

The temperature dependence of greenhouse gas production from Central African savannah soils

Nicholas T. Girkin^{a,*}, Hannah V. Cooper^{a,1}, Alice S. Johnston^b, Martha Ledger^a, G.R. Mouanda Niamba^c, Christopher H. Vane^d, Vicky Moss-Hayes^d, Dafydd Crabtree^e, Greta C. Dargie^e, Saul Vasquez^a, Yannick Bocko^f, Emmanuel Mampouya Wenina^f, Mackline Mbemba^f, Arnoud Boom^g, Suspense Averti Ifo^f, Simon L. Lewis^e, Sofie Sjögersten^a

^a School of Biosciences, University of Nottingham, UK

^b Faculty of Engineering and Applied Sciences, Cranfield University, UK

^c Wildlife Conservation Society, Epena, Congo

^d British Geological Survey, Keyworth, UK

^e School of Geography, University of Leeds, Leeds, UK

^f École Normale Supérieure, Université Marien Ngouabi, 99324 Brazzaville, Congo

^g School of Geography, Geology & the Environment, University of Leicester, UK

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ABSTRACT

Savannahs cover 20 % of the global land surface, but there have been few studies of greenhouse gas (GHG) dynamics from savannah soils. Here, we assess potential turnover of carbon dioxide (CO₂), methane (CH₄) and nitrous oxide (N₂O) from surface (0–10 cm) and subsurface (20–30 cm) soils from two contrasting tropical savannah sites in the Republic of Congo, Central Africa, under dry (40 % water-filled-pore-space, WFPS) and wet (70 % WFPS) conditions. Under baseline conditions (25 °C), we found soils were sources of CO₂ and N₂O, but a sink for CH₄. Assessment of the temperature response of GHG fluxes between 20 and 35 °C revealed variable temperature dependences. That is, CO₂ fluxes showed a strong temperature response, whereas the temperature response of N₂O fluxes was only significant under dry conditions, and no significant temperature response of CH₄ fluxes was observed. The temperature quotient (Q₁₀) of soil respiration increased from 1.58 ± 0.004 to 1.92 ± 0.006 at sites with lower soil organic carbon contents. The relative increase in N₂O with CO₂ fluxes across temperatures was significantly influenced by moisture conditions at both sites. No temperature or soil moisture response was observed for CH₄ fluxes, collectively implying divergent GHG responses to changing climatic conditions. Using Rock-Eval pyrolysis we assessed the organic chemistry of all soil types, which indicated contrasting degrees of stability of carbon sources between sites and with depth which, alongside significant differences in a range of other soil parameters (including organic matter content, total carbon, total nitrogen, electrical conductivity, and pH), may account for site-specific differences in baseline GHG emissions. Taken together, our results are amongst the first measures of GHG temperature sensitivity of tropical savannah soils, and demonstrate that soil CO₂ emissions are more sensitive to warming and changes in moisture than the emissions of other GHGs, although relatively low compared to responses reported for soils from other tropical ecosystems. This implies that GHG fluxes from savannah soils in the region may be at least partially resilient to climate-induced soil warming compared to other ecosystems.

1. Introduction

Savannah ecosystems cover approximately 20 % of the global land surface (Pennington et al., 2018), with tropical savannahs alone

covering 27.6 million km² (Hutley and Setterfield, 2008). Savannahs are generally dominated by grass and small shrub species, have distinct wet and dry seasons (Pennington et al., 2018), and have frequent fire disturbance (Govender et al., 2006). In addition to removing living and

* Corresponding author.

E-mail address: nicholas.girkin@nottingham.ac.uk (N.T. Girkin).

¹ Contributed equally.

dead aboveground biomass, fires can significantly affect soil properties (Amoako and Gambiza, 2019), which may in turn affect the rates of greenhouse gas (GHG) production.

Savannahs will undergo significant changes by 2100 due to a combination of warming, altered precipitation patterns, fire regimes, land use change, and agricultural intensification. Current estimates of air temperature changes across the tropics are of 3–4 °C warming by 2100 but potentially up to 6 °C for Central Africa, alongside alterations in precipitation (IPCC, 2021). These changes are likely to drive significant alterations in the rates of GHG production and net emissions, with the potential for significant feedback loops from land-atmosphere exchange.

Governing the response of soil respiration and decomposition rates are thermodynamic and kinetic relationships, as described by the Arrhenius and Michaelis-Menten equations, respectively. The Arrhenius equation (Eq. (1)), broadly predicts a substrate of low-quality will require higher amounts of energy to be degraded, and therefore its rate of decomposition is slower (Fierer et al., 2005; Lloyd and Taylor, 1994). As the net activation energy (E_a) required for decomposition (R_T) increases, the temperature dependence of decomposition should also increase as a result, leading to an inverse relationship between organic matter quality and temperature sensitivity (Gershenson et al., 2009).

$$\ln(R_T) = B e^{-E_a/kT} \quad (1)$$

Where R_T is the reaction rate at a specified temperature (T , in Kelvins), B is a normalization constant independent of temperature and the Boltzmann-Arrhenius factor, $e^{-E_a/kT}$, describes the temperature-dependence of reaction rate, where E_a is the activation energy (eV) and k is Boltzmann's constant ($8.62 \times 10^{-5} \text{ eV K}^{-1}$) (Davidson and Janssens, 2006). While many previous studies support this, other studies have reported equivalent values for both thermally stable ("refractory") and labile pools or a lower temperature dependence for recalcitrant organic carbon (Fang et al., 2005; Fierer et al., 2005; Giardina and Ryan, 2000; Leifeld and Fuhrer, 2005). Underpinning the relationship between substrate availability and the temperature dependence of decomposition pathways is the Michaelis-Menten equation (Davidson and Janssens, 2006; Michaelis et al., 1913):

$$R = \frac{V_{\max}[C]}{K_m + [C]} \quad (2)$$

Where R is the rate of soil respiration, V_{\max} is the maximum rate of enzyme activity, $[C]$ is the concentration of available carbon, and K_m is the dissociation constant. Both V_{\max} and K_m are temperature dependent. When systems are not kinetically limited i.e. there is no substrate limitation, the temperature dependence of soil respiration reduces to that of V_{\max} but under substrate limiting conditions, the temperature dependence of K_m becomes increasingly important, and the overall sensitivity of respiration decreases (Gershenson et al., 2009). Based solely on the carbon quality hypothesis, temperature dependence should increase with increasingly recalcitrant substrates, and decrease with depth due to reduced labile carbon availability.

Previous studies of the temperature dependency of GHG emissions from tropical soils are limited. Prevailing expectations are that the temperature dependency of GHG production is lower in the tropics than in cooler higher latitudes (Bekku et al., 2003). However, recent findings suggest the potential for substantial feedback in GHG emissions following warming in the tropics. Recent in situ warming of a Central American forest soil showed 4 °C of soil warming increased CO₂ emissions by 55 % (Nottingham et al., 2020).

A range of Q_{10} values (which describes the relative change in soil respiration due to an increase in temperature of 10 °C and is used as a proxy to quantify the feedback intensity of soil respiration to increasing temperatures) have been reported for tropical soils. For example, studies in tropical peats, which feature a large labile carbon pool report Q_{10} values of approximately 1.4–2 for CO₂ production, 1.4–6.8 for methanogenesis, and 2.7 for methanotrophy (Girkin et al., 2020a, 2020b;

Sjögersten et al., 2018). Q_{10} values for CO₂ emissions in tropical mineral soils range from 2.1 to 2.7 (Bekku et al., 2003), while equivalent Q_{10} values for CO₂ production in savannah soils specifically, include 1.9 for a North Australian savannah soil (Chen et al., 2002), 2.1 in Senegalese savannahs, (Elberling et al., 2003), 4.9 from a woody Brazilian savannah (da Rocha et al., 2002), and up to 5.0 for a burnt Brazilian savannah soil (Poth et al., 1995). Q_{10} values for N₂O fluxes are much less frequently assessed, but include Q_{10} values of 2.8–3.6 for a temperate forest (Bagherzadeh et al., 2008), and 0.8–1.2 for mountain forest and meadow ecosystems in subtropical China (Zhang et al., 2016).

In this study, we assess the temperature dependency of GHG emissions from two tropical savannah soils from the Central African basin under wet and dry moisture states. We subsequently assess the extent to which soil properties predict the temperature dependency of GHG dynamics. We hypothesised that i) Baseline soil GHG emissions would differ between savannah soil types with greater rates in soils with greater labile carbon stocks; and that ii) temperature dependency would differ between sites and surface and subsurface soils, with greater temperature dependency in soils with more recalcitrant carbon.

2. Methods

2.1. Study sites

This study was conducted using soils collected from two savannah sites in Likouala province, Republic of Congo. The soils were collected during a field campaign between January – March 2019. Soils were collected near the villages of Ekolongouma (1.204 N, 17.909E) and Itanga (1.2025 N 17.4345E) (Fig. 1).

