



***In vitro* and *in vivo* evaluation of a sustained-release once-a-day formulation of the novel antihypertensive drug MT-1207**

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10 3 **1207**

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7 22 **a-day formulation of the novel antihypertensive drug MT-**
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14 24 **Abstract**

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17 25 Hypertension is one of the most common chronic cardiovascular disorders. Sustained-
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19 26 release formulations are developed to maintain drug therapeutic levels throughout the
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21 27 treatment of hypertension, to promote patient compliance and improve patient outcomes. We
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23 28 have developed and tested in *in vivo* trials a once-a-day tablet formulation for the novel
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25 29 antihypertensive drug MT-1207. The tablets based upon a hydrophilic polymer matrix
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27 30 underwent post-compression parameter and physicochemical characterisations, along with *in*
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29 31 *vitro* drug release testing. The most promising formulation containing 31% w/w HPMC
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31 32 K15M gave a 24-hour release of MT-1207 with an almost constant release rate up to 20
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33 33 hours. Follow in *in vivo* studies were carried out in Beagle dogs for the optimised sustained-
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35 34 release tablets in comparison to immediate-release tablets. The results showed that a
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37 35 sustained release of MT-1207 from the new formulation was achieved with a drug $t_{1/2}$ 2-2.5
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39 36 times longer than the immediate-release tablets. Moreover, the $AUC_{0-\infty}$ values of both
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41 37 sustained- and immediate-release tablets were identical at the same dose of 30 mg, indicating
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43 38 that the same amount of drug was absorbed in each case. For treatments based upon MT-1207
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45 39 this development is significant for future commercial exploitation via scale-up and further
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47 40 trials, and for improved patient outcomes.

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57 42 **Keywords:** MT-1207, Sustained-release tablets, Hydrophilic matrix, Drug release,
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59 43 Pharmacokinetics

1. Introduction

Hypertension is the most common cardiovascular disease, with 1.39 billion adults worldwide having the condition in 2010 (Katzung et al. 2012; Mills et al. 2016). Many variables influence the prevalence of this disease, including race, age and education (Katzung et al. 2012). Hypertension presents a significant challenge in therapeutics as it is usually a lifelong disease that causes few symptoms until its advanced stage. There is a need for improved formulations that can make more effective use of current drugs, as even relatively small improvements in factors such as patient compliance and therapeutic efficacy are important and would have a very significant impact on human health.

There are many drugs that are used to treat hypertension through different mechanisms of action, such as beta-blockers, angiotensin-converting enzyme (ACE) inhibitors, angiotensin II receptor antagonists, alpha-receptor antagonists, diuretics. It is mainly the level of blood pressure and the presence, and severity of, end organ damage that dictate the selection of a drug or drug combination for a patient. The potential use of multiple drugs targeting different receptors leads to the polypharmacy paradigm in the treatment of hypertension (Tsioufis and Thomopoulos 2017; Williams et al. 2018).

Since hypertension is a chronic disorder, a sustained concentration of the drug(s) is required throughout the time of treatment (Nair et al. 2010). Sustained-release formulations are frequently used to achieve this, where a portion of the drug is released immediately and the rest of the amount is released at a predetermined rate for an extended period of time (usually 12 to 18 hours) (Wen and Park 2010). For the extended-release enteric-coated tablets and capsules are the most common forms of such formulations, where the drug release starts in the neutral environment of the small intestine, rather than in the acidic environment of the stomach (Prakash and Markham 1999). In fixed-dose combination formulations, either the same drug is incorporated in both parts of the dosage form, i.e. the immediate-release and

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2
3 69 sustained-release formulation parts or one drug is incorporated into an immediate-release part
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5 70 and a second into a sustained-release part (Alaux et al. 2000; Simon 2005; Wen and Park
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7
8 71 2010).

9
10 72 The use of sustained-release formulations comes with advantages and certain
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12 73 disadvantages. Drug concentration can be maintained at an optimum level so that its
13
14 74 therapeutic effect is prolonged (Klein 2002). This may result in an improved treatment
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16 75 efficiency with a less total amount of drug, less frequent administration, minimised adverse
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18 76 effects and enhanced patient compliance (Michelson 1991; Hutton and Morris 1992;
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20 77 Wagstaff and Goa 2001; Pieper 2002; Michel 2002; McCarberg 2007). In contrast, due to the
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22 78 use of higher amounts of the drug in the formulation, damage of the drug reservoir of a
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24 79 sustained-release formulation can result in dose dumping and a drug blood concentration
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27 80 above the toxic level (Wen and Park 2010).

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32 81 Although there are many sustained-release pharmaceutical products available on the
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34 82 market, the mechanisms by which controlled release of a drug from a formulation can be
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36 83 achieved are relatively few. Dissolution-controlled formulations have the drug in a reservoir
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38 84 or dispersed in a matrix and the drug release rate is determined either by the dissolution rate
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40 85 and thickness of the polymer membrane, or the dissolution rate of the polymer matrix,
41
42 86 respectively. Likewise, in a diffusion-controlled formulation, drug release is controlled by the
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44 87 rate at which the drug molecules diffuse through a polymer membrane (reservoir systems) or
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46 88 a matrix. The matrix can be formed *in situ* from a physical mixture of the drug and the
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48 89 excipients or be in the form of a solid dispersion where the matrix is usually formed by a
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50 90 soluble polymer (Shergill et al. 2016; Baghel et al. 2016). Additionally, osmosis-based
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52 91 formulation is an important category of controlled-release products, where drug release
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54 92 occurs at a rate dependent on the rate of water penetration through a semi-permeable
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56 93 membrane surrounding the drug core and on the thickness of the membrane. Zero-order

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3 94 release of the drug can be achieved from these systems largely independent of environmental
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5 95 factors, such as temperature, pH and presence of food (Emara et al. 2012). Finally, ion-
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8 96 exchange-based formulations consist of ion-exchange resins that are water-insoluble
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10 97 polymers containing ionic groups. Drug molecules attach to the surface of the polymer
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12 98 through electrostatic interactions, provided that their charge is opposite to that of the
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14 99 polymer. The drug release takes place through replacement of the drug molecules with other
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17 100 ions with the same charge on the polymer surface (Malinovskaja et al. 2013). Here, we chose
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19 101 to exploit a sustained-release system based on a hydrophilic polymer matrix.

22 102 Hydrophilic polymer matrix systems have been extensively used in oral controlled-
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24 103 release delivery. The desired controlled-release profile is achieved easily through their use,
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26
27 104 they are cost-effective and have the United States (US) Food and Drug Administration (FDA)
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29 105 acceptance (Merchant et al. 2006). A typical hydrophilic polymer matrix system consists of
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31 106 drug, hydrophilic polymer(s) and other excipients distributed throughout the matrix. This
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34 107 system is dependent on polymer wetting, hydration and dissolution with regards to achieving
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36 108 controlled release of the drug (Merchant et al. 2006).

39 109 Many different hydrophilic polymers can be used to prepare controlled-release matrix
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41 110 systems. The most commonly used ones are cellulose ether derivatives which include
42
43 111 hydroxypropylmethylcellulose (HPMC) (Melia 1991). HPMC is a partly O-methylated and
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45 112 O-(2-hydroxypropylated) cellulose (Rowe et al. 2009). Its hydration rate depends on the
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48 113 molecular structure and degree of substitution of these substituents. More specifically, the
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50 114 hydration rate of HPMC increases with increasing hydroxypropyl content (Dipti et al. 2014).
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53 115 It forms swellable matrices that lead to the formation of viscous solutions, instead of true gels
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55 116 (Aulton and Taylor 2017). [High viscosity grades of HPMC can be used in tablets to retard](#)
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57 117 [drug release from matrices at a percentage of 10-80% w/w \(Rowe et al. 2009\).](#)
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3 118 MT-1207 hydrochloride is a novel antihypertensive drug that presents combinatory
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6 119 pharmacological actions. More specifically, it exhibits highly inhibitory activities
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8 120 simultaneously towards adrenergic α_{1A} , α_{1B} and serotonin (5-Hydroxytryptamine) $5-HT_{2A}$
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10 121 receptors (2019 email from Shenyang Haiwang Biotechnology Co. Ltd to us; unreferenced).
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12 122 In a hypertension animal model, MT-1207 had an evident dose-effect relationship. It can
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14 123 reduce the blood pressure without accelerating the heart rate while improving the auditory
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16 124 brainstem response (ABR) and kidney protection functions. Therefore, it is considered
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18 125 promising and efficacious in the treatment of hypertension. It is expected to prevent the first-
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20 126 dose effect and postural hypotension effectively (2019 email from Shenyang Haiwang
21
22 127 Biotechnology Co. Ltd to us; unreferenced). As is now well known, the drug is largely
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24 128 eliminated via hepatic clearance, mainly S-methylation. MT-1207 is a poorly soluble and
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26 129 highly lipophilic (logP of 4.15) basic compound with a pK_a value of 7.31. The most stable
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28 130 polymorph (anhydrous) has been chosen to be formulated in immediate-release tablets at a
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30 131 dose of 10 mg. Immediate-release tablets of MT-1207 hydrochloride were manufactured at an
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32 132 industrial scale by Shenyang Haiwang Biotechnology Co., Limited (Shenyang, China) and
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34 133 are being tested in terms of efficacy in phase II clinical trials. These tablets are designed to be
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36 134 administered three times a day. However, due to the challenges that come with hypertension,
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38 135 it is desirable to reduce the dosing frequency as much as possible to improve patient
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40 136 compliance.

