

Abstract

 Parasitoids have the potential to suppress populations of their hosts and thus may play an important role in influencing the temporal and spatial dynamics of pest arthropods. Behavioural interactions between foraging females, collectively constituting 'mutual interference', can reduce host suppression. We use laboratory microcosms to assess the prevalence and consequences of mutual interference behaviour in a bethylid wasp, *Goniozus nephantidis* (Muesebeck) (Hymenoptera: Bethylidae), which is known to brood guard and to engage in agonistic contests for individual hosts and which is also an agent of biological pest

control. We hold host and parasitoid numbers constant and vary the degree of female-female

contact that can occur. Mutual interference is manifest in a considerable reduction in the

number of offspring produced when females are not fully isolated from each other, due to

effects operating at the early stages of offspring production. This mutual interference may

contribute towards the limited degree of host population suppression achieved when some

species of bethylids are deployed as agents of biological pest control and also has clear

potential to influence the efficiency of mass rearing of parasitoids prior to field release.

Introduction

 Parasitoid wasps are free-living as adults: females search for and attack hosts, usually the immature stage of another arthropod, laying their eggs on or in these hosts which then form the only food resource for their developing, parasitic, offspring, leading to the death of the host (Godfray, 1994; Jervis, 2005). Parasitoid efficiency in host attack, and consequently offspring production, may be tempered by behavioural interactions with other foraging females, termed 'mutual interference' (Hassell & Varley, 1969; Hassell, 1971, 2000; Rogers & Hassell, 1974; Beddington, 1975; Free et al., 1977; Begon et al., 1996; Driessen & Visser, 1997; Fellowes et al., 2005; Jervis et al., 2005; Kidd & Jervis, 2005). Interference interactions can take a variety of forms; seen broadly they include not only the rate of parasitism of the current generation of hosts but also influences on the size and sexual composition of the next generation of parasitoids (Visser & Driessen, 1991; Visser et al., 1999). Examples include time-wasting disruption of foraging for hosts without explicitly agonistic interactions between foraging females (Hassell, 1971, 2000; Cronin & Strong, 1993; Field et al., 1998; Wajnberg et al., 2004; Le Lann et al., 2011; Yazdani & Keller, 2015), aggressive patch or brood guarding (Hassell, 1971; Waage, 1982; Field et al., 1997; Goubault et al., 2005; Nakamatsu et al., 2009; Venkatesan et al., 2009a,b; de Jong et al., 2011; Hardy et al., 2013; Mohamad et al., 2015), clutch size and superparasitism 19 decisions differing in the presence, or anticipated presence, of competitors (van Alphen $\&$ Visser, 1990; Visser & Driessen, 1991; Visser et al., 1992; Visser, 1996; Field et al., 1998; Goubault et al., 2007a), and sex allocation decisions contingent on the number of ovipositing 'foundress' females present (Hamilton, 1967; Waage, 1982; Meunier & Bernstein, 2002; Irvin & Hoddle, 2006; Ode & Hardy, 2008; Luo et al., 2014).

 Empirical investigations of factors contributing to mutual interference have often been conducted from the perspective of behavioural ecology but there has also been great interest in their consequences for host–parasitoid population dynamics, in part because this contributes towards explaining population phenomena on the basis of natural selection (Hassell & May, 1973, 1989; Anderson, 1989; Cronin & Strong, 1993; Driessen & Visser, 1997; Visser et al., 1999; Bernstein, 2000; Meunier & Bernstein, 2002). Typically, mutual interference interactions reduce the per-host production of female offspring (the sex that attacks future generations of hosts) and will also be more prevalent at higher parasitoid densities: this density-dependent effect may be expected to contribute to the dynamic stability of host–parasitoid populations, but extreme effects of interference can also lead to predictions 34 that host populations will not be regulated (sufficiently suppressed) by parasitism (Rogers $\&$

