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- 4 Mutual interference reduces offspring production in a brood-
- 5 guarding bethylid wasp

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17 **Running title:** Mutual interference in a bethylid wasp

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Abstract

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3 important role in influencing the temporal and spatial dynamics of pest arthropods. 4 Behavioural interactions between foraging females, collectively constituting 'mutual 5 interference', can reduce host suppression. We use laboratory microcosms to assess the 6 prevalence and consequences of mutual interference behaviour in a bethylid wasp, *Goniozus* 7 nephantidis (Muesebeck) (Hymenoptera: Bethylidae), which is known to brood guard and to 8 engage in agonistic contests for individual hosts and which is also an agent of biological pest 9 control. We hold host and parasitoid numbers constant and vary the degree of female-female 10 contact that can occur. Mutual interference is manifest in a considerable reduction in the

Parasitoids have the potential to suppress populations of their hosts and thus may play an

number of offspring produced when females are not fully isolated from each other, due to

effects operating at the early stages of offspring production. This mutual interference may

contribute towards the limited degree of host population suppression achieved when some

species of bethylids are deployed as agents of biological pest control and also has clear

potential to influence the efficiency of mass rearing of parasitoids prior to field release.

Introduction

- 2 Parasitoid wasps are free-living as adults: females search for and attack hosts, usually the
- 3 immature stage of another arthropod, laying their eggs on or in these hosts which then form
- 4 the only food resource for their developing, parasitic, offspring, leading to the death of the
- 5 host (Godfray, 1994; Jervis, 2005). Parasitoid efficiency in host attack, and consequently
- 6 offspring production, may be tempered by behavioural interactions with other foraging
- females, termed 'mutual interference' (Hassell & Varley, 1969; Hassell, 1971, 2000; Rogers
- 8 & Hassell, 1974; Beddington, 1975; Free et al., 1977; Begon et al., 1996; Driessen & Visser,
- 9 1997; Fellowes et al., 2005; Jervis et al., 2005; Kidd & Jervis, 2005).
- Interference interactions can take a variety of forms; seen broadly they include not
- only the rate of parasitism of the current generation of hosts but also influences on the size
- and sexual composition of the next generation of parasitoids (Visser & Driessen, 1991; Visser
- et al., 1999). Examples include time-wasting disruption of foraging for hosts without
- explicitly agonistic interactions between foraging females (Hassell, 1971, 2000; Cronin &
- 15 Strong, 1993; Field et al., 1998; Wajnberg et al., 2004; Le Lann et al., 2011; Yazdani &
- Keller, 2015), aggressive patch or brood guarding (Hassell, 1971; Waage, 1982; Field et al.,
- 17 1997; Goubault et al., 2005; Nakamatsu et al., 2009; Venkatesan et al., 2009a,b; de Jong et
- al., 2011; Hardy et al., 2013; Mohamad et al., 2015), clutch size and superparasitism
- decisions differing in the presence, or anticipated presence, of competitors (van Alphen &
- 20 Visser, 1990; Visser & Driessen, 1991; Visser et al., 1992; Visser, 1996; Field et al., 1998;
- Goubault et al., 2007a), and sex allocation decisions contingent on the number of ovipositing
- 22 'foundress' females present (Hamilton, 1967; Waage, 1982; Meunier & Bernstein, 2002;
- 23 Irvin & Hoddle, 2006; Ode & Hardy, 2008; Luo et al., 2014).
- 24 Empirical investigations of factors contributing to mutual interference have often been
- conducted from the perspective of behavioural ecology but there has also been great interest
- 26 in their consequences for host–parasitoid population dynamics, in part because this
- 27 contributes towards explaining population phenomena on the basis of natural selection
- 28 (Hassell & May, 1973, 1989; Anderson, 1989; Cronin & Strong, 1993; Driessen & Visser,
- 29 1997; Visser et al., 1999; Bernstein, 2000; Meunier & Bernstein, 2002). Typically, mutual
- 30 interference interactions reduce the per-host production of female offspring (the sex that
- 31 attacks future generations of hosts) and will also be more prevalent at higher parasitoid
- densities: this density-dependent effect may be expected to contribute to the dynamic stability
- of host–parasitoid populations, but extreme effects of interference can also lead to predictions
- 34 that host populations will not be regulated (sufficiently suppressed) by parasitism (Rogers &