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47 137 In the present study, a new sustained-release matrix tablet formulation of MT-1207
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49 138 hydrochloride was prepared. HPMC polymers of different viscosities were used as release
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51 139 retardants. Different tablet formulations were prepared using wet granulation and were
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53 140 characterised in terms of their physical properties and *in vitro* drug release. An optimised
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55 141 tablet formulation which provided a 24-hour release of MT-1207 in dissolution medium was
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3 142 developed. Pharmacokinetic studies were performed in Beagle dogs, after oral administration
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5 143 of the optimised sustained-release and immediate-release tablets.
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9 144 **2. Materials and Methods**

10 11 12 145 **2.1 Materials**

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14 146 MT-1207 hydrochloride reference standard and micronised MT-1207
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16 147 hydrochloride:colloidal silicon dioxide mixture with a mass ratio of 5:1 were donated by
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18 148 Shenyang Haiwang Biotechnology Co., Ltd (Shenyang, China). Sodium phosphate
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20 149 monobasic dihydrate, sodium acetate anhydrous, polyvinylpyrrolidone K30,
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22 150 hydroxypropylmethyl cellulose average molecular weight 86000 (K4M),
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24 151 hydroxypropylmethyl cellulose average molecular weight 90000 (K15M) and magnesium
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26 152 stearate were purchased from Sigma-Aldrich (Gillingham, United Kingdom). Alpha-D-
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28 153 lactose monohydrate, sodium dodecyl sulfate (SDS), hydrochloric acid 37%, methanol HPLC
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30 154 grade and propan-2-ol HPLC grade were purchased from Fisher Scientific (Loughborough,
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32 155 United Kingdom).
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39 156 **2.2 HPLC-UV method for the quantitation of MT-1207 samples**

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41 157 An HPLC-UV method was developed to quantify MT-1207 samples for solubility and
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43 158 dissolution testing. A Dionex Ultimate 3000 HPLC system was used (Thermo Scientific®,
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45 159 United Kingdom) with an ACE® Generix 3 C18, 150 × 4.6 mm, 3 µm column. The mobile
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47 160 phase consisted of 0.1 M sodium acetate buffer pH 4.0 and methanol at a ratio of 40:60 (%
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49 161 v/v). The mobile phase flow rate was set at 0.7 mL/min. The wavelength maximum of MT-
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51 162 1207 was found to be 230 nm, therefore the UV wavelength was set at that value. The
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53 163 injection volume was 20 µL. The retention time of MT-1207 was approximately 5 minutes.
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55 164 The quantitation of MT-1207 samples was carried out using a calibration curve which
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57 165 covered a concentration range of 1-50 µg/mL. The HPLC method was validated in terms of
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3 166 linearity, the limit of detection (LOD) and limit of quantitation (LOQ) as per International
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5 167 Council for Harmonisation (ICH) and FDA guidelines. The method LOD was 0.1 µg/mL and
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7 168 the LOQ was 0.4 µg/mL. The validation of the HPLC method for specificity, accuracy, intra-
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9 169 day and inter-day precision was carried out along with that of the dissolution method.
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13 170 **2.3 Scanning electron microscopy (SEM) analysis of MT-1207 reference standard and** 14 15 **micronised MT-1207** 16 171

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18 172 The MT-1207 hydrochloride reference standard and micronised MT-1207:silicon
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20 173 dioxide 5:1 mixture were examined by scanning electron microscopy (Philips® XL30 SEM,
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22 174 Netherlands) using 10 kV accelerating voltage. Before imaging, the samples were sputtered
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24 175 with gold using an SC7640 Sputter Coater (Polaron®, United Kingdom). Coated samples
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26 176 were then observed in the SEM at 10 kV under vacuum conditions at ambient temperature.
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30 177 **2.4 Solubility measurements of MT-1207:colloidal silicon dioxide 5:1 mixture** 31 32

33 178 Solubility testing was carried out for MT-1207 reference standard and the micronised
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35 179 MT-1207:colloidal silicon dioxide 5:1 mixture. The aim was to determine the solubility of
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37 180 MT-1207 in different pH values and to define whether micronisation had an effect on the
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39 181 solubility of MT-1207 agent. The testing was carried out in hydrochloric acid pH 1.2 and 0.1
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41 182 M phosphate buffer pH 6.8 simulating the pH values in the fasted stomach and small
42
43 183 intestine, respectively. Solubility studies were carried out using the shake-flask method (Baka
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45 184 et al. 2008). A known excess quantity of the drug was weighed each time and dispersed in 5
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47 185 mL of medium. Then, the dispersions were placed in a Stuart® orbital incubator and were
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49 186 shaken for 24 hours at 37°C. Afterwards, 3 mL were placed in Sarstedt® tubes and
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51 187 centrifuged in an Eppendorf® Centrifuge 5810 R (2700 g, 15 minutes). A two-step dilution
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53 188 was carried out for the supernatant. For the samples in hydrochloric acid, 500 µL of the
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55 189 supernatant were each time diluted to 1 mL with water:methanol 50:50 (% v/v) and, then, 100
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3 190 μL of the diluted solution were further diluted to 1 mL with water:methanol 50:50 (% v/v).
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5 191 For the samples in 0.1 M phosphate pH 6.8, 500 μL were diluted to 1 mL with methanol and,
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7 192 then, 100 μL from the diluted solution were further diluted to 1 mL with 0.1 M phosphate pH
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9 193 6.8:Methanol 50:50 (% v/v). Each time, the final diluent was transferred into an amber HPLC
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11 194 vial for HPLC analysis.
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15 195 **2.5 Preparation of MT-1207 tablet formulations**

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18 196 The HPMC matrix tablets were prepared using a wet granulation method. The
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20 197 micronised MT-1207:silicon dioxide mixture was geometrically diluted with alpha-D-lactose
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22 198 monohydrate and then the rest of the excipients were added. Mixing was carried out for 15
23
24 199 minutes manually. The powder mixture was then transferred into a beaker and the required
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26 200 amount of isopropyl alcohol (IPA) was added as droplets under manual mixing that was
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28 201 conducted using a spatula. IPA has low toxicity in humans and is, therefore, classified as
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30 202 Class 3 solvent, as per United States Pharmacopeia (USP) <467> (USP 2017). Afterwards,
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32 203 there was a wet massing time of 30 seconds. The wet mass was manually pressed through a
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34 204 12-mesh stainless steel sieve. The wet granules were then left in the fume hood to dry for two
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36 205 hours and were then placed in the tray drier for two hours at 60°C. The dried granules were
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38 206 then sieved through a 25-45-170-mesh sieve array and the few fines (particle size < 90 μm)
39
40 207 were discarded. Magnesium stearate was added as a lubricant in the granulated mixture and
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42 208 mixing was carried out for 5 minutes. The granules were then compressed into tablets using a
43
44 209 Piccola rotary tablet press machine (Riva[®], Argentina). 8 mm flat-faced round punches and
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46 210 dies were used during the compression process. The compression force applied during the
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48 211 preparation of the different tablet formulations was 6.5 kN. The target tablet weight was
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50 212 224.5 mg for formulations F1-F4 and 226.5 mg for formulation A1. Formulation F1
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52 213 contained 18% w/w HPMC K4M, while formulations F2, F3 and F4 contained 18% w/w,
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54 214 22% w/w and 31% w/w HPMC K15M, respectively. The compositions of the different tablet
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3 215 formulations can be seen in Table 1. Three different batches of the optimised formulation
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5 216 were prepared and characterised to ensure the reproducibility of the manufacturing method.
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7 217 Finally, formulation A1 used in the pharmacokinetic study was prepared based on the
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9 218 optimised formulation containing 30 mg MT-1207, instead.

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13 219 [Table 1]
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16 220 2.6 Flowability characterisation of the tablet blends

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19 221 The flow properties of the granulated mixture are most important. The flowability
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21 222 directly affects the filling uniformity of the die in the tableting machine and, therefore, the
22
23 223 corresponding uniformity of tablet weight and drug content (USP 2016). As per the USP, the
24
25 224 flowability was assessed in terms of bulk/tapped density, compressibility index, Hausner ratio
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27 225 and angle of repose.

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31 226 The bulk/tapped density testing was carried out using tapped density tester (Copley[®],
32
33 227 Nottingham, United Kingdom) as per USP <616> (USP 2015). Each time, a certain amount
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35 228 of granulated mixture was weighed and then added inside a 100-mL graduated measuring
36
37 229 cylinder. The initial volume (bulk volume, V_0) was recorded. Then, the mixture was tapped
38
39 230 10, 500 and 1250 times and the respective volumes were recorded (V_{10} , V_{500} , V_{1250}). If the
40
41 231 difference between V_{500} and V_{1250} was not more than 2 mL then V_{1250} was considered as the
42
43 232 final tapped volume (USP 2015). The V_0 and V_{1250} were then used to calculate the
44
45 233 compressibility (Carr) index and Hausner ratio. These values were used to characterise the
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47 234 flowability of the different mixtures.

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52 235 The angle of repose testing was also used to characterise the flowability of powder
53
54 236 mixtures. The fixed funnel method was carried out, as per USP <1174> (USP <1174> 2012).
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56 237 Each time, an amount of powder was added inside a funnel and left to flow through it. As the
57
58 238 tip of the powder cone reached the bottom of the funnel, the funnel was carefully moved
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3 239 upwards. When the diameter of the cone reached slightly more than 6 cm the diameter of the
4
5 240 base and the height of the cone were measured using a ruler. These values were then used to
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7
8 241 calculate the angle of repose. Based on the value of the angle of repose, the flowability of the
9
10 242 mixture was characterised, as per USP (USP <1174> 2012).

13 243 **2.7 Hardness and friability of tablets**

16 244 The HPMC matrix tablets prepared were assessed in terms of their hardness using a
17
18 245 C50 Tablet Hardness tester (Engineering Systems (Nottm)[®], Nottingham, United Kingdom).
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20 246 The test was carried out as per USP <1217> (USP <1217> 2012). 10 tablets were tested for
21
22
23 247 each formulation batch. Each time, the tablet dimensions (diameter, thickness) were
24
25 248 measured. Then, the tablet was placed in the hardness tester and the hardness value was
26
27 249 recorded in kg units. Afterwards, the tablet tensile strength was calculated, as per USP (USP
28
29 250 <1217> 2012).

32 251 The friability of the HPMC matrix tablets was assessed, as per USP <1216> (USP
33
34 252 <1216> 2012). The tablets were weighed before being placed into the friability tester
35
36
37 253 (Erweka[®], Heusenstamm, Germany). The friabilator was set at a revolution rate of 25 rpm for
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39 254 4 minutes. The tablets were then removed from the device and were smoothly wiped using a
40
41
42 255 brush. Then, they were weighed and the % friability was calculated. As per USP, the
43
44 256 friability should be less than 1.0%.

47 257 **2.8 Uniformity of weight**

50 258 The tablet weight variation was assessed, as per USP <905> (USP 2016). 10 tablets were
51
52 259 weighed individually for each formulation. The mean tablet weight was calculated and
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54 260 expressed as % of the target tablet weight. Then, the acceptance value was calculated. The
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57 261 tablet weight variation was considered to be within the acceptable range set in USP if the
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59 262 acceptance value was less than 15.