 Hassell, 1974; Begon et al., 1996; Bernstein, 2000; Hassell, 2000; Kidd & Jervis, 2005). To practitioners of biological control, who deploy parasitoids against agricultural pests, the importance of mutual interference is thus that it reduces the likelihood that pest populations will be depressed below thresholds of economic damage (Anderson, 1989; Hassell & May, 1989). Although there have been many successful biocontrol programmes, attempts have also frequently failed because parasitoids have not established and because established parasitoids have not depressed the pest populations sufficiently (e.g., Mills, 2000; Kapranas & Hardy, 2014). In particular, parasitoids are often mass reared and innundatively released at high densities, which may artificially promote the occurrence of economically detrimental mutual interference interactions (Irvin & Hoddle, 2006; Ode & Hardy, 2008).

 In this study we use simple laboratory microcosms to assess the prevalence and consequences of mutual interference behaviour in a bethylid wasp, *Goniozus nephantidis* (Muesebeck) (Hymenoptera: Bethylidae). This parasitoid species is known to guard individual hosts and developing broods, to engage in agonistic contests for hosts prior to and following oviposition (Hardy et al., 2013), and is also mass reared for release in biological control programmes against coconut pests (Shameer et al., 2002; Mohan & Shameer, 2003; Venkatesan et al., 2003, 2007, 2009a,b; Lyla et al., 2006). Our purpose is to understand better the competitive and reproductive biology of brood-guarding parasitoids for which prior most investigations have been confined to dyadic interactions (Hardy et al., 2013), and to in turn suggest approaches towards enhancing the efficiency of their mass rearing and field release when utilized as agents of biological pest control. Venkatesan et al. (2009b) have already reported that individual *G. nephantidis* females produced more progeny (per host and also in total) when provided with just one host rather than multiple hosts simultaneously within a single chamber (a glass vial). They also reported that when the number of females within the chamber to which a single host was presented was varied, progeny production was greatest when just one female wasp was present: offspring production was thus greatest (per female and also in total) when one host and one female *G. nephantidis* were present (Venkatesan et al., 2009b). Here, we hold the number of hosts available per female (resource-weighted density; Lewontin & Levins, 1989) constant, at unity, and experimentally vary both spatial density (individuals per unit area) and the degree of physical sub-division between chambers within microcosms (the latter allowing parasitoid behaviour to determine the organism- weighted density; Lewontin & Levins, 1989). The overall hypothesis is that parasitoids will produce fewer offspring when there is more contact possible between reproductive adult females, even though in all cases one host per female is provided.

Biology of **Goniozus nephantidis**

 Goniozus nephantidis is a gregarious larval ectoparasitoid for which many aspects of behaviour and life history have been documented (Hardy et al., 1992; Cook, 1993; Hardy & Cook, 1995; Humphries et al., 2006; Goubault et al., 2007a,b, 2008; Venkatesan et al., 2009a,b). It is naturally associated with the coconut pest, *Opisina arenosella* Walker (Lepidoptera: Oecophoridae) in the Indian sub-continent (Mohan et al., 2004; Venkatesan et al., 2007, 2009a,b). *Opisina arenosella* larvae feed on the vascular tissues of coconut fronds by chewing, leading to affected leaves showing a burnt appearance, and also construct protective tunnels and galleries with silk, leaf parts, and frass (Mohan et al., 2004; Venkatesan et al., 2009a). Coconut crop yield losses due to *O. arenosella* can be as high as 45% and it can take up to 4 years for palms to regain normal yield after heavy infestation (Mohan et al., 2010). Adult *G. nephantidis* females forage for hosts by entering the galleries. On encountering a host larva, the adult female attacks it by injecting venom whereupon the host becomes paralysed within minutes and their development is arrested (idiobiontism). Eggs are laid onto the host 1-4 days later (Jayaratnam, 1941; Goubault et al., 2007b), the intervening period possibly being used to mature eggs (Stokkebo & Hardy, 2000; but see Goubault et al., 2007b). The number of eggs laid on a host is positively correlated with host size, with clutches ranging up to ca. 18 (Hardy et al., 1992). Eggs hatch about 1 day after oviposition and the larvae begin to feed on the host through punctures in its integument. During the period between paralysis and oviposition, females remain in close physical association with their hosts, aggressively guarding them against intruding conspecific females 23 which would otherwise utilize the unguarded host (Hardy & Blackburn, 1991; Petersen & Hardy, 1996). Mothers also remain with their broods after laying eggs, typically until the

offspring pupate around 7 days later, and during this period guard their eggs and developing

larvae against intruders (Hardy & Blackburn, 1991; Goubault et al., 2007b; Venkatesan et al.,