The Ekolongouma village site is a seasonally flooded savannah adjacent to the Ubangi River, dominated by *Poaceae* sp. The site was adjacent to seasonally flooded forest where vegetation was predominantly confined to higher ground *Trichilia welwitschi*, *Cola* sp., *Strombosloopsis tetrandra*, *Guibourtia demeusei* and *Dialium pachyphilum* (Dargie, 2015).

The Itanga village site bordered the Likouala-aux-Herbes River. Close to the river, the site comprised seasonally flooded savannah and was dominated by *Poaceae* sp. and featured a largely flat microtopography. After approximately 250 m, the site transitioned to a dominance of *Hyparenia diplandra*. After approximately 750 m, the site transitions to a seasonally flooded forest with an increasing abundance of *Raphia hookeri* canopy palms (Dargie, 2015).

At each site, four paired replicate surface (0–10 cm) and sub-surface (20–30 cm) soil samples were collected, with sub-surface samples collected directly under surface samples. Due to logistical limitations arising from the remote nature of the sites and the need to carry all collected material, soils from 10 to 20 cm were excluded. All collected samples were transported to the University of Nottingham, UK for analysis, where they were stored at 4 °C. Samples were collected by hand, with surface litter removed first, before soils were placed in zip-lock bags. Samples were located up to 100 m apart and collected from the approximate centre of both savannah sites to reduce edge effects from contrasting vegetation inputs and local village activities.

2.2. Soil properties

Gravimetric water content was determined from the mass of water lost from 10 g wet weight soil oven dried at 105 °C for 24 h. Soil organic matter content was determined as the mass lost after ignition for 7 h at 550 °C. Total carbon and total nitrogen were measured using a LEICO total elemental analyser. Electrical conductivity and pH were measured in soils extracts at a 1:5 ratio with deionised water.

Soil sub-samples were analysed using a Rock-Eval 6 analyser. Freeze-dried powdered soil samples (60 mg) were heated at 300 °C for three minutes before an increase in temperature to 650 °C at a rate of 25 °C per minute in an inert N₂ atmosphere. Residual carbon was oxidized from

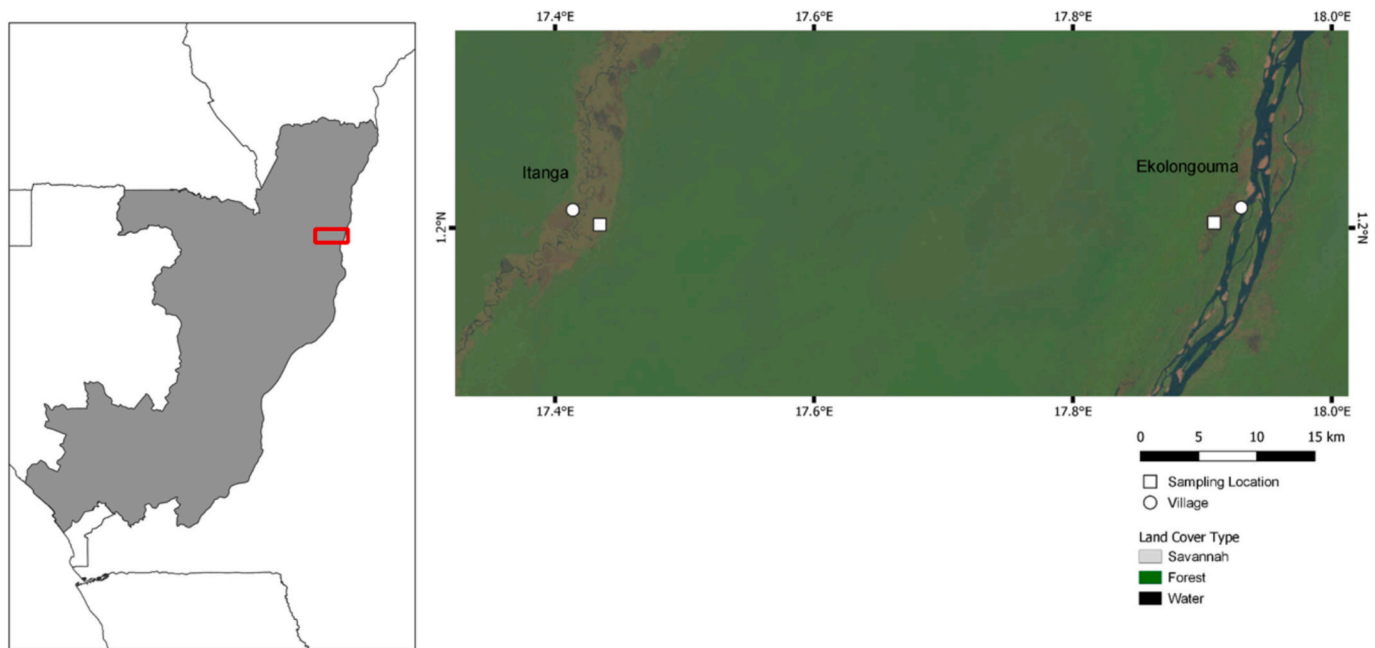


Fig. 1. Locations of Ekolongouma and Itanga savannah sites in the Republic of Congo.

300 °C to 850 °C at a rate of 20 °C per minute with the release of hydrocarbons during the two-stage pyrolysis process detected by a flame ionization detector, with an infrared cell detecting the release of CO and CO₂ during the thermal cracking of the organic matter.

Rock-Eval analysis produces a range of standard parameters including S1 (a measure of free hydrocarbons released on heating to 300 °C), S2 (hydrocarbons released on the thermal cracking of organic matter for temperatures up to 650 °C), S3CO and S3CO₂ (the CO and CO₂ yielded from the breakdown of kerogen), Tpk2 (the temperature associated with the highest yield of bound hydrocarbons) and total organic carbon (TOC_{RE}). The Hydrogen Index (HI, mg HC g⁻¹ TOC_{RE}), a measure of hydrocarbons released relative to TOC was calculated as:

$$S2 \times 100 / \text{TOC}_{\text{RE}} \quad (3)$$

The Oxygen Index (OI, mg O₂ g⁻¹ TOC_{RE}), corresponding to the amount of oxygen released as CO₂ relative to TOC_{RE} was calculated as:

$$S3 \times 100 / \text{TOC}_{\text{RE}} \quad (4)$$

A set of indices, I (immature) and R (refractory), describing the preservation of thermally labile and highly thermostable organic matter respectively, were calculated using the approach proposed by Sebag et al. (2016) using the integrated areas under the S2 curve between given temperature nodes. Areas were calculated as 200–340 °C (A1, labile biopolymers), 340–400 °C (A2, resistant biopolymers), 400–460 °C (A3, immature geopolymers), and > 460 °C (A4, mature geopolymers). The I-index was calculated as $\log((A1 + A2)/A3)$ and the R-index was calculated as $(A3 + A4)/100$ and represents the contribution of mature organic matter to the S2 signal (Sebag et al., 2016). This set of indices has previously been applied to assessments of carbon thermostability in tropical forest peat soils (Cooper et al., 2019; Girkin et al., 2019; Girkin et al., 2020a, 2020b) but not tropical mineral savannah soils. Fresh and immature organic matter is characterised by a low R-index (i.e. low levels of refractory carbon), and a high I-index (increased labile carbon), with the inverse true for older more recalcitrant organic matter (Sebag et al., 2016).

2.3. Potential greenhouse gas emissions

Soil GHG emissions were measured across four temperatures (20, 25,

30 and 35 °C) and at two moisture states using an adapted approach previously described in Sjögersten et al. (2018), Girkin et al. (2020a, 2020b) and Dhandapani et al. (2020) developed for other tropical soils. In brief, soil samples (10 g dry weight equivalent of homogenised, sieved and repacked soil compacted to field bulk density (approximately 1.1 g cm⁻³) were placed in 64 (2 sites × 2 depths × 4 replicates × 4 temperatures) glass serum bottles, each with a volume of 120 mL (Kinesis, St. Neots, UK). Soils were rewetted so all reached 20 % gravimetric moisture content, corresponding to approximately 40 % water-filled pore space (WFPS). These conditions represented the mean gravimetric moisture content of collected soil samples (Table 1). Soils were then maintained at this moisture content by regular weighing, with weights monitored daily during the sampling period. Gas sampling was carried out before rewetting of soil samples (Somers et al., 2019). Four replicates of each soil type were placed in 20–35 °C incubators for one month for acclimation prior to the first sampling event.