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3 263 **2.9 Differential scanning calorimetry analysis of MT-1207 and the sustained-release**
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5 264 **tablets**

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8 265 The thermal behaviours of MT-1207 hydrochloride and pulverised tablets were
9
10 266 evaluated using DSC Q2000 (TA[®] Instruments, Centennial Park, United Kingdom). Samples
11
12 267 (1-5 mg) were each time weighed and sealed inside an aluminum pan with pierced aluminum
13
14 268 lids. The analysis took place over a temperature range of 40-240°C at a rate of 5°C/min in an
15
16 269 atmosphere of nitrogen, the flow rate of which was set at 50 mL/min. The analysis for the
17
18 270 API took place over a temperature range of 40-300°C to spot any potential thermal
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20 271 degradation at temperature values beyond 240°C.
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25 272 **2.10 Fourier-Transform Infrared (FTIR) spectrophotometric studies**

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27
28 273 Fourier-Transform Infrared (FTIR) spectra were obtained for MT-1207
29
30 274 hydrochloride and the sustained-release tablets that were prepared using Cary 630 FTIR
31
32 275 spectrophotometer (Agilent Technologies[®], Craven Arms, United Kingdom). Each time, the
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34 276 solid sample was placed on the lid and scanned at the range 4000-650 cm⁻¹.
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38 277 **2.11 X-ray powder diffraction (XRPD) analysis of MT-1207 and the sustained-release**
39
40 278 **tablets**

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43 279 The diffractograms of MT-1207 hydrochloride and pulverised tablets were obtained
44
45 280 using an X'Pert Pro MPD X-ray diffractometer (Malvern Panalytical[®], Malvern, United
46
47 281 Kingdom). Cu K-alpha1 radiation was used. The samples were scanned at an angle range of
48
49 282 4-50°(2θ) with a step size of 0.01°(2θ) at 40 mV, 40 mA and a sample rotation speed of 15
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51 283 rpm.
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55 284 **2.12 In vitro release of MT-1207 from sustained-release tablets**

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58 285 *In vitro* dissolution studies were carried out using a USP II dissolution apparatus
59
60 286 (Copley[®], Nottingham, United Kingdom) equipped with paddles. The apparatus was set at a

1
2
3 287 constant rotation speed of 75 rpm. Dissolution studies were carried out in 900 mL of 0.1 M
4
5 288 phosphate buffer pH 6.8 with 0.2% w/v SDS at $37 \pm 0.5^\circ\text{C}$. Sinkers were used to prevent
6
7 289 floating of tablets and their adhesion to the bottom of the vessels. Sampling was carried out
8
9
10 290 manually at certain time intervals (0.08, 0.25, 0.5, 1, 1.5, 2, 3, 4, 6, 8, 10, 12, 14, 16, 18, 20,
11
12 291 22 and 24 hours), followed by centrifugation of the samples (2300 g, 10 minutes). Then, each
13
14 292 time 1 mL from the supernatant was transferred into an HPLC vial and 20 μL injected into
15
16 293 the HPLC for analysis. The *in vitro* release study was carried out in triplicate for each
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18 294 formulation.

23 295 **2.13 Swelling and erosion studies**

25 296 Swelling and erosion studies were carried out for the F4 tablets in dissolution medium,
26
27 297 0.1 M phosphate pH 6.8 with 0.2% w/v SDS. The USP II Dissolution Apparatus was used.
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29 298 Each time, the pre-weighed tablet (initial weight, W_0) was immersed in 900 mL dissolution
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31 299 medium within a dissolution vessel. The temperature was maintained at $37 \pm 0.5^\circ\text{C}$. The
32
33 300 paddle speed was set at 75 rpm. At certain time intervals (1, 2, 4, 6, 8, 12, 16, 20 and 24
34
35 301 hours) the tablet was taken out of the dissolution vessel and excess water was carefully
36
37 302 plotted from its surface. The wet tablet was weighed and the wet weight value, W_t , was
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39 303 obtained. The % swelling index of the tablets was calculated using Equation 1:
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$$47 305 \quad \% \text{ Swelling index} = (W_t - W_0) \times 100/W_0 \quad (\text{Equation 1})$$

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52
53 307 The same tablets that were used to calculate the swelling index were also used to
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55 308 calculate the % erosion value. After weighing, the wet tablets were placed inside an oven and
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57 309 the temperature was set at 60°C . The tablets were each time dried until constant weight. The
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3 310 dry weight value, W_f , was recorded. The % erosion of the tablets at each timepoint was
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5 311 calculated using Equation 2:
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8 312

$$11 \quad 313 \quad \% \text{ Erosion} = (W_0 - W_f) \times 100/W_0 \quad (\text{Equation 2})$$

14 314

17 315 All experiments were done in triplicate and the results were recorded as mean \pm
18
19 316 standard deviation.
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23 317 **2.14 Kinetics and mechanism of drug release**

25 318 Based on the results obtained from *in vitro* dissolution testing for the optimised
26
27 319 formulation F4, the release kinetics of MT-1207 from that formulation were studied. For this
28
29 320 purpose, the data obtained from the *in vitro* drug release testing were plotted in various
30
31 321 kinetic models. Those were first-order (Equation 3) as log percentage of MT-1207 remaining
32
33 322 undissolved vs time, zero-order (Equation 4) as the cumulative amount of drug released vs
34
35 323 time and Higuchi's models (Equation 5) as cumulative percentage of MT-1207 released vs
36
37 324 square root of time. Finally, the data were plotted using the Hixson-Crowell cube root law to
38
39 325 evaluate the drug release with changes in the particle/tablet surface area and diameter
40
41 326 (Equation 6) (Merchant et al. 2006).
42
43
44
45
46
47
48
49

327

$$50 \quad 328 \quad \text{Log}Q = \text{Log}Q_0 - kt/2.303 \quad (\text{Equation 3})$$

52 329

55 330 where Q is the amount of the drug in the tablet at time t, Q_0 is the total amount of the drug
56
57 331 and k is the first-order constant.
58
59
60

332

333

$$C = k_0 t \quad (\text{Equation 4})$$

334

335 where C is the concentration of the drug at time t and k_0 is the zero-order constant.

336

337

$$Q = kt^{1/2} \quad (\text{Equation 5})$$

338

339 where Q is the cumulative % of drug released at time t and k is a constant the value of which
340 depends on the system design variables (Merchant et al. 2006).

341

342

$$\sqrt[3]{Q_0} - \sqrt[3]{Q_t} = k_{HC} \times t \quad (\text{Equation 6})$$

343

344 where Q_t is the amount of drug released in time t , Q_0 is the initial total amount of drug in the
345 tablet and k_{HC} is the rate constant for the Hixson-Crowell equation which refers to the plot of
346 the cube root of the drug remaining in the tablet vs time.

347 The mechanism of drug release from F4 tablets was determined using the Korsmeyer-
348 Peppas model (Korsmeyer et al. 1983). The data from the *in vitro* dissolution testing were
349 plotted in the Korsmeyer-Peppas equation (Equation 7) as log cumulative percentage of drug
350 released vs log time. The exponent n was calculated by measuring the slope of the straight
351 line.

352

1
2
3 353
$$M_t/M_\infty = kt^n$$
 (Equation 7)
4
5

6 354
7
8

9 355 where M_t is the amount of drug released at time t , M_∞ is the total amount of drug and k is a
10
11 356 kinetic constant the value of which is characteristic of the drug/polymer system and n is an
12
13 357 exponent that is used to characterise the mechanism of drug release (Korsmeyer et al. 1983).
14
15
16 358 For cylinder tablets, $n = 0.45$ corresponds to drug release controlled by Fickian diffusion,
17
18 359 $0.45 < n < 0.89$ corresponds to anomalous (non-Fickian diffusion) which refers to a
19
20 360 combination of Fickian diffusion and polymer relaxation mechanisms, while $n > 0.89$ defines
21
22 361 super Case-II transport where drug release is controlled by polymer relaxation (Peppas 1985;
23
24 362 Siepmann and Peppas 2001; Nguyen et al. 2020).
25
26
27

28 363 **2.15 *In vivo* animal study**

29
30
31 364 An *in vivo* animal study was carried out as per the principles of the Declaration of
32
33 365 Helsinki in Beagle dogs (body weight 10 ± 2 kg) for MT-1207 30 mg sustained-release (A1)
34
35 366 and 10 mg immediate-release tablets. Eight healthy beagle dogs were used, half male and half
36
37 367 female. According to gender and weight, they were randomly divided into two groups, 4 in
38
39 368 each group (2 males and 2 females). After fasting for 12 hours (h), the two groups of animals
40
41
42 369 were given MT-1207 tablets orally. In the first half of the first group (Group 1-A), the
43
44 370 sustained-release tablets were administered once a day, one tablet at a time, and blank blood
45
46 371 was taken before administration. After administration, at 0.25 h, 0.5 h, 1 h, 1.5 h, 2 h, 3 h, 4
47
48 372 h, 6 h, 8 h, 12 h, 16 h, 24 h and 32 h timepoints approximately 1.5 mL of venous blood were
49
50 373 each time collected. In the second half of the first group (Group 1-B) immediate-release
51
52 374 tablets were administered three times a day, one tablet at a time, with an interval of 8 hours.
53
54 375 Blank blood was collected before administration and then at 0.25 h, 0.5 h, 1 h, 2 h, 4 h, 8 h
55
56 376 after the first administration (second administration), 8.25 h, 8.5 h, 9 h, 11 h, 13 h, 16 h (third
57
58
59
60

1
2
3 377 dose), 16.25 h, 16.5 h, 17 h, and 24 h. Approximately 1.5 mL of venous blood were collected
4
5 378 at each timepoint. Two groups of animals were cross-administered with another formulation
6
7 379 after 7 days of wash-out period. In the second group (Group 2), the immediate-release tablets
8
9 380 were administered once a day, three tablets at a time, and blank blood was taken before
10
11 381 administration, and at 0.25 h, 0.5 h, 1 h, 1.5 h, 2 h, 3 h, 4 h, 8 h, and 24 h timepoints.
12
13 382 Approximately, 1.5 mL of venous blood was collected at each timepoint. In all the cases,
14
15 383 each sample was placed in a test tube containing heparin, centrifuged and the plasma was
16
17 384 separated and stored at -20°C. MT-1207 in plasma samples was determined using UPLC-
18
19 385 MS/MS.
20
21
22
23
24

25 386 **2.16 Bioanalytical method**

26
27 387 MT-1207 in plasma samples was determined by a validated UPLC-MS/MS method using
28
29 388 verapamil hydrochloride as an internal standard. Each time 10 µL of plasma sample were
30
31 389 pipetted in 1.5 mL Eppendorf® tube. 200 µL of verapamil hydrochloride 2 ng/mL in
32
33 390 acetonitrile were added and vortex was carried out for 5 minutes. Centrifugation was then
34
35 391 carried out at 15000 rpm for 5 minutes and 100 µL of supernatant were collected for UPLC-
36
37 392 MS/MS analysis. The ion source was an electrospray ionisation source (ESI). A positive ion
38
39 393 scanning method was used for detection. The solvent gas (nitrogen) flow rate was 1000 L/h,
40
41 394 the temperature of the solvent gas was 500°C, and the capillary voltage was 3.0 kV. The
42
43 395 scanning method was Multiple Response Monitoring (MRM). The cone voltage was set at 40
44
45 396 V, while the collision energy was 20 eV. For quantitative analysis, the ion pairs used had m/z
46
47 397 393.26 → 274.04 (MT-1207) and m/z 455.25 → 156.06 (internal standard). The samples
48
49 398 were applied to an ACQUITY Ultra Performance Liquid Chromatography system with Xevo
50
51 399 TQ-XS Triple Quadrupole Mass Spectrometer with operating software MassLynx V4.2
52
53 400 (Waters Technology Limited Company). The column used was an ACQUITY UPLC BEH
54
55 401 C18 liquid chromatography column (2.1 × 50 mm, 1.7 µm). The mobile phase consisted of
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57
58
59
60

