1	Aroma binding and stability in brewed coffee: a case study of 2-furfurylthiol
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17	Abstract: The aroma stability of fresh coffee brew was investigated during storage over
18	60 minutes, there was a substantial reduction in available 2-furfurylthiol (2-FFT) (84%),
19	methanethiol (72%), 3-methyl-1H-pyrole (68%) and an increase of 2-pentylfuran
20	(65%). It is proposed that 2-FFT was reduced through reversible chemical binding and
21	irreversible losses. Bound 2-FFT was released after cysteine addition, thereby
22	demonstrating that a reversible binding reaction was the dominant mechanism of 2-FFT
23	loss in natural coffee brew. The reduction in available 2-FFT was investigated at
24	different pH and temperatures. At high pH, the reversible binding of 2-FFT was shown
25	to protect 2-FFT from irreversible losses, while irreversible losses led to the reduction
26	of total 2-FFT at low pH. A model reaction system was developed and a potential
27	conjugate, hydroxyhydroquinone, was reacted with 2-FFT. Hydroxyhydroquinone also
28	showed 2-FFT was released after cysteine addition at high pH.
29	Key words: coffee brew aroma stability; 2-furfurylthiol (2-FFT); reversible and
30	irreversible degradation; binding site stability
31	Chemical compounds studies in article: 2-furfurylthiol (PubChem CID 7363); L-
32	cysteine (PubChem CID 5862); hydroxyhydroquinone (PubChem CID 10787); Sodium
33	dihydrogen phosphate (PubChem CID: 23672064); disodium hydrogen phosphate
34	(PubChem CID: 24203); hydrochloric acid (PubChem CID: 313); sodium hydroxide

35 (PubChem CID: 14798); 3-heptanone (PubChem CID: 7802)

36

37 1. Introduction

After coffee roasting, furans, pyrazines, thiols, aldehydes and many other volatiles 38 39 are present in the headspace (Yang et al., 2016) and together form the complex aroma profile of coffee brew (Semmelroch & Grosch, 1995). However, some of these coffee 40 aroma compounds are unstable during storage of roasted coffee and coffee brew 41 (Dulsat-Serra, Quintanilla-Casas, & Vichi, 2016). This aroma deterioration results from 42 the interaction with the matrix (Fisk, Boyer, & Linforth, 2012; Yu et al., 2012) and 43 aroma degradation, and is termed aroma staling (Hofmann & Schieberle, 2002, 2004; 44 45 Müller & Hofmann, 2007). 2-Furfurylthiol (2-FFT), as a sulfur compound in coffee, has been established as 46 one of the key aromas that contribute to the characteristic flavour of coffee based on 47 48 sensory studies and model dilute experiments (Blank, Sen, & Grosch, 1992; Hofmann & Schieberle, 2002; Semmelroch & Grosch, 1995). However, 2-FFT rapidly reduces 49 during coffee brew processing or storage due to these staling reactions. This loss could 50 cause a significant reduction of sulfury-roasty aroma and is partially responsible for the 51

52 inferior sensory quality of aged coffee brews (Hofmann, Czerny, & Schieberle, 2001;

53 Mayer, Czerny, & Grosch, 2000; Semmelroch & Grosch, 1996).

The reduction of available 2-FFT during coffee staling can be divided into reversible and irreversible staling events (Charles-Bernard, Kraehenbuehl, Rytz, & Roberts, 2005; Guichard, 2002). The irreversible losses of 2-FFT are presumed to be due to physical diffusion/volatile loss and chemical degradation reactions, such as polymerization or oxidation (Blank et al., 2002; Charles-Bernard, Kraehenbuehl, et al., 59 2005). This irreversible fraction is very hard to regenerate. Reversible losses are 60 believed to be mainly through covalent bonding to non-volatile components in the 61 coffee matrix, it is proposed that this 2-FFT lost due to reversible reactions could be 62 subsequently released again by cysteine addition (Müller & Hofmann, 2007; Mestdagh,