2009a). Mothers continue to mature eggs within their abdomens whilst guarding and

 surviving mothers have an increasing tendency to leave their broods, possibly to forage for further hosts (Goubault et al., 2007b).

 Although prior-ownership is an advantage in host-ownership contests, other factors, especially body size differences, contribute to determining the outcomes of agonistic encounters (Petersen & Hardy, 1996; Humphries et al., 2006; Hardy et al., 2013) and prior owners can thus be driven away from hosts they have paralysed (fighting is rarely fatal). Intruders that displace mothers will often destroy the brood and then lay their own eggs, but

 the probability of destruction followed by oviposition declines as the developmental stage of 2 the original brood increases (Hardy & Blackburn, 1991; Goubault et al., 2007b).

 Some offspring fail to mature, even in clutches that are not destroyed by intruders, with mortality of guarded offspring estimated at around 28% (Hardy & Cook, 1995), mainly due to low juvenile survival during earlier developmental stages. Surviving offspring eclose as adults approximately 2 weeks after oviposition: males eclose first and mate with their sisters, then adults disperse from the natal site over the following few days (Hardy et al., 8 1999). The sex ratios of broods at eclosion and dispersal are usually female biased (Hardy $\&$ Cook, 1995; Hardy et al., 1999; Venkatesan et al., 2009a,b).

Materials and methods

 Goniozus nephantidis cultures were maintained using the rice moth, *Corcyra cephalonica* Stainton (Lepidoptera: Pyralidae), as a factitious host (following Hardy & Blackburn, 1991; Hardy et al., 1992; Petersen & Hardy, 1996). *Corcyra cephalonica* was reared on a diet of glycerol, honey, corn meal, wheat bran, and yeast (Lizé et al., 2012). All cultures and experiments were carried out in a climate room at 27 °C, L12:D12, with relative humidity maintained at *ca*. 35% by evaporation from a water bath.

 We assessed the effects of host and parasitoid density and of spatial sub-division on parasitoid reproduction using experimental arenas excavated from polyethylene blocks and with transparent Plexiglas lids, with designs derived from the simpler contest arenas used and illustrated in Petersen & Hardy (1996) and Goubault et al. (2006). We used four types of arenas, termed block A-D (Figure 1), all excavated to a depth of 6 mm. Block A contained 25 chambers and each chamber was 1.7 cm in diameter. The total excavated area (floor area) 24 within the block of the chamber was 56.8 cm^2 . As wasps could not pass between the chambers, block A served as a control treatment in which interactions between adult females wasps were prevented. Block B was the same as A except that each chamber was connected to the others around it by a 1 cm long and 0.1 cm wide slot, through which adult *G. nephantidis* can pass with ease. Block B thus had an extra 4 cm^2 of floor area, an increase of 7% compared to A, but retained the same chamber size as block A and also as in previous studies (Petersen & Hardy, 1996; Goubault et al., 2006). Block C had the same total excavated area as A but in the form of a single chamber with no spatial sub-division. Block D 32 also had a single chamber but with half the total excavated area (28.4 cm^2) . We selected larval *C. cephalonica* weighing between 30-40 mg (mean = 31.4) from

our cultures and presented them individually to an adult female *G. nephantidis* until the larva

 was paralysed (but with no eggs laid on its integument), and then the paralysing female wasp was discarded. Twenty-five paralysed *C. cephalonica* larvae were placed into each replicate block, one per chamber in blocks A and B, and all in the single chamber in C and D. In the latter block types we attempted to arrange the larvae into a regular array within the single chamber. The total weights of the host larvae at the start of the experiment did not differ 6 significantly across block types (ANOVA: $F_{3,44} = 0.54$, $P = 0.66$, $r^2 = 0.035$).