1 Hassell, 1974; Begon et al., 1996; Bernstein, 2000; Hassell, 2000; Kidd & Jervis, 2005). To

2 practitioners of biological control, who deploy parasitoids against agricultural pests, the

3 importance of mutual interference is thus that it reduces the likelihood that pest populations

4 will be depressed below thresholds of economic damage (Anderson, 1989; Hassell & May,

5 1989). Although there have been many successful biocontrol programmes, attempts have also

frequently failed because parasitoids have not established and because established parasitoids

7 have not depressed the pest populations sufficiently (e.g., Mills, 2000; Kapranas & Hardy,

2014). In particular, parasitoids are often mass reared and innundatively released at high

densities, which may artificially promote the occurrence of economically detrimental mutual

interference interactions (Irvin & Hoddle, 2006; Ode & Hardy, 2008).

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In this study we use simple laboratory microcosms to assess the prevalence and consequences of mutual interference behaviour in a bethylid wasp, Goniozus nephantidis (Muesebeck) (Hymenoptera: Bethylidae). This parasitoid species is known to guard individual hosts and developing broods, to engage in agonistic contests for hosts prior to and following oviposition (Hardy et al., 2013), and is also mass reared for release in biological control programmes against coconut pests (Shameer et al., 2002; Mohan & Shameer, 2003; Venkatesan et al., 2003, 2007, 2009a,b; Lyla et al., 2006). Our purpose is to understand better the competitive and reproductive biology of brood-guarding parasitoids for which prior most investigations have been confined to dyadic interactions (Hardy et al., 2013), and to in turn suggest approaches towards enhancing the efficiency of their mass rearing and field release when utilized as agents of biological pest control. Venkatesan et al. (2009b) have already reported that individual G. nephantidis females produced more progeny (per host and also in total) when provided with just one host rather than multiple hosts simultaneously within a single chamber (a glass vial). They also reported that when the number of females within the chamber to which a single host was presented was varied, progeny production was greatest when just one female wasp was present: offspring production was thus greatest (per female and also in total) when one host and one female G. nephantidis were present (Venkatesan et al., 2009b). Here, we hold the number of hosts available per female (resource-weighted density; Lewontin & Levins, 1989) constant, at unity, and experimentally vary both spatial density (individuals per unit area) and the degree of physical sub-division between chambers within microcosms (the latter allowing parasitoid behaviour to determine the organismweighted density; Lewontin & Levins, 1989). The overall hypothesis is that parasitoids will produce fewer offspring when there is more contact possible between reproductive adult females, even though in all cases one host per female is provided.

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Biology of Goniozus nephantidis

- 3 Goniozus nephantidis is a gregarious larval ectoparasitoid for which many aspects of
- 4 behaviour and life history have been documented (Hardy et al., 1992; Cook, 1993; Hardy &
- 5 Cook, 1995; Humphries et al., 2006; Goubault et al., 2007a,b, 2008; Venkatesan et al.,
- 6 2009a,b). It is naturally associated with the coconut pest, *Opisina arenosella* Walker
- 7 (Lepidoptera: Oecophoridae) in the Indian sub-continent (Mohan et al., 2004; Venkatesan et
- 8 al., 2007, 2009a,b). Opisina arenosella larvae feed on the vascular tissues of coconut fronds
- 9 by chewing, leading to affected leaves showing a burnt appearance, and also construct
- protective tunnels and galleries with silk, leaf parts, and frass (Mohan et al., 2004;
- 11 Venkatesan et al., 2009a). Coconut crop yield losses due to O. arenosella can be as high as
- 12 45% and it can take up to 4 years for palms to regain normal yield after heavy infestation
- 13 (Mohan et al., 2010). Adult *G. nephantidis* females forage for hosts by entering the galleries.
- On encountering a host larva, the adult female attacks it by injecting venom whereupon the
- 15 host becomes paralysed within minutes and their development is arrested (idiobiontism).
- 16 Eggs are laid onto the host 1-4 days later (Jayaratnam, 1941; Goubault et al., 2007b), the
- intervening period possibly being used to mature eggs (Stokkebo & Hardy, 2000; but see
- Goubault et al., 2007b). The number of eggs laid on a host is positively correlated with host
- size, with clutches ranging up to ca. 18 (Hardy et al., 1992). Eggs hatch about 1 day after
- 20 oviposition and the larvae begin to feed on the host through punctures in its integument.

