the lung eQTL study at Laval University. Lung tissue eQTL study was funded by Merck Research Laboratories. M.O. is a Postdoctoral Fellow of the Michael Smith Foundation for Health Research (MSFHR) and the Canadian Institute for Health Research (CIHR) Integrated and Mentored Pulmonary and Cardiovascular Training program (IMPACT). Y.B. is the recipient of a Junior 2 Research Scholar award from the Fonds de recherche Québec – Santé (FRQS).

Supplementary Methods:

Lung eQTL analyses

Variants in 2Mb regions (new sentinel variant +/- 1Mb) which were in LD ($r^2 > 0.3$) with the sentinel variant, for each of the 16 new sentinel variants, were assessed in a lung eQTL resource based on lung tissues of 1,111 individuals. Variants were selected if they (i) met an FDR threshold of 10%, and (ii) had eQTL *P* value < 5 x 10⁻⁵. In addition, for each probeset we ranked variants according to their correlation with the sentinel SNP (as measured by r^2) first and to their eQTL *P* value second, and we only selected results for the top ranked variant for each probeset.

The descriptions of the lung eQTL dataset and subject demographics have been published previously⁷⁻⁹. Briefly, non-tumor lung tissues were collected from patients who underwent lung resection surgery at three participating sites: Laval University (Quebec City, Canada), University of Groningen (Groningen, The Netherlands), and University of British Columbia (Vancouver, Canada). Whole-genome gene expression and genotyping data were obtained from these specimens. Gene expression profiling was performed using an Affymetrix custom array (GPL10379) testing 51,627 noncontrol probe sets and normalized using RMA¹⁰⁶. Genotyping was performed using the Illumina Human1M-Duo BeadChip array. Genotype imputation was undertaken using the 1000 Genomes Project¹⁰⁷ reference panel. Following standard microarray and genotyping quality controls, 1111 patients were available including 409 from Laval, 363 from Groningen, and 339 from UBC. Association testing for each variant with mRNA expression in either *cis* (within 1 Mb of transcript start site) or in *trans* (all other eQTLs) was undertaken separately in each study, then their results were meta-analysed using inverse variance weighting meta-analysis and a 10% FDR genome-wide was applied. For the X chromosome only HapMap imputed data was available and the association testing was undertaken separately for males and females in each study.

Blood eQTL analyses

Variants in 2Mb regions (new sentinel variant +/- 1Mb) which were in LD ($r^2 > 0.3$) with the sentinel variant, for each of the 16 new sentinel variants, and were in HapMap were assessed in a publicly available blood eQTL dataset⁶ with results from the analysis of 5,311 individuals. Association testing was undertaken both for *cis* (distance between the SNP and the probe midpoint < 250kb) and *trans* (distance between the SNP and the probe midpoint < 250kb) and *trans* (distance between the SNP and the probe midpoint >5Mb) eQTL signals. No X chromosome data was available. Variants were selected if they (i) met an FDR threshold of 10%, and (ii) had eQTL *P* value < 5 x 10⁻⁵. In addition, for each probeset we ranked variants according to their correlation with the sentinel SNP (as measured by r^2) first and to their eQTL *P* value second, and we only selected results for the top ranked variant for each probeset.

DNase hypersensitivity analysis

DNase hypersensitivity sites as defined by ENCODE¹⁰⁸ within 2Mb regions centred in the sentinel variant for the 16 new sentinel variants were accessed using the Uniform DNasel HS track in the UCSC table browser¹⁰⁹ in the following cell types: A549, epithelial cell line derived from a lung carcinoma tissue; IMR90, fetal lung fibroblasts; AG04450, fetal lung fibroblast; BC_Lung_01-11002, lung, donor 01-11002, age 83, caucasian, DNA and RNA extract; BC_Lung_H12817N, lung, donor H12817N, age 71, caucasian, DNA and RNA extra; FibroP_AG08396, lung fibroblasts taken from individuals with Parkinson's disease; HMVEC-LBI, blood microvascular endothelial cells, lung-derived; HMVEC-LLy, lymphatic microvascular endothelial cells, lung-derived; HPF, pulmonary fibroblasts isolated from lung tissue; HPAEC, pulmonary artery endothelial cells; HPAEpiC, pulmonary alveolar epithelial cells; HPAF, pulmonary artery fibroblasts; Lung_BC, lung, parental cell type to donors 01-11002 and H12817N; Lung_OC, primary frozen lung tissue from NICHD donor IDs 1104 (Rep B1
DNase), 602 (Rep B2 DNase), 1442 (Rep B3 DNase) and CF Center donor IDs DD006I (Rep B1 FAIRE) and DD007I (Rep B2 FAIRE); NHLF, lung fibroblasts; TBEC, trachea and bronchial epithelial cells from frush lung tissue provided by National Disease Research Interchange, passage 2 primary, non-immortalized cells, donor was 21 years old; WI-38, embryonic lung fibroblast cells, hTERT immortalized, includes Raf1 construct. More details on cell type definitions can be found here <u>http://genome.ucsc.edu/ENCODE/cellTypes.html</u>. Sites for any cell which contained at least one variant strongly correlated ($r^2 > 0.8$) with any of the new 16 sentinel variants were selected.