 We then selected female *G. nephantidis* aged 3-5 days since emergence as adults in broods containing at least one male (females had thus mated) and which had had no 9 experience of attacking or ovipositing on hosts; we anesthetized them with $CO₂$ gas and then placed them into the chambers within the blocks (one female per chamber in A and B, and all 25 females in the centre of the chamber in C and D). The Plexiglas lid was then placed over the block. Anesthetized wasps regained their activity within 2 min. Each block was inspected daily until adult offspring matured (adult emergence from cocoons).

 From each replicate block we recorded the numbers of hosts on which eggs were laid, the clutch sizes laid onto each host, and the numbers of offspring reaching the larval, pupal, and adult stages. Due to ectoparasitic development, immature *G. nephantidis* are observable at all stages; data on numbers of eggs laid are, however, likely to have been underestimates as some eggs could have been eaten between oviposition and observation. Adult female wasps (mothers) were removed from each block once parasitoid offspring had reached the pupal stage; this avoided confusion between generations during subsequent weight measurements. The numbers of adult male and female offspring were recorded and we obtained measures of 22 the mean dry weight of adult male and of adult female offspring. There were 12 replicates for each of the four block types.

Statistical analysis

 Data were mainly analysed using generalized linear modelling (GLM) techniques (Crawley, 1993; Zuur et al., 2009). In most analyses, the sampling unit was taken as the summed observations deriving from an individual replicate block, but we also carried out mixed effects modelling (GLMM) on clutch size, in which the replicate within which individual observations derived was fitted as a random factor (Zuur et al., 2009). We adopted quasi- Poisson errors and a log-link function for log-linear analyses of count data and quasi- binomial errors and a logit-link function for logistic analyses of data on proportions, in both cases using empirically estimated scale parameters to correct for overdispersion (Crawley, 1993; Wilson & Hardy, 2002; Zuur et al., 2009; Warton & Hui, 2011). When significant

differences across treatments were found from fitting an initial GLM, the parsimonious

model was identified by aggregation of factor levels (Crawley, 1993). For the analysis of

adult dry weight, errors were non-normally distributed so we employed non-parametric

Kruskal-Wallis ANOVAs (Siegel & Castellan, 1988). All statistical testing was 2-tailed. All

analyses were carried out using the GENSTAT statistical package (v. 15.1; VSN

- International, Hemel Hempstead, UK).
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Results

The proportion of hosts on which eggs were observed varied significantly between block

- 10 types (logistic ANOVA: $F_{3,40} = 8.02$, P<0.001; Figure 2): the probability of a host being
- oviposited on was highest in blocks of type A and did not differ significantly among blocks
- 12 of the other three types (model simplification by aggregation of factor levels: $F_{2,42} = 0.85$, P =
- 0.43). Among those hosts on which eggs were observed, clutch sizes varied significantly

14 across block types (log-linear GLMM: Wald statistic = 70.08 , $F_{3,42.8} = 23.36$, P<0.001), with

larger clutches in A [mean (+SE, -SE); A: 7.98 (+0.263, -0.254); B, 5.78 (+0.352, -0.331); C:

4.42 (+0.348, -0.323); D: 4.35 (+0.344, -0.319)]. In terms of the total numbers of eggs,

larvae, pupae, and adults produced per replicate, there were no significant differences among

- 18 B, C, and D, but production was significantly greater in A (Table 1). Ultimately, around $4\times$ as
- many adults emerged from type A than from the other block types (Figure 3).

 The mean (+SE, -SE) proportion of offspring that survived from the egg to the adult stage was 0.59 (+0.033, -0.032) and was not significantly influenced by block type (logistic 22 ANOVA: $F_{3,42} = 2.11$, $P = 0.11$), but note that this estimate is constrained to use the observed maximum number of eggs per replicate and the true probability of survival in some replicates could have been lower due to ovicide (see above). However, the probability of (observed) 25 eggs surviving to the larval stage was affected by block type $(F_{3,42} = 4.05, P = 0.013)$ and was greatest in A and not significantly different among B, C, and D (aggregation of factor levels: 27 F_{2,44} = 0.50, P = 0.61). The probability of larvae surviving to the pupal stage was not 28 significantly influenced by block type $(F_{3,42} = 1.20, P = 0.32)$, nor was the pupal to adult

29 survival $(F_{3,42} = 2.74, P = 0.055)$.

 The mean (+SE, -SE) overall sex ratio of adult offspring (proportion that were male) 31 was 0.32 (+0.021, -0.020) and was not influenced by block type (logistic ANOVA: $F_{3,42}$ = $32 \quad 1.24$, $P = 0.31$). The mean dry weights of adult male offspring, female offspring, or males and 33 females pooled did not differ across the block types (Kruskal-Wallis ANOVA; males: $H =$ 34 7.013, P = 0.071; females: H = 5.338, P = 0.15; pooled: H = 4.717, P = 0.19; all d.f. = 3).

Discussion

 Offspring production by *G. nephantidis* females is negatively affected by direct or indirect contact with conspecific females: only around one quarter as many adults developed under incomplete separation of broods compared to complete separation. Given that one host per female was always provided, the effect is not dependent on resource-weighted density, nor is it due to variation in spatial density (which was either held constant or had non-significant effect), but can be viewed as due to variation in organism-weighted density (Lewontin & Levins, 1989). That is, when not isolated, all females can experience higher parasitoid densities due to behavioural and/or chemical (Goubault et al., 2008) interactions with other females. It is notable that interference interactions occurred even when each female was initially placed in a separate chamber containing a host. A similar regular distribution of females across host patches was observed as the end result of agonistic interactions followed by dispersal of the loser in *Asobara citri* (Fischer), a braconid parasitoid of *Drosophila* spp. larvae, when females were experimentally released at a central point (de Jong et al., 2011). The fact that *G. nephantidis* females interacted with each other despite being placed in a regular array (block type B) suggests that the perceived host-patch is larger than a single experimental chamber. This accords with prior observation that although brood-guarding *G. nephantidis* females spend most of their time in the close vicinity of their hosts, they also patrol beyond the confines of the chamber (Goubault et al., 2007b) and it is during such patrolling behaviour that they are likely to encounter other females. Direct physical encounters can lead to the release of volatile chemicals (Goubault et al., 2008) which may in turn influence the behaviour of further females indirectly.

 The differences in *G. nephantidis* offspring production were principally due to effects operating at the early stages of offspring production: far fewer hosts were oviposited onto and also fewer eggs per host were observed when more than one female had access to each host. We pose three candidate explanations, which are not mutually exclusive. The first, a mechanistic explanation, is that females that were not completely isolated were continually disturbed, and possibly displaced from hosts, prior to and during oviposition. This is supported by observations of mutual attack and disruption of oviposition when one host was presented to several *G. nephantidis* females within a chamber (Venkatesan et al., 2009b). The second, an adaptive explanation, is that game-theoretic reasoning predicts females should lay smaller clutches after having encountered competitors, due to anticipation of a more competitive future environment for their offspring coupled with an increased size of offspring