21 During the period between paralysis and oviposition, females remain in close physical

association with their hosts, aggressively guarding them against intruding conspecific females

- 23 which would otherwise utilize the unguarded host (Hardy & Blackburn, 1991; Petersen &
- Hardy, 1996). Mothers also remain with their broods after laying eggs, typically until the
- offspring pupate around 7 days later, and during this period guard their eggs and developing
- larvae against intruders (Hardy & Blackburn, 1991; Goubault et al., 2007b; Venkatesan et al.,
- 27 2009a). Mothers continue to mature eggs within their abdomens whilst guarding and
- 28 surviving mothers have an increasing tendency to leave their broods, possibly to forage for
- 29 further hosts (Goubault et al., 2007b).
- 30 Although prior-ownership is an advantage in host-ownership contests, other factors,
- 31 especially body size differences, contribute to determining the outcomes of agonistic
- 32 encounters (Petersen & Hardy, 1996; Humphries et al., 2006; Hardy et al., 2013) and prior
- owners can thus be driven away from hosts they have paralysed (fighting is rarely fatal).
- 34 Intruders that displace mothers will often destroy the brood and then lay their own eggs, but

the probability of destruction followed by oviposition declines as the developmental stage of the original brood increases (Hardy & Blackburn, 1991; Goubault et al., 2007b).

Some offspring fail to mature, even in clutches that are not destroyed by intruders, with mortality of guarded offspring estimated at around 28% (Hardy & Cook, 1995), mainly due to low juvenile survival during earlier developmental stages. Surviving offspring eclose as adults approximately 2 weeks after oviposition: males eclose first and mate with their sisters, then adults disperse from the natal site over the following few days (Hardy et al., 1999). The sex ratios of broods at eclosion and dispersal are usually female biased (Hardy & Cook, 1995; Hardy et al., 1999; Venkatesan et al., 2009a,b).

Materials and methods

- Goniozus nephantidis cultures were maintained using the rice moth, Corcyra cephalonica Stainton (Lepidoptera: Pyralidae), as a factitious host (following Hardy & Blackburn, 1991; Hardy et al., 1992; Petersen & Hardy, 1996). Corcyra cephalonica was reared on a diet of
- glycerol, honey, corn meal, wheat bran, and yeast (Lizé et al., 2012). All cultures and experiments were carried out in a climate room at 27 °C, L12:D12, with relative humidity maintained at *ca*. 35% by evaporation from a water bath.

We assessed the effects of host and parasitoid density and of spatial sub-division on parasitoid reproduction using experimental arenas excavated from polyethylene blocks and with transparent Plexiglas lids, with designs derived from the simpler contest arenas used and illustrated in Petersen & Hardy (1996) and Goubault et al. (2006). We used four types of arenas, termed block A-D (Figure 1), all excavated to a depth of 6 mm. Block A contained 25 chambers and each chamber was 1.7 cm in diameter. The total excavated area (floor area) within the block of the chamber was 56.8 cm². As wasps could not pass between the chambers, block A served as a control treatment in which interactions between adult females wasps were prevented. Block B was the same as A except that each chamber was connected to the others around it by a 1 cm long and 0.1 cm wide slot, through which adult *G. nephantidis* can pass with ease. Block B thus had an extra 4 cm² of floor area, an increase of 7% compared to A, but retained the same chamber size as block A and also as in previous studies (Petersen & Hardy, 1996; Goubault et al., 2006). Block C had the same total excavated area as A but in the form of a single chamber with no spatial sub-division. Block D

We selected larval *C. cephalonica* weighing between 30-40 mg (mean = 31.4) from our cultures and presented them individually to an adult female *G. nephantidis* until the larva

also had a single chamber but with half the total excavated area (28.4 cm²).

1 was paralysed (but with no eggs laid on its integument), and then the paralysing female wasp

was discarded. Twenty-five paralysed C. cephalonica larvae were placed into each replicate

block, one per chamber in blocks A and B, and all in the single chamber in C and D. In the

latter block types we attempted to arrange the larvae into a regular array within the single

chamber. The total weights of the host larvae at the start of the experiment did not differ

significantly across block types (ANOVA: $F_{3,44} = 0.54$, P = 0.66, $r^2 = 0.035$).

We then selected female *G. nephantidis* aged 3-5 days since emergence as adults in broods containing at least one male (females had thus mated) and which had had no experience of attacking or ovipositing on hosts; we anesthetized them with CO₂ gas and then placed them into the chambers within the blocks (one female per chamber in A and B, and all 25 females in the centre of the chamber in C and D). The Plexiglas lid was then placed over the block. Anesthetized wasps regained their activity within 2 min. Each block was inspected daily until adult offspring matured (adult emergence from cocoons).

From each replicate block we recorded the numbers of hosts on which eggs were laid, the clutch sizes laid onto each host, and the numbers of offspring reaching the larval, pupal, and adult stages. Due to ectoparasitic development, immature *G. nephantidis* are observable at all stages; data on numbers of eggs laid are, however, likely to have been underestimates as some eggs could have been eaten between oviposition and observation. Adult female wasps (mothers) were removed from each block once parasitoid offspring had reached the pupal stage; this avoided confusion between generations during subsequent weight measurements. The numbers of adult male and female offspring were recorded and we obtained measures of the mean dry weight of adult male and of adult female offspring. There were 12 replicates for each of the four block types.

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Statistical analysis

- Data were mainly analysed using generalized linear modelling (GLM) techniques (Crawley,
- 27 1993; Zuur et al., 2009). In most analyses, the sampling unit was taken as the summed
- observations deriving from an individual replicate block, but we also carried out mixed
- 29 effects modelling (GLMM) on clutch size, in which the replicate within which individual
- 30 observations derived was fitted as a random factor (Zuur et al., 2009). We adopted quasi-
- 31 Poisson errors and a log-link function for log-linear analyses of count data and quasi-
- 32 binomial errors and a logit-link function for logistic analyses of data on proportions, in both
- cases using empirically estimated scale parameters to correct for overdispersion (Crawley,
- 34 1993; Wilson & Hardy, 2002; Zuur et al., 2009; Warton & Hui, 2011). When significant

- differences across treatments were found from fitting an initial GLM, the parsimonious
- 2 model was identified by aggregation of factor levels (Crawley, 1993). For the analysis of
- 3 adult dry weight, errors were non-normally distributed so we employed non-parametric
- 4 Kruskal-Wallis ANOVAs (Siegel & Castellan, 1988). All statistical testing was 2-tailed. All
- 5 analyses were carried out using the GENSTAT statistical package (v. 15.1; VSN
- 6 International, Hemel Hempstead, UK).

8 Results

- 9 The proportion of hosts on which eggs were observed varied significantly between block
- types (logistic ANOVA: $F_{3,40} = 8.02$, P<0.001; Figure 2): the probability of a host being
- oviposited on was highest in blocks of type A and did not differ significantly among blocks
- of the other three types (model simplification by aggregation of factor levels: $F_{2,42} = 0.85$, P =
- 13 0.43). Among those hosts on which eggs were observed, clutch sizes varied significantly
- across block types (log-linear GLMM: Wald statistic = 70.08, $F_{3,42.8} = 23.36$, P<0.001), with
- larger clutches in A [mean (+SE, -SE); A: 7.98 (+0.263, -0.254); B, 5.78 (+0.352, -0.331); C:
- 4.42 (+0.348, -0.323); D: 4.35 (+0.344, -0.319)]. In terms of the total numbers of eggs,
- 17 larvae, pupae, and adults produced per replicate, there were no significant differences among
- 18 B, C, and D, but production was significantly greater in A (Table 1). Ultimately, around $4\times$ as
- many adults emerged from type A than from the other block types (Figure 3).
- 20 The mean (+SE, -SE) proportion of offspring that survived from the egg to the adult
- stage was 0.59 (+0.033, -0.032) and was not significantly influenced by block type (logistic
- ANOVA: $F_{3,42} = 2.11$, P = 0.11), but note that this estimate is constrained to use the observed
- 23 maximum number of eggs per replicate and the true probability of survival in some replicates
- could have been lower due to ovicide (see above). However, the probability of (observed)
- eggs surviving to the larval stage was affected by block type ($F_{3,42} = 4.05$, P = 0.013) and was
- 26 greatest in A and not significantly different among B, C, and D (aggregation of factor levels:
- F_{2,44} = 0.50, P = 0.61). The probability of larvae surviving to the pupal stage was not
- significantly influenced by block type ($F_{3,42} = 1.20$, P = 0.32), nor was the pupal to adult
- 29 survival ($F_{3.42} = 2.74$, P = 0.055).
- The mean (+SE, -SE) overall sex ratio of adult offspring (proportion that were male)
- was 0.32 (+0.021, -0.020) and was not influenced by block type (logistic ANOVA: $F_{3.42}$ =
- 32 1.24, P = 0.31). The mean dry weights of adult male offspring, female offspring, or males and
- females pooled did not differ across the block types (Kruskal-Wallis ANOVA; males: H =
- 7.013, P = 0.071; females: H = 5.338, P = 0.15; pooled: H = 4.717, P = 0.19; all d.f. = 3).

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Discussion

3 Offspring production by G. nephantidis females is negatively affected by direct or indirect 4 contact with conspecific females: only around one quarter as many adults developed under 5 incomplete separation of broods compared to complete separation. Given that one host per 6 female was always provided, the effect is not dependent on resource-weighted density, nor is 7 it due to variation in spatial density (which was either held constant or had non-significant 8 effect), but can be viewed as due to variation in organism-weighted density (Lewontin & 9 Levins, 1989). That is, when not isolated, all females can experience higher parasitoid 10 densities due to behavioural and/or chemical (Goubault et al., 2008) interactions with other 11 females. It is notable that interference interactions occurred even when each female was 12 initially placed in a separate chamber containing a host. A similar regular distribution of 13 females across host patches was observed as the end result of agonistic interactions followed 14 by dispersal of the loser in Asobara citri (Fischer), a braconid parasitoid of Drosophila spp. 15 larvae, when females were experimentally released at a central point (de Jong et al., 2011). 16 The fact that G. nephantidis females interacted with each other despite being placed in a 17 regular array (block type B) suggests that the perceived host-patch is larger than a single 18 experimental chamber. This accords with prior observation that although brood-guarding G. 19 nephantidis females spend most of their time in the close vicinity of their hosts, they also 20 patrol beyond the confines of the chamber (Goubault et al., 2007b) and it is during such 21 patrolling behaviour that they are likely to encounter other females. Direct physical 22 encounters can lead to the release of volatile chemicals (Goubault et al., 2008) which may in 23 turn influence the behaviour of further females indirectly. 24 The differences in G. nephantidis offspring production were principally due to effects 25 operating at the early stages of offspring production: far fewer hosts were oviposited onto and 26 also fewer eggs per host were observed when more than one female had access to each host. 27 We pose three candidate explanations, which are not mutually exclusive. The first, a 28 mechanistic explanation, is that females that were not completely isolated were continually 29 disturbed, and possibly displaced from hosts, prior to and during oviposition. This is 30 supported by observations of mutual attack and disruption of oviposition when one host was 31 presented to several G. nephantidis females within a chamber (Venkatesan et al., 2009b). The 32 second, an adaptive explanation, is that game-theoretic reasoning predicts females should lay 33 smaller clutches after having encountered competitors, due to anticipation of a more 34 competitive future environment for their offspring coupled with an increased size of offspring

1 that develop from smaller clutches on given-sized hosts (Petersen & Hardy, 1996; Mesterton-2 Gibbons & Hardy, 2004). This gains some support from prior observation of the required 3 effects in G. nephantidis: when females with hosts weighing <30 mg experienced 1-4 4 competitors for 30 min, their subsequent clutch sizes were reduced by ca. 18% and offspring 5 sizes were consequently enhanced (Goubault et al., 2007a). In our present experiment, the 6 reduction in observed clutch size in response to encountering competitors was more marked 7 (from ca. 8 to ca. 4-6 eggs per host) but so was the likely occurrence of female-female 8 encounters. However, we found no evidence for effects on the size of maturing offspring – 9 we used hosts of 30-40 mg, and Goubault et al. (2007a) also found no effects using hosts >30 10 mg. The lack of difference in sex ratios across experimental treatments also suggests that 11 mothers did not adaptively adjust their reproductive decisions according to the density of 12 ovipositing females (Hamilton, 1967; Legner & Warkentin, 1988; Irvin & Hoddle, 2006; Ode 13 & Hardy, 2008), probably because aggressive host and brood guarding naturally lead to 14 broods being produced by single foundresses (Hardy & Cook, 1995; Hardy et al., 2013) 15 resulting in little selection for sex-ratio adjustment to foundress number (Herre, 1987). 16 The third explanation is that oviposition occurred to a similar extent in all 17 experimental conditions but fewer eggs were observed on non-isolated hosts because some 18 were eaten prior to being counted. Ovicidal behaviour is well documented in G. nephantidis, 19 its congeners, and some other bethylids (Venkatraman & Chacko, 1961; Goertzen & Doutt, 20 1975; Hardy & Blackburn, 1991; Mayhew, 1997; Infante et al., 2001; Pérez-Lachaud et al., 2002; Goubault et al., 2007b; Bentley et al., 2009; Venkatesan et al., 2009b), occurring when 21 22 clutches of eggs are not attended by a guarding mother, and the lower probability of egg-to-23 larval survival in blocks of type B, C, and D supports this. As with the mechanistic disruption 24 hypothesis above, females that oviposited may have been intruded upon by such a number of 25 other females that they were unable to defend successfully their clutches (for analogous 26 arguments see Waage, 1982; van Alphen & Visser, 1990; Moore & Greeff, 2003; de Jong et 27 al., 2011). The similar probabilities of the survival of offspring between the larval stages and 28 adulthood, across experimental treatments, accords with prior reports of conspecific larvicide 29 being absent or rare in G. nephantidis and congeners and, if it occurs, to be limited to the 30 smaller larval stadia (Venkatraman & Chako, 1961; Goertzen & Doutt, 1975; Hardy & 31 Blackburn, 1991; Goubault et al., 2007b; Bentley et al., 2009; Venkatesan et al., 2009a). 32 Irrespective of the particular mechanism of mutual interference, the number of G. 33 *nephantidis* offspring produced is negatively affected by contact between adult females

during the host- and clutch-guarding phases. We now consider the potential consequences of

- 1 this for biological pest control programmes using G. nephantidis, its congeners, and other
- 2 bethylids. Members of the genus *Goniozus* have been used in several biocontrol programs
- 3 (Dharmaraju, 1963; Legner & Silveira-Guido, 1983; Gothilf & Mazor, 1987; Legner &
- 4 Warkentin, 1988; Legner & Gordh, 1992; Mohan & Shameer, 2003; Lyla et al., 2006;
- 5 Venkatesan et al., 2007, 2009a,b) and others are regarded as having pest control potential
- 6 (Witethom & Gordh, 1994; Conlong, 1994; Berry, 1998; Abbas et al., 2008; Etiam, 2001;
- 7 Sadeghi et al., 2012). For instance, estimates of the percentage of O. arenosella parasitized
- 8 by G. nephantidis in coconut plantations range from around 4% to nearly 60% (Lyla et al.,
- 9 2006; Venkatesan et al., 2007) and almond pests have been shown to be regulated by
- 10 Goniozus legneri Gordh following classical biocontrol introduction (Legner & Gordh, 1992).
- Across all biocontrol programmes using bethylids, about half have resulted in parasitoid
- population establishment and around 10% have achieved partial control of the target pests but
- records typically indicate that pest suppression is not sufficient to no longer require further
- 14 control measures (Greathead & Greathead, 1992; Hardy & Goubault, 2007). Mutual
- interference behaviour could clearly be a contributory factor to a limited suppression of hosts
- once bethylid populations are established and parasitoid density increases such that foraging
- females frequently encounter each other at hosts. Even if mutual interference effects in the
- 18 field are weaker than indicated by our experiment (given that laboratory studies typically
- over-estimate mutual interference due to the use of artificially high parasitoid densities; Free
- et al., 1977), our data do suggest that field release of mass-reared G. nephantidis females may
- be more effective (on a per parasitoid basis) when females are released at lower densities,
- 22 with lower female-female encounter rates, than at higher densities. However, as females
- spend much time brood-guarding during which they could be foraging for further hosts, their
- 24 ability to supress large numbers of hosts will be limited (Venkatesan et al., 2009b). A
- contrasting suggestion is that, by leading to a regular distribution of parasitism across hosts,
- 26 female-female fighting behaviour in A. citri might promote host population suppression (de
- 27 Jong et al., 2011). It also contrasts with expectation when deploying bethylids of the genus
- 28 Sclerodermus (Yang et al., 2014): in Sclerodermus, the presence of other females can
- 29 enhance the probability of offspring production because females do not interact with each
- 30 other aggressively and exhibit cooperative brood care after collectively suppressing and
- ovipositing on an individual host (Hu et al., 2012; Tang et al., 2014; Kapranas et al., 2016).
- Mutual interference interactions are also likely to influence the efficiency of mass rearing of *G. nephantidis* and its congeners. Our data, and the prior results of Venkatesan et
- al. (2009b), clearly suggest that mass-rearing programmes will not maximize the production

of female offspring if multiple adult females are in contact with each other once presented with hosts: using one vial for each female wasp plus one host will thus likely be more effective than using larger communal containers. Irvin & Hoddle (2006) reached the same conclusion when considering the economics of mass rearing mymarid parasitoids which adjust their progeny sex ratios when ovipositing in the presence of conspecifics, and several other studies have similarly concluded that mass production will not be optimized by using high parasitoid densities (e.g., Luo et al., 2014). In contrast to our results, Abbas (1999) reported that in the congener G. legneri there was no reduction in offspring production when multiple hosts were provided to multiple females rather than individual females being presented with hosts separately; we have no ready explanation for this, especially as the reproductive and competitive behaviours of G. nephantidis and G. legneri are generally very similar (Hardy et al., 2013). More in accord with our results is that Legner & Warkentin (1988) found that for G. legneri the production of offspring per adult female 'foundress' was reduced by an increase in foundress number and increased by an increase in the number of hosts provided. They further found that the largest number of offspring per host was produced when just one foundress was present (Legner & Warkentin, 1988).

The use of simple laboratory microcosms to suggest population-level consequences of behaviour has its drawbacks, given that there is neither the fine control achieved by experiments on the behaviours of one or a few individuals, nor the realism of field-scale evaluations. Nonetheless, the value of microcosms to examine the summed effects of spatial complexity and/or agonistic interactions has been recognised at least since the work of Huffaker (1958; Kareiva, 1989; Hassell, 2000; Infante et al., 2001; Batchelor et al., 2006; de Jong et al., 2011). Adopting the microcosm approach in this study has shown that interactions between *G. nephantidis* females extend beyond the immediate vicinity of provided hosts, and generate increased organism-weighted density when females are not completely separated. These interactions constitute mutual interference, manifest as a considerable reduction in the number of offspring produced. This mutual interference may contribute towards the limited degree of host population suppression achieved when some species of bethylids are deployed as agents of biological pest control and also has clear potential to influence the efficiency of mass rearing of female parasitoids prior to field release.

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Conlong DE (1994) A review and perspectives for the biological control of the African

1	sugarcane stalkborer Eldana saccharina Walker (Lepidoptera: Pyralidae). Agriculture,
2	Ecosystems and Environment 48: 9-17.
3	Cook JM (1993) Experimental test of sex determination in Goniozus nephantidis. Heredity
4	71: 130-137.
5	Crawley MJ (1993) GLIM for Ecologists. Blackwell Scientific Publications, Oxford, UK.
6	Cronin JT & Strong DR (1993) Superparasitism and mutual interference in the egg parasitoid
7	Angrus delicates (Hymenoptera: Mymaridae). Ecological Entomology 18: 293-32.
8	Dharmaraju E (1963) Biological control of coconut leaf caterpillar (Nephantis serinopa
9	Meyrick) in Ceylon. Bulletin of the Coconut Research Institute Ceylon 21: 1-46.
10	Driessen G & Visser ME (1997) Components of parasitoid interference. Oikos 79: 209-218.
11	Etiam A (2001) Oviposition behavior and development of immature stages of Parasierola
12	swirskiana, a parasitoid of the lesser moth Batrachedra amydraula. Phytoparasitica 29:
13	405-412.
14	Fellowes MDE, van Alphen JJM & Jervis MA (2005) Foraging behaviour. Insects as Natural
15	Enemies: A Practical Perspective (ed. by MA Jervis), pp. 1-71. Springer, Dordrecht,
16	The Netherlands.
17	Field SA, Keller MA & Calbert F (1997) The pay-off from superparasitism in the egg
18	parasitoid Trissolcus basalis, in relation to patch defence. Ecological Entomology 22:
19	142-149.
20	Field SA, Calbert G & Keller MA (1998) Patch defence in the parasitoid wasp Trissolcus
21	basalis (Insecta: Scelionidae): the time structure of pairwise contests, and the 'waiting
22	game'. Ethology 104: 821-840.
23	Free CA, Beddington JR & Lawton JH (1977) On the inadequacy of simple models of mutual
24	interference for parasitism and predation. Journal of Animal Ecology 46: 543-554.
25	Godfray HCJ (1994) Parasitoids: Behavioral and Evolutionary Ecology. Princeton University
26	Press, Princeton, NJ, USA.
27	Goertzen R & Doutt RL (1975) Ovicidal propensity of Goniozus. Annales of the
28	Entomological Society of America 68: 869–870.
29	Gothilf S & Mazor M (1987) Release and recovery of imported parasites of the carob moth,
30	Spectorbates ceratonia (Lepidoptera: Pyralidae) in Israel. Israel Journal of Entomology
31	21: 19-23.
32	Goubault M, Mack AFS & Hardy ICW (2007a) Encountering competitors reduces clutch size
33	and increases offspring size in a parasitoid with female-female fighting. Proceedings of

the Royal Society of London B 274: 2571-2577.

- 1 Goubault M, Scott D & Hardy ICW (2007b) The importance of offspring value: maternal
- defence in parasitoid contests. Animal Behaviour 74: 437-446.
- 3 Goubault M, Outreman Y, Poinsot D & Cortesero AM (2005) Patch exploitation strategies of
- 4 parasitic wasps under intraspecific competition. Behavioral Ecology 16: 693–701.
- 5 Goubault M, Batchelor TP, Linforth RST, Taylor AJ & Hardy ICW (2006) Volatile emission
- by contest losers revealed by real-time chemical analysis. Proceedings of the Royal
- 7 Society of London B 273: 2853-2859.
- 8 Goubault M, Batchelor TP, Romani R, Linforth RST, Fritzsche M et al. (2008) Volatile
- 9 chemical release by bethylid wasps: identity, phylogeny, anatomy and behaviour.
- Biological Journal of the Linnean Society 94: 837-852.
- 11 Greathead P & Greathead AH (1992) Biological control of insect pests by parasitoids and
- predators. The BIOCAT data base. Biocontrol News and Information 13: 61-68.
- Hamilton WD (1967) Extraordinary sex ratios. Science 156: 477-488.
- 14 Hardy ICW & Blackburn TM (1991) Brood guarding in a bethylid wasp. Ecological
- 15 Entomology 16: 55-62.
- Hardy ICW & Cook JM (1995) Brood sex ratio variance, developmental mortality and
- virginity in a gregarious parasitoid wasp. Oecologia 103: 162-169.
- Hardy ICW & Goubault M (2007) Wasp fights: understanding and utilizing agonistic
- bethylid behaviour. Biocontrol News and Information 28: 11-15.
- 20 Hardy ICW, Goubault M & Batchelor TP (2013) Hymenopteran contests and agonistic
- behaviour. Animal Contests (ed. by ICW Hardy ICW & M Briffa), pp. 147-177.
- Cambridge University Press, Cambridge, UK.
- Hardy ICW, Griffiths NT & Godfray HCJ (1992) Clutch size in a parasitoid wasp: a
- 24 manipulation experiment. Journal of Animal Ecology 61: 121-129.
- 25 Hardy ICW, Pedersen JB, Sejr MK & Linderoth UH (1999) Local mating, dispersal and sex
- ratio in a gregarious parasitoid wasp. Ethology 105: 57-72.
- 27 Hassell MP & May RM (1973) Stability in host-parasite models. Journal of Animal Ecology
- 28 42: 693-726.
- 29 Hassell MP & May RM (1989) The population biology of host-parasite and host-parasitoid
- 30 associations. Perspectives in Ecological Theory (ed. by J Roughgarden, RM May & SA
- 31 Levin), pp. 319-347. Princeton University Press, Princeton, NJ, USA.
- Hassell MP & Varley GC (1969) New inductive population model for insect parasites and its
- bearing on biological control. Nature 223: 1133-1137.
- 34 Hassell MP (1971) Mutual interference between searching insect parasites. Journal of Animal

1 Ecology 40: 473-486. 2 Hassell MP (2000) The Spatial and Temporal Dynamics of Host-Parasitoid Interactions. 3 Oxford University Press, Oxford, UK. 4 Herre EA (1987) Optimality, plasticity and selective regime in fig wasp sex-ratios. Nature 5 329: 627-629. 6 Hu Z, Zhao X, Li Y, Liu X & Zhang Q (2012) Maternal care in the parasitoid Sclerodermus 7 harmandi (Hymenoptera: Bethylidae). PLoS ONE 12: e51246. 8 Huffaker C (1958) Experimental studies on predation: dispersion factors and predator-prey 9 oscillations. Hilgardia 27: 343-383. 10 Humphries EL, Hebblethwