

1 **Soil microstructure alterations induced by land use change**
2 **for sugarcane expansion in Brazil**

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16 Running Title: Soil microstructure alterations by land use change.

17

18 **Summary**

19 Land use change (LUC) alters soil structure and consequently, the functions and
20 services provided by these soils. Conversion from extensive pasture to sugarcane is one
21 the most large-scale land transitions in Brazil due to the growth of the domestic and
22 global demands of bioenergy. However, the impacts of sugarcane expansion on the soil
23 structure under extensive pasture remains unclear, especially when considering changes
24 at the microscale. We investigated if LUC for sugarcane cultivation impacted on soil
25 microstructure quality. Undisturbed soil samples were taken from two soil layers (0-10
26 and 10-20 cm) under three contrasting land uses (native vegetation – NV, pasture – PA
27 and sugarcane – SC) in three different locations in the central area of southern Brazil.
28 Oriented thin sections (30 μm) were used for micromorphological analysis. The total
29 area of pores decreased following the LUC in the following order; NV > PA > SC in
30 both soil layers. The area of large complex packing pores (>0.01 mm^2) also decreased
31 with the LUC sequence: NV>PA>SC. Qualitative and semi-quantitative
32 micromorphological analysis confirmed porosity reduction was driven by the decrease
33 in complex packing pores and that biological features decreased in the same LUC
34 sequence as the quantitative parameters. Therefore, LUC for sugarcane expansion
35 reduced microscale soil porosity, irrespectively of soil type and site-specific conditions,
36 indicating the adoption of more sustainable management practices is imperative to
37 preserve soil structure and sustain soil functions in Brazilian sugarcane fields.

38 **Keywords:** Soil micromorphology, sugarcane, soil physical quality, complex packing
39 pores, bioenergy production

40 **Introduction**

41 The growing interest in biofuels has resulted in a new demand for arable land for
42 bioenergy crop production. Land use change (LUC) is one of the greatest threats to
43 soil quality (Cherubin *et al.*, 2016a; Bonilla-Bedoya *et al.*, 2017), as it can have
44 significant impacts on soil biodiversity (Franco *et al.*, 2016), carbon storage (Mello *et*
45 *al.*, 2014) and ecosystem services (Foley *et al.*, 2011). Brazil is the world' largest
46 sugarcane producer with 8.59 million ha of cultivation area and production of 29 Mt
47 of sugar and 33 billion L of ethanol (CONAB, 2019). Conversion from extensive and
48 degraded pastureland to sugarcane production is the main scenario of LUC used to
49 support sugarcane expansion in Brazil (Adami *et al.*, 2012; Strassburg *et al.*, 2014).

50 However, the intensive mechanization used in sugarcane fields, including soil
51 tillage by ploughing and disking and heavy machinery traffic during mechanical
52 harvesting degrades soil structure, affecting multiples processes and functions in these
53 soils (Cherubin *et al.*, 2016; Robot *et al.*, 2018). Soil structure is typically defined by
54 the arrangement of soil particles and aggregates and the pores among the structural
55 units, which regulates multiple processes and services such as: water retention and
56 conductivity, soil aeration, soil organic matter turnover, nutrient cycling (Six *et al.*,
57 2004), soil erodibility (Barthès & Roose, 2002) and plant growth. Therefore, parameters
58 related to soil structure are considered key indicators of soil quality (Bünemann *et al.*,
59 2018). Soil microstructure relates to the compositional arrangement of soil at a smaller
60 scale (i.e. at the micron scale)) and can be assessed by the use of thin sections, also
61 known as micromorphology (Bullock *et al.* 1985). Although microstructure assessment
62 by thin section can be time-consuming and generally does not provide 3D structural
63 information, it provides more detail than other approaches where visualization of the
64 soil micro-fabric is concerned.

65 Although traditional soil physical properties (e.g. bulk density, soil porosity, soil
66 penetration resistance, soil aggregation etc.) along with visual assessment methods can
67 efficiently infer the stability, and even resilience of soil structure (Cherubin *et al.*,
68 2016b, 2017; Castioni *et al.*, 2018), these methods cannot reveal the precise spatial
69 arrangement of soil structure and the geometrical form of pores and aggregates.
70 Imaging methods, such as micromorphology, can be used to further study the dynamics
71 of soil structural development across the time and/or space and help improve
72 understanding concerning the impact of soil structure on soil functioning (Guimarães
73 *et al.*, 2013; Silva *et al.*, 2015; Souza *et al.*, 2015; Pires *et al.*, 2017). Whilst other
74 imaging methods such as X-ray Computed Tomography (CT) have become more
75 popular for the analysis of soil pore space in recent years, particularly as they facilitate
76 faster acquisition of images and 3D visualisation, micromorphology is still an important
77 technique for the analysis of soil structure as it permits the microscopic visualization of
78 some soil properties, such as those derived from organic matter, e.g. fecal deposits, that
79 are currently not straightforward to image by X-ray CT (Helliwell *et al.*, 2013).