1
2
3 402 0.1% formic acid in water (mobile phase A) and acetonitrile (mobile phase B). Verapamil
4
5 403 hydrochloride was used as the internal standard for determination. The gradient elution was:
6
7 404 0-1.2 min: 20-45% B, 1.2-1.5 min: 45-95% B, 1.5-1.8 min: 95% B, 1.8-2.5 min: 95-20% B.
8
9
10 405 The flow rate was set at 0.5 mL/min. The column temperature was set at 45°C.
11
12

13 406 Quantification was carried out using a calibration curve. 10 mg MT-1027 were
14
15 407 accurately weighed and dissolved in DMSO to prepare a stock solution of MT-1207 with a
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17 408 concentration of 10 mg/mL. This was gradually diluted with a 50% acetonitrile solution to
18
19 409 obtain standard working solution concentrations of 10, 20, 40, 100, 200, 400, 1000, 2000,
20
21 410 4000 ng/mL. The concentrations of the quality control samples were 30, 320, 3200 ng/mL.
22
23
24 411 To prepare a quality control working solution 10 µL were added to 190 µL Beagle dog blank
25
26 412 plasma. Vortex mixing was carried out to obtain the standard curve with drug concentration
27
28 413 values of 0.5, 1, 2, 5, 10, 20, 50, 100, and 200 ng/mL and plasma with drug concentration
29
30 414 values of 0.5, 1.5, 16, and 160 ng/mL. 20 µL of medicated plasma were transferred into a 1.5
31
32 415 mL Eppendorf® tube and 200 µL of 2 ng/mL verapamil hydrochloride in acetonitrile were
33
34 416 added. Vortex was carried out for 5 minutes, followed by centrifugation at 15,000 rpm for 5
35
36 417 minutes. 100 µL of supernatant were collected for UPLC-MS/MS measurement. The lower
37
38 418 limit of quantitation (LLOQ) was 0.500 ng/mL. The linear regression coefficient of the
39
40 419 standard curve was around 0.9978. The accuracy and precision values of the quality control
41
42 420 samples were 101.0% and 3.2% CV, respectively.
43
44
45
46
47
48

49 421 **2.17 *In vitro and in vivo data analysis***

50
51 422 The peak plasma concentration of MT-1207 (C_{max}), the time to reach C_{max} (T_{max}) and the
52
53 423 area under the concentration-time curve from 0 hours to 24 hours (AUC_{0-24h}) and to infinity
54
55 424 ($AUC_{0-\infty}$) values were calculated from the plasma concentration vs time profile using
56
57 425 WinNonlin® version 6.4. Furthermore, the half-life ($t_{1/2}$), clearance (CL), apparent volume of
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59
60

1
2
3 426 distribution (V_d), and mean residence time (MRT) values of MT-1207 after administration of
4
5 427 the tablets following the different dosing regimens were also calculated using WinNonlin®
6
7 428 version 6.4. Finally, the bioequivalence between the sustained-release and immediate-release
8
9 429 tablets was investigated by calculating the ratio of AUC_{0-24h} values of the sustained-release
10
11 430 tablet dosing regimen and the two separate immediate-release tablet dosing regimens.
12
13
14

15 431 T-test and one-way analysis of variance (ANOVA) were carried out for the statistical
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17 432 analysis of results using Microsoft Excel® software. A statistically significant difference was
18
19 433 reflected by a P value less than 0.05.
20
21
22

23 434 **3. Results and Discussion**

24 25 26 435 **3.1 Scanning electron microscopy and solubility characterisation of MT-1207**

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28
29 436 Solubility testing was carried out in hydrochloric acid pH 1.2 and phosphate buffer pH
30
31 437 6.8 to obtain information on the solubility of the drug at different pH values. MT-1207 was
32
33 438 more soluble in hydrochloric acid than in phosphate buffer (Table S1). Since the latter was
34
35 439 chosen as the dissolution medium in the present study, a small amount of SDS was added into
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37 440 phosphate buffer to ensure sink conditions during dissolution testing.
38
39
40

41 441 There was no statistically significant difference between the recorded solubility values
42
43 442 of non-micronised MT-1207 and micronised MT-1207:silicon dioxide mixture in the
44
45 443 different media (t-test, $p > 0.05$) (Table S1). These results agree with literature findings since
46
47 444 particle size reduction of drugs at this scale was not expected to lead to increased aqueous
48
49 445 solubility unless it caused amorphisation of the API (Jinno et al. 2006; Loh et al. 2015).
50
51 446 Particle size reduction could, however, enhance the dissolution rate of MT-1207, due to
52
53 447 increased drug surface area for interaction with liquid media (Chu et al. 2012; Marinko and
54
55 448 Zámotný 2020). In Figure 1, it can be seen that the drug crystals have an elongated shape.
56
57 449 The long-axis particle size values of the non-micronised drug and micronised drug that was
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59
60

1
2
3 450 used to make the tablets were $(11.97 \pm 7.42) \mu\text{m}$ and $(4.02 \pm 1.90) \mu\text{m}$, respectively (n=100
4
5 451 particles for each batch). The particle size distribution was narrower in the micronised batch
6
7 452 than in the non-micronised drug and was used for the preparation of both immediate and
8
9 453 sustained release tablets.

10
11
12
13 454 **[Figure 1]**

14
15
16 455 **3.2 Characterisation of flowability and compressibility of the tablet powder blends**

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18
19 456 Before compression, the granulated powder blends were assessed in terms of
20
21 457 compressibility and flowability through bulk/tapped density and angle of repose testing, as
22
23 458 per USP.

24
25
26 459 The flowability of a powder blend is most important since it determines whether the
27
28 460 filling of the powder in the dies of the tableting machine is uniform, thus ensuring uniform
29
30 461 tablet weight and dimensions and drug content uniformity (Abd-Elbary et al. 2012). The
31
32 462 flowability of a powder blend can be assessed through Carr index, Hausner ratio or angle of
33
34 463 repose. Compressibility is also important since it defines the ability of a powder to compress
35
36 464 upon application of pressure (Jamzad and Fassihi 2007). Carr index is a good indicator of the
37
38 465 powder flowability and compressibility (Carr 1965b). Therefore, a Carr index value between
39
40 466 5% and 16% is desirable to achieve both good flowability and compressibility (Jamzad and
41
42 467 Fassihi 2007). F1, F3, F4 and A1 powder blends had Carr index values of 14.7, 15.4, 12.2
43
44 468 and 13.3%, respectively (Table S2), which indicates a combination of good flowability and
45
46 469 compressibility. The F2 powder blend, however, had a relatively high compressibility index
47
48 470 value of 23.5%, but still, its flowability was characterised as fair, which ensured its free flow
49
50 471 in the feeder of the tableting machine and the dies.

51
52
53
54
55
56
57 472 Hausner ratio is another measure of the flow properties of a powder blend (Hausner
58
59 473 1967). It provides information on the extent of the powder densification upon compression.

1
2
3 474 Powders with a higher tendency to densify have higher Hausner ratio values. All powder
4
5 475 blends had Hausner ratio values that indicated fair to good flowability and were in complete
6
7 476 alignment with the respective Carr index values. For each tablet batch, both of these values
8
9 477 were calculated through bulk/tapped density testing, as per USP. The Hausner ratio values
10
11 478 ranged between 1.14 and 1.18 for the powder mixtures of formulations F1, F3, F4 and A1
12
13 479 (Table S2) which confirmed their good flowability and compressibility, along with the
14
15 480 respective Carr index values. F2 powder blend, however, gave a relatively high Hausner ratio
16
17 481 of 1.31, but, still, its flowability was characterised as fair and in alignment with the respective
18
19 482 Carr index value.

20
21
22
23
24 483 The angle of repose can also be used to characterise the flow properties of solids. It is
25
26 484 a characteristic related to powder cohesion or resistance to movement between particles (USP
27
28 485 <1174> 2012). High angle of repose values correspond to high powder cohesiveness and,
29
30 486 thus, poor flow properties. The angle of repose values for the F1, F3, F4 Repeat 2, F4 Repeat
31
32 487 3 and A1 powder mixtures were 34.2°, 33.2°, 38.7°, 35.6° and 36.4°, respectively (Table S2).
33
34 488 These test findings align with the bulk/tapped density results for flowability of the blends. In
35
36 489 F2 and F4 Repeat 1 batches, there was a deviation of the angle of repose value from
37
38 490 bulk/tapped density results. For F2 the angle of repose value (39.0°) defined a good
39
40 491 flowability, while the rest of the results indicated a fair flowability. For F4, the angle of
41
42 492 repose value, 42.0°, defined a passable flowability of the powder whilst the bulk/tapped
43
44 493 density results indicated a good flowability of the same blend. Such subtle variations **can**
45
46 494 **occur** within these empirical type tests but they remain broadly consistent.

495 ***3.3 Post-compression parameters of tablet formulations***

496 The hardness of the prepared tablet formulations was assessed (Table S3). Increasing
497 HPMC content and viscosity grade from formulations F1 to F4 resulted in higher hardness

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2
3 498 values. This could be due to the binding properties of HPMC, so increasing its content
4
5 499 probably resulted in stronger solid bridges within the tablets and, thus, higher hardness
6
7 500 (Chowhan 1980; Dipti et al. 2014). The hardness test was also carried out for F4 Repeats 2
8
9 501 and 3 (Table S3). There was no statistically significant difference in tablet hardness between
10
11 502 the three F4 batches (one-way ANOVA, $p > 0.05$). Also, the hardness of A1 tablets bound to
12
13 503 be used for animal studies was similar to that of F4 tablets (one-way ANOVA, $p > 0.05$).
14

15
16
17 504 All tablet batches gave friability values that were well within the acceptable range of
18
19 505 1.0% set in USP (Table S4). This confirmed their good quality and their ability to withstand
20
21 506 the expected, abovementioned mechanical shocks.
22

23
24
25 507 All tablet batches gave acceptable mean weight values and acceptance values, with
26
27 508 regards to uniformity of weight (Table S5). Therefore, for all formulations, the tablets had a
28
29 509 uniform weight of drug substance per weight of dosage unit.
30

31 32 33 510 **3.4 Differential scanning calorimetry (DSC) analysis** 34

35
36 511 The thermal behaviour of an MT-1207 reference standard and that of the tablet
37
38 512 formulations was assessed using DSC.
39

40
41 513 DSC thermograms of MT-1207 and the different formulations can be seen in Figures 2
42
43 514 and 3. MT-1207 hydrochloride crystal (Figure 2) showed a sharp endothermic peak at around
44
45 515 210°C which was attributed to its melting point, while degradation seemed to take place at
46
47 516 around 275°C. In the thermograms of the tablet formulations (Figure 3) different peaks were
48
49 517 noted. The peaks at around 150-160°C correspond to the dehydration of alpha-D-lactose
50
51 518 monohydrate towards beta-lactose and melting point of PVP K30 and the peak at around
52
53 519 220°C corresponds to the melting point of beta-lactose. A relatively broad endothermic peak
54
55 520 was seen at the temperature range 178-197°C. This is proposed to correspond to the melting
56
57 521 point of the MT-1207 hydrochloride crystal. A broadening of the peak was noted, though,
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59
60

1
2
3 522 along with a reduction in the heat of fusion, compared to the reference standard. This could
4
5 523 potentially be due to the mixing process which results in lower purity of each component,
6
7
8 524 thus leading to lower melting point of the drug crystal (Smith 1982; Dürig and Fassihi 1993;
9
10 525 Pani et al. 2011). Additionally, the strong physical interactions taking place between the drug
11
12 526 and the excipients during mixing, granulation and, particularly, compression into tablets
13
14 527 could significantly contribute to the broadening of the MT-1207 endothermic peak (Mura et
15
16 528 al. 1995). Finally, the dispersion of the drug crystal in the HPMC and PVP K30 polymer
17
18 529 matrix could account for the reduction in both the melting point and the heat of fusion of the
19
20
21 530 drug (Mura et al. 1998).

22
23
24 531 **[Figure 2]**

25
26
27 532 **[Figure 3]**

31 533 **3.5 Fourier-Transform Infrared (FTIR) analysis**

32
33 534 In Figure 4 the Fourier-Transform Infrared (FTIR) spectra of MT-1207 hydrochloride
34
35 535 reference standard (Figure 4A), HPMC K15M (Figure 4B), colloidal silicon dioxide (Figure
36
37 536 4C) and pulverised tablets (Figure 4D) can be seen.

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39
40
41 537 FTIR characterisation is useful in determining whether chemical reactions between the
42
43 538 API and the excipients occur during mixing, granulation and tableting (Pani et al. 2011). The
44
45 539 chemical structure of MT-1207 does not contain functional groups that would readily react
46
47 540 with the excipients. The peaks in the 2800-3000 cm^{-1} region (Figure 4A) that correspond to
48
49 541 aliphatic and aromatic C-H stretch can also be seen unchanged in the pulverised tablet
50
51 542 spectrum (Figure 4D). Furthermore, the peaks in the 1250-1300 cm^{-1} region that correspond
52
53 543 to C-N stretch can be seen in both the spectra of both the MT-1207 reference standard and
54
55 544 pulverised tablets. Finally, the fingerprint region of the pulverised tablets is a simple mixture
56
57
58 545 of the fingerprint regions of MT-1207 hydrochloride (Figure 4A), colloidal silicon dioxide

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2
3 546 (Figure 4B) and HPMC K15M (Figure 4C) with no additional peaks observed. These results
4
5 547 illustrate that there were no significant chemical interactions between MT-1207 and the
6
7
8 548 excipients.

9
10
11 549 **[Figure 4]**

12 13 14 550 **3.6 X-ray powder diffraction (XRPD) analysis**

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16 551 The diffractograms of MT-1207 hydrochloride reference standard and formulations F1
17
18 552 and F4 were acquired and can be seen in Figures 5A and 5B, respectively. The peaks of MT-
19
20
21 553 1207 anhydrous crystal are assigned in Figure 5C. The diffractograms of the sustained-
22
23
24 554 release tablet formulations were identical. In Figure 5C, it can be seen that the peaks of the
25
26 555 F4 diffractogram over the angle($^{\circ}2\theta$) values of 11.5, 15 and 18 corresponded to the
27
28 556 respective peaks of the MT-1207 hydrochloride reference standard, while all the other peaks
29
30
31 557 of the F4 diffractogram corresponded to the peaks of alpha-D-lactose monohydrate. The
32
33 558 reduction in peak height and broadening of the MT-1207 peaks in the F1 and F4
34
35 559 diffractograms (Figures 5B and 5C) can be attributed to the mixing of the drug with the rest
36
37 560 of the excipients, most of which are amorphous. This resulted in a lower percentage of the
38
39
40 561 API in the mixture analysed and, therefore, to broader and less intense peaks. All the peaks
41
42 562 present in the diffractogram of F4 could be attributed to either MT-1207 anhydrous crystal or
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44 563 alpha-D-lactose monohydrate. Therefore, it can be asserted that within the sensitivity of the
45
46 564 analysis that the drug was present in the tablets solely in the form of the anhydrous crystal.

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48
49 565 **[Figure 5]**

50 51 52 566 **3.7 In vitro release of MT-1207 from sustained-release tablets**

53
54
55 567 The *in vitro* release profiles of the immediate-release tablets and the different
56
57
58 568 sustained-release tablet batches can be seen in Figure 6. HPMC has been used as a release
59
60 569 retarding polymer due to its ability to swell when in contact with water. In the present study,

1
2
3 570 HPMC polymers of different molecular weight and viscosity values were used in the different
4
5 571 formulations. HPMC K4M was the initial choice of release retarding polymer. In the
6
7 572 literature, this viscosity grade of HPMC has been effective in retarding the *in vitro* release of
8
9 573 different drugs from sustained-release formulations (Abd-Elbary et al. 2012; Hiremath and
10
11 574 Saha, 2008; Savaşer et al. 2013). However, when HPMC K4M was used as a sole release
12
13 575 retardant at a percentage of 18% w/w (formulation F1) it was unable to significantly retard
14
15 576 the release of MT-1207. For this reason, in the rest of the batches, HPMC K4M was replaced
16
17 577 by HPMC K15M, a polymer of higher molecular weight and viscosity.
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19
20
21

22 578 [Figure 6]

23
24
25 579 In Figure 6 it can be seen that a sustained release of MT-1207 was achieved from
26
27 580 tablets with HPMC K15M. In the literature, it has been noted that drug release rate from
28
29 581 matrix tablets decreases with increasing HPMC concentration and viscosity, due to increasing
30
31 582 strength and thickness of the gel layer that is formed upon swelling of the polymer and the
32
33 583 diffusion of the drug through the polymer chains being retarded (Jamzad and Fassihi 2006;
34
35 584 Hiremath and Saha 2008). Lactose, in contrast, promotes drug release through promoting
36
37 585 matrix hydration, gel formation and free volume (Gao et al. 1996). Increasing the
38
39 586 concentration of HPMC K15M from F2 to F4 resulted in a slower release of MT-1207 which
40
41 587 was maintained linear over a longer time. The results of the present study are, therefore, fully
42
43 588 aligned with the literature (Jamzad and Fassihi 2006). The desired release profile of the drug
44
45 589 was acquired from formulation F4 (31% w/w HPMC K15M), reaching approximately 93%
46
47 590 release after 24 hours of dissolution with the linearity of release being maintained for 20
48
49 591 hours. This is most desirable since it could potentially ensure constant, effective levels of
50
51 592 MT-1207 in plasma following absorption of the drug through the gastrointestinal tract
52
53 593 (Merchant et al. 2006). Therefore, formulation F4 was selected as the optimised formulation
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55 594 and for *in vivo* studies. Furthermore, no statistically significant difference was noted between
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2
3 595 the release profiles from the three different F4 batches (one-way ANOVA, $p > 0.05$) (Figure
4
5 596 S1). Therefore, it was concluded that the manufacturing process and the formula gave rise to
6
7
8 597 tablets with reproducible properties. Finally, formulation A1 was prepared based on F4 with a
9
10 598 change in the drug dose to 30 mg. There was no statistically significant difference in the *in*
11
12 599 *vitro* release profile of MT-1207 between the F4 and A1 formulations (t-test, $p > 0.05$).
13
14 600 Therefore, the properties of F4 and A1 tablets were identical.
15
16
17

18 601 **3.8 Swelling and erosion study**

19
20 602 The swelling and erosion behaviour of tablets is most important since it can affect
21
22 603 drug release kinetics. In Figure 7, the results from the swelling and erosion studies of F4
23
24 604 tablets can be seen. The tablets showed significant swelling in the dissolution medium with
25
26 605 the tablet weight increasing by 170.6% w/w after 8 hours of dissolution. A rapid swelling
27
28 606 phase was initially noted, since 110.3% w/w swelling took place within 2 hours of
29
30 607 dissolution, followed by a relatively slower increase between 2 and 8 hours of dissolution.
31
32 608 After 8 hours of dissolution, the tablet erosion reached 64.0% w/w indicating that most of the
33
34 609 tablet mass had eroded at the time where maximum swelling of the tablets was achieved.
35
36 610 After that timepoint, a noteworthy reduction of the erosion rate was recorded, possibly
37
38 611 because maximum swelling was reached and therefore the thickness of the hydrogel and the
39
40 612 viscosity of the surrounding viscous solution were maximised. These results, along with the
41
42 613 *in vitro* release findings, indicate that diffusion and swelling may have contributed, to some
43
44 614 extent, to the release of MT-1207 from the F4 tablets, but drug release was predominantly
45
46 615 erosion-controlled.
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52

53 616 **[Figure 7]**
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617 3.9 Kinetics and mechanism of MT-1207 release from formulation F4

618 Through the selection of an appropriate molecular weight of HPMC in the matrix and
619 the rate of matrix hydration, the rate of polymer disentanglement can be controlled.
620 Therefore, the mechanism of drug release is dependent on the sum of drug diffusion and
621 polymer relaxation and erosion. The release profile of F4 tablets was plotted against different
622 models to elucidate the kinetics of drug release, while the Korsmeyer-Peppas model was used
623 to obtain information on the mechanism of drug release. All models were used to analyse the
624 data of up to 60% of the release curve (Peppas 1985; Sauri et al. 2014).

625 The release data of formulation F4 were plotted in zero-order, first-order, Higuchi,
626 Hixson-Crowell and Korsmeyer-Peppas models. The regression coefficient values of the
627 different model plots can be seen in Table 12 for both formulations. The data fitted best to the
628 zero-order model ($R^2 = 0.9992$). When the data were plotted against the Korsmeyer-Peppas
629 model, the n value (calculated from the slope of the curve) was 0.998 (Table 2) which
630 corresponded to a zero-order mechanism of release of MT-1207 from F4. Therefore, it can be
631 assumed that both the kinetics and mechanism of the release of MT-1207 from F4 tablets
632 were zero-order. The kinetic analysis findings were aligned with the swelling and erosion
633 study results which, being well correlated with the *in vitro* release profiles, also indicated an
634 erosion-controlled drug release from the F4 tablets.