 that develop from smaller clutches on given-sized hosts (Petersen & Hardy, 1996; Mesterton- Gibbons & Hardy, 2004). This gains some support from prior observation of the required effects in *G. nephantidis*: when females with hosts weighing <30 mg experienced 1-4 competitors for 30 min, their subsequent clutch sizes were reduced by ca. 18% and offspring sizes were consequently enhanced (Goubault et al., 2007a). In our present experiment, the reduction in observed clutch size in response to encountering competitors was more marked (from ca. 8 to ca. 4-6 eggs per host) but so was the likely occurrence of female-female encounters. However, we found no evidence for effects on the size of maturing offspring – we used hosts of 30-40 mg, and Goubault et al. (2007a) also found no effects using hosts >30 mg. The lack of difference in sex ratios across experimental treatments also suggests that mothers did not adaptively adjust their reproductive decisions according to the density of ovipositing females (Hamilton, 1967; Legner & Warkentin, 1988; Irvin & Hoddle, 2006; Ode & Hardy, 2008), probably because aggressive host and brood guarding naturally lead to broods being produced by single foundresses (Hardy & Cook, 1995; Hardy et al., 2013) resulting in little selection for sex-ratio adjustment to foundress number (Herre, 1987).

 The third explanation is that oviposition occurred to a similar extent in all experimental conditions but fewer eggs were observed on non-isolated hosts because some were eaten prior to being counted. Ovicidal behaviour is well documented in *G. nephantidis,* its congeners, and some other bethylids (Venkatraman & Chacko, 1961; Goertzen & Doutt, 1975; Hardy & Blackburn, 1991; Mayhew, 1997; Infante et al., 2001; Pérez-Lachaud et al., 2002; Goubault et al., 2007b; Bentley et al., 2009; Venkatesan et al., 2009b), occurring when clutches of eggs are not attended by a guarding mother, and the lower probability of egg-to- larval survival in blocks of type B, C, and D supports this. As with the mechanistic disruption hypothesis above, females that oviposited may have been intruded upon by such a number of other females that they were unable to defend successfully their clutches (for analogous arguments see Waage, 1982; van Alphen & Visser, 1990; Moore & Greeff, 2003; de Jong et al., 2011). The similar probabilities of the survival of offspring between the larval stages and adulthood, across experimental treatments, accords with prior reports of conspecific larvicide being absent or rare in *G. nephantidis* and congeners and, if it occurs, to be limited to the smaller larval stadia (Venkatraman & Chako, 1961; Goertzen & Doutt, 1975; Hardy & Blackburn, 1991; Goubault et al., 2007b; Bentley et al., 2009; Venkatesan et al., 2009a). Irrespective of the particular mechanism of mutual interference, the number of *G. nephantidis* offspring produced is negatively affected by contact between adult females during the host- and clutch-guarding phases. We now consider the potential consequences of

 this for biological pest control programmes using *G. nephantidis*, its congeners, and other bethylids. Members of the genus *Goniozus* have been used in several biocontrol programs (Dharmaraju, 1963; Legner & Silveira-Guido, 1983; Gothilf & Mazor, 1987; Legner & Warkentin, 1988; Legner & Gordh, 1992; Mohan & Shameer, 2003; Lyla et al., 2006; Venkatesan et al., 2007, 2009a,b) and others are regarded as having pest control potential (Witethom & Gordh, 1994; Conlong, 1994; Berry, 1998; Abbas et al., 2008; Etiam, 2001; Sadeghi et al., 2012). For instance, estimates of the percentage of *O. arenosella* parasitized by *G. nephantidis* in coconut plantations range from around 4% to nearly 60% (Lyla et al., 2006; Venkatesan et al., 2007) and almond pests have been shown to be regulated by *Goniozus legneri* Gordh following classical biocontrol introduction (Legner & Gordh, 1992). Across all biocontrol programmes using bethylids, about half have resulted in parasitoid population establishment and around 10% have achieved partial control of the target pests but records typically indicate that pest suppression is not sufficient to no longer require further control measures (Greathead & Greathead, 1992; Hardy & Goubault, 2007). Mutual interference behaviour could clearly be a contributory factor to a limited suppression of hosts once bethylid populations are established and parasitoid density increases such that foraging females frequently encounter each other at hosts. Even if mutual interference effects in the field are weaker than indicated by our experiment (given that laboratory studies typically over-estimate mutual interference due to the use of artificially high parasitoid densities; Free et al., 1977), our data do suggest that field release of mass-reared *G. nephantidis* females may be more effective (on a per parasitoid basis) when females are released at lower densities, with lower female-female encounter rates, than at higher densities. However, as females spend much time brood-guarding during which they could be foraging for further hosts, their ability to supress large numbers of hosts will be limited (Venkatesan et al., 2009b). A contrasting suggestion is that, by leading to a regular distribution of parasitism across hosts, female-female fighting behaviour in *A. citri* might promote host population suppression (de Jong et al., 2011). It also contrasts with expectation when deploying bethylids of the genus *Sclerodermus* (Yang et al., 2014): in *Sclerodermus,* the presence of other females can enhance the probability of offspring production because females do not interact with each other aggressively and exhibit cooperative brood care after collectively suppressing and ovipositing on an individual host (Hu et al., 2012; Tang et al., 2014; Kapranas et al., 2016). Mutual interference interactions are also likely to influence the efficiency of mass rearing of *G. nephantidis* and its congeners. Our data, and the prior results of Venkatesan et al. (2009b), clearly suggest that mass-rearing programmes will not maximize the production