On each sampling day, bottles were removed from their incubators and placed on an orbital shaker for one hour to allow the exchange of headspace gases. Ten replicate air samples (20 mL) were collected from randomly selected unsealed bottles by syringe and injected at over-pressure into pre-evacuated 12 mL glass silica gas vials (LABCO, UK). Bottles were then sealed with rubber butyl stoppers (13 × 19 × 12 mm; Rubber B.V., Hilversum, NL). Bottles were then returned to their incubators for 1 h after which a 20 mL gas sample was collected for analysis using a gas chromatograph (Shimadzu, 2014). CO₂, CH₄ and N₂O concentrations of gas samples were measured using a single injection system with a 1 mL sample loop using N₂ as a carrier gas through a non-polar methyl silicone capillary column (CBP1-W12–100, 0.53 mm I. D., 12 m, 5 mm) and a porous polymer packed column (HayeSep Q 80/100). Thermal conductivity, flame ionization, and an electron capture detector were used to measure CO₂, CH₄ and N₂O, respectively (Girkin et al., 2020b).

After measuring fluxes at 20 % gravimetric moisture content on three occasions over four weeks, soils were further rewetted to 30 % gravimetric moisture, corresponding to approximately 70 % WFPS. Following rewetting, soils were left open to the atmosphere for one month (while being maintained at constant soil moisture content) before measurement of GHGs, to allow acclimation and to minimise any impact of the Birch effect (increased carbon and nitrogen mineralisation following

Table 1Surface and subsurface soil biogeochemical properties for Ekolongouma and Itanga. Means ± 1 SE ($n = 4$). * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$.

	Ekolongouma		Itanga		Depth	Site	Depth*Site
	Surface	Subsurface	Surface	Subsurface			
Gravimetric moisture (%)	28.7 \pm 1.6	29.3 \pm 0.7	79.9 \pm 4.7	73.5 \pm 1.9		***	
Organic matter (%)	16.3 \pm 0.5	15.3 \pm 0.5	52.8 \pm 8.2	48.3 \pm 9.8		***	
pH	4.1 \pm 0.04	4.3 \pm 0.1	3.1 \pm 0.03	3.1 \pm 0.02		***	
Conductivity (μ S)	8.6 \pm 0.6	7.8 \pm 0.2	100.2 \pm 17.5	75.4 \pm 6.9		***	
Total carbon (%)	6.2 \pm 0.5	5.2 \pm 0.4	35.4 \pm 1.0	33.5 \pm 1.7		***	
Total nitrogen (%)	0.3 \pm 0.02	0.3 \pm 0.02	1.5 \pm 0.1	1.3 \pm 0.1	*	***	
TOC (%)	5.3 \pm 0.4	4.7 \pm 0.4	23.2 \pm 6.6	28.6 \pm 2.3		***	
HI (Hydrogen Index; mg HC g ⁻¹ TOC _{REG})	174 \pm 10.2	155 \pm 6.7	135.3 \pm 20.2	125.5 \pm 13.2		*	
OI (Oxygen Index; mg O ₂ g ⁻¹ TOC _{REG})	232 \pm 11.5	242.8 \pm 12.1	152 \pm 3.7	130.3 \pm 3.6		***	
I (Immature) Index	0.3 \pm 0.01	0.3 \pm 0.03	0.2 \pm 0.02	0.1 \pm 0.03		*	***
R (Refractory) Index	0.6 \pm 0.01	0.5 \pm 0.02	0.6 \pm 0.01	0.6 \pm 0.02			

rewetting) on fluxes (Lopez-Sangil et al., 2018).

Fluxes were calculated using the concentration difference between the initial headspace concentrations compared to those measured after one-hour incubation, according to the ideal gas law (Girkin et al., 2018). These calculations also accounted for the volume of soil occupied by soil and water within the bottle. In total 375 greenhouse gas measurements were made during this study.

2.4. Data and statistical analysis

The temperature dependence of CO₂, CH₄ and N₂O fluxes were first assessed by applying general linear models to flux measurements across the temperature range to solve the parameters in Eq. (1) (Johnston and Sibly, 2018). GHG fluxes were standardised prior to analyses in R v4.3.2 (R Core Team, 2023) to enable application of Eq. (1) to negative GHG flux measurements for CH₄ and N₂O, alongside CO₂ following: $\ln(R_T/\text{mean}(R_T))$. Arrhenius temperatures were similarly standardised followed: $1/kT_c - 1/kT$, where T_c is the mean temperature (300.65 K: 27.5 °C).

First, all GHG flux measurements were analysed according to Eq. (1) to establish the temperature dependence and sensitivity across site, soil depth and moisture conditions. Relationships between GHG fluxes were also analysed, using one-way ANOVA and $p < 0.05$ to test their significance. For each GHG flux, we then explored whether temperature responses interacted significantly with additional predictors in the order: site, moisture, depth. For all models, a better goodness of fit to the GHG flux measurements depended on additional degrees of freedom (df) resulting in a difference in Akaike Information Criterion (Δ AIC) value >5 compared to the model with temperature as the only predictor. This condition ensures parsimony when additional predictor variables are selected (Johnston et al., 2021). Following regression analyses, we used B and E_a parameters for those sites and GHG fluxes that yielded significant Arrhenius temperature interactions ($p > 0.05$) to estimate Q_{10} values. As Q_{10} values are the proportional increase in non-logged GHG flux (R) with a 10 °C increase in temperature ($T + 10$), we calculated R values between the observed temperature range of 20 and 35 °C (T_{20-35}) and estimated Q_{10} as:

$$Q_{10} = R(T_{+10})/R(T_{20-35}) \quad (5)$$

We also tested relationships between non-standardised GHG fluxes and their interaction with predictor variables such as site, depth, moisture and temperature groupings. Differences between soil properties and GHG exchange between 0 and 35 °C were assessed using a mixed effects model fitted using Residual Maximum Likelihood (REML) to account for variable dependence. The model included sampling time point and soil replicates as random effects, and depth and site as fixed effects. Differences in baseline GHG fluxes (defined as the rate of emissions at ambient temperature (25 °C) under both moisture states were investigated using Tukey t -tests for pairwise comparisons between the two sites, depths and moisture conditions. Relationships between

key environmental variables and mean GHG emissions were assessed using Principal Component Analysis (PCA), based on correlation matrices.

3. Results

3.1. Soil properties

A range of soil properties varied significantly between Ekolongouma and Itanga, but to a lesser extent between the surface and subsurface (Table 1). Soils from Ekolongouma had relatively low water content (approximately 29 %), with high organic matter content (15.2–16.3 %), acid pH (4.1–4.3), and low electrical conductivity (51.7–6.2 μ S). Total carbon and nitrogen were in the range of 5.2–6.2 % and 0.3 %, respectively. In contrast, soils from Itanga had significantly greater gravimetric moisture (73.5–79.9 %), more acidic pH (3.1–3.1), higher electrical conductivity (75.4–100.2 μ S), and total carbon and nitrogen (33.5–35.4 %, 1.3–1.5 % respectively). Ekolongouma soils had higher HI (hydrogen index) and OI (oxygen index) values, compared to soils from Itanga.

3.2. Baseline greenhouse gas emissions

Baseline CO₂ emissions (CO₂ emissions from dry and wet soils at 25 °C) ranged from 4 to 29 μ g CO₂ g⁻¹ h⁻¹ across both sites and depths. CO₂ emissions were substantially higher from surface compared to subsurface soils from Itanga but was broadly comparable between Ekolongouma soils ($p = 0.046$, Fig. 2a).

Both sites were consistent sinks for CH₄ under both dry and wet conditions, indicating CH₄ oxidation (Fig. 2b), with uptake ranging from 0.7 to 4.1 ng CH₄ g⁻¹ h⁻¹. For Ekolongouma soils, oxidation was greater in surface than subsurface soils, but for Itanga, mean subsurface oxidation was greater ($p = 0.043$). Overall, CH₄ oxidation was higher in Itanga soils ($p = 0.002$).

Soils ranged from a source to a small sink for N₂O (−0.2–2.5 ng N₂O g⁻¹ h⁻¹). Emissions varied significantly by depth ($p = 0.049$) and between sites ($p < 0.001$, Fig. 2c). Emissions also varied significantly in the interaction between depth and location ($p = 0.04$). Ekolongouma soils were consistently a small sink under dry conditions and small source when wet, with somewhat greater emissions from surface soils, although soil moisture and interactions were not significant ($p = 0.557$). In contrast, soils from Itanga were consistently a source of N₂O, with substantial subsurface production, particularly following rewetting.

3.3. Temperature dependence of greenhouse gas exchange

Temperature responses of GHG fluxes were only significant for CO₂ emissions across the two sites, depths and moisture treatments (Fig. 3a), whereas there was no significant temperature response for CH₄ oxidation or N₂O fluxes (Figs. 3b & d). While CH₄ oxidation showed no

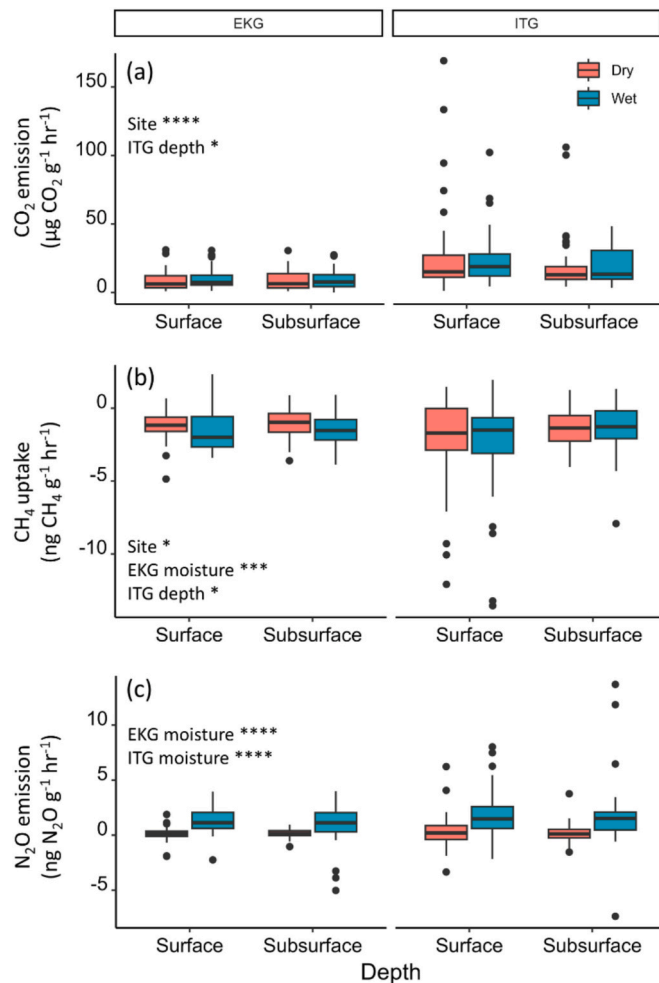


Fig. 2. Baseline (a) CO₂ emission, (b) CH₄ uptake, and (c) N₂O fluxes at 25 °C at 30 % (dry) and 70 % (wet) gravimetric moisture content in surface and subsurface savannah soils at Ekolongouma (EKG) and Itanga (ITG) village sites. Asterisks indicate significance levels for pairwise comparisons (site, depth, moisture): * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$.

relationship with other GHG fluxes (Fig. 3c), N₂O fluxes increased significantly with CO₂ emissions across all measurements (Fig. 3e).

Interactions between the temperature response of GHG fluxes were tested for site, soil depth and moisture interactions, with the condition that additional explanatory variables have a greater goodness of fit with additional degrees of freedom of $\Delta AIC_{df} < -5$. Few regression models were selected on this basis and were limited to significant relationships between CO₂ flux temperature dependence and site (Figs. 4a & b) and between standardised N₂O and CO₂ fluxes with moisture (Fig. 4d & e). Greater values for the regression slopes (E_a and b) in Fig. 4 indicate a greater increase in CO₂ flux with an increase in temperature in Fig. 4a & b, and a greater increase in N₂O flux with an increase in CO₂ flux in Fig. 4c & d. That is, CO₂ emissions were more temperature dependent in Ekolongouma ($E_a = 0.509$, Fig. 4a) than Itanga ($E_a = 0.357$, Fig. 4b) while N₂O fluxes were less sensitive to increasing CO₂ emissions in wet soils ($b = 0.148$, Fig. 4d) and more sensitive in dry soils ($b = 0.879$, Fig. 4e).

For the regression models that yielded a significant temperature response we estimated Q₁₀ values for CO₂ and N₂O fluxes (Table 2). That is, Q₁₀ values were not estimated for CH₄ fluxes or N₂O fluxes in wet conditions due to the non-significant temperature response across treatments (Fig. 2). Q₁₀ estimates for soil respiration were broadly comparable for EKG and ITG (1.92 and 1.58). The estimated Q₁₀ for N₂O fluxes in dry conditions were negative, representing a decrease in N₂O

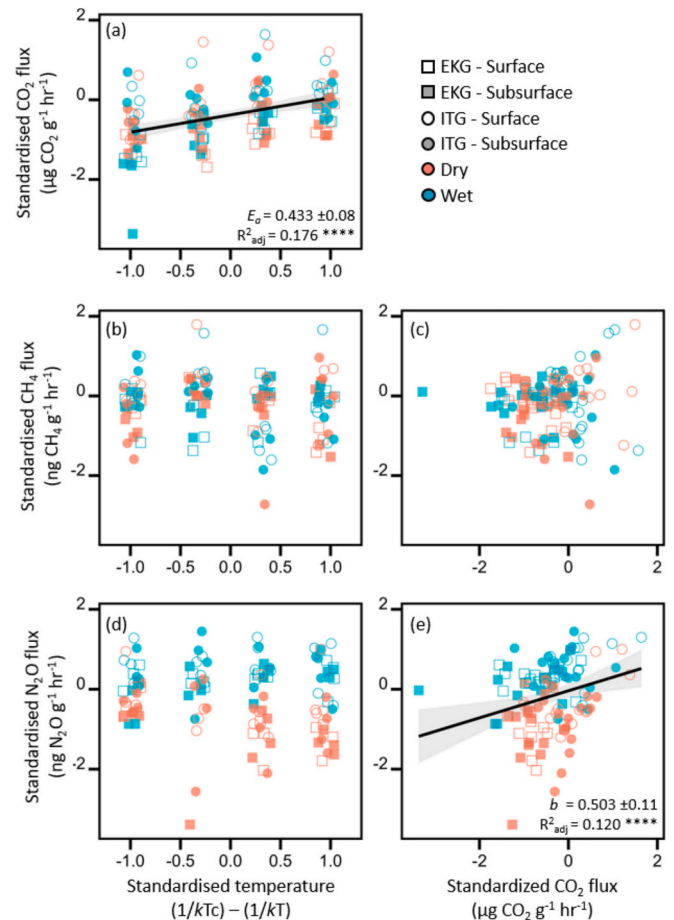


Fig. 3. The temperature response of GHG fluxes across two savannah sites (Ekolongouma (EKG): square symbols, Itanga (ITG): circle symbols). Plots show standardised GHG fluxes against standardised Arrhenius temperature (a, b & d) or standardised CO₂ flux (c & e) for measurements across the two sites, depths (Surface: open symbols; Subsurface: filled symbols) and moisture (Dry: coral symbols; Wet: turquoise symbols) conditions. Symbols in a, b and d are jittered for visibility across static temperatures (unstandardised values of 20, 25, 30 and 35 °C). Significant linear regression relationships are indicated by black solid lines with the activation energy (E_a) or slope (b) values presented alongside adjusted R^2 fits and significance levels ($p < 0.0001$: ****).

fluxes with an increase in temperature.

We investigated non-standardised GHG flux relationships against one another over the experimental temperature range to identify relationships between CO₂, and N₂O and CH₄ according to additional explanatory variables. We found the relationship between N₂O and CO₂ fluxes held for non-standardised measurements (Fig. 5a) and that the key explanatory variable for differences in this relationship was temperature range (Fig. 5b). In contrast to the standardised GHG fluxes (Fig. 3), non-standardised CH₄ oxidation showed a significant relationship with CO₂ emissions across temperature treatments (Fig. 5c) and showed a significant interaction with soil depth (Fig. 5d).

3.4. Principle component analysis

Separate PCAs were run for wet and dry soils. For wet soils, the selected parameters accounted for 79 % of variance (Fig. 6a). The first component separated soils by properties (OI, I index, pH, electrical conductivity, and C:N), while the second component separated soils by GHGs (N₂O, CO₂, and CH₄). Similarly, in dry soils (Fig. 6c), the selected variables accounted for 80 % of variability, with the first principle component separating soils by soil properties (OI, I index, pH and

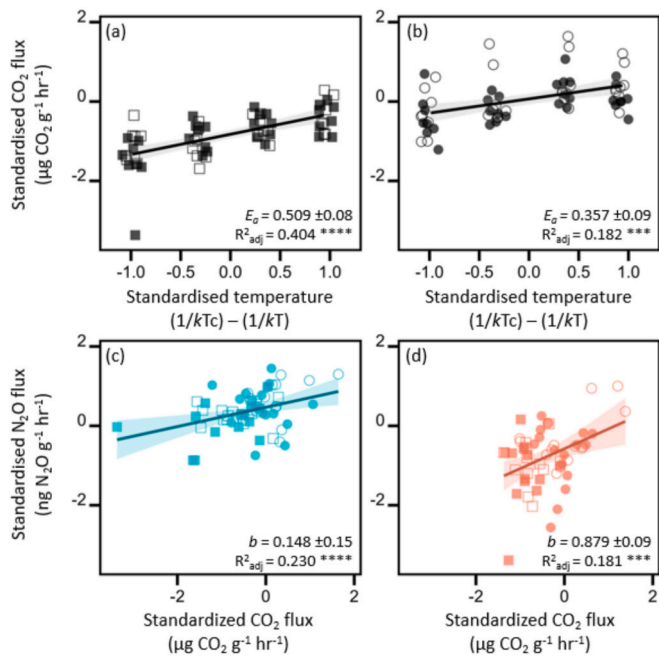


Fig. 4. Significantly different GHG flux relationships across sites and moisture treatments. Plots show (a & b) the temperature dependence of standardised CO₂ fluxes in Ekolongouma (a) and Itanga (b) and (c & d) relationships between standardised N₂O and CO₂ fluxes across both sites (symbol shapes and colours as in Fig. 3) in wet (c) and dry (d) conditions. Slope and significance values as in Fig. 3. Symbols in a and b are jittered for visibility across static temperatures (unstandardised values of 20, 25, 30 and 35 °C).

electrical conductivity), and the second component separating C:N, and GHG exchange (CH₄ and CO₂).