635 [Table 12]

636 [Table 2]

637 3.10 In vivo animal study

638 The sustained-release and immediate-release tablets of MT-1207 were cross-
639 administered orally in four Beagle dogs. Figure 8 illustrates the pharmacokinetic profiles of
640 MT-1207 in the dog plasma after oral administration of one MT-1207 30 mg sustained-

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3 641 release tablet, three 10 mg immediate-release tablets with 8-hour interval between each and
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5 642 three 10 mg immediate-release tablets administered at one time. The pharmacokinetic
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7 643 parameters recorded on the respective dosing regimens in Beagle dogs can be seen in Table 3.
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9
10 644 A sustained pharmacokinetic profile of MT-1207 can be seen after oral administration of the
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12 645 sustained-release tablet, compared to the pharmacokinetic profiles of the IR tablets. There
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14 646 was a statistically significant decline (t-test, $p < 0.05$) in the C_{\max} value from 62.5 ng/mL
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16 647 (recorded in 3×10 mg IR o.d.) to 37.01 ng/mL for 30 mg SR o.d. This, along with the
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18 648 increase recorded in T_{\max} value from 1 and 1.63 hours (10 mg IR t.i.d. and 3×10 mg IR o.d.,
19
20 649 respectively) to 4.38 hours in 30 mg SR indicates a sustained release of the drug, along with
21
22 650 no evidence of dose dumping after the oral administration of sustained-release tablets
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24 651 (Defang et al. 2005). Hence, HPMC K15M was able, as indicated in the *in vitro* data, to
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26 652 provide a sustained release of MT-1207 from the tablets *in vivo*. The drug level remained at
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28 653 similar concentration values for about 3 hours (3-6 hours after administration) before
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30 654 declining, which indicates the prevalence of drug elimination over absorption. With regards
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32 655 to the extent of absorption, it was noted that in all the above-mentioned cases the AUC_{0-24h}
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34 656 and $AUC_{0-\infty}$ values recorded were statistically the same (one-way ANOVA, $p > 0.05$) (Table
35
36 657 3). In addition to this, the bioequivalence between the sustained-release and immediate-
37
38 658 release tablets was investigated by calculating the AUC_{0-24h} ratio of the sustained-release
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40 659 tablet dosing regimen and the immediate-release tablet dosing regimens. As per the EMA
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42 660 guidelines, the prerequisite for bioequivalence is that the AUC_{0-t} ratio of the test and
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44 661 reference products should fall within the range 80.00-125.00% (CPMP/EWP/QWP/1401/98
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46 662 Rev. 1/ Corr 2010). The AUC_{0-24h} ratios of the sustained-release tablet and the t.i.d. and o.d.
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48 663 immediate-release tablet dosing regimens were 98.12% and 90.05%, respectively. Based on
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50 664 these findings, it can be assumed that the whole amount of drug was released from the
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3 665 sustained-release tablets *in vivo* and was absorbed to the same extent as when the same dose
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5 666 was administered in immediate-release tablets, irrespective of the dosing regimen.
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9 667 **[Figure 8]**

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11 668 The distribution of MT-1207 in the body is an important parameter considered during
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13 669 pharmacokinetic analysis. The apparent volume of distribution, V_d , reflects the space of the
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15 670 body seemed to be occupied by the drug (Shargel et al. 2012). V_d increases with increasing
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17 671 distribution of the drug to peripheral tissues. The mean residence time, MRT, reflects the
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19 672 average time during which the drug resides in the body. In the present case, even though the
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21 673 SR tablet showed the highest value for the apparent volume of distribution which was about
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23 674 2.5 times higher than the IR tablet, there seemed to be no statistically significant difference
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25 675 (one-way ANOVA, $p > 0.05$) between the V_d values of MT-1207 recorded in SR o.d., IR
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27 676 t.i.d. and $3 \times$ IR o.d (Table 3). Furthermore, with regards to MRT, the value obtained for $3 \times$
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29 677 IR o.d. increased more than 2 times for the same dose of MT-1207 administered as a
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31 678 sustained-release tablet (Table 3) and the difference between the two values was deemed to
32
33 679 be statistically significant (t-test, $p < 0.05$). This indicates that the time during which the drug
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35 680 resides in the body was increased significantly when MT-1207 was administered in the form
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37 681 of a sustained-release tablet, compared to immediate-release tablets. There was a statistically
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39 682 significant difference in MRT between all three groups (one-way ANOVA, $p < 0.05$).
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46 683 The half-life, $t_{1/2}$, of MT-1207 was prolonged to 3.44 hours when the sustained-release
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48 684 tablet was administered in the dogs, 2 to 2.5 times higher than when three immediate-release
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50 685 tablets administered at the same time (Table 3). When immediate-release tablets were
51
52 686 administered at 8-hour intervals the MT-1207 $t_{1/2}$ was similar to that recorded after
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54 687 administration of the sustained-release tablet. With regards to clearance, there was no
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3 688 statistically significant difference (one-way ANOVA, $p > 0.05$) between the clearance values
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5 689 that were recorded in Beagle dogs for the three different dosing regimens (Table 3).
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8 690 Hepatic metabolism is generally higher in Beagle dogs than in humans (Boxenbaum
9 691 1982; Shargel et al. 2012). Phase I clinical trials were carried out for MT-1207 at a certain
10 692 dose range. At the 30 mg dose, the $t_{1/2}$ value of MT-1207 was 7.29 hours in humans (2020
11 693 email from Shenyang Haiwang Biotechnology Co. Ltd to us; unreferenced). Considering that
12 694 formulation A1 gave a $t_{1/2}$ in Beagle dogs that was 2-2.5 times higher than that of the
13 695 immediate-release tablets, the formulation could be considered as a once-a-day treatment,
14 696 provided that a similar increase in $t_{1/2}$ occurs in humans.
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25 697 **[Table 3]**

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28 698 **4. Conclusions**

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30 699 In the present study, a new sustained-release tablet for the novel antihypertensive drug
31 700 MT-1207 was successfully developed. HPMC K15M was able to provide a sustained release
32 701 of MT-1207 *in vitro* and in an animal model. Amongst the different formulations, the
33 702 optimised formulation (F4) containing 31% w/w HPMC K15M was chosen for a study in
34 703 Beagle dogs since it provided a 24-hour *in vitro* release of MT-1207 that demonstrated a
35 704 sustained release of the drug. The half-life of the drug was extended, while the amount of
36 705 drug absorbed was the same as that of the immediate-release tablets. Based on the
37 706 pharmacokinetic data from Beagle dogs and phase I clinical trials, this formulation can be
38 707 considered for once-a-day treatment of hypertension, an advance which could have
39 708 significant commercial and patient health benefits.
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54
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56
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59
60

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12
13 717 scanning and electron microscope that was used in the present study.
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18 718 **Declaration of interest**

19
20 719 The authors declare that Shenyang Haiwang Biotechnology Co., Limited which funded the
21
22 720 present study is the patent holder of MT-1207 agent.
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Ingredients (mg)	Formulation batch				
	F1	F2	F3	F4	A1
MT-1207 Hydrochloride	20	20	20	20	30
Alpha-D-lactose monohydrate	140	140	130	110	100
Polyvinylpyrrolidone K30	20	20	20	20	20
HPMC K15M	-	40	50	70	70
HPMC K4M	40	-	-	-	-
Colloidal silicon dioxide	4	4	4	4	6
Magnesium stearate	0.5	0.5	0.5	0.5	0.5
Tablet weight	224.5	224.5	224.5	224.5	226.5

Models	Kinetic constant	Regression coefficient
First-order	0.142	0.9835
Zero-order	0.001	0.9992
Higuchi	22.479	0.9849
Hixson-Crowell	0.103	0.9689
Korsmeyer-Peppas		
Kinetic constant	Regression coefficient	Exponent (<i>n</i>)
4.586	0.9985	0.98

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Table 3: Pharmacokinetic parameters of MT-1207 after oral administration of a single MT-1207 30 mg sustained-release tablet (30 mg SR o.d.), 10 mg immediate-release tablets (one tablet at a time) three times a day (10 mg IR t.i.d.) and three 10 mg immediate-release tablets administered at the same time (3 × 10 mg IR o.d.) to Beagle dogs (n=4).

Dosing Regimen	$t_{1/2}$ (h)	T_{max} (h)	C_{max} (ng/mL)	AUC_{0-24h} (h*ng/mL)	$AUC_{0-\infty}$ (h*ng/mL)	V_d (L/kg)	CL (L/h/kg)	MRT (h)
30 mg SR o.d.	3.44±1.49	4.38±2.14	37.01±8.29	213.6±59.8	222.1±56.4	723.3±448.1	140.9±31.1	6.40±1.09
10 mg IR t.i.d.	3.47±1.49	1.00±0.71	32.22±23.28	217.7±81.6	229.7±86.7	540.1±247.0	116.8±57.3	13.05±3.77
3 × 10 mg IR o.d.	1.69±0.20	1.63±0.25	62.50±18.65	237.2±53.8	250.8±57.8	299.7±58.8	124.1±26.5	2.84±0.27

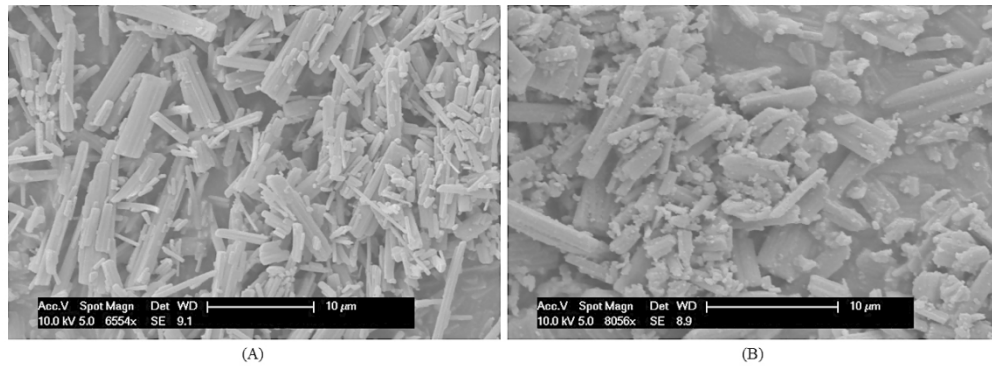


Figure 1: SEM image of (A) MT-1207 reference standard and (B) micronised MT-1207:silicon dioxide 5:1 mixture.

32x12mm (1000 x 1000 DPI)

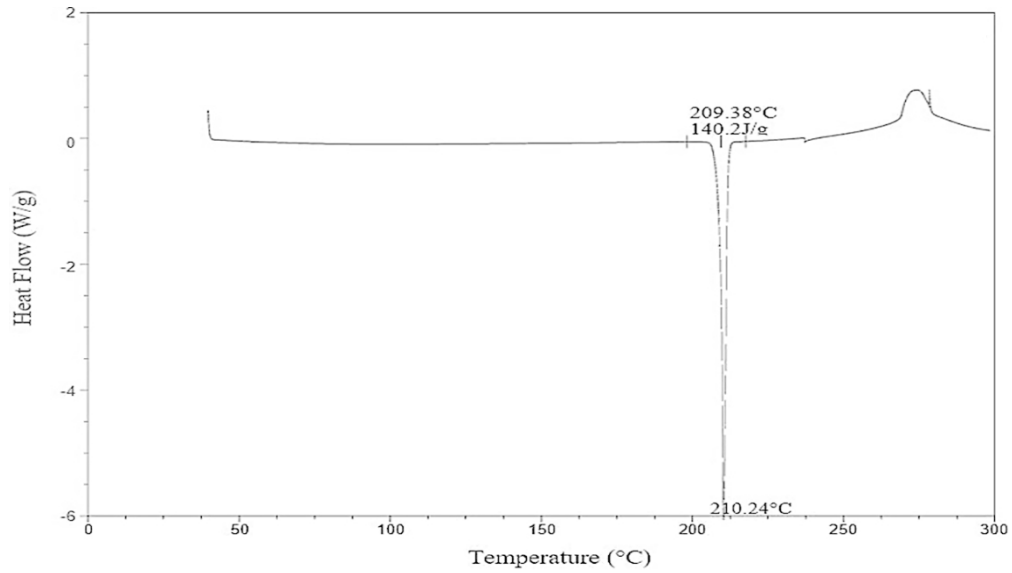


Figure 2: DSC thermogram of MT-1207 hydrochloride reference standard.

32x18mm (1000 x 1000 DPI)

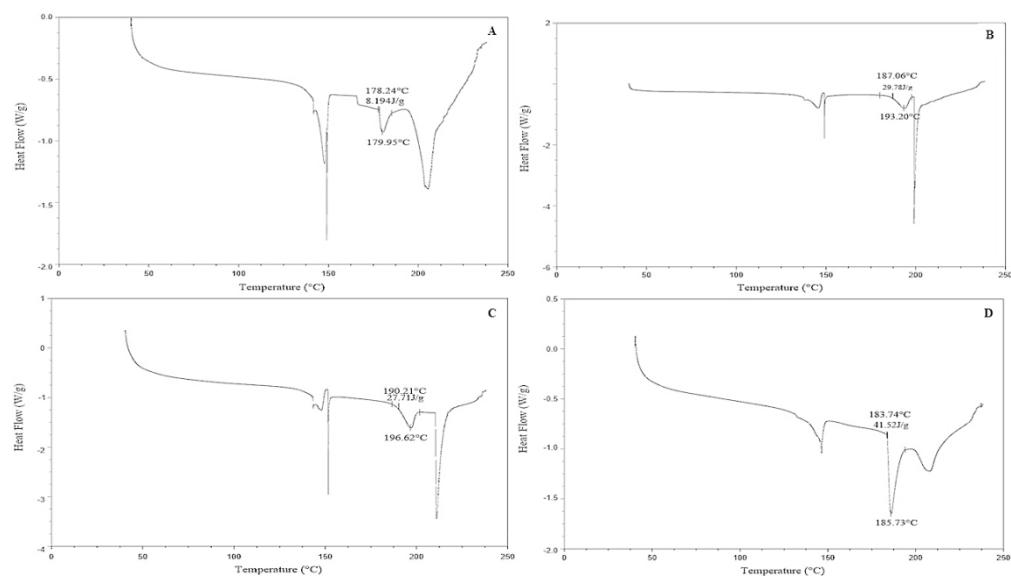


Figure 3: DSC thermograms of (A) Formulation F1, (B) Formulation F2, (C) Formulation F3 and (D) Formulation F4.

32x18mm (1000 x 1000 DPI)

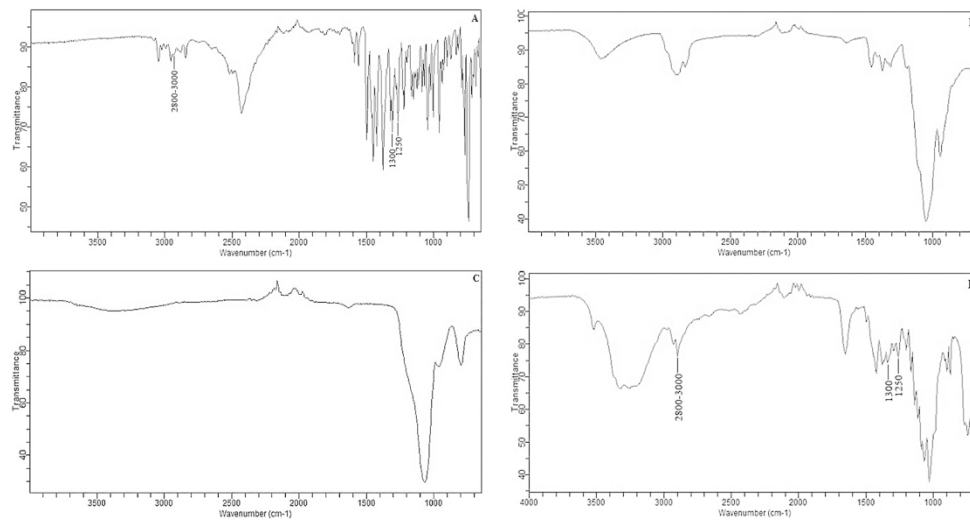


Figure 4: Fourier-Transform Infrared (FTIR) spectra of (A) MT-1207 hydrochloride reference standard, (B) HPMC K15M, (C) colloidal silicon dioxide and (D) pulverised tablets [Transmittance vs Wavenumber (cm⁻¹)].

51x26mm (1000 x 1000 DPI)

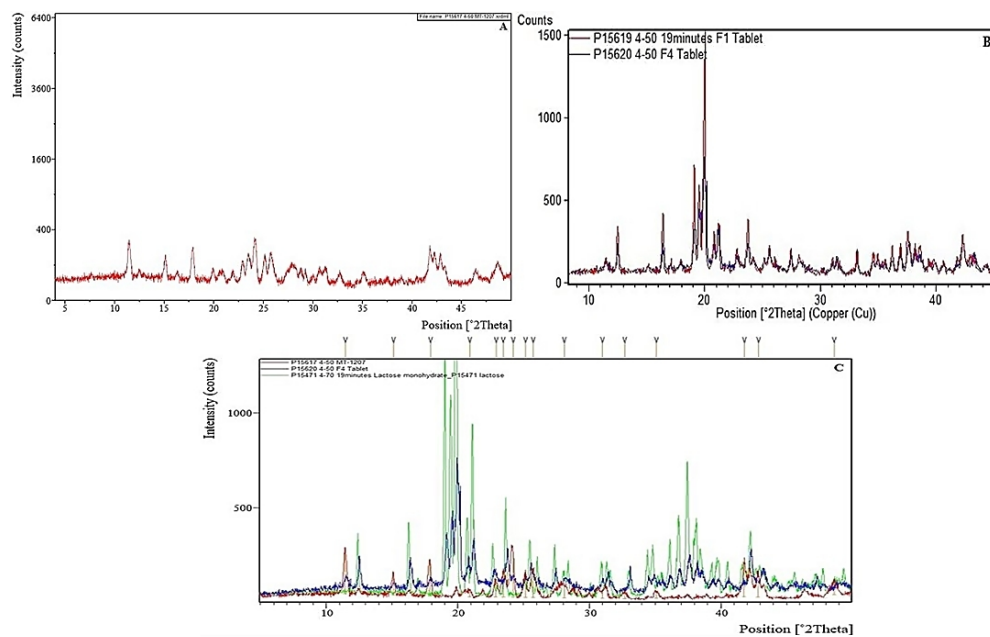


Figure 5: Diffractograms of (A) MT-1207 hydrochloride reference standard, (B) formulations F1 (red) and F4 (blue) and (C) MT-1207 hydrochloride reference standard (red), F4 (blue) and alpha-D-lactose monohydrate (green) combined [Intensity (Counts) vs Position($^{\circ}$ 2Theta)].

27x17mm (1000 x 1000 DPI)

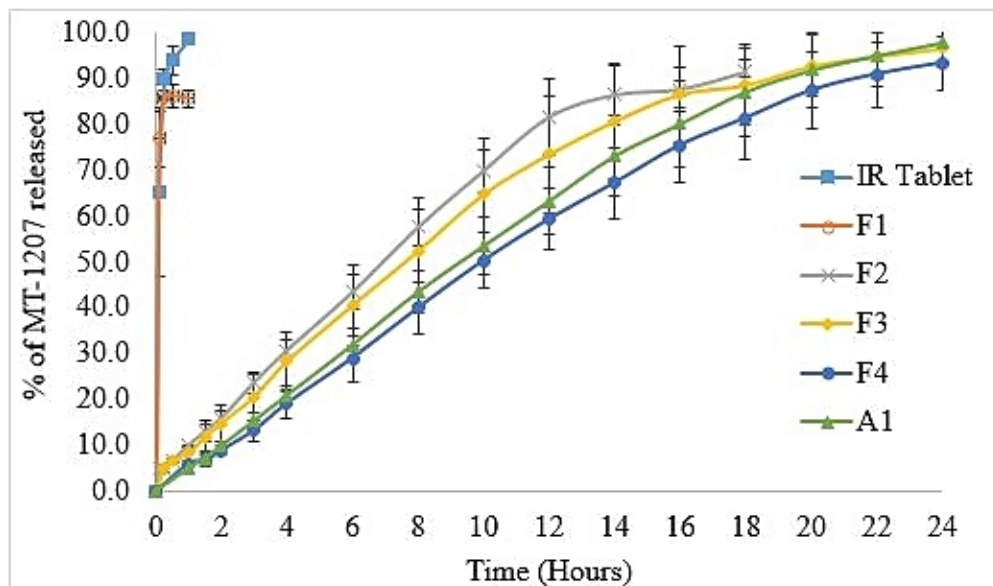


Figure 6: *In vitro* release profiles of MT-1207 from immediate-release tablets, formulations F1, F2, F3, F4 and A1 in 0.1 M phosphate pH 6.8 with 0.2% w/v SDS (n=3). The results are presented as mean \pm standard deviation.

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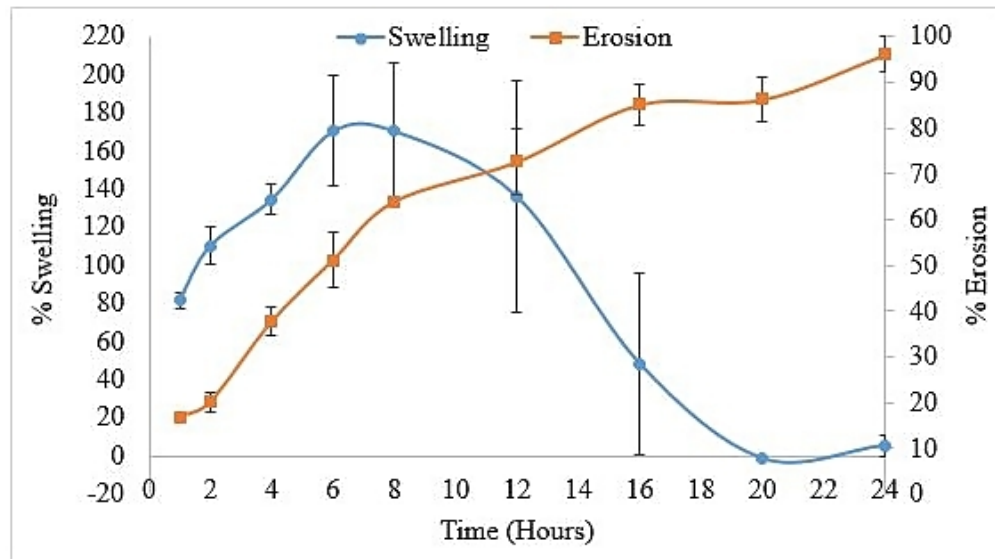


Figure 7: *In vitro* swelling and erosion profiles of formulation F4 in 0.1 M phosphate pH 6.8 with 0.2% w/v SDS (% swelling and % erosion vs time, n=3). The results are presented as mean \pm standard deviation.