 of female offspring if multiple adult females are in contact with each other once presented with hosts: using one vial for each female wasp plus one host will thus likely be more effective than using larger communal containers. Irvin & Hoddle (2006) reached the same conclusion when considering the economics of mass rearing mymarid parasitoids which adjust their progeny sex ratios when ovipositing in the presence of conspecifics, and several other studies have similarly concluded that mass production will not be optimized by using high parasitoid densities (e.g., Luo et al., 2014). In contrast to our results, Abbas (1999) reported that in the congener *G. legneri* there was no reduction in offspring production when multiple hosts were provided to multiple females rather than individual females being presented with hosts separately; we have no ready explanation for this, especially as the reproductive and competitive behaviours of *G. nephantidis* and *G. legneri* are generally very similar (Hardy et al., 2013). More in accord with our results is that Legner & Warkentin (1988) found that for *G. legneri* the production of offspring per adult female 'foundress' was reduced by an increase in foundress number and increased by an increase in the number of hosts provided. They further found that the largest number of offspring per host was produced when just one foundress was present (Legner & Warkentin, 1988).

 The use of simple laboratory microcosms to suggest population-level consequences of behaviour has its drawbacks, given that there is neither the fine control achieved by experiments on the behaviours of one or a few individuals, nor the realism of field-scale evaluations. Nonetheless, the value of microcosms to examine the summed effects of spatial complexity and/or agonistic interactions has been recognised at least since the work of Huffaker (1958; Kareiva, 1989; Hassell, 2000; Infante et al., 2001; Batchelor et al., 2006; de Jong et al., 2011). Adopting the microcosm approach in this study has shown that interactions between *G. nephantidis* females extend beyond the immediate vicinity of provided hosts, and generate increased organism-weighted density when females are not completely separated. These interactions constitute mutual interference, manifest as a considerable reduction in the number of offspring produced. This mutual interference may contribute towards the limited degree of host population suppression achieved when some species of bethylids are deployed as agents of biological pest control and also has clear potential to influence the efficiency of mass rearing of female parasitoids prior to field release.

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Table 1 Effects of experimental treatment on total offspring production at progressive life-

history stages

¹At every life-history stage, the overall significant difference in offspring production was due

to greater numbers in block type A than in the other block types (see Figure 1): model

simplification by aggregation of factor levels showed that there were no significant

differences between block types B, C, and D.

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Figure captions

Figure 1 Illustration of treatment blocks to vary sub-division and parasitoid density.

Figure 2 The effect of treatment on the proportion of hosts within a replicate that had eggs

laid onto them. Bars represent standard errors of the means and are asymmetrical due to back

transformation from the logit scale. Differences between block types B, C and D were not

- significant.
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Figure 3 The effect of treatment on the number of adult progeny produced per replicate. Bars

represent standard errors of the means and are asymmetrical due to back transformation from

the log scale. Differences between block types B, C and D were not significant.

Figure1.

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- Figure 2.

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- Figure 3.
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