63 Davidek, Chaumonteuil, Folmer, & Blank, 2014; Sun et al., 2018).

Previous studies have shown that the thiol group of 2-FFT is a good nucleophile 64 and could be involved in nucleophilic and radical reactions (Rowe, 2009). Through the 65 binding reaction, 2-FFT could be reversibly bound to conjugates in coffee brew, such 66 67 as 1, 4-bis (5-amino-5-carboxy-1-pentyl) pyrazinium radical cation (CROSSPY) that is found in coffee melanoidins (Hofmann & Schieberle, 2002; Tominaga, Blanchard, 68 Darriet, & Dubourdieu, 2000). More recent studies have reported that 69 70 hydroxyhydroquinone (HHQ), one of the chlorogenic acid degradation products, was the dominant conjugate to bind 2-FFT (Müller & Hofmann, 2007). In this binding 71 reaction, 2-FFT is bound by HHQ through the reactive quinone converted from HHQ, 72 leading to a rapid reduction of 2-FFT (Fig. 1) (Müller, Hemmersbach, van't Slo, & 73 Hofmann, 2006; Müller & Hofmann, 2007). Hofmann and Schieberle showed that this 74 75 covalent binding could be established within 15 min (Hofmann & Schieberle, 2002) Cysteine has been shown previously to release 2-FFT that is bound by coffee 76 matrix (Darriet, Tominaga, Lavigne, Boidron, & Dubourdieu, 1995; Mestdagh et al., 77 2014). After cysteine addition, the bound form of 2-FFT is competitively replaced by 78 cysteine due to its mercapto structure and high reducing properties. High cysteine 79

concentration could also prevent 2-FFT from forming dimers (Rowe, 2009). Cysteine

addition has also been shown to reversibly release bound 2-FFT in coffee brew. This 81 has been used to determinate the total: bound 2-FFT in coffee brews. Hereinafter "total 82 2-FFT" refers to the whole available 2-FFT in the coffee brew at any time point, 83 including both free 2-FFT and reversibly bound 2-FFT. "Free 2-FFT" means the 2-FFT 84 fraction that exists in free form in a coffee brew (Sun et al., 2018). 85 pH is important for 2-FFT formation and loss in coffee brew. High pH conditions 86 favor the formation of 2-FFT during coffee roasting and also increased 2-FFT loss 87 during heating in model systems (Hofmann & Schieberle, 1998; Kumazawa & Masuda, 88 2003). However, the impact of pH on coffee aroma binding in coffee brews, and the 89 relative understanding of free: bound 2-FFT in the coffee brew and their relative 90 stabilities are not well understood and is therefore the focus of this study. 91

92 In this study, coffee aroma staling was evaluated and the mechanism behind 2-FFT loss investigated. The stability of free versus total 2-FFT was evaluated. Reversibly 93 bound 2-FFT was released by the addition of cysteine enabling the calculation of the 94 reversibly bound and irreversibly lost 2-FFT fractions with the further calculation of 95 losses of 2FFT through volatilization. The effects of pH and temperature on 2-FFT 96 binding capacity was evaluated for coffee brew. To elucidate the pH effect on potential 97 conjugate, HHQ, a model system was developed to explain the interaction of HHQ and 98 2-FFT with or without cysteine addition. Aroma loss through binding reactions is a 99 common costly problem for the soluble coffee industry, Robusta coffee is commonly 100 101 used in this field, therefore Robusta coffee was selected as the target for this study.

102 2. Materials and methods

2.1 Materials and reagents 103

104	2-furfurylthiol (purity \geq 98%), hydroxyhydroquinone (purity \geq 99%),
105	homogenous series of <i>n</i> -alkanes (C6-C26) standard were purchase from Sigma-Aldrich
106	company (Poole, UK). Sodium dihydrogen phosphate (purity ≥ 99%), disodium
107	hydrogen phosphate (purity $\geq 99\%$), sodium hydroxide (purity $\geq 97\%$), methanol
108	(purity \geq 99%), concentrated hydrochloric acid (purity \geq 36.5%), L-cysteine (purity \geq
109	98%), 3-heptanone (purity \geq 99%) were purchased from Acros Organic company (New
110	Jersey, USA). Robusta washed coffee beans from Vietnam were purchased from
111	Pennine Tea and Coffee <mark>C</mark> ompany (Halifax, UK).
112	2.2 Coffee sample preparation
113	Green Robusta washed coffee beans (250 g) were roasted by a convection oven
114	(Mono Equipment, Swansea, UK). Before roasting, the oven was pre-heated for 5 min
115	to reach 225 °C. The coffee beans were roasted for 15 min at 225 \pm 2 °C and, after
116	cooling to 20 °C by convective cold air (around 10 °C cooling for 5 min), were ground
117	using a coffee grinder for 11 seconds (KG 49, Delonghi, Australia). The ground coffee
118	was screened by a metal sieve (700 μ m, Endecotts, Essex, UK) to control particle size.
119	The color of roasted ground coffee was analyzed by an UltraScan Pro (Hunterlab, US)
120	in a 1 cm Petri plate. The result was: L^* (lightness) = 41 ± 0.3, a^* (red, green) = 3.74 ±
121	0.2, b^* (yellow, blue) = 3.81 ± 0.3. The L^* , a^* and b^* values were in accordance with
122	the optimal roasting degree of medium roasting (Mendes, de Menezes, Aparecida, &
123	Da Silva, 2001). All ground samples were stored at -80 °C under nitrogen gas. Before
124	brewing, ground samples were defrosted at 15 °C for 15 min.

125	The ground coffee (9 g) was brewed using a French press (3 Cup Black Cafetière,
126	Argos, UK). Before brewing, the French press was rinsed by deionized water (around
127	92 °C) to preheated it for 10 seconds. Then the ground coffee (9 g) was mixed by
128	deionized water (180 mL, 92 °C) in the French press coffee maker. The plunger was
129	pressed down until the filter touched the coffee liquid surface and stood for 4 min before
130	depressing the plunger. The resulting fresh coffee liquid was shaken gently in French
131	press for 5 seconds to make coffee brew uniform.
132	In order to minimize the volume of headspace and exclude the aroma physical
133	diffusion, the coffee brew was poured from French press to fill full an amber vial of 40
134	mL which was sealed by screw top with PTFE-lined silicone septa (Supelco, Sigma
135	Aldrich, UK). The sealed amber vials with fresh coffee were cooled to 40 $^{\circ}$ C by water

- bath and following experiments were started from these "sealed amber-vial samples"
- 137 respectively.
- 138 2.3 Coffee aroma stability
- The sealed amber-vial samples were stored in a water bath (FB60307, Fisher Brand, UK) up to 60 min (0, 15, 30, 45, 60 min) at 40 °C prior to solid-phase microextraction-gas chromatography mass spectrometry (SPME-GC-MS).
- 142 2.4 Effect of storage time on free and total 2-furfurylthiol concentration under open143 vial condition
- 144 Five milliliter of fresh coffee brew was transferred from the sealed amber-vial
- samples (prepared from item 2.2) to GC headspace vials (20 mL, 22.5 mm × 75.5 mm,
- 146 Sigma-Aldrich, UK) by pipette (Argos, UK). The GC headspace vials were left open

- 147 (open-vial condition) to allow aroma volatilization so that the physical diffusion loss
- 148 was evaluated. The headspace vials were stored for up to 60 min at 40 °C in a water
- bath before sealing for free and total 2-FFT SPME-GC-MS analysis.
- 150 2.5 *Effect of pH and storage temperature on free and total* 2-*FFT in coffee brew*
- 151 The sealed amber-vial samples were adjusted to pH 3 to 9 with 1 M hydrochloric
- acid solution and 1 M sodium hydroxide solution. Samples were stored at 20, 55 or
- ¹⁵³ 90 °C in a water bath for 1 h respectively. After cooling down to 40 °C by water bath,
- 154 the free 2-FFT concentration of samples was analyzed by SPME-GC-MS. Another
- aliquot of 40 mL amber-vial sample was subjected to the same pH incubation (without
- 156 different temperature storage). Then the total 2-FFT concentration was analyzed by
- 157 SPME-GC-MS.
- To further study the pH effect on 2-FFT binding and release, additional experiments were also performed:
- 160 **Cysteine addition after pH incubation:** The sealed amber-vial samples (40 mL) were
- adjusted to pH 3 or 9, and stored for 1 h in water bath at 55 °C. After that, the pH of
- samples was adjusted back to 6 prior to total 2-FFT analysis by SPME-GC-MS.

163 **Cysteine addition before pH incubation**: Cysteine of 0.64 g was added into the sealed

- amber-vial samples (40 mL) after cooling down to 40 °C by water bath (to prevent
- 165 cysteine from undergoing Maillard reaction at high temperature) (Sun et al., 2018) and
- stirred at 1000 rpm for 5 min on a magnetic stirrer (IKA RET control-visc, UK). The
- 167 pH of samples was adjusted to 3 and 9. After 1 h storage in a water bath at 55 °C, the
- total 2-FFT was analyzed via SPME-GC-MS.

The buffer solution was prepared using disodium phosphate solution (0.2 M), 170 sodium dihydrogen phosphate solution (0.2 M) and sodium hydroxide solution (0.1 M) 171 and hydrochloric acid solution (0.1 M) to adjust the pH to 3 or 9. An aqueous solution 172 of hydroxyhydroquinone (HHQ) (0.004 g HHQ diluted in 30 mL Milipore water) was 173 added (300 μ L) into phosphate buffer solution (40 mL) and placed in amber vials (40 174 mL), the samples was then stored for 1 h in a water bath at 40 °C to allow incubation 175 (Müller & Hofmann, 2007). 2-FFT (36 µg in 31.7 µL of methanol) was added into HHQ 176 buffer solution. 2-FFT was reacted with HHQ for 1 h at 40 °C prior, to evaluate free 177 and total 2-FFT by SPME-GC-MS. 178

179 2.7 Quantification of volatile compounds

180 The quantification of 2-FFT was carried out using the internal standard quantification method (Sun et al., 2018). 3-Heptanone (4.1 μ g in 5 μ L of methanol) was 181 added into calibration solution as an internal standard (IS) to accommodate for 182 instrument drift. For coffee brew, the calibration curve was established by adding 2-183 FFT (0, 1.8, 3.6, 7.2, 14.4 µg in 31.7 µL of methanol) into a prefabricated coffee model 184 which had a similar matrix to coffee brew. To prepare this coffee model, cysteine (1.6 185 g) was presented into fresh coffee brew (200 mL) at 40 °C. The coffee brew was dried 186 by rotary vacuum evaporation at 40 °C. The dried sample was then dissolved in 200 187 mL of ultra-pure water. This part of experiment (from adding cysteine to samples 188 dissolved into water) was repeated once more to release maximum 2-FFT from the 189 coffee brew (Sun, et al., 2018). For the model experiment, the calibration curve was 190

191	established by adding 2-FFT (0, 9, 18, 36, 44 μ g in 31.7 μ L of methanol) into the
192	phosphate buffer solution. The concentration of 2-furfurylthiol could be calculated by
193	the calibration curve made by relative peak area (peak area of standard 2-FFT to that of
194	internal standard) to the concentration of standard 2-FFT (y = 0.0055 x + 0.0381, R^2 =
195	<mark>0.998)</mark> .
196	Approximate quantification of all volatiles for the aroma stability study (Item 2.3)
197	were calculated by comparing the GC peak areas of volatiles with the peak area of 3-
198	heptanone internal standard, using a response factor of 1 to give relative concentrations
199	(Liu, Yang, Linforth, Fisk, & Yang, 2019). The percentage of volatile was calculated by
200	the comparison of their relative concentration after storage to their initial relative
201	concentration in fresh coffee. All samples were analyzed in one run in randomized order
202	2.8 SPME-GC-MS
203	Five milliliters of samples were placed into GC headspace vials (20 mL, 22.5 mm
204	× 75.5 mm, Sigma-Aldrich, UK) from the amber vials by pipette. 3-heptanone (4.09 μ g
205	in 5 µL of methanol) (Caporaso, Whitworth, Cui, & Fisk, 2018; Liu et al., 2019) was
206	added into the samples as the internal standard before vials were sealed by screw top
207	with PTFE-lined silicone septa. For total 2-FFT analysis, cysteine powder was added
208	to each vial before closure at 16 g/L (Sun et al., 2018) so that the bound form of 2-FFT
209	was released to the free form. With cysteine, the free 2-FFT currently detected by
210	SPME-GC-MS was the amount of total 2-FFT. Aroma analysis for free 2-FFT in coffee

- 211 brew had no cysteine addition.
- 212 The analysis was carried out using a gas chromatography coupled with the Single-

213 Quadrupole Mass Spectrometer (Thermo Fisher Scientific, Hemel Hemptead, UK). A 214 50/30 μm DVB/CAR/PDMS SPME fiber (Supelco, Sigma Aldrich, UK) was used to 215 extract volatile aroma compounds from the samples headspace. Samples were 216 incubated at 40 °C for 2 min. The SPME extraction procedure was presented at 40 °C 217 for 10 min. The fiber desorption was at 250 °C for 3 min in the injector and then 218 analyzed by GC-MS.

A ZB-WAX column (30 m \times 0.25 mm I.D., 1 μ m film thickness; Phenomenex Inc, 219 Macclesfield, UK) was used to separate constituents. Analytical GC conditions in GC-220 MS were as follows: Helium was used as carrier gas (1 mL/min). Injector temperature 221 was 250 °C. Splitless injector mode was used. The oven temperature program was held 222 at 40 °C for 5 min, then raised at a rate of 3 °C /min to 160 °C. After that, oven 223 224 temperature was raised at 20 °C /min to 240 °C and held for 2 min; Energy voltage was 70 eV. Single ion monitoring (SIM) and Scan mode was used (30 - 300 m/z). Samples 225 were run in triplicate randomized order. 226

2-FFT was identified by comparing the detected 2-FFT mass spectra and linear 227 retention indices (LRI) with 2-FFT standard mass spectra and LRI value from reference 228 data libraries (NIST 11 and WILEY 07 databases). 114 m/z was used as the 229 characteristic ion for 2-FFT quantification (Mestdagh et al., 2014; Tominaga & 230 Dubourdieu, 2006). The identification of other volatiles was identified by comparing 231 the detected volatiles mass spectra and LRI value with the mass spectra and LRI value 232 from data libraries (NIST 11 and WILEY 07 databases). The linear retention indices 233 values were calculated with homogenous series of n-alkanes (C6-C26) standard in the 234

235 same GC conditions.

236 2.9 Statistical Analysis

Data were presented as mean values \pm standard deviation. Statistical analysis was conducted by SPSS 19.0 (SPSS Inc., Chicago, USA). Duncan's multiple range tests was used, and p < 0.05 was considered as significant. All samples were measured in triplicate.

- 241 **3. Results and discussion**
- 242 *3.1 Coffee aroma stability*

To identify which coffee volatiles are directly impacted by chemical changes over 243 storage, samples were stored in fully filled vials. This allowed the investigation of 244 storage time on aroma, while excluding the effects of physical diffusion and losses into 245 246 the headspace. Eighty-nine volatile aroma compounds were detected in the fresh coffee brew and after 1 h storage at 40 °C, there were substantial changes over storage time in 247 the aroma concentration of four aroma compounds, shown in Fig. 2 A to D. 85 of the 248 249 volatile aroma compounds did not change their relative headspace concentration by more than 50%. The headspace relative concentration of 2-furfurylthiol, methanethiol 250 and 3-methyl-1H-pyrrole decreased by 84%, 72%, 68% respectively; 2-pentylfuran 251 increased to 165% compared to its relative concentration in coffee brew before storage. 252 Of all the compounds evaluated, 2-FFT showed the fastest decrease, with a 68% 253 reduction in free 2-FFT within 15 min. It should also be noted that of the four 254 compounds that changed, 2-FFT has the highest odour activity value (OAV) and highest 255 flavor dilution factor (FD factor) by aroma extract dilution analysis (Semmelroch & 256

257	Grosch, 1995; Vermeulen, Gijs, & Collin, 2005), and a sulfury-roasty aroma, which is
258	highly characteristic of coffee aroma (Akiyama et al., 2008; Dulsat-Serra et al., 2016).
259	Therefore, 2-FFT was selected as the key instable aroma compound for further studies.
260	3.2 Effect of storage time on reversible and irreversible binding of 2-FFT under open-
261	vial condition
262	Total available 2-FFT can be lost by irreversible losses and reversible binding
263	reactions (Charles-Bernard, Kraehenbuehl, et al., 2005; Guichard, 2002). These two
264	losses affect the total and potential for free 2-FFT in coffee. As the total 2-FFT includes
265	free 2-FFT and reversibly bound 2-FFT, the difference between the total 2-FFT amount
266	and free 2-FFT amount is defined as "reversibly bound 2-FFT"; On the other hand,
267	irreversible losses occurred and increased during coffee storage. So, the difference of
268	total 2-FFT between original and stored coffee is defined as "irreversibly lost 2-FFT".
269	Thus, the total and free 2-FFT of the stored coffee were determined to investigate the
270	extent of reversibly bound and irreversibly lost 2-FFT during storage. Over 1h storage,
271	the concentration of both total and free 2-FFT decreased significantly (Table 1). The
272	biggest change happened within the first 30 minutes and then plateaued. Total 2-FFT
273	decreased from $\frac{153}{153}$ to $\frac{138}{\mu g/L}$ (10% loss) (p<0.05), and free 2-FFT decreased from
274	$\frac{1}{3.4}$ to 0.4 µg/L (88% loss) over 1 h storage (p<0.05). The total 2-FFT concentration was
275	much higher than free 2-FFT concentration, indicating that a large amount of 2-FFT
276	was reversibly bound to coffee components within the fresh coffee brew.
277	Using the free and total 2-FFT amounts, the reversibly bound and irreversibly lost

278 2-FFT amount could be calculated using the equation below. After a defined period of

279 storage (X min).

- Reversibly bound 2-FFT = Total X Free X 280 Irreversibly lost 2-FFT = Total 0 - Total X281 X: the concentration of aroma in coffee brew stored at time X min; 282 *O*: the concentration of aroma in original fresh coffee brew at time 0 min. 283 Reversibly lost 2-FFT and irreversibly lost 2-FFT are presented in Fig. 3. Over the 284 1 h storage period, the proportion of reversibly bound 2-FFT reduced from 149.6 to 285 137.1 μg/L). The irreversibly lost 2-FFT increased from 0 to 15.5 μg/L. These low 286 $(\sim 10\%)$ losses suggest that the volatilization and irreversible chemical degradation have 287 a limited effect on 2-FFT loss compared to the predominant reversible binding in 288 natural coffee brew (~pH 6.2). Due to this, in natural coffee (~pH 6.2), reversible loss 289 290 is proposed to be the main reason for the loss of free 2-FFT, which is expected to play a significant role in the loss of the characteristic aroma of coffee during staling 291 (Hofmann & Schieberle, 2002; Mayer & Grosch, 2001). 292 3.3 The effect of pH and storage temperature on free 2-FFT concentration in coffee 293 brew and the model system 294 Coffee brew samples were stored at different pH and temperatures to investigate 295
- 296 the impact on free 2-FFT stability. Both pH and temperature (Fig. 4 A) had a significant
- ²⁹⁷ impact on free 2-FFT concentration. After 1 h incubation at elevated temperatures, free
- 298 2-FFT concentration decreased significantly (2.7 to 0.4 μg/L at 20 °C; 1.9 to 0.5 μg/L
- 299 at 55 °C; 1.6 to 0.5 μg/L at 90 °C), the loss of free 2-FFT was greatest at highest pH
- values at all temperature (Fig. 4 A). This resulted in almost all free 2-FFT being

reversibly bound by the coffee matrix at pH 6-9. The same trend was also found in the model system (Fig. 4 B) where the concentration of free 2-FFT decreased from 383 to $0.02 \mu g/L$ when the pH of buffer solution was increased from 3 to 9.

- This pH sensitivity can be explained by the impact on quinone. As discussed 304 previously, during the reversible binding reaction between 2-FFT and HHQ, the 305 reactive quinone is formed from HHQ first, then it reacts with 2-FFT (Fig. 1) (Müller 306 & Hofmann, 2007). The highly reactive quinone is unstable at low pH due to its 307 carbonyl property (Li & Chen, 2005). So, under low pH conditions, the reversible 308 binding reactions are inhibited resulting in a greater concentration of free 2-FFT at low 309 pH as shown in Fig. 4 A and B. The inverse of this means that at high pH the conversion 310 of quinone is not be inhibited and more free 2-FFT is reversibly bound. 311 312 The 2-FFT in coffee brew stored at 20 °C had the highest free 2-FFT concentration (Fig. 3 A), while at 90 °C there was less free 2-FFT compared to samples stored at 20 313 and 55 °C. This result suggests that the high temperature may increase the efficiency of 314 315 the reversible binding reaction leading to more efficient 2-FFT binding.
- 316 *3.4.* The effect of pH on total 2-FFT concentration in coffee brew and model system
- 317 pH also impacted the total 2-FFT level. After 1 h storage, total 2-FFT showed a
- significant increase when the incubation pH was increased (Fig. 4 C), increasing from
- 2.6 to $159 \ \mu g/L$. In the model system, the similar increase was found, but to a lesser
- 320 extent (351 to 524 μ g/L) (Fig. 4 D).
- 321 To study the reason for the low level of total 2-FFT at low pH and high level at
- 322 high pH value, 2-FFT releasing ability of cysteine was further studied as cysteine might