4. Discussion

4.1. Soil properties

We identified significant differences in a range of biogeochemical properties between sites, with the exception of the R index, with only limited differences between soil depths. Soils from Itanga were characterised by high organic matter content in both the surface and subsurface and very acidic pH, possibly indicating significant time since the last major fire event (Úbeda et al., 2009), or due to relatively high soil moisture levels (Table 1). Conversely, soils from Ekolongouma have substantially lower organic matter content, and higher (although still acidic) pH. A higher I index and OI may have been driven by relatively recent fresh inputs of litter. High HI and low OI values (e.g. in soils from Ekolongouma) indicate the presence of hydrogen rich compounds which can include celluloses, whereas low HI and high OI values in Itanga soils are broadly indicative of increased presence of aromatic carbon (i.e. increased recalcitrant black carbon and humic substances) (Saenger et al., 2013). Such differences may be accounted for by contrasts in fire regime, which are frequent in savannahs in the region, or through

contrasts in soil moisture resulting from flooding regime. In general, fires in savannahs deplete labile soil organic carbon fractions (for example relatively fresh litter inputs), although precise effects are determined by the inherent properties of the litter itself. Oxygen-containing functional groups are particularly vulnerable to loss on heating. In turn, this is associated with a relative increase in humic fractions, with soils heavily affected by fire featuring high quantities of black carbon (González-Pérez et al., 2004). pH increases following burning due to the incorporation of ash are generally reported (Úbeda et al., 2009), although conversely, pH can decrease through nitrification, which is enhanced in the short term following burning due to increased temperatures, and the availability of pyrolysable material (Mohamed et al., 2007), or due to high concentrations of aluminium found in ash produced at low temperatures (Pereira et al., 2017). Conversely, differences may be associated with contrasting moisture regimes between sites, with Itanga featuring higher gravimetric moisture compared to soils from Ekolongouma (Table 1). Increased moisture may be indicative of flooding (and neighbouring peatland sites in the area periodically flooded) (Crezee et al., 2022), which would reduce rates of aerobic decomposition and thus drive the accumulation of organic matter in the profile, and also result in lower pH (Girkin et al., 2019).

4.2. Temperature response of greenhouse gas emissions

Baseline rates of soil respiration (4–29 μg CO₂ g⁻¹ h⁻¹), CH₄ oxidation, and N₂O fluxes were broadly comparable to values previously reported in the literature (Castaldi et al., 2010; Hao et al., 1988; Livesley et al., 2011; Sanhueza and Santana, 1994; Wachiye et al., 2020), with

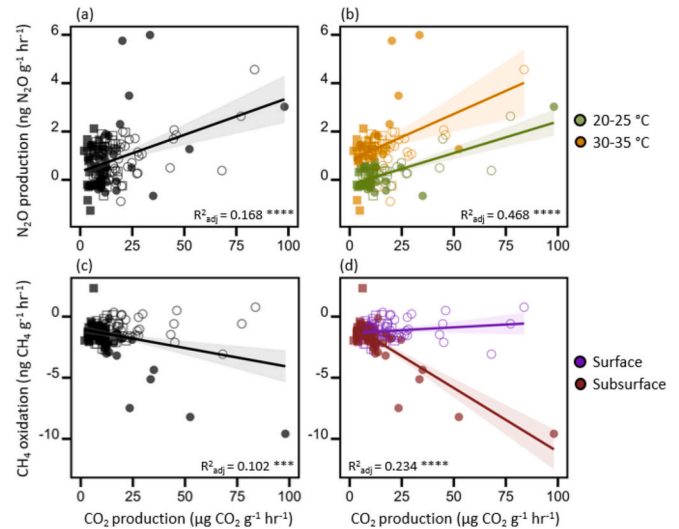


Fig. 5. GHG flux relationships with one another across the experimental temperature range, showing non-standardised measurements for (a & b) N₂O and CO₂ and (c & d) CH₄ and CO₂ fluxes. Significant interactions between GHG flux relationships were found for (b) N₂O and temperature range and (d) CH₄ and soil depth.

Table 2

Regression analysis for the two savannah sites and GHG flux responses to temperature (note CH₄ did not show a significant temperature response). Q₁₀ values were estimated using the parameters *B* and *E_a* from Eq. (1) and calculated for temperatures ranging between 20 and 35 °C. NS indicates a non-significant temperature response for N₂O in wet conditions.

Site /Moisture treatment	GHG	<i>B</i>	<i>E_a</i>	<i>F</i>	<i>R</i> ² _{adj}	<i>p</i>	Estimated Q ₁₀
EKG	CO ₂	−0.824 ± 0.055	0.509 ± 0.077	43.73	0.404	****	1.92 ± 0.006
ITG	CO ₂	0.066 ± 0.066	0.357 ± 0.092	15.00	0.182	***	1.58 ± 0.004
Wet	N ₂ O	ns					
Dry	N ₂ O	−0.709 ± 0.108	−0.343 ± 0.144	5.70	0.077	*	−0.64 ± 0.001

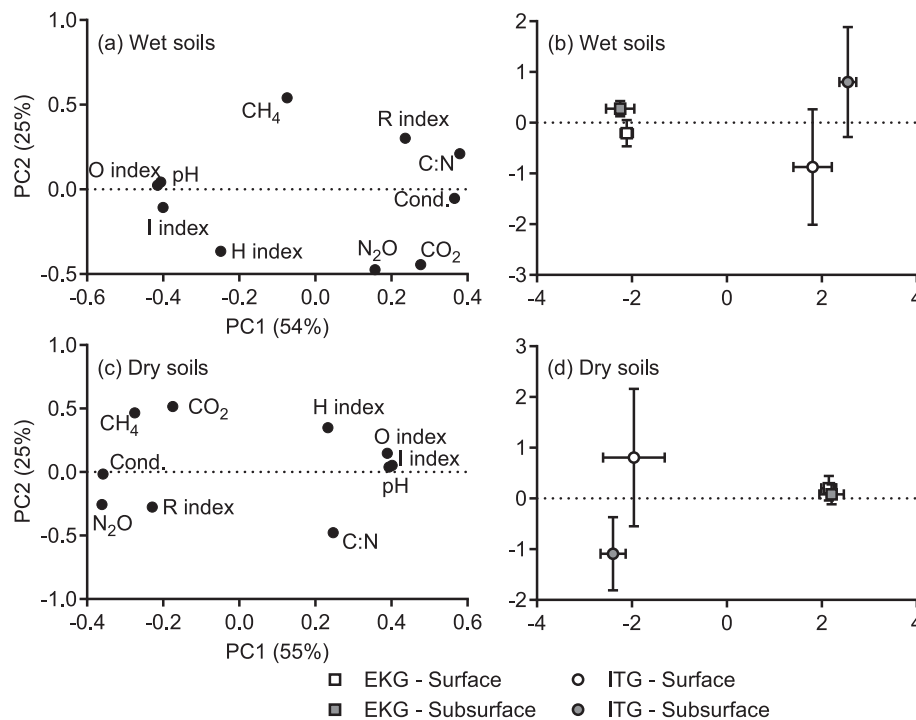


Fig. 6. (a & c) PCA loadings and (b & d) PCA scores for wet and dry soils for GHG emissions and soil properties.

the exception that, in this study, soils were consistently a sink for CH_4 (i.e. significant methanotrophic activity). Previous studies of the processes underlying the exchange of GHGs have indicated important roles for changes in moisture and burning, although many of these effects can be transitory (Castaldi et al., 2010).