80 Considering the intense mechanization applied to sugarcane soils, we conducted a
81 field study to evaluate the impact of LUC for sugarcane expansion on soil
82 microstructure characteristics using soil thin sections. The hypothesis was that the
83 intensity of sugarcane cultivation had a significant impact of the alteration on soil
84 microstructure and subsequent soil quality.

85

86 **Materials and methods**

87 *Study sites*

88 Undisturbed soil samples for 0 – 10 and 10 – 20 cm soil depth were taken in the
89 central region of southern Brazil at three different locations within the main sugarcane-

90 producing region of the country, as follows: Lat_17S near the city of Jataí – Goáis State
91 (17°56'16"S 51°38'31"W), Lat_21S near the city of Valparaíso – São Paulo State
92 (21°14'48"S 50°47'04"W) and Lat_23S near the city of Ipaussu – São Paulo State
93 (23°05'08"S 49°37'52"W) with soil orders was classified as Oxisol, Alfisol/Ultisol and
94 Oxisol by the USDA Soil Taxonomy (Soil Survey Staff, 2014), respectively. The
95 climate was classified according to Köppen-Geiger's system as mesothermal tropical
96 (Awa), humid tropical (Aw) and tropical (Cwa), respectively. The mean annual
97 temperature and precipitation is 24.0 °C and 1600 mm (Awa) at Lat_17S, 23.4 °C and
98 1240 mm (Aw) at Lat_21S and 21.7 °C and 1479 mm (Cwa) at Lat_23S, with the rainy
99 season in the Spring-Summer (October to April) and the dry season during the Autumn-
100 Winter (May to September). More detailed climate information (mean monthly
101 temperature and precipitation) are available in Cherubin *et al.* (2015).

102 In each site, we sampled a LUC sequence, including native vegetation (NV,
103 baseline), pasture (PA) and sugarcane (SC) areas. Selected physical and chemical soil
104 properties are found in Table 1. The land use and management history of each site, as
105 well as chemical and physical characterization of the soils are further described in
106 Cherubin *et al.* (2015; 2016). For all sugarcane areas, the soil was prepared by
107 ploughing and disking previously to cropping. The SC fields at Lat_17S, Lat_21S and
108 Lat_23S was in the third, third and fourth ratoon, respectively. In SC fields fertilizer
109 was applied annually and harvesting was performed using a 20 Mg harvester and
110 transported by a tractor and wagon (10 + 30 Mg). A controlled traffic system was not
111 used in these areas.

112 113 *Soil sampling and preparation*

114 One undisturbed soil sample (7 x 12 x 6 cm) was collected in the Lat_17S, Lat_21S
115 e Lat_23S for NV, PA and SC in two soil layers (0-10 and 10-20 cm), totaling 18

116 samples (3 sites x 3 land uses x 2 soil depths). For sugarcane, the soil was sampled in
117 the inter-rows. The soils were air dried for 35 days and then placed in an oven at 40 °C
118 for 48h. The dry samples were impregnated with a polyester resin, styrene monomer
119 and fluorescent dye (Tinopal BASF®) by capillarity in a vacuum chamber. After
120 impregnation, vertically oriented soil thin sections (c. 30 µm thick) were obtained for
121 qualitative and semi-quantitative description (Bullock *et al.*, 1985; Stoops, 2003;
122 Cooper *et al.*, 2017) and quantitative image analysis (Cooper *et al.* 2016). Figure 1
123 illustrates the sampling procedure adopted in the field.

124

125 *Micromorphological analysis*

126 The thin sections were analyzed using a Zeiss petrographic microscope. The
127 qualitative description of thin section was made following the classifications described
128 in Bullock *et al.* (1985) and Stoops (2003) only for thin sections from Lat_23S. This
129 method provides reference images for a semiquantitative assessment of porosity and
130 the description of pore morphology. The pores were classified as packing pores, i.e.
131 those that result from the loose packing of soil components; channel pores, i.e. tubular
132 smooth pores with a cylindrical or arched cross section which are uniform over much
133 of the length; vughs, i.e. more or less equidimensional, irregularly shaped, smooth or
134 rough, usually not interconnected; and planar pores, i.e. flat, accommodating or not,
135 smooth or rough, resulting from shrinkage or compaction (Stoops, 2003). The soil
136 coarse/fine (c/f) fabric was classified as either porphyric (i.e. coarse grains embedded
137 in fine material), enaulic (i.e. fine material appears as micro-aggregates between coarser
138 components) or combinations of these as described in Stoops (2003).