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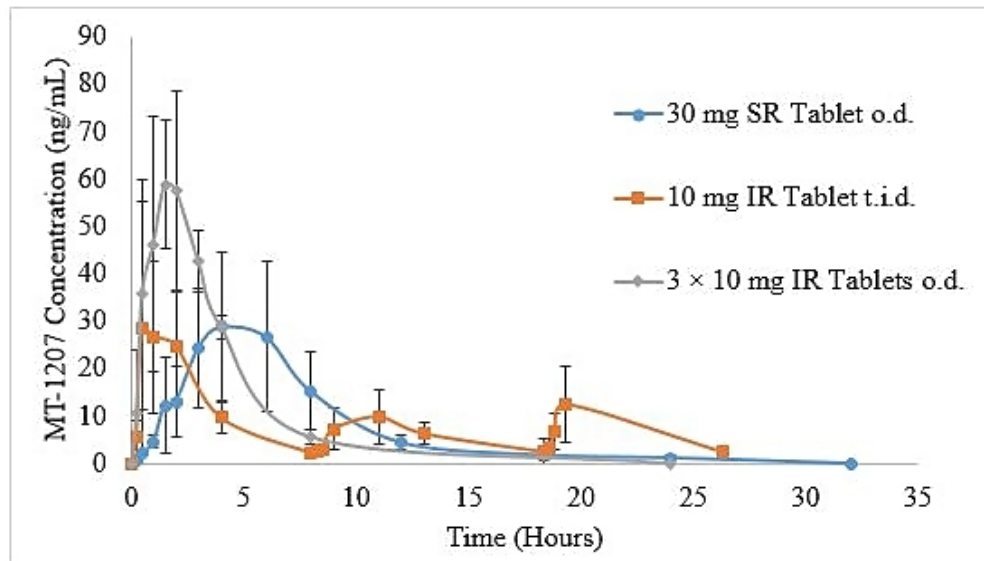


Figure 8: Plasma concentration of MT-1207 in Beagle dog after oral administration of 30 mg SR tablet o.d., 10 mg IR tablet t.i.d. and 3 x 10 mg IR tablet o.d. The results are presented as mean \pm standard deviation.

14x7mm (1000 x 1000 DPI)

Supplementary materials for

***In vitro* and *in vivo* evaluation of a sustained-release once-a-day formulation of the novel antihypertensive drug MT-1207**

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This file includes:

Table S1: Mean solubility values of non-micronised MT-1207 hydrochloride reference standard and micronised MT-1207 hydrochloride:colloidal silicon dioxide 5:1 mixture in hydrochloric acid (HCl) pH 1.2 (n=9) and 0.1 M phosphate buffer pH 6.8 (n=9). Results are expressed as mean ± standard deviation.

Table S2: Flowability characterisation results.

Table S3: Tablet hardness testing results (n=10 tablets for each formulation). Results are presented as mean ± standard deviation.

Table S4: Tablet friability results.

Table S5: Uniformity of weight results (n=10 tablets per formulation).

Figure S1: *In vitro* release profiles of MT-1207 (% of MT-1207 released vs time) from the three F4 batches in 0.1 M phosphate pH 6.8 with 0.2% w/v SDS (n=3).

Table S1: Mean solubility values of non-micronised MT-1207 hydrochloride reference standard and micronised MT-1207 hydrochloride:colloidal silicon dioxide 5:1 mixture in hydrochloric acid (HCl) pH 1.2 (n=9) and 0.1 M phosphate buffer pH 6.8 (n=9). Results are expressed as mean \pm standard deviation.

Sample Name	MT-1207 Solubility (mg/mL)
Non-micronised MT-1207 hydrochloride in HCl pH 1.2	1.22 \pm 0.05
Micronised MT-1207 hydrochloride:colloidal silicon dioxide 5:1 mixture in HCl pH 1.2	1.34 \pm 0.08
Non-micronised MT-1207 hydrochloride in 0.1M phosphate pH 6.8	0.01
Micronised MT-1207 hydrochloride:colloidal silicon dioxide 5:1 mixture in 0.1M phosphate pH 6.8	0.03 \pm 0.01

Table S2: Flowability characterisation results.

Tablet Formulation	Compressibility (Carr) Index (%)	Hausner Ratio	Angle of repose (°)	Flowability (as per USP)
F1	14.7	1.17	34.2	Good
F2	23.5	1.31	33.2	Fair
F3	15.4	1.18	38.7	Fair
F4	12.2	1.14	42.0	Passable to Good
F4 Repeat 2	12.9	1.15	39.0	Good
F4 Repeat 3	14.0	1.16	35.6	Good
A1	13.3	1.15	36.4	Good

Table S3: Tablet hardness testing results (n=10 tablets for each formulation). Results are presented as mean \pm standard deviation.

Tablet Formulation	Diameter (mm)	Thickness (mm)	Hardness (kg)	Tensile Strength (kPa)
F1	8.01 \pm 0.03	3.51 \pm 0.03	9.18 \pm 0.86	2057.2 \pm 186.7
F2	7.98 \pm 0.04	3.49 \pm 0.02	10.79 \pm 0.50	2413.7 \pm 114.0
F3	7.99 \pm 0.03	3.52 \pm 0.04	11.69 \pm 0.58	2595.9 \pm 134.6
F4	7.99 \pm 0.03	3.50 \pm 0.07	13.89 \pm 1.70	3100.2 \pm 336.6
F4 Repeat 2	8.00 \pm 0.03	3.50 \pm 0.05	14.94 \pm 1.41	3332.0 \pm 287.4
F4 Repeat 3	7.99 \pm 0.03	3.50 \pm 0.02	13.18 \pm 1.51	2943.7 \pm 339.6
A1	7.98 \pm 0.04	3.52 \pm 0.06	14.81 \pm 0.40	3295.0 \pm 125.2

Table S4: Tablet friability results.

Tablet Formulation	% Friability
F1	0.18
F2	0.09
F3	0.06
F4	0.01
F4 Repeat 2	0.01
F4 Repeat 3	0.14
A1	0.04

Table S5: Uniformity of weight results (n=10 tablets per formulation).

Formulation	Mean Tablet		Acceptance Value
	Weight (% of label claim)	Standard Deviation	
F1	100.4	2.8	6.7
F2	100.1	3.8	9.2
F3	99.9	0.8	2.0
F4	100.2	1.4	3.4
F4 Repeat 2	99.8	1.3	3.1
F4 Repeat 3	99.6	3.8	9.2
A1	100.0	1.2	9.2

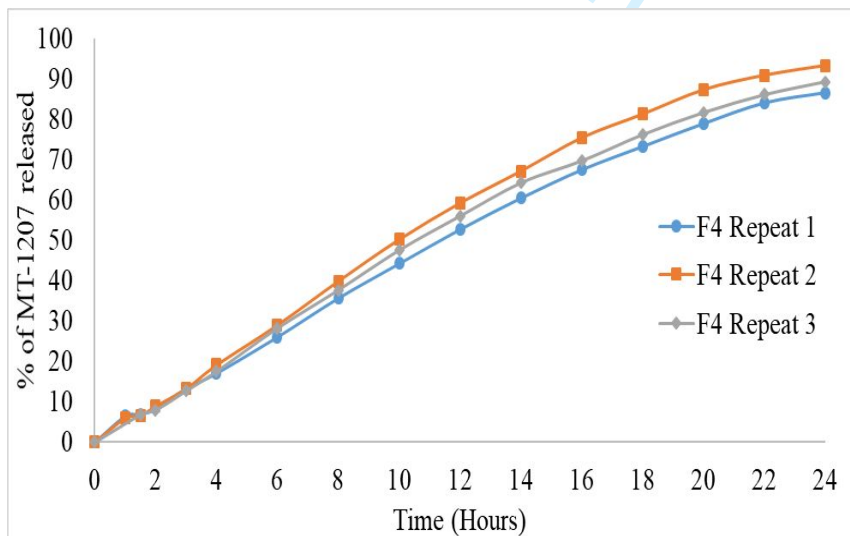
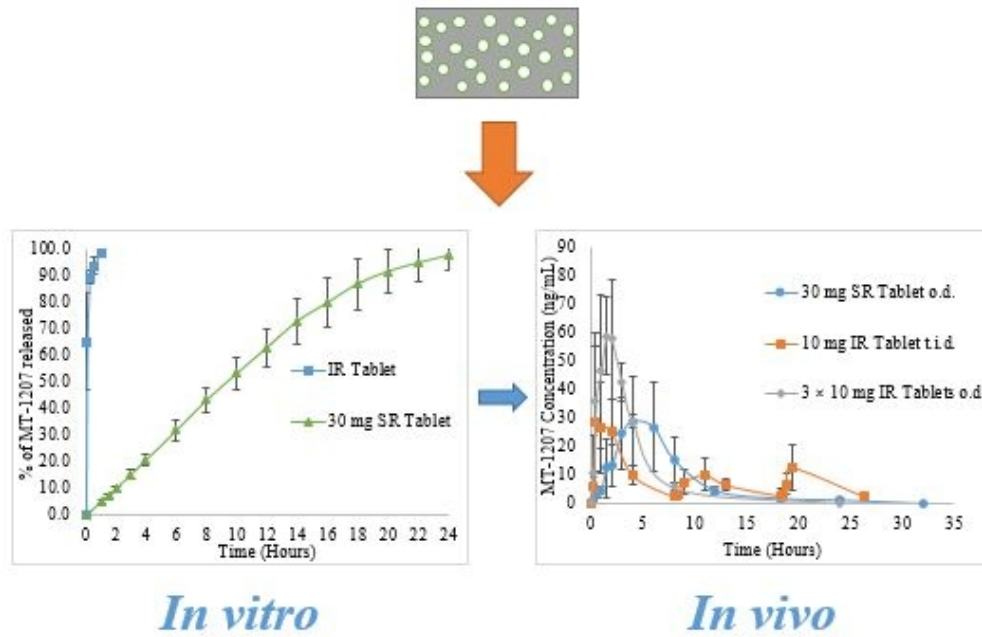


Figure S1: *In vitro* release profiles of MT-1207 (% of MT-1207 released vs time) from the three F4 batches in 0.1 M phosphate pH 6.8 with 0.2% w/v SDS (n=3).



139x89mm (96 x 96 DPI)