CO_2 emissions was strongly influenced by temperature across both sites and all conditions (Fig. 3a), while CH_4 oxidation (Fig. 3b) and N_2O production in wet conditions (Fig. 3d, showing wet and dry conditions together) showing a non-significant temperature response. N_2O fluxes were, however, related to CO_2 fluxes across experimental temperatures and wet and dry soil conditions (Fig. 3e). Site had a large effect on the temperature dependence (how much of the variation in GHG flux was explained by temperature) and sensitivity (how much the GHG flux increases with temperature) (Fig. 4a & b). That is, temperature explained greater variation in CO_2 fluxes ($R^2_{\text{adj}} = 0.40$) and CO_2 was more temperature dependent ($E_a = 0.51$) for Ekolongouma than Itanga soils ($R^2_{\text{adj}} = 0.18$; $E_a = 0.36$) (Table 2). Many of the soil biogeochemical properties measured varied significantly between sites, and in particular soil organic matter and carbon which were substantially higher in both surface and subsurface soils at Itanga; soils from Ekolongouma also featured a higher I index (Table 1). Results from our PCA indicated that O, I and R indices (i.e. the availability of labile and recalcitrant carbon) are important for mediating observed responses. In contrast to the carbon quality hypothesis, the increased presence of aromatic/recalcitrant carbon at the ITG site was not associated with higher Q_{10} values (1.58 for ITG versus 1.92 for EKG). This somewhat surprising finding may reflect one or more of several processes. Physical protection may differ between soils (Davidson and Janssens, 2006; Cooper et al., 2021), with the potential for increased protection of carbon within aggregates at ITG, resulting in apparent substrate limitation at microsites, despite greater total carbon. Other environmental constraints can also be significant, for example the increased presence of clay-sized minerals that can adsorb organic matter, and help retain soil moisture (Davidson and Janssens, 2006).

Soil moisture exerts a strong limitation on GHG fluxes in the dry season (Thomas et al., 2011; Wood et al., 2013), and although we found no evidence for a soil moisture interaction with CO_2 or CH_4 fluxes, we

did observe a significant interaction between N_2O with CO_2 in wet vs dry conditions or both Ekolongouma and Itanga soils (Fig. 4c & d). Fig. 4c shows higher N_2O fluxes relative to CO_2 fluxes in wet conditions and Fig. 4d shows a greater increase (sensitivity) of N_2O fluxes to increasing CO_2 fluxes in dry conditions. This is important in a wider context, as soil moisture is closely coupled to regional climate, particularly precipitation patterns. Based on our findings, extended wet seasons will likely result in higher N_2O fluxes, but N_2O will increase more rapidly in dry conditions. The relationship between N_2O and CO_2 fluxes is also higher, although not as sensitive to higher temperature ranges (Fig. 5a & b) making temperature and soil moisture interactions an important consideration for predicting future soil GHG-climate feedbacks.

Under wetter conditions, methanogenesis is likely to have increased in anaerobic microsites, although oxidation remains the dominant process, particularly under dry conditions (Fest et al., 2017). Savannah soils have previously been reported to act as CH_4 sources when fully inundated, with emission rates exponentially related to soil temperature (Otter and Scholes, 2000). Savannah ecosystems more broadly can also become substantial CH_4 sources through biomass burning (Laris et al., 2021), although in our study we found no evidence of temperature dependence for CH_4 oxidation and did not observe CH_4 production (Whalen and Reeburgh, 1996). Previous studies have demonstrated substantial temperature dependence of methanogenesis, particularly for high carbon peat soils (Girkin et al., 2020a; Sjögersten et al., 2018). Within this study, subsurface soils are likely to have been exposed to higher concentrations of headspace CH_4 than they would have been exposed to in situ due to the role of surface soils as a diffusion barrier for atmospheric CH_4 (Murguía-Flores et al., 2021). As a consequence, our estimates for CH_4 uptake for subsurface soils are likely an over-estimation, as soils will have had increased substrate availability for methanotrophy.

The relatively low N_2O flux rates (and sometimes negative fluxes, particularly under dry, basal conditions) across all temperatures likely reflect the acidic and nutrient-poor soils (Andersson et al., 2004; Castaldi et al., 2006) (Table 1), and match relatively low emissions reported in tropical forest soils elsewhere in the region (Barthel et al., 2022). In the tropics, high production and flux rates in situ are generally only reported

in tropical forests and in soils converted to agricultural use (Weitz et al., 2001). Under moisture limited conditions, low soil water content limits the presence of anoxic microsites required for denitrification (Firestone and Davidson, 1989). Increased WFPS in soils reduces oxygen availability, increasing fluxes of N_2O . However, extensive flooding beyond the experimental treatments applied in this study are likely to substantially reduce production because high moisture contents favours the production of N_2 rather than N_2O (Ciarlo et al., 2007). During the wet season, in situ soils are likely to emit pulses of N_2O as rainfall will stimulate mineralisation and nitrification (Castaldi et al., 2010). Previous studies of the temperature dependence of N_2O fluxes in temperate environments have strong exponential responses to increasing temperatures (Schindlbacher et al., 2004), but studies in the tropics and in savannahs specifically, are more limited. In general, increasing temperatures drive enhanced nitrification and denitrification, with optima between 25 and 35 °C (Melling et al., 2007). Our results, however, show a comparatively limited response to soil warming. In dry soils, N_2O fluxes are likely greater due to canonical nitrification-denitrification resulting in N_2O , but at higher soil moistures there is increased production of N_2 as the product of denitrification, resulting in lower N_2O fluxes, although this may be low at 70 % WFPS (Girkin and Cooper, 2023). Soil denitrification can be inhibited under acidic conditions, with denitrification rates decreasing with increasing soil acidity (Ciarlo et al., 2007). pH has previously been identified as important in predicting Q_{10} s both within and between land uses (Meyer et al., 2018; Min et al., 2014; Zhou et al., 2013), likely the result of pH-dependent microbial community structure and function (Blagodatskaya and Anderson, 1998). We only identified a significant temperature response for N_2O net production from EKG and not for ITG; moreover, the temperature response for EKG soils was low, collectively suggesting that these soils may be relatively insensitive to changes in soil temperature across the measured ranges, and implying a certain level of resilience to soil warming.

Our Q_{10} values for soil respiration (Table 2) are broadly comparable to findings globally (e.g. 1.9 for a North Australian savannah soil (Chen et al., 2002), and 2.1 in Senegalese savannahs (Elberling et al., 2003), and 1.4 to 2.7 during a 21 day incubation experiment with soils from the Brazilian cerrado (Espíndola et al., 2018). Makhado and Scholes (2011) reported a range of Q_{10} values for soil respiration (0.59–3.07) in *Acacia* and *Combretum* dominated savannah areas in the Kruger National Park across a series of 10 °C temperature gradients. Overall, these values are generally lower than those reported for other tropical ecosystems, potentially indicating some degree of resilience of microbial communities in tropical savannah soils in the region to rising temperatures (Girkin et al., 2020a, 2020b; Sjögersten et al., 2018).

5. Conclusion

Here, we show the response of GHG production in Central African savannah soils to changes in temperature and soil moisture. We identified that soil CO_2 production can be sensitive to warming under certain circumstances. In contrast, the temperature response of N_2O fluxes was only significant in dry conditions and no significant temperature response of CH_4 fluxes was observed. Significant differences in soil properties and GHG production between sites are likely driven by differences in site history and were strong drivers of production, mediating effects through contrasting soil organic matter properties. Overall, our results show that savannah soils can exhibit comparatively limited responses to increasing soil temperatures, which may indicate the potential resilience of soil processes to climate warming.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.geodrs.2025.e00934>.