139 *Micromorphometrical analysis*

140 Ultraviolet light was used to enhance the contrast between the pore space and soil
 141 matrix, and images were obtained using a charged couple device photographic camera
 142 (DFW-X700, Sony®). For each soil sample, fifteen images of 180 mm² were randomly
 143 obtained (Figure 1). The images were digitalized with a resolution of 1024 x 768 pixels
 144 in 256 shades of gray in a 10x amplification giving a pixel size of 12.5 µm. Pore
 145 segmentation was undertaken in Noesis Visilog version 5.4 by means of a user defined
 146 threshold (maintained throughout the study), opening and closing filtering, and
 147 labelling, which correspond to the individualization of each object followed by its
 148 identification. The smallest segmented pore had a diameter of 37.5 µm, which is
 149 classified in the meso/macro-pore size range; the size class most sensitive to soil
 150 compaction (Richard *et al.*, 2001).

151 The total area of pores (Tap) for each image was calculated as the percentage of the
 152 sum of the areas of the individual pores divided by the total area of the assessed image
 153 (Hallaire & Cointepas, 1993). Pore shape was classified into three groups as in Cooper
 154 *et al.* (2016): rounded, elongated and complex. Two indexes were used to determine
 155 the pore shape, as described in Eq. 1 and Eq. 2:

$$156 \quad I1 = \frac{P^2}{4\pi A} \quad (\text{Eq. 1})$$

157 Where P is the perimeter of the pore and A is the area.

$$158 \quad I2 = \frac{\frac{1}{m} \sum_i (NI)_i}{\frac{1}{n} \sum_j (DF)_j} \quad (\text{Eq. 2})$$

159 NI is the number of intercepts of the object in direction i ($i = 0^\circ, 45^\circ, 90^\circ$, and
 160 135°), DF is the Feret diameter of the object in the direction j ($j = 0^\circ$ and 90°), m
 161 correspond to the number of i directions and n to the number of j directions. The I2
 162 index was used complementary to I1 for a better pore segregation according to shape.

163 When morphometric shapes are compared with the micromorphological
164 classification, rounded pores correspond to vughs, elongated pores to channel and
165 planar pores, and complex pores to packing pores.

166

167 *Data analysis*

168 The mean soil porosity of each site was derived from 15 subsamples (every
169 image from a single thin section), which were used as pseudo replicates (Hurlbert,
170 1984) to compare the difference in LUC porosity for each site; to compare the LUC
171 effect on soil porosity for the central-southern region each site was considered as a
172 replicate (n=3). Data normality was tested by Shapiro-Wilk's test ($p > 0.05$), followed
173 by an analysis of variance (ANOVA) and *post hoc* via a Duncan's test ($p < 0.05$).

174

175 **Results**

176 *Micromorphological analysis*

177 Regardless of land use, the soils presented a dominant porphyric relative
178 distribution with secondary areas presenting as porphyric-enauclic, enauclic-porphyric
179 and enauclic related distributions. The porphyric-enauclic related distribution areas only
180 occurred in agricultural land uses (PA and SC) whilst the enauclic-prophyric areas were
181 only observed in NV soils (Table 3).

182 The soil micromorphological descriptions also showed a reduction in soil porosity
183 in both layers due to the LUC from native vegetation to pasture (Table 3). Also, the
184 pore morphology observed for native vegetation showed more complex packing pores
185 than in the pasture. In the pasture soils, there was a reduction in complex packing pores
186 and an increase in policoncave vughs in both layers whereas planar pores were
187 generally identified in the subsurface layer (Table 3).

188 The porosity of soil under sugarcane was lower than pasture only for the 0-10 cm
189 layer. The pore morphology analysis showed a further reduction of complex packing
190 pores from pasture to sugarcane and an increase in spherical and policoncave vughs and
191 channels (Table 3). When pedofeatures were analyzed, a reduction in biological
192 features from native vegetation soil to pasture was observed. However, the bio-pores,
193 characterized by the infilling of pores, and aggregates had no clear differences in
194 diameter. The LUC from pasture to sugarcane also led to a reduction in biological
195 features (pores, aggregates and coprolites) and the size of biological-derived aggregates
196 in the 0.1-0.2 m layer (Table 3).

197 *Micromorphometrical analysis*

198 Considering all sites, the total area of pores (Tap) was 1.2 to 2.1 times higher in
199 the surface layer (0-10 cm) of NV soils than pasture soils, whereas, sugarcane soil had
200 a Tap 1.5 to 2.2 times lower than pasture soils (Table 2). The same pattern of change
201 induced by LUC (Table 2) was observed at site scale, except for Lat_21S where PA did
202 not differ from NV. For the subsurface layer (10-20 cm), LUC did not induce changes
203 in Tap (Table 2) when considered at the regional scale. However, for Lat_23S, the NV
204 had a higher porosity than PA and SC, and the Tap of NV was higher than PA, which
205 was higher than SC at Lat_17S (Table 2).

206 For the top soil layer (0-10 cm), the soil pores at NV were rounded, elongated and
207 predominantly, complex pores. A reduction in complex and larger pores was observed
208 in accordance with a reduction in Tap with the LUC sequence; NV > PA > SC. This
209 indicates the reduction of the Tap was driven by large and complex pores representing
210 a loss of in the portion of complex packing pores, which is observed in Figure 2, where
211 the 10-20 cm soil layer was less sensitive to this alterations at Lat_21S and Lat_23S
212 (Figure 3 and 4).