Conditional analysis: functional annotation

In order to identify suggestive functional variants responsible for the new signals we observed or already known signals from previous GWAS studies^{13,15-17}, 2Mb windows (1Mb either side of the sentinel SNP for the novel and known signals) were searched for functional variants. Variants were annotated using ENSEMBL's Variant Effect Predictor¹⁶ and functional effects were predicted with SIFT¹⁷, PolyPhen-2¹⁸, CADD¹⁹, and GWAVA¹¹⁰ databases. If a variant was annotated as 'deleterious' by SIFT, 'probably damaging' or 'potentially damaging' by PolyPhen-2, had a CADD scaled score ≥ 20 (CADD PHRED \geq 20), or had a GWAVA score > 0.5, it was treated as a functional variant. CADD (Combined Annotation-Dependent Depletion) is a method for integrating many diverse annotations, namely conservation metrics, functional genomic data, transcript information, and protein level scores into a single score for each coding and noncoding variant. Scaled CADD score (CADD_PHRED) ranks each variant relative to all possible substitutions of the human genome (~8.6 billion SNVs of the GRCh37/hg19 reference genome). A scaled CADD score greater or equal to 20 for a variant, indicates that the variant is within the 1% most deleterious variants in the human genome ¹⁹. GWAVA (genome-wide annotation of variants) is a tool that combines information from a wide range of annotations to predict the functional impact of noncoding variants. We used a GWAVA score

threshold of 0.5, as proposed by authors, above which noncoding variants were considered as 'deleterious' ¹¹⁰. Annotation results were filtered with VEP's --pick flag, which selects only one consequence per variant based on the canonical biotype status and length of the transcript as well as the ranking of the consequence type. For variants with multiple annotations, we selected the most deleterious annotation (i.e. if a variant was annotated as frameshift variant and intronic variant, the variant was considered to be frameshift). For each locus, functional variants that had a stage 1 *P* value < 5×10^{-4} or were in LD with the sentinel SNP ($r^2 > 0.3$) were selected. Each functional variant was then tested jointly with the sentinel variant in the region, using GCTA²⁰ and data from 5,000 individuals from UK BiLEVE to estimate LD, to see whether the functional variant would explain for the signal. We considered that a signal was explained by a functional variant if the joint *P* value for the sentinel variant when fitted with the functional variant was > 0.01.

Selection of priority genes

For each of the 16 loci that comprised the novel signals a small number of genes were selected as priority genes for subsequent analyses. These genes were selected according to the following criteria, based on eQTL results from blood and lung tissue for the sentinel variant in each region and for variants in LD ($r^2>0.3$) with the sentinel (**Supplementary Table 3 a**), and the results of the conditional analysis on functional variants (**Supplementary Table 3 f**). eQTL results were restricted to variants that met a 10% FDR threshold and had eQTL *P*<5x10⁻⁵. For each probeset, variants were ranked first according to r^2 with the sentinel variant in the region, and second by the eQTL *P* value. Only the highest ranking variant was kept for each probeset. If the association of a sentinel variant was explained by a missense variant according to the conditional analysis, the gene where the missense variant lies was selected (this was the case for three regions: *GPR126, RIN3* and *LTBP4*). If no missense variant explained the association and no eQTL results met the FDR and *P* value

thresholds for a region, the gene closest to the sentinel variant was chosen (this was the case for three regions: *KCNS3*, *TBX3*, and *MN1*). If there was only evidence from blood eQTL up to two of the most significant genes that met the FDR and *P* value thresholds were selected (this was the case for one region: *NPNT*). In the *NPNT* region, *NPNT* was added to the list of priority genes as well as the genes implicated by the eQTL analysis, given the prior evidence of association with lung function for *NPNT*². If there was evidence from lung tissue eQTL up to two of the most significant genes that met the FDR and *P* value thresholds.

Messenger RNA sequencing

Passage 3 normal human bronchial epithelial cells (NHBECs) (Lonza, UK), were cultured in growth factor-supplemented medium (BEGM, Lonza as described previously¹¹¹). Cells were grown under these conditions and four different experimental conditions as part of a related RNA interference (RNAi) project each in three independent biological replicates (12 samples in total). Total RNA was extracted using established methods for RNA isolation (Sigma-Aldrich GenElute Mammalian Total RNA Miniprep Kit) and RNA quality was assessed for degradation on Agilent 2100 Bioanalyzer with all twelve samples having a RNA Integrity Number (RIN) at ~8 or above 8. The sequencing library was prepared with Illumina TruSeq RNA Sample Prep Kit v2. mRNA was poly-A selected by capturing total RNA samples with oligo-dT coated magnetic beads. The mRNA was then fragmented and randomly primed. cDNA was synthesised using random primers. Finally ready-for-sequencing library was prepared by end-repair, phosphorylation, A-tailing, adapter ligation and PCR amplification. Paired-end sequencing was performed on the Illumina HiSeq2000 platform using TruSeq v3

chemistry over 100 cycles yielding approximately 40 million reads per sample. The generated raw reads FastQ files (100 base pairs; Sanger / Illumina 1.9 encoding) were quality evaluated using FastQC. Mean quality scores across the bases for all reads in all twelve samples were above 28. Unmodified reads were used for subsequent analysis on Ubuntu 12.04 LTS operating system. Unspliced alignments onto human genome build GRCh37 were performed for each sample individually using Bowtie2 tool utilized by TopHat v2.0.12¹¹². Reads aligning to more than 20 positions were discarded. The subset of reads that were not aligned uniquely were used by TopHat¹¹² to identify splice junctions. Cufflinks v2.2.1 programme ^{113,114} was used to assemble transcriptome for each individual sample. Transcriptomes from all the samples were merged using Cuffmerge v1.0.0 feature ^{113,114} in order to identify low-expression transcripts requiring deep sequencing coverage. The Cuffmerge ^{113,114} generated novel gene transfer format (GTF) annotation file was compared to Ensembl GTF annotation of GRCh37 genome build by using Cuffcompare v2.2.1 ^{113,114}. All 12 NHBEC samples were used to identify reported and novel transcripts. The three untreated basal NHBEC samples were used to determine abundance under basal culture conditions, mean FPKM expressions of individual isoforms were calculated and their percentage of total transcripts expression determined. Splicing graphs depicting novel and known splice transcripts were generated using SpliceGrapher v0.2.4¹¹⁵.

Fetal differential expression

Publically available Affymetrix U133 Plus 2 array data (Gene expression omnibus: GSE14334) of 38 fetal lung samples spanning the Pseudoglandular (7–16 weeks) and Canalicular (17–22 weeks) stages of lung development was mined as previously reported¹³. *P* values were adjusted for multiple testing using the Benjamini & Hochberg method¹⁴. Probes for each of a subset of genes in the 16 loci

(see criteria for selection in the "Selection of priority genes" section in the **Supplementary Methods**) were searched for within the results.

COPD differential expression

Publically available microarray data (GSE37147¹⁵) was mined using GEO2R on the gene expression omnibus website (<u>http://www.ncbi.nlm.nih.gov/geo/info/geo2r.html</u>). Two sample groups were defined. Affymetrix Human ST1.0 array expression data, which includes one probe per gene, for 87 bronchial brushings in the lungs of individuals with COPD was defined as the first group, whilst the second group had the expression profiles of 151 bronchial brushings from ever smoker individuals without COPD. Differential expression between the 2 groups was identified using the default array statistics. *P* values were adjusted for multiple testing using the Benjamini & Hochberg method ¹⁴. Probes for each of a subset of genes in the 16 loci (see criteria for selection in the "Selection of priority genes" section in the **Supplementary Methods**) were searched for within the results.

MAGENTA

Stage 1 GWAS results were enriched for known biological pathways using MAGENTA v2¹¹⁶. MAGENTA defines a *P* value for each gene that is the lowest *P* value within 110kb upstream and 40kb downstream of the gene and is corrected for gene size, number of SNPs per gene and linkage disequilibrium within the region. For each gene set, the null hypothesis that there is a random distribution of gene association score ranks within the gene set is tested against the alternative hypothesis that there are more gene association score ranks above a given rank cut-off (75th percentile cut-off is recommended for polygenic traits) compared to random sampling of 10,000 gene sets of identical size. For each gene set, a false discovery rate (FDR) is calculated as the fraction of 113 all randomly sampled gene sets (10,000 x number of gene sets tested) that have more genes with P value below the cut off (75th percentile) than in the gene set being tested, divided by the fraction of real gene sets that have more genes with P value below the cut off (75th percentile) than in the gene set being tested.

Six databases of biological pathways were tested: including Ingenuity Pathway (June 2008, number of pathways n=81), KEGG (2010, n=186), PANTHER Molecular Function (January 2010, n=216), PANTHER Biological Processes (January 2010, n=217), PANTHER Pathways (January 2010, n=94) and Gene Ontology (April 2010, n=1778). Significance thresholds were Bonferroni corrected for each database.

Genes within 500kb either side from the sentinel variants were flagged in the analysis. Sensitivity analyses were run removing genes in the HLA region on chromosome 6.

Supplementary References:

- 1. Wilk, J.B. *et al.* A genome-wide association study of pulmonary function measures in the Framingham Heart Study. *PLoS Genet* 5, e1000429 (2009).
- 2. Hancock, D.B. *et al.* Meta-analyses of genome-wide association studies identify multiple loci associated with pulmonary function. *Nat Genet* 42, 45-52 (2010).
- 3. Soler Artigas, M. *et al.* Genome-wide association and large-scale follow up identifies 16 new loci influencing lung function. *Nature Genetics* 43, 1082-1090 (2011).
- 4. Loth, D.W. *et al.* Genome-wide association analysis identifies six new loci associated with forced vital capacity. *Nat Genet* 46, 669-77 (2014).
- 5. Repapi, E. *et al.* Genome-wide association study identifies five loci associated with lung function. *Nat Genet* 42, 36-44 (2010).
- 6. Westra, H.J. *et al.* Systematic identification of trans eQTLs as putative drivers of known disease associations. *Nat Genet* 45, 1238-43 (2013).
- 7. Hao, K. *et al.* Lung eQTLs to help reveal the molecular underpinnings of asthma. *PLoS Genet* 8, e1003029 (2012).
- 8. Lamontagne, M. *et al.* Refining susceptibility loci of chronic obstructive pulmonary disease with lung eqtls. *PLoS One* 8, e70220 (2013).
- 9. Obeidat, M.e. *et al.* GSTCD and INTS12 Regulation and Expression in the Human Lung. *Plos One* 8(2013).
- 10. Encode Project Consortium. An integrated encyclopedia of DNA elements in the human genome. *Nature* 489, 57-74 (2012).
- 11. Thurman, R.E. *et al.* The accessible chromatin landscape of the human genome. *Nature* 489, 75-82 (2012).
- 12. Uhlen, M. *et al.* Towards a knowledge-based Human Protein Atlas. *Nat Biotechnol* 28, 1248-50 (2010).
- 13. Melen, E. *et al.* Expression analysis of asthma candidate genes during human and murine lung development. *Respir Res* 12, 86 (2011).
- 14. Benjamini, Y., Hochberg, Y. Controlling the false discovery rate: a practical and powerful approach to multiple testing. *J R Stat Soc B* 57, 289-300 (1995).
- 15. Steiling, K. *et al.* A dynamic bronchial airway gene expression signature of chronic obstructive pulmonary disease and lung function impairment. *Am J Respir Crit Care Med* 187, 933-42 (2013).
- 16. McLaren, W. *et al.* Deriving the consequences of genomic variants with the Ensembl API and SNP Effect Predictor. *Bioinformatics* 26, 2069-70 (2010).
- 17. Kumar, P., Henikoff, S. & Ng, P.C. Predicting the effects of coding non-synonymous variants on protein function using the SIFT algorithm. *Nat Protoc* 4, 1073-81 (2009).
- 18. Adzhubei, I.A. *et al.* A method and server for predicting damaging missense mutations. *Nat Methods* 7, 248-9 (2010).
- 19. Kircher, M. *et al.* A general framework for estimating the relative pathogenicity of human genetic variants. *Nat Genet* 46, 310-5 (2014).
- 20. Yang, J. *et al.* Conditional and joint multiple-SNP analysis of GWAS summary statistics identifies additional variants influencing complex traits. *Nat Genet* 44, 369-75, S1-3 (2012).
- 21. Ahram, D. *et al.* A homozygous mutation in ADAMTSL4 causes autosomal-recessive isolated ectopia lentis. *Am J Hum Genet* 84, 274-8 (2009).
- 22. Groenman, F., Rutter, M., Caniggia, I., Tibboel, D. & Post, M. Hypoxia-inducible factors in the first trimester human lung. *J Histochem Cytochem* 55, 355-63 (2007).
- 23. Otsubo, K. *et al.* ETV6-ARNT fusion in a patient with childhood T lymphoblastic leukemia. *Cancer Genet Cytogenet* 202, 22-6 (2010).

- 24. Laviad, E.L. *et al.* Characterization of ceramide synthase 2: tissue distribution, substrate specificity, and inhibition by sphingosine 1-phosphate. *J Biol Chem* 283, 5677-84 (2008).
- 25. Erez-Roman, R., Pienik, R. & Futerman, A.H. Increased ceramide synthase 2 and 6 mRNA levels in breast cancer tissues and correlation with sphingosine kinase expression. *Biochem Biophys Res Commun* 391, 219-23 (2010).
- 26. Saftig, P. *et al.* Impaired osteoclastic bone resorption leads to osteopetrosis in cathepsin-Kdeficient mice. *Proc Natl Acad Sci U S A* 95, 13453-8 (1998).
- 27. Gelb, B.D., Shi, G.P., Chapman, H.A. & Desnick, R.J. Pycnodysostosis, a lysosomal disease caused by cathepsin K deficiency. *Science* 273, 1236-8 (1996).
- 28. Costantino, C.M., Ploegh, H.L. & Hafler, D.A. Cathepsin S regulates class II MHC processing in human CD4+ HLA-DR+ T cells. *J Immunol* 183, 945-52 (2009).
- 29. Reiser, J., Adair, B. & Reinheckel, T. Specialized roles for cysteine cathepsins in health and disease. *J Clin Invest* 120, 3421-31 (2010).
- 30. Heron, L., Virsolvy, A., Apiou, F., Le Cam, A. & Bataille, D. Isolation, characterization, and chromosomal localization of the human ENSA gene that encodes alpha-endosulfine, a regulator of beta-cell K(ATP) channels. *Diabetes* 48, 1873-6 (1999).
- 31. Ng, M.M., Dippold, H.C., Buschman, M.D., Noakes, C.J. & Field, S.J. GOLPH3L antagonizes GOLPH3 to determine Golgi morphology. *Mol Biol Cell* 24, 796-808 (2013).
- 32. Kunigou, O. *et al.* Role of GOLPH3 and GOLPH3L in the proliferation of human rhabdomyosarcoma. *Oncol Rep* 26, 1337-42 (2011).
- 33. Chen, Y.T. *et al.* Identification of CT46/HORMAD1, an immunogenic cancer/testis antigen encoding a putative meiosis-related protein. *Cancer Immun* 5, 9 (2005).
- 34. Zhan, Q., Bieszczad, C.K., Bae, I., Fornace, A.J., Jr. & Craig, R.W. Induction of BCL2 family member MCL1 as an early response to DNA damage. *Oncogene* 14, 1031-9 (1997).
- 35. Okita, H. *et al.* Acute myeloid leukemia possessing jumping translocation is related to highly elevated levels of EAT/mcl-1, a Bcl-2 related gene with anti-apoptotic functions. *Leuk Res* 24, 73-7 (2000).
- 36. Ryu, H. *et al.* ESET/SETDB1 gene expression and histone H3 (K9) trimethylation in Huntington's disease. *Proc Natl Acad Sci U S A* 103, 19176-81 (2006).
- 37. Petrache, I. *et al.* Ceramide synthases expression and role of ceramide synthase-2 in the lung: insight from human lung cells and mouse models. *PLoS One* 8, e62968 (2013).
- 38. Buhling, F. et al. Pivotal role of cathepsin K in lung fibrosis. Am J Pathol 164, 2203-16 (2004).
- 39. Quadri, M. *et al.* Mutations in SLC30A10 cause parkinsonism and dystonia with hypermanganesemia, polycythemia, and chronic liver disease. *Am J Hum Genet* 90, 467-77 (2012).
- 40. Hao, K., Niu, T., Xu, X. & Fang, Z. Single-nucleotide polymorphisms of the KCNS3 gene are significantly associated with airway hyperresponsiveness. *Hum Genet* 116, 378-83 (2005).
- 41. Nyholt, D.R. *et al.* A high-density association screen of 155 ion transport genes for involvement with common migraine. *Hum Mol Genet* 17, 3318-31 (2008).
- 42. Wintermeyer, W. *et al.* Mechanisms of elongation on the ribosome: dynamics of a macromolecular machine. *Biochem Soc Trans* 32, 733-7 (2004).
- 43. Balasubramaniam, S. *et al.* Infantile Progressive Hepatoencephalomyopathy with Combined OXPHOS Deficiency due to Mutations in the Mitochondrial Translation Elongation Factor Gene GFM1. *JIMD Rep* 5, 113-22 (2012).
- 44. Winteringham, L.N., Kobelke, S., Williams, J.H., Ingley, E. & Klinken, S.P. Myeloid Leukemia Factor 1 inhibits erythropoietin-induced differentiation, cell cycle exit and p27Kip1 accumulation. *Oncogene* 23, 5105-9 (2004).
- 45. Yoneda-Kato, N. *et al.* The t(3;5)(q25.1;q34) of myelodysplastic syndrome and acute myeloid leukemia produces a novel fusion gene, NPM-MLF1. *Oncogene* 12, 265-75 (1996).

- 46. Winteringham, L.N. *et al.* Myeloid leukemia factor 1 associates with a novel heterogeneous nuclear ribonucleoprotein U-like molecule. *J Biol Chem* 281, 38791-800 (2006).
- 47. Sahab, Z.J. *et al.* Tumor suppressor RARRES1 interacts with cytoplasmic carboxypeptidase AGBL2 to regulate the alpha-tubulin tyrosination cycle. *Cancer Res* 71, 1219-28 (2011).
- 48. Ellinger, J. *et al.* Hypermethylation of cell-free serum DNA indicates worse outcome in patients with bladder cancer. *J Urol* 179, 346-52 (2008).
- 49. Berndt, S.I. *et al.* Genome-wide meta-analysis identifies 11 new loci for anthropometric traits and provides insights into genetic architecture. *Nat Genet* 45, 501-12 (2013).
- 50. Potkin, S.G. *et al.* Gene discovery through imaging genetics: identification of two novel genes associated with schizophrenia. *Mol Psychiatry* 14, 416-28 (2009).
- 51. Board, P.G. & Menon, D. Glutathione transferases, regulators of cellular metabolism and physiology. *Biochim Biophys Acta* 1830, 3267-88 (2013).
- 52. Hayes, J.D. & Strange, R.C. Glutathione S-transferase polymorphisms and their biological consequences. *Pharmacology* 61, 154-66 (2000).
- 53. Baillat, D. *et al.* Integrator, a multiprotein mediator of small nuclear RNA processing, associates with the C-terminal repeat of RNA polymerase II. *Cell* 123, 265-76 (2005).
- 54. Huang, J.T. & Lee, V. Identification and characterization of a novel human nephronectin gene in silico. *Int J Mol Med* 15, 719-24 (2005).
- 55. Inagaki, F.F. *et al.* Nephronectin is upregulated in acute and chronic hepatitis and aggravates liver injury by recruiting CD4 positive cells. *Biochem Biophys Res Commun* 430, 751-6 (2013).
- 56. Soranzo, N. *et al.* Meta-analysis of genome-wide scans for human adult stature identifies novel Loci and associations with measures of skeletal frame size. *PLoS Genet* 5, e1000445 (2009).
- 57. Lango Allen, H. *et al.* Hundreds of variants clustered in genomic loci and biological pathways affect human height. *Nature* 467, 832-8 (2010).
- 58. Lettre, G. *et al.* Identification of ten loci associated with height highlights new biological pathways in human growth. *Nat Genet* 40, 584-91 (2008).
- 59. van der Valk, R.J. *et al.* A novel common variant in DCST2 is associated with length in early life and height in adulthood. *Hum Mol Genet* (2014).
- 60. Kou, I. *et al.* Genetic variants in GPR126 are associated with adolescent idiopathic scoliosis. *Nat Genet* 45, 676-9 (2013).
- 61. Mogha, A. *et al.* Gpr126 functions in Schwann cells to control differentiation and myelination via G-protein activation. *J Neurosci* 33, 17976-85 (2013).
- 62. Moriguchi, T. *et al.* DREG, a developmentally regulated G protein-coupled receptor containing two conserved proteolytic cleavage sites. *Genes Cells* 9, 549-60 (2004).
- 63. Waller-Evans, H. *et al.* The orphan adhesion-GPCR GPR126 is required for embryonic development in the mouse. *PLoS One* 5, e14047 (2010).
- 64. Freilinger, T. *et al.* Genome-wide association analysis identifies susceptibility loci for migraine without aura. *Nat Genet* 44, 777-82 (2012).
- 65. Frosk, P. *et al.* Limb-girdle muscular dystrophy type 2H associated with mutation in TRIM32, a putative E3-ubiquitin-ligase gene. *Am J Hum Genet* 70, 663-72 (2002).
- 66. Netchine, I. *et al.* Mutations in LHX3 result in a new syndrome revealed by combined pituitary hormone deficiency. *Nat Genet* 25, 182-6 (2000).
- 67. Wittke, I. *et al.* Neuroblastoma-derived sulfhydryl oxidase, a new member of the sulfhydryl oxidase/Quiescin6 family, regulates sensitization to interferon gamma-induced cell death in human neuroblastoma cells. *Cancer Res* 63, 7742-52 (2003).
- 68. Sprooten, E. *et al.* Common genetic variants and gene expression associated with white matter microstructure in the human brain. *Neuroimage* 97, 252-61 (2014).
- 69. Bamshad, M. *et al.* Mutations in human TBX3 alter limb, apocrine and genital development in ulnar-mammary syndrome. *Nat Genet* 16, 311-5 (1997).

- 70. Bogarapu, S. *et al.* Phenotype of a patient with contiguous deletion of TBX5 and TBX3: expanding the disease spectrum. *Am J Med Genet A* 164A, 1304-9 (2014).
- 71. Li, J. *et al.* The Anti-proliferative Function of the TGF-beta1 Signalling Pathway Involves the Repression of the Oncogenic TBX2 by its Homologue TBX3. *J Biol Chem* (2014).
- 72. Li, J., Weinberg, M.S., Zerbini, L. & Prince, S. The oncogenic TBX3 is a downstream target and mediator of the TGF-beta1 signaling pathway. *Mol Biol Cell* 24, 3569-76 (2013).
- 73. Kawaguchi, Y. *et al.* CAG expansions in a novel gene for Machado-Joseph disease at chromosome 14q32.1. *Nat Genet* 8, 221-8 (1994).
- 74. Albig, A.R. & Schiemann, W.P. Fibulin-5 antagonizes vascular endothelial growth factor (VEGF) signaling and angiogenic sprouting by endothelial cells. *DNA Cell Biol* 23, 367-79 (2004).
- 75. Loeys, B. *et al.* Homozygosity for a missense mutation in fibulin-5 (FBLN5) results in a severe form of cutis laxa. *Hum Mol Genet* 11, 2113-8 (2002).
- 76. Brandsma, C.A. *et al.* A large lung gene expression study identifying fibulin-5 as a novel player in tissue repair in COPD. *Thorax* 70, 21-32 (2015).
- 77. Dabovic, B. *et al.* Function of Latent TGFbeta Binding Protein 4 and Fibulin 5 in Elastogenesis and Lung Development. *J Cell Physiol* 230, 226-36 (2015).
- 78. Follit, J.A. *et al.* The Golgin GMAP210/TRIP11 anchors IFT20 to the Golgi complex. *PLoS Genet* 4, e1000315 (2008).
- 79. Smits, P. *et al.* Lethal skeletal dysplasia in mice and humans lacking the golgin GMAP-210. *N Engl J Med* 362, 206-16 (2010).
- 80. Yanagisawa, H. *et al.* Fibulin-5 is an elastin-binding protein essential for elastic fibre development in vivo. *Nature* 415, 168-71 (2002).
- 81. Nakamura, T. *et al.* Fibulin-5/DANCE is essential for elastogenesis in vivo. *Nature* 415, 171-5 (2002).
- 82. Kajiho, H. *et al.* RIN3: a novel Rab5 GEF interacting with amphiphysin II involved in the early endocytic pathway. *J Cell Sci* 116, 4159-68 (2003).
- 83. Albagha, O.M. *et al.* Genome-wide association identifies three new susceptibility loci for Paget's disease of bone. *Nat Genet* 43, 685-9 (2011).
- 84. Cho, M.H. *et al.* Risk loci for chronic obstructive pulmonary disease: a genome-wide association study and meta-analysis. *Lancet Respir Med* 2, 214-25 (2014).
- 85. Lambert, J.C. *et al.* Meta-analysis of 74,046 individuals identifies 11 new susceptibility loci for Alzheimer's disease. *Nat Genet* 45, 1452-8 (2013).
- 86. Wadehra, M. *et al.* Expression of epithelial membrane protein-2 is associated with endometrial adenocarcinoma of unfavorable outcome. *Cancer* 107, 90-8 (2006).
- 87. Street, V.A. *et al.* Mapping of Charcot-Marie-Tooth disease type 1C to chromosome 16p identifies a novel locus for demyelinating neuropathies. *Am J Hum Genet* 70, 244-50 (2002).
- 88. Cao, W., Ijiri, T.W., Huang, A.P. & Gerton, G.L. Characterization of a novel tektin member, TEKT5, in mouse sperm. *J Androl* 32, 55-69 (2011).
- 89. Onouchi, Y. *et al.* ITPKC functional polymorphism associated with Kawasaki disease susceptibility and formation of coronary artery aneurysms. *Nat Genet* 40, 35-42 (2008).
- 90. Ryan, D.M. *et al.* Smoking dysregulates the human airway basal cell transcriptome at COPD risk locus 19q13.2. *PLoS One* 9, e88051 (2014).
- 91. Zhong, W., Jiang, M.M., Weinmaster, G., Jan, L.Y. & Jan, Y.N. Differential expression of mammalian Numb, Numblike and Notch1 suggests distinct roles during mouse cortical neurogenesis. *Development* 124, 1887-97 (1997).
- 92. He, H. *et al.* Identification and characterization of nine novel human small GTPases showing variable expressions in liver cancer tissues. *Gene Expr* 10, 231-42 (2002).

- 93. Sterner-Kock, A. *et al.* Disruption of the gene encoding the latent transforming growth factorbeta binding protein 4 (LTBP-4) causes abnormal lung development, cardiomyopathy, and colorectal cancer. *Genes Dev* 16, 2264-73 (2002).
- 94. Buijs, A. *et al.* Translocation (12;22) (p13;q11) in myeloproliferative disorders results in fusion of the ETS-like TEL gene on 12p13 to the MN1 gene on 22q11. *Oncogene* 10, 1511-9 (1995).
- 95. Tarpey, P.S. *et al.* Mutations in the gene encoding the Sigma 2 subunit of the adaptor protein 1 complex, AP1S2, cause X-linked mental retardation. *Am J Hum Genet* 79, 1119-24 (2006).
- 96. Cacciagli, P. *et al.* AP1S2 is mutated in X-linked Dandy-Walker malformation with intellectual disability, basal ganglia disease and seizures (Pettigrew syndrome). *Eur J Hum Genet* 22, 363-8 (2014).
- 97. Saillour, Y. *et al.* Mutations in the AP1S2 gene encoding the sigma 2 subunit of the adaptor protein 1 complex are associated with syndromic X-linked mental retardation with hydrocephalus and calcifications in basal ganglia. *J Med Genet* 44, 739-44 (2007).
- 98. Shan, L. *et al.* Bombesin-like peptide receptor gene expression, regulation, and function in fetal murine lung. *Am J Physiol Lung Cell Mol Physiol* 286, L165-73 (2004).
- 99. Sunday, M.E., Choi, N., Spindel, E.R., Chin, W.W. & Mark, E.J. Gastrin-releasing peptide gene expression in small cell and large cell undifferentiated lung carcinomas. *Hum Pathol* 22, 1030-9 (1991).
- 100. Yoshida, K. *et al.* Frequent pathway mutations of splicing machinery in myelodysplasia. *Nature* 478, 64-9 (2011).
- 101. Hindorff LA, M.J., Morales J , Junkins HA, Hall PN, Klemm AK, and Manolio TA. A. Catalog of Published Genome-Wide Association Studies. Accessed 05-01-2015.
- 102. Golding, J., Pembrey, M., Jones, R. & Team, A.S. ALSPAC--the Avon Longitudinal Study of Parents and Children. I. Study methodology. *Paediatr Perinat Epidemiol* 15, 74-87 (2001).
- 103. Wilk, J.B. *et al.* Evidence for major genes influencing pulmonary function in the NHLBI family heart study. *Genet Epidemiol* 19, 81-94 (2000).
- 104. Lewitter, F.I., Tager, I.B., McGue, M., Tishler, P.V. & Speizer, F.E. Genetic and environmental determinants of level of pulmonary function. *Am J Epidemiol* 120, 518-30 (1984).
- 105. Palmer, L.J. *et al.* Familial aggregation and heritability of adult lung function: results from the Busselton Health Study. *Eur Respir J* 17, 696-702 (2001).
- 106. Irizarry, R.A. *et al.* Exploration, normalization, and summaries of high density oligonucleotide array probe level data. *Biostatistics* 4, 249-64 (2003).
- 107. Abecasis, G.R. *et al.* An integrated map of genetic variation from 1,092 human genomes. *Nature* 491, 56-65 (2012).
- 108. Rosenbloom, K.R. *et al.* ENCODE data in the UCSC Genome Browser: year 5 update. *Nucleic Acids Res* 41, D56-63 (2013).
- 109. Karolchik, D. *et al.* The UCSC Table Browser data retrieval tool. *Nucleic Acids Res* 32, D493-6 (2004).
- 110. Ritchie, G.R., Dunham, I., Zeggini, E. & Flicek, P. Functional annotation of noncoding sequence variants. *Nat Methods* 11, 294-6 (2014).
- 111. Stewart, C.E., Nijmeh, H.S., Brightling, C.E. & Sayers, I. uPAR regulates bronchial epithelial repair in vitro and is elevated in asthmatic epithelium. *Thorax* 67, 477-87 (2012).
- 112. Trapnell, C., Pachter, L. & Salzberg, S.L. TopHat: discovering splice junctions with RNA-Seq. *Bioinformatics* 25, 1105-11 (2009).
- 113. Trapnell, C. *et al.* Transcript assembly and quantification by RNA-Seq reveals unannotated transcripts and isoform switching during cell differentiation. *Nat Biotechnol* 28, 511-5 (2010).
- 114. Roberts, A., Pimentel, H., Trapnell, C. & Pachter, L. Identification of novel transcripts in annotated genomes using RNA-Seq. *Bioinformatics* 27, 2325-9 (2011).

- 115. Rogers, M.F., Thomas, J., Reddy, A.S. & Ben-Hur, A. SpliceGrapher: detecting patterns of alternative splicing from RNA-Seq data in the context of gene models and EST data. *Genome Biol* 13, R4 (2012).
- 116. Segrè, A.V. *et al.* Common inherited variation in mitochondrial genes is not enriched for associations with type 2 diabetes or related glycemic traits. *PLoS Genet* 6(2010).
- 117. Strachan, D.P. *et al.* Lifecourse influences on health among British adults: effects of region of residence in childhood and adulthood. *Int J Epidemiol* 36, 522-31 (2007).
- 118. Marossy, A.E., Strachan, D.P., Rudnicka, A.R. & Anderson, H.R. Childhood chest illness and the rate of decline of adult lung function between ages 35 and 45 years. *Am J Respir Crit Care Med* 175, 355-9 (2007).
- 119. Vitart, V. *et al.* SLC2A9 is a newly identified urate transporter influencing serum urate concentration, urate excretion and gout. *Nat Genet* 40, 437-42 (2008).
- 120. Zemunik, T. *et al.* Genome-wide association study of biochemical traits in Korcula Island, Croatia. *Croat Med J* 50, 23-33 (2009).
- 121. Rudan, I. *et al.* "10001 Dalmatians:" Croatia launches its national biobank. *Croat Med J* 50, 4-6 (2009).
- 122. Day, N. *et al.* EPIC-Norfolk: study design and characteristics of the cohort. European Prospective Investigation of Cancer. *Br J Cancer* 80 Suppl 1, 95-103 (1999).
- 123. Canoy, D. *et al.* Abdominal obesity and respiratory function in men and women in the EPIC-Norfolk Study, United Kingdom. *Am J Epidemiol* 159, 1140-9 (2004).
- 124. Cox, B., Huppert, F. & Whichelow, M. *The Health and Lifestyle Survey: seven years on*, (Dartmouth Publishing Company, Aldershot, United Kingdom, 1993).
- 125. Smith, B.H. *et al.* Cohort Profile: Generation Scotland: Scottish Family Health Study (GS:SFHS). The study, its participants and their potential for genetic research on health and illness. *Int J Epidemiol* 42, 689-700 (2013).
- 126. Heistaro, S. Methodology report. Health 2000 survey. in *Publications of National Public Health Institute* (ed. Heistaro, S.) (2000).
- 127. Kristiansson, K. *et al.* Genome-wide screen for metabolic syndrome susceptibility Loci reveals strong lipid gene contribution but no evidence for common genetic basis for clustering of metabolic syndrome traits. *Circ Cardiovasc Genet* 5, 242-9 (2012).
- 128. Holle, R., Happich, M., Lowel, H., Wichmann, H.E. & Group, M.K.S. KORA--a research platform for population based health research. *Gesundheitswesen* 67 Suppl 1, S19-25 (2005).
- 129. Wichmann, H.E., Gieger, C., Illig, T. & Group, M.K.S. KORA-gen--resource for population genetics, controls and a broad spectrum of disease phenotypes. *Gesundheitswesen* 67 Suppl 1, S26-30 (2005).
- 130. Miller, M.R. et al. Standardisation of spirometry. Eur Respir J 26, 319-38 (2005).
- 131. Peters, A. *et al.* [Multimorbidity and successful aging: the population-based KORA-Age study]. *Z Gerontol Geriatr* 44 Suppl 2, 41-54 (2011).
- 132. Burney, P.G., Luczynska, C., Chinn, S. & Jarvis, D. The European Community Respiratory Health Survey. *Eur Respir J* 7, 954-60 (1994).
- 133. Main Protocol for The European Community Respiratory Health Survey (ECRHS) I, http://www.ecrhs.org/ECRHS%20I/Main%20protocol.pdf.
- 134. Deary, I.J. *et al.* The Lothian Birth Cohort 1936: a study to examine influences on cognitive ageing from age 11 to age 70 and beyond. *BMC Geriatr* 7, 28 (2007).
- 135. Aulchenko, Y.S., Ripke, S., Isaacs, A. & van Duijn, C.M. GenABEL: an R library for genomewide association analysis. *Bioinformatics* 23, 1294-6 (2007).
- Martin, B.W. *et al.* SAPALDIA: methods and participation in the cross-sectional part of the Swiss Study on Air Pollution and Lung Diseases in Adults. *Soz Praventivmed* 42, 67-84 (1997).

- 137. Ackermann-Liebrich, U. *et al.* Follow-up of the Swiss Cohort Study on Air Pollution and Lung Diseases in Adults (SAPALDIA 2) 1991-2003: methods and characterization of participants. *Soz Praventivmed* 50, 245-63 (2005).
- 138. John, U. *et al.* Study of Health In Pomerania (SHIP): a health examination survey in an east German region: objectives and design. *Soz Praventivmed* 46, 186-94 (2001).
- 139. Volzke, H. *et al.* Reference intervals of serum thyroid function tests in a previously iodinedeficient area. *Thyroid* 15, 279-85 (2005).
- 140. Nelson, S.B., Gardner, R.M., Crapo, R.O. & Jensen, R.L. Performance evaluation of contemporary spirometers. *Chest* 97, 288-97 (1990).
- 141. Standardization of spirometry--1987 update. Statement of the American Thoracic Society. *Am Rev Respir Dis* 136, 1285-98 (1987).
- 142. Raitakari, O.T. *et al.* Cohort profile: the cardiovascular risk in Young Finns Study. *Int J Epidemiol* 37, 1220-6 (2008).
- 143. Boyd, A. *et al.* Cohort Profile: the 'children of the 90s'--the index offspring of the Avon Longitudinal Study of Parents and Children. *Int J Epidemiol* 42, 111-27 (2013).
- 144. Knox, J., Jarvis, D., Walter, E.H. & Committee, E.I.S. The European Community Respiratory Health Survey II. *Eur Respir J* 21, 556; author reply 556 (2003).
- 145. Moffatt, M.F. *et al.* A large-scale, consortium-based genomewide association study of asthma. *N Engl J Med* 363, 1211-21 (2010).
- 146. Chinn, S. *et al.* Smoking cessation, lung function, and weight gain: a follow-up study. *Lancet* 365, 1629-35; discussion 1600-1 (2005).
- Lind, L., Fors, N., Hall, J., Marttala, K. & Stenborg, A. A comparison of three different methods to evaluate endothelium-dependent vasodilation in the elderly: the Prospective Investigation of the Vasculature in Uppsala Seniors (PIVUS) study. *Arterioscler Thromb Vasc Biol* 25, 2368-75 (2005).
- 148. Zhou, X. & Stephens, M. Genome-wide efficient mixed-model analysis for association studies. *Nat Genet* 44, 821-4 (2012).
- 149. Wain, L.V., . & et al. Utilising UK Biobank to generate novel insights into the genetics of lung health and disease, and smoking behaviour. *The Lancet Respiratory Medicine* Manuscript in preparation(2015).