Data availability

Data is available in supplementary materials

References

- Amoako, E.E., Gambiza, J., 2019. Effects of Anthropogenic Fires on Soil Properties and the Implications of Fire Frequency for the Guinea Savanna Ecological Zone. *Scientific African*, Ghana. <https://doi.org/10.1016/j.sciaf.2019.e00201>.
- Andersson, M., Michelsen, A., Jensen, M., Kjeller, A., 2004. Tropical savannah woodland: effects of experimental fire on soil microorganisms and soil emissions of carbon dioxide. *Soil Biol. Biochem.* <https://doi.org/10.1016/j.soilbio.2004.01.015>.
- Bagherzadeh, A., Brumme, R., Beese, F., 2008. Temperature dependence of carbon mineralization and nitrous oxide emission in a temperate forest ecosystem. *J. For. Res.* <https://doi.org/10.1007/s11676-008-0018-3>.
- Barthel, M., Bauters, M., Baumgartner, S., Drake, T.W., Bey, N.M., Bush, G., Six, J., 2022. Low N_2O and variable CH_4 fluxes from tropical forest soils of the Congo Basin. *Nat. Commun.* 13 (1), 330.
- Bekku, Y.S., Nakatsubo, T., Kume, A., Adachi, M., Koizumi, H., 2003. Effect of warming on the temperature dependence of soil respiration rate in arctic, temperate and tropical soils. *Appl. Soil Ecol.* [https://doi.org/10.1016/S0929-1393\(02\)00158-0](https://doi.org/10.1016/S0929-1393(02)00158-0).
- Blagodatskaya, E.V., Anderson, T.-H., 1998. Interactive effects of pH and substrate quality on the fungal-to-bacterial ratio and qCO_2 of microbial communities in forest soils. *Soil Biol. Biochem.* 30, 1269–1274. [https://doi.org/10.1016/S0038-0717\(98\)00050-9](https://doi.org/10.1016/S0038-0717(98)00050-9).
- Castaldi, S., Ermice, A., Strumia, S., 2006. Fluxes of N_2O and CH_4 from soils of savannas and seasonally-dry ecosystems. *J. Biogeogr.* <https://doi.org/10.1111/j.1365-2699.2005.01447.x>.
- Castaldi, S., De Grandcourt, A., Rasile, A., Skiba, U., Valentini, R., 2010. CO_2 , CH_4 and N_2O fluxes from soil of a burned grassland in Central Africa. *Biogeosciences*. <https://doi.org/10.5194/bg-7-3459-2010>.
- Chen, X., Eamus, D., Hutley, L.B., 2002. Seasonal patterns of soil carbon dioxide efflux from a wet-dry tropical savanna of northern Australia. *Aust. J. Bot.* <https://doi.org/10.1071/BT01049>.
- Ciarlo, E., Conti, M., Bartoloni, N., Rubio, G., 2007. The effect of moisture on nitrous oxide emissions from soil and the $N\ 2O/(N_2O+N_2)$ ratio under laboratory conditions. *Biol. Fertil. Soils*. <https://doi.org/10.1007/s00374-006-0147-9>.
- Cooper, H.V., Vane, C.H., Evers, S., Aplin, P., Girkin, N.T., Sjögersten, S., 2019. From peat swamp forest to oil palm plantations: the stability of tropical peatland carbon. *Geoderma* 342, 109–117.
- Cooper, H.V., Sjögersten, S., Lark, R.M., Girkin, N.T., Vane, C.H., Calonego, J.C., Mooney, S.J., 2021. Long-term zero-tillage enhances the protection of soil carbon in tropical agriculture. *Eur. J. Soil Sci.* 72 (6), 2477–2492.
- Crezee, B., Dargie, G.C., Ewango, C.E.N., et al., 2022. Mapping peat thickness and carbon stocks of the Central Congo Basin using field data. *Nat. Geosci.* 15, 639–644. <https://doi.org/10.1038/s41561-022-00966-7>.
- da Rocha, H.R., Freitas, H.C., Rosolem, R., Juárez, R.I.N., Tannus, R.N., Ligo, M.A., Cabral, O.M.R., Dias, M.A.F.S., 2002. Measurements of CO_2 exchange over a woodland savanna (Cerrado Sensus stricto) in southeast Brazil. *Biota Neotropica*. <https://doi.org/10.1590/s1676-06032002000100009>.
- Dargie, G.C., 2015. Quantifying and Understanding the Tropical Peatlands of the Central Congo Basin. University of Leeds.
- Davidson, E.A., Janssens, I.A., 2006. Temperature sensitivity of soil carbon decomposition and feedbacks to climate change. *Nature*. <https://doi.org/10.1038/nature04514>.
- Elberling, B., Fensholt, R., Larsen, L., Petersen, A.I.S., Sandholt, I., 2003. Water content and land use history controlling soil CO_2 respiration and carbon stock in savanna soil and groundnut fields in semi-arid Senegal. *Geografisk Tidsskrift*. <https://doi.org/10.1080/00167223.2003.10649491>.
- Espíndola, S.P., Bobuřská, L., de Siqueira Ferreira, A., 2018. Impact of nitrogen additions on soil microbial respiration and temperature sensitivity in native and agricultural ecosystems in the Brazilian Cerrado. *J. Therm. Biol.* 75, 120–127.