213

214 **Discussion**

215 *Impacts of conversion from native vegetation to pasture on soil microstructure*

216 Land transition from native vegetation to pasture promoted reduction in porosity in
217 surface and subsurface soil layers at Lat_23S e Lat_17S. However, considering the data
218 at the regional scale, this conversion induced a reduction of the soil porosity only for
219 the superficial layers (Table 2). These results are in agreement with a higher soil bulk
220 density (BD), reduced macroporosity (MaP) and hydraulic conductivity (K_{fs}) of these
221 same pasture soils found by Cherubin *et al.*, (2016b). In addition, despite the
222 contrasting scales of evaluation, our micromorphometric analysis confirmed the results
223 obtained by on-farm visual evaluation by Cherubin *et al.* (2017), using the Visual
224 Evaluation of Soil Structure (VESS) method (Guimarães *et al.*, 2011). Based on VESS
225 assessment, pasture soils presented larger, harder and less porous aggregates than native
226 vegetation soils, resulting in lower overall soil physical quality in the 0-25 cm layer
227 (Cherubin *et al.*, 2017).

228 Cattle trampling may be the main driver of soil porosity reduction in pastures.
229 Mulholland & Fullen (1991) observed higher BD and penetration resistance in
230 pastureland soil after trampling using a thin section evaluation. Also, soils under native
231 vegetation can have higher organic matter inputs than the anthropic land uses,
232 increasing organic matter content (Franco *et al.*, 2015), which is responsible for
233 aggregate formation and stabilization (Six *et al.*, 2004), providing better soil physical
234 conditions (Cherubin *et al.*, 2016b).

235 The quantitative pore shape results showed a reduction in larger complex pores (Figures
236 3 and 4). This reduction did not alter the soil microstructure between these LUC's, but
237 changes were identified in the qualitative pore morphology analysis showing a decrease

238 in complex packing pores and an increase in spherical and policoncave vughs and
239 fissures from NV to PA (Table 3). These changes in the quantitative and qualitative
240 pore morphology assessments are also reflected in the changes in the related c/f
241 distribution with a transformation of enaulic and enaulic-porphyric related distribution
242 in NV to a porphyric-enaulic related distribution in PA. This morphological evidence
243 suggests an incipient compaction process in PA that caused by animal trampling and
244 poor pasture management that may reduce the benefits of soil macrofauna bioturbation,
245 which is partly responsible for the formation of these morphological features.
246 Compaction causes a reduction in the total volume of pores, and this reduction not only
247 alters pore morphology but changes the pore size distribution (Boivin *et al.*, 2006).
248 Therefore, the pore size and shape results obtained in this study can be useful indicators
249 or proxies for pore connectivity and tortuosity properties, which are important for the
250 evaluation of changes in key soil functions and services (Silva *et al.*, 2015; Rabot *et al.*,
251 2018), such as regulation of water fluxes and soil aeration, induced by land use change
252 and soil management practices. Although, the observation in 2D is a limitation in this
253 instance as assessment of pore connectivity in 3D is more appropriate for prediction of
254 some soil functions e.g. soil hydraulic behaviour. Further investigations combining both
255 the data from thin sections and X-ray imaging would improve our understanding
256 concerning the soil structure changes induced by agricultural land uses, as well as to
257 better establish the linkage between soil structure dynamics and the provision of soil
258 functions and ecosystem services.

259 *Impacts of conversion from pasture to sugarcane on soil microstructure*

260 Our results indicated a reduction on total porosity, mainly in the surface soil layer
261 (0-10 cm), when sugarcane was converted from pasture (Figure 2). The decrease of

262 packing pores observed in the micromorphological analyses (Table 3) confirms the
263 reduction of porosity and complex pores observed in the quantitative image analyses.

264 Overall, land transition from pasture to sugarcane increases the mechanical
265 compressive stresses applied on the soil surface, causing microstructural degradation
266 due to the coalescence of aggregates by compaction. The effect of this microstructural
267 degradation in this study is evidenced by the significant reduction in the complex pore
268 areas due to LUC, and in some sites, by the increase of less connected and more rounded
269 pores (Figures 3 and 4). This pore morphology change was also observed in the
270 decrease in the percentage of complex packing pores and increased percentage of
271 spherical and policoncave pores from PA to SC (Table 3). Microstructure changes from
272 a microgranular to blocky structure, both with well developed aggregates, and an
273 increase in porphyric c/f distributions, were also observed. These modifications in
274 microstructure, c/f distribution and pore morphology occur due to mechanical stress
275 (Silva *et al.*, 2015), and reduce soil aeration, water and nutrient uptake and crop yield
276 (Lipiec *et al.*, 1996). Soil compaction creates a restrictive environment for plant growth
277 due the physical impediment for roots development (Lipiec & Hatano, 2003) and the
278 reduction of soil aeration and consequentially, the redox potential (Eh) (Czyz, 2004),
279 creating a poor bio-chemical environment (Husson, 2013). Otto *et al.* (2011) showed
280 the inverse relationship between soil penetration resistance and diverse root parameters
281 (root length, area and density). The background for these limitations for plant and root
282 growth could lie in changes in microstructure and pore morphology due to LUC as we
283 have shown in this study.

284 Our results highlighted the urgent need for more sustainable management practices
285 to improve soil physical quality, especially those related to the improvement of soil
286 microstructure and pore morphology, mitigating the negative impact of biofuel

287 production. As sugarcane planting typically occurs between September and March (in
288 the central region of southern region in Brazil), which is also the rainy season, it is
289 important to avoid, or at least restrict, machinery traffic under high soil moisture
290 conditions and to encourage the introduction of conservation agriculture cropping
291 systems that reduce or eliminate soil tillage (Barbosa et al., 2019) and recommend the
292 use of cover crops as an alternative to prevent soil structure degradation and mitigate
293 other agronomic issues, such as weeds, pests and soil fertility. In this context cover
294 crops can also be used to improve soil structure at scales as fine as considered here
295 through root modification of the soil porous architecture (Bacq-Labreuil et al. 2019).
296 As there is an increasing interest in sugarcane straw to cogenerate bioelectricity or
297 produce 2G ethanol, maintaining part of the sugarcane straw in the field is an important
298 practice to improve several soil physical quality properties, such as soil structure, pore
299 size and morphology, BD, resistance to penetration, among others (Castioni *et al.*, 2018;
300 Castioni et al., 2019).

301 Other soil parameters, such as soil organic matter, soil fauna and soil texture
302 (Vreeken-Buijs *et al.*, 1998; Six *et al.*, 2004; Porre *et al.*, 2016; Bonetti *et al.*, 2017),
303 are important for soil structuring, and may contribute to the differences in changes in
304 pore morphology and size observed in this study. However, irrespectively of the site-
305 specific conditions (climatic, biological, chemical and physical), the results of the
306 micromorphological and micromorphometrical analysis, together with the physical
307 attributes provided by Cherubin *et al.* (2016b and 2017), show that the soil compaction
308 process occurs following LUC. More sustainable management practices are necessary
309 to maintain the soil physical properties (e.g. soil structure, pore morphology and size,
310 pore connectivity, etc.) that influence soil functions, (e.g. hydraulic conductivity, air

311 permeability, C storage, physical stability to resist against degradation, etc.) in Brazilian
312 sugarcane fields to achieve the expected productivities.

313

314 **Conclusions**

315 Land use change from native vegetation to pasture to sugarcane degraded the soil
316 microstructure, reducing the porosity of the soil and negatively influencing the pore
317 shape and size distribution, irrespectively of the soil texture and site environmental
318 conditions. As changes in soil microstructure and pore morphology affect important
319 soil hydrological and physical attributes, which in turn can negatively affect crop yield,
320 the adoption of more sustainable management practices in sugarcane fields (e.g.
321 reduced soil tillage, cover crop incorporation, straw retention and machinery traffic
322 control) is imperative to preserve and/or enhances soil structure, and consequently
323 sustain soil function in a productive capacity.

324

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487

489 **Table 1.** Soil physical and chemical characteristics of Lat_21S, Lat_17S and Lat_23S.

Soil attributes	Soil layer	Lat_21S			Lat_17S			Lat_23S		
		NV ¹	PA ²	SC ³	NV ¹	PA ²	SC ³	NV ¹	PA ²	SC ³
Sand (g/kg)	0-20	738	760	767	612	827	587	195	231	230
Silt (g/kg)	0-20	82	66	76	70	24	83	150	192	118
Clay (g/kg)	0-20	180	175	157	318	149	350	655	578	651
BD ⁴ (g/cm ³)	0-10	0.99	1.22	1.21	0.97	1.18	1.26	0.71	1.05	1.07
	10-20	1.08	1.34	1.29	1.01	1.26	1.19	0.83	1.03	1.06
C (g/kg)	0-10	21.8	13.3	11.1	15.6	9.5	10.8	36.7	36.4	18.9
	10-20	16.0	9.5	9.9	12.9	8.4	10.4	33.7	27.6	18.4

490 Values represent the mean of each land use. ¹ – Native Vegetation; ² – Pasture; ³ - Sugarcane; ⁴ – Bulk
 491 density. Adapted from Franco *et al.* (2015).

492

493

494 **Table 2.** Mean comparison of the total area of pores (Tap) in three land use in region scale (all evaluate sites) and for each location.

Location	Layer (cm)	Tap (%)		
		Land use		
		Native vegetation	Pasture	Sugarcane
Region Scale	0 – 10	36.5 ^{aA} ± 8.7	22.3 ^{bA} ± 1.3	12.8 ^{cA} ± 2.3
	10 – 20	36.3 ^{aA} ± 22.0	23.9 ^{aA} ± 6.4	18.9 ^{aA} ± 5.4
Lat_23S	0 – 10	37.2 ^{aA} ± 14.8	22.2 ^{bA} ± 8.2	10.2 ^{cB} ± 9.8
	10 – 20	33.4 ^{aA} ± 12.8	25.1 ^{bA} ± 12.2	24.8 ^{bA} ± 12.8
Lat_21S	0 – 10	27.3 ^{aA} ± 8.5	23.7 ^{aA} ± 6.5	14.5 ^{bA} ± 6.0
	10 – 20	15.8 ^{aB} ± 6.0	16.9 ^{aB} ± 6.8	14.7 ^{aA} ± 8.6
Lat_17S	0 – 10	45.1 ^{aB} ± 8.3	21.1 ^{bB} ± 3.1	13.8 ^{cB} ± 3.0
	10 – 20	59.6 ^{aA} ± 10.6	29.6 ^{bA} ± 6.0	16.5 ^{cA} ± 6.0

495 Different lowercase letter indicates statistical difference between the land use, and uppercase letter indicates the statistical difference
 496 between layers by Duncan test with 5% probability.

Table 3 Micromorphological description of the different land uses of two soil layers at Lat_23S.

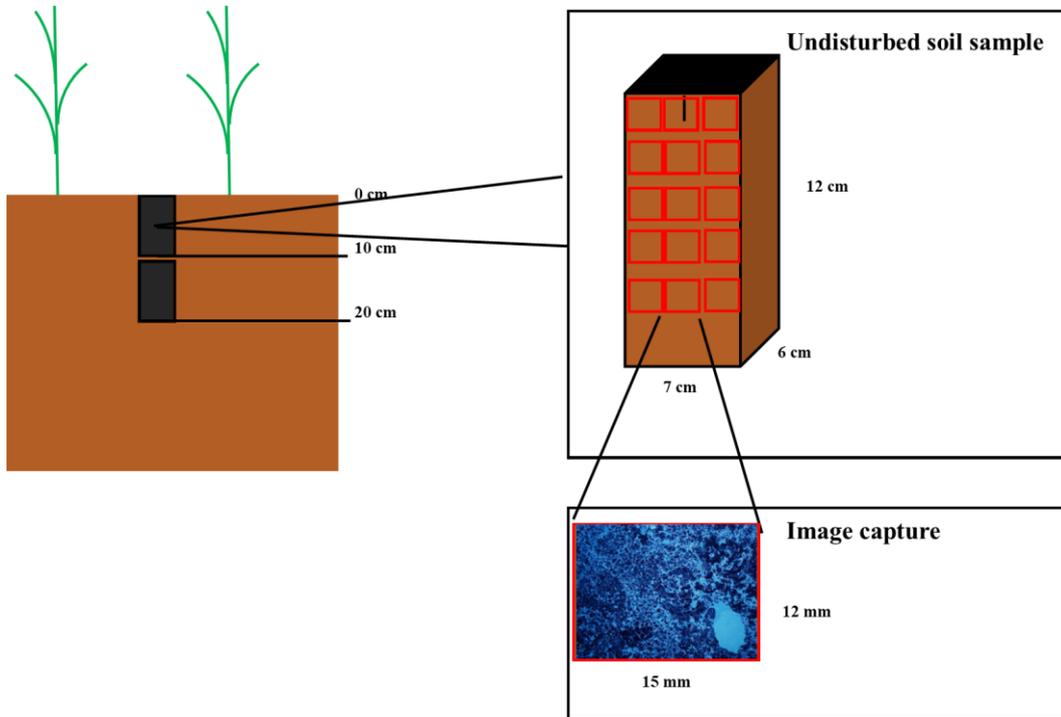
		Native Vegetation		Pasture		Sugarcane	
		0-10 cm	10-20 cm	0-10 cm	10-20 cm	0-10 cm	10-20 cm
Soil matrix Composition	Coarse material	25 %	25 %	30 %	35 %	35 %	30 %
	Fine Material	35 %	40 %	40 %	35 %	40 %	40 %
	Porosity	40 %	35 %	30 %	30 %	25 %	30 %
c/f Related Distribution*	Porphyric	97 %	97 %	95 %	98 %	99 %	95 %
	Porphyric-enaulic	-	-	5 %	2 %	1 %	5 %
	Enaulic-porphyric	2 %	2 %	-	-	-	-
	Enaulic	1 %	1 %	-	-	-	-
Coarse Material	The coarse material is composed by polycrystalline quartz, sub accommodated and poorly selected.						
Fine Material	The fine material is composed by clay and iron oxides.						
Pores	Complex packing	60 %	60 %	50 %	40 %	30 %	40 %
	Spherical and policoncave vughs	15 %	20 %	25 %	30 %	30 %	30 %
	Channels	15 %	10 %	15 %	15 %	20 %	10 %
	Fissures	10 %	10 %	10 %	15 %	10 %	20 %
Microstructure	Predominantly micro-granular with strong to moderate pedality and partially accommodated.					Complex microstructure composed by one predominantly micro granular with strong to moderate pedality and partially accommodated zone; and the other zone	

composed by subangular blocks with strong pedality and partially accommodated.

Pedofeatures	Biological features	30 %	30 %	25 %	25 %	15 %.	20 %
	Charcoal	5 %	5 %	1-5 %	1-5 %	1-5 %	1-5 %
Basic Organic Material	Biological pores diameter (mm)	0.6 to 3	0.2 to 2.2	0.1 to 3.7	0.4 to 2.5	0.5 to 3.4	0.4 to 1.9
	Biological aggregates diameter (mm)	0.1 to 0.7	0.1 to 0.5	0.2 to 0.6	0.1 to 0.7	0.1 to 0.5	0.2 to 0.4
	Coprolite				Present		

498 *c/f: Ratio between coarse (c) and fine (f) material.

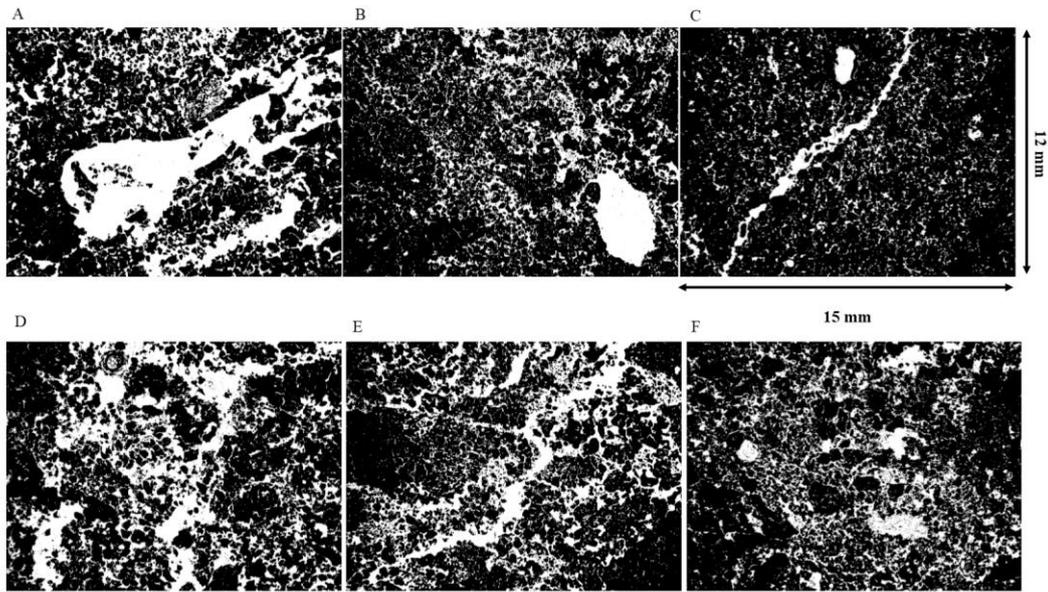
499 **FIGURE CAPTIONS**



500

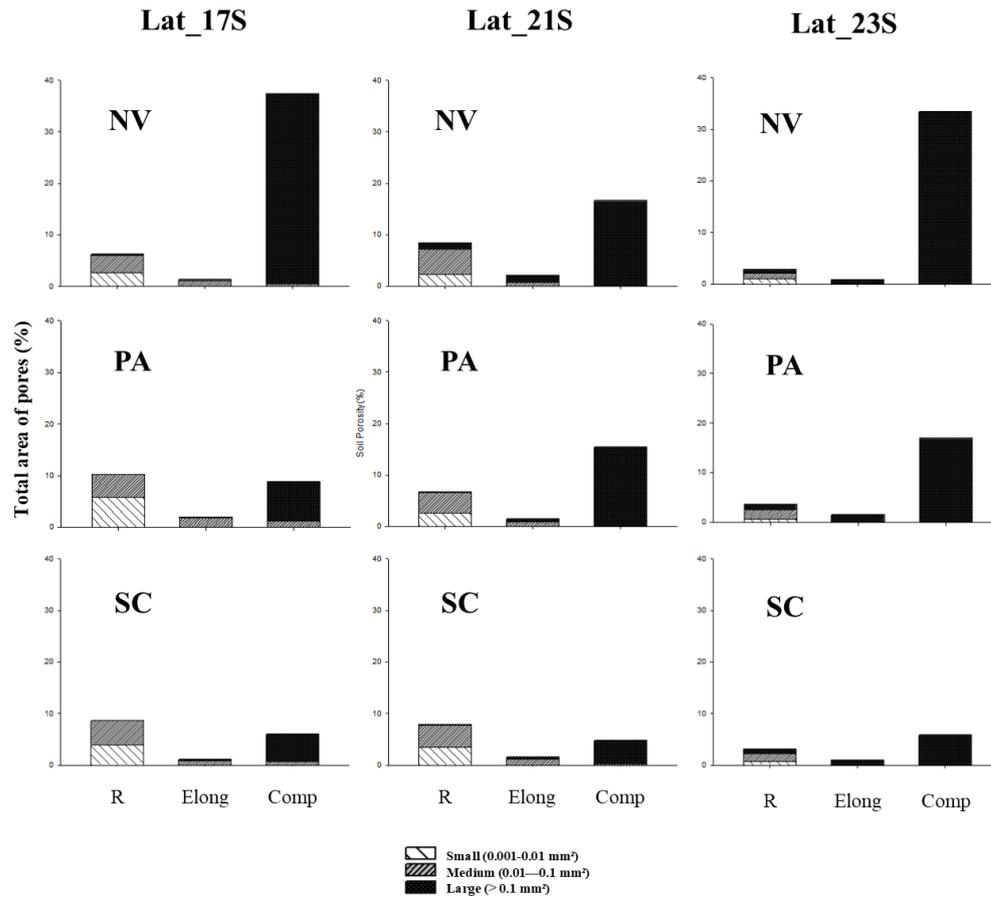
501 **Figure 1.** Illustration of soil sampling procedure adopted in the field and details of
502 orientation and scale of samples.

503



504

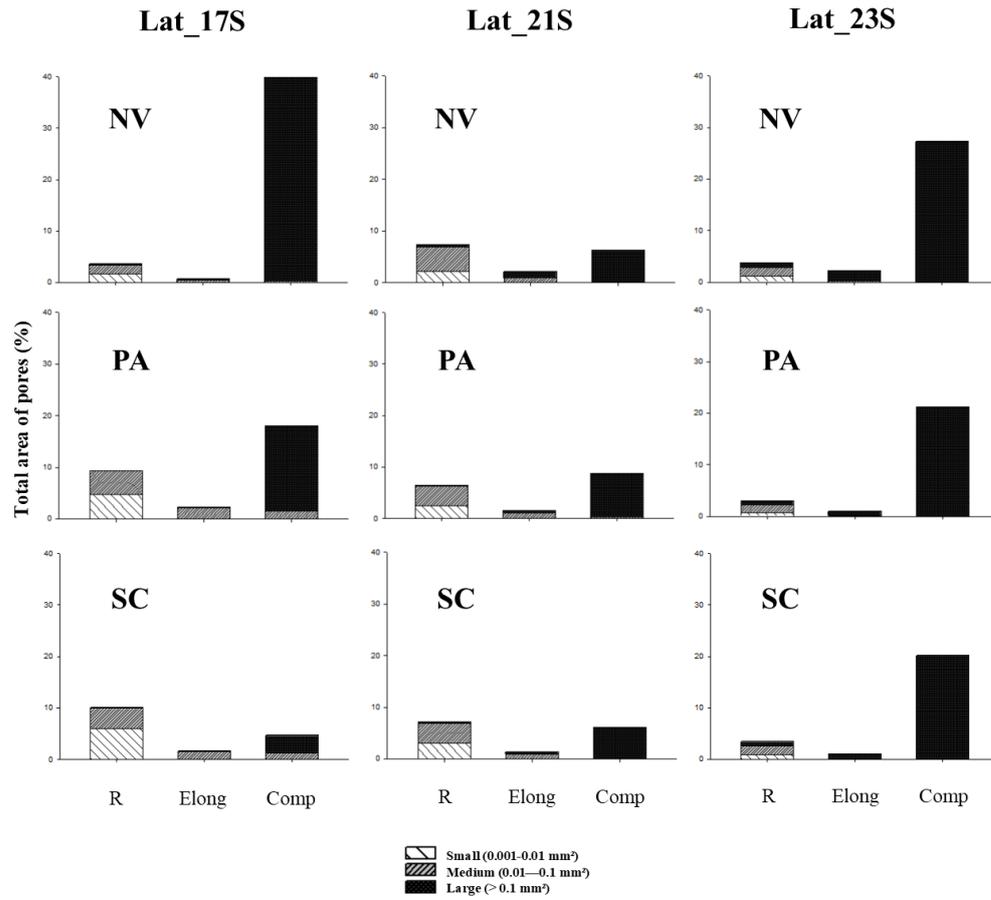
505 **Figure 2.** Binary microphotographs of representative thin section's areas (180 mm²) of
506 the 0 – 10 cm soil layer of native vegetation (A), pasture (B) and sugarcane (C); and
507 the 10 – 20 cm soil layer of native vegetation (D), pasture (E) and sugarcane (F) where
508 black is soil matrix and white is the pore space of microaggregates coalescence.



509

510 **Figure 3.** Pore shape and size distribution for 0-10 cm soil layer. R, Rounded; Elong,

511 Elongated; Comp, Complex; SC, sugarcane; PA, pasture; NV, native vegetation.



512

513 **Figure 4** Pore shape and size distribution for 10-20 cm soil layer. R, Rounded; Elong,

514 Elongated; Comp, Complex; SC, sugarcane; PA, pasture; NV, native vegetation.