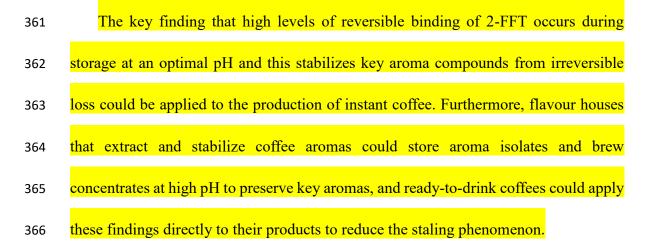
323	be sensitive to pH. Before cysteine addition, samples were incubated at different pH
324	and were adjusted back to pH 6, as cysteine has previously been shown to effectively
325	release reversibly bound 2-FFT at pH 6 (Mestdagh et al., 2014; Sun et al., 2018). After
326	control of the pH, the result still showed a low total 2-FFT level when stored at low pH
327	and high total 2-FFT level when stored at high pH (Fig. 5: Pre pH incubation). The total
328	2-FFT at pH 9 (105 μ g/L) was over twice as the amount retained when the coffee brew
329	was stored at pH 3 (50 μ g/L), suggesting that the 2-FFT releasing ability by cysteine
330	was not susceptible to pH.
331	With the addition of cysteine, total 2-FFT at pH 3 was much lower than pH 9 as
332	shown in Fig. 4 C. This low amount of total 2-FFT might be a result of irreversible
333	losses as the reversible binding is inhibited at low pH (Discussed in Item 3.3). To further
334	verify this irreversible loss effect on total 2-FFT, cysteine was added into coffee brew
335	before pH incubation (Fig. 5.). After pH incubation, total 2-FFT under both pH 3 and 9
336	incubation presented a much lower level (56.8 μg/L and 49.9 μg/L). This result
337	indicated that when the bound 2-FFT was released out of the "protection" of conjugates
338	(such as HHQ) prior to incubation, the total 2-FFT was equally lost by the irreversible
339	losses occurring during incubation. Suggesting that 2-FFT is protected by reversible
340	binding during incubation.
341	It should also be noted that the total 2-FFT concentration in the model system at
342	pH 3 (Fig. 4 D) was not degraded as much as the total 2-FFT in the coffee brew (Fig. 4

- C). Suggesting other effects caused by the matrix difference between the coffee and the
- 344 model system. In coffee brew, the matrix is much more complex and would increase

irreversible losses through other mechanisms such as radical delivery from Fenton
reaction (Charles-Bernard, Roberts, & Kraehenbuehl, 2005). This suggests that
additional coffee non-volatiles do contribute to 2-FFT irreversible losses (CharlesBernard, Kraehenbuehl, et al., 2005; Charles-Bernard, Roberts, et al., 2005).

349 4. Conclusion

This study identified four unstable aroma compounds in coffee brew, the 350 availability of which changed significantly over a 1h holding period, these were 2-351 furfurylthiol, methanethiol, 3-methyl-1-pyrole and 2-pentylfuran. 2-furfurylthiol 352 353 suffered the greatest losses and was selected for further investigation. It was shown that reversible binding with HHQ was the dominant reason for 2-furfurylthiol staling in 354 natural coffee. To further explain this reversible binding reaction, 2-FFT binding was 355 356 studied and showed that at low pH, the 2-FFT binding reaction to hydroxyhydroquinone (HHQ) is inhibited and the availability of free 2-FFT level increased compared to free 357 2-FFT at higher pH values. This work also showed that without the protection of 358 conjugates (reversibly bound to HHQ) in coffee brew, free 2-furfurylthiol could be 359 irreversibly lost. 360



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