- Fang, C., Smith, P., Moncrieff, J.B., Smith, J.U., 2005. Similar response of labile and resistant soil organic matter pools to changes in temperature. *Nature* 433, 57–59.
- Fest, B.J., Hinko-Najera, N., Wardlaw, T., Griffith, D.W., Livesley, S.J., Arndt, S.K., 2017. Soil methane oxidation in both dry and wet temperate eucalypt forests shows a near-identical relationship with soil air-filled porosity. *Biogeosciences* 14 (2), 467–479.
- Fierer, N., Craine, J.M., McLaughlan, K., Schimel, J.P., 2005. Litter quality and the temperature sensitivity of decomposition. *Ecology*. <https://doi.org/10.1890/04-1254>.
- Firestone, M.K., Davidson, E.A., 1989. Microbiological basis of NO and N₂O production and consumption in soil. *Exchan.Trace Gases Between Terrest. Ecosyst. Atmosph.* <https://doi.org/10.1017/CBO9781107415324.004>.
- Gershenson, A., Bader, N.E., Cheng, W., 2009. Effects of substrate availability on the temperature sensitivity of soil organic matter decomposition. *Glob. Chang. Biol.* <https://doi.org/10.1111/j.1365-2486.2008.01827.x>.
- Giardina, C.P., Ryan, M.G., 2000. Evidence that decomposition rates of organic carbon in mineral soil do not vary with temperature. *Nature* 404, 858–861.
- Girkin, N.T., Cooper, H.V., 2023. Nitrogen and ammonia in soils. In: *Encyclopedia of Soils in the Environment*, pp. 142–151.
- Girkin, N.T., Turner, B.L., Ostle, N., Craigon, J., Sjögersten, S., 2018. Root exudate analogues accelerate CO₂ and CH₄ production in tropical peat. *Soil Biol. Biochem.* 117, 48–55. <https://doi.org/10.1016/j.soilbio.2017.11.008>.
- Girkin, N.T., Vane, C.H., Cooper, H.V., Moss-Hayes, V., Craigon, J., Turner, B.L., Ostle, N., Sjögersten, S., 2019. Spatial variability of organic matter properties determines methane fluxes in a tropical forested peatland. *Biogeochemistry* 142. <https://doi.org/10.1007/s10533-018-0531-1>.
- Girkin, N.T., Dhandapani, S., Evers, S., Ostle, N., Turner, B.L., Sjögersten, S., 2020a. Interactions between labile carbon, temperature and land use regulate carbon dioxide and methane production in tropical peat. *Biogeochemistry* 147, 87–97.
- Girkin, N.T., Vane, C.H., Turner, B.L., Ostle, N.J., Sjögersten, S., 2020b. Root oxygen mitigates methane fluxes in tropical peatlands. *Environ. Res. Lett.* 15 (6), 064013.
- González-Pérez, J.A., González-Vila, F.J., Almendros, G., Knicker, H., 2004. The effect of fire on soil organic matter - a review. *Environ. Int.* <https://doi.org/10.1016/j.envint.2004.02.003>.
- Govender, N., Trollope, W.S.W., Van Wilgen, B.W., 2006. The effect of fire season, fire frequency, rainfall and management on fire intensity in savanna vegetation in South Africa. *J. Appl. Ecol.* <https://doi.org/10.1111/j.1365-2664.2006.01184.x>.
- Hao, W.M., Scharffe, D., Crutzen, P.J., Sanhueza, E., 1988. Production of N₂O, CH₄, and CO₂ from soils in the tropical savanna during the dry season. *J. Atmos. Chem.* <https://doi.org/10.1007/BF00048256>.
- Hutley, L.B., Setterfield, S.A., 2008. Savannas. In: *Encyclopedia of Ecology*. Elsevier, Oxford, pp. 3143–3154.
- IPCC, 2021. Climate Change 2021: The Physical Science Basis. In: *Contribution of Working Group I to the Sixth Assessment Report of the Intergovernmental Panel on Climate Change*.
- Johnston, A.S.A., Sibly, R.M., 2018. The influence of soil communities on the temperature sensitivity of soil respiration. *Nat. Ecol. & Evolut.* 2, 1597–1602.
- Johnston, A.S.A., Meade, A., Ardö, J., Arriga, N., Black, A., Blanken, P.D., Bonal, D., Brümmer, C., Cescatti, A., Dusek, J., Graf, A., Gioli, B., Godek, I., Gough, C.M., Ikawa, H., Jassal, R., Kobayashi, H., Magliulo, V., Manca, G., Montagnani, L., Moyano, F.E., Olesen, J.E., Sachs, T., Shao, C., Tagesson, T., Wohlfahrt, G., Wolf, S., Woodgate, W., Varlagin, A., Venditti, C., 2021. Temperature thresholds of ecosystem respiration at a global scale. *Nat. Ecol. & Evolut.* 1–8. <https://doi.org/10.1038/s41559-021-01398-z>.
- Laris, P., Koné, M., Dembélé, F., Rodrigue, C.M., Yang, L., Jacobs, R., Laris, Q., 2021. Methane gas emissions from savanna fires: what analysis of local burning regimes in a working west African landscape tell us. *Biogeosciences* 18 (23), 6229–6244.
- Leifeld, J., Fuhrer, J., 2005. The temperature response of CO₂ production from bulk soils and soil fractions is related to soil organic matter quality. *Biogeochemistry* 75, 433–453.
- Livesley, S.J., Grover, S., Hutley, L.B., Jamali, H., Butterbach-Bahl, K., Fest, B., Beringer, J., Arndt, S.K., 2011. Seasonal variation and fire effects on CH₄, N₂O and CO₂ exchange in savanna soils of northern Australia. *Agric. For. Meteorol.* <https://doi.org/10.1016/j.agrformet.2011.02.001>.
- Lloyd, J., Taylor, J., 1994. On the temperature dependence of soil respiration. *Funct. Ecol.* <https://doi.org/10.2307/2389824>.
- Lopez-Sangil, L., Hartley, I.P., Rovira, P., Casals, P., Sayer, E.J., 2018. Drying and rewetting conditions differentially affect the mineralization of fresh plant litter and extant soil organic matter. *Soil Biol. Biochem.* <https://doi.org/10.1016/j.soilbio.2018.06.001>.
- Makhado, R.A., Scholes, R.J., 2011. Determinants of soil respiration in a semi-arid savanna ecosystem, Kruger National Park, South Africa. *Koedoe: African Protect. Area Conservat. Sci.* 53 (1), 1–8.
- Melling, L., Hatano, R., Goh, K.J., 2007. Nitrous Oxide Emissions from Three Ecosystems in Tropical Peatland of Sarawak. *Soil Science and Plant Nutrition, Malaysia*. <https://doi.org/10.1111/j.1747-0765.2007.00196.x>.
- Meyer, N., Welp, G., Amelung, W., 2018. The temperature sensitivity (Q₁₀) of soil respiration: controlling factors and spatial prediction at regional scale based on environmental soil classes. *Glob. Biogeochem. Cycles*. <https://doi.org/10.1002/2017GB005644>.
- Michaelis, L., Mand, M., Menten, L., 1913. Die Kinetik der Invertiirwirkung. *Biochem. Z.* 49, 333–369.
- Min, K., Lehmeier, C.A., Ballantyne, F., Tatarko, A., Billings, S.A., 2014. Differential effects of pH on temperature sensitivity of organic carbon and nitrogen decay. *Soil Biol. Biochem.* <https://doi.org/10.1016/j.soilbio.2014.05.021>.
- Mohamed, A., Härdtle, W., Jirjahn, B., Niemeyer, T., Von Oheimb, G., 2007. Effects of prescribed burning on plant available nutrients in dry heathland ecosystems. *Plant Ecol.* <https://doi.org/10.1007/s11258-006-9183-7>.
- Murguia-Flores, F., Ganesan, A.L., Arndt, S., Hornibrook, E.R., 2021. Global uptake of atmospheric methane by soil from 1900 to 2100. *Glob. Biogeochem. Cycles* 35 (7), e2020GB006774.
- Nottingham, A.T., Meir, P., Velasquez, E., Turner, B.L., 2020. Soil carbon loss by experimental warming in a tropical forest. *Nature*. <https://doi.org/10.1038/s41586-020-2566-4>.
- Otter, L.B., Scholes, M.C., 2000. Methane sources and sinks in a periodically flooded south African savanna. *Glob. Biogeochem. Cycles* 14 (1), 97–111.
- Pennington, R.T., Lehmann, C.E.R., Rowland, L.M., 2018. Tropical savannas and dry forests. *Curr. Biol.* <https://doi.org/10.1016/j.cub.2018.03.014>.
- Pereira, P., Cerda, A., Martin, D., Úbeda, X., Depellegrin, D., Novara, A., Martínez-Murillo, J.F., Brevik, E.C., Menshov, O., Comino, J.R., Miesel, J., 2017. Short-term low-severity spring grassland fire impacts on soil extractable elements and soil ratios in Lithuania. *Sci. Total Environ.* 578, 469–475. <https://doi.org/10.1016/j.scitotenv.2016.10.210>.
- Poth, M., Anderson, I.C., Miranda, H.S., Miranda, A.C., Riggan, P.J., 1995. The magnitude and persistence of soil NO, N₂O, CH₄, and CO₂ fluxes from burned tropical savanna in Brazil. *Glob. Biogeochem. Cycles*. <https://doi.org/10.1029/95GB02086>.
- R Core Team, 2023. R: The R Project for Statistical Computing [WWW Document]. URL. <https://www.r-project.org/> (accessed 1.9.24).
- Saenger, A., Cecillon, L., Sebag, D., Brun, J.J., 2013. Soil organic carbon quantity, chemistry and thermal stability in a mountainous landscape: a rock-Eval pyrolysis survey. *Org. Geochem.* 54, 101–114. <https://doi.org/10.1016/j.orggeochem.2012.10.008>.
- Sanhueza, E., Santana, M., 1994. CO₂ emissions from tropical savanna soil under first year of cultivation. *Interciencia* 20–23.
- Schindlbacher, A., Zechmeister-Boltenstern, S., Butterbach-Bahl, K., 2004. Effects of soil moisture and temperature on NO, NO₂, and N₂O emissions from European forest soils. *J. Geophys. Res. D: Atmospher.* <https://doi.org/10.1029/2004JD004590>.
- Sebag, D., Verrecchia, E.P., Cecillon, L., Adatte, T., Albrecht, R., Aubert, M., Bureau, F., Cailleau, G., Copard, Y., Decaens, T., Disnar, J.R., Hetenyi, M., Nyilas, T., Trombino, L., 2016. Dynamics of soil organic matter based on new rock-Eval indices. *Geoderma* 284, 185–203. <https://doi.org/10.1016/j.geoderma.2016.08.025>.
- Sjögersten, S., Aplin, P., Gauci, V., Peacock, M., Siegenthaler, A., Turner, B.L., 2018. Temperature response of ex-situ greenhouse gas emissions from tropical peatlands: interactions between forest type and peat moisture conditions. *Geoderma*. <https://doi.org/10.1016/j.geoderma.2018.02.029>.
- Somers, C., Girkin, N.T., Rippey, B., Lanigan, G.J., Richards, K.G., 2019. The effects of urine nitrogen application rate on nitrogen transformations in grassland soils. *J. Agric. Sci.* 157 (6), 515–522.
- Thomas, A.D., Hoon, S.R., Dougill, A.J., 2011. Soil respiration at five sites along the Kalahari transect: effects of temperature, precipitation pulses and biological soil crust cover. *Geoderma*. <https://doi.org/10.1016/j.geoderma.2011.07.034>.
- Úbeda, X., Pereira, P., Outeiro, L., Martin, D.A., 2009. Effects of fire temperature on the physical and chemical characteristics of the ash from two plots of Cork oak (*Quercus Suber*). *Land Degrad. Dev.* <https://doi.org/10.1002/ldr.930>.
- Wachiye, S., Merbold, L., Vesala, T., Rinne, J., Räsänen, M., Leitner, S., Pellikka, P., 2020. Soil greenhouse gas emissions under different land-use types in savanna ecosystems of Kenya. *Biogeosciences* 17 (8), 2149–2167.
- Weitz, A.M., Linder, E., Frolking, S., Crill, P.M., Keller, M., 2001. N₂O emissions from humid tropical agricultural soils: effects of soil moisture, texture and nitrogen availability. *Soil Biol. Biochem.* [https://doi.org/10.1016/S0038-0717\(01\)00013-X](https://doi.org/10.1016/S0038-0717(01)00013-X).
- Whalen, S.C., Reeburgh, W.S., 1996. Moisture and temperature sensitivity of CH₄ oxidation in boreal soils. *Soil Biol. Biochem.* 28 (10–11), 1271–1281.
- Wood, T.E., Detto, M., Silver, W.L., 2013. Sensitivity of soil respiration to variability in soil moisture and temperature in a humid tropical forest. *PLoS One* 8, e80965. <https://doi.org/10.1371/journal.pone.0080965>.
- Zhang, J., Peng, C., Zhu, Q., Xue, W., Shen, Y., Yang, Y., Shi, G., Shi, S., Wang, M., 2016. Temperature sensitivity of soil carbon dioxide and nitrous oxide emissions in mountain forest and meadow ecosystems in China. *Atmos. Environ.* <https://doi.org/10.1016/j.atmosenv.2016.08.011>.
- Zhou, Z., Guo, C., Meng, H., 2013. Temperature sensitivity and basal rate of soil respiration and their determinants in temperate forests of North China. *PLoS One*. <https://doi.org/10.1371/journal.pone.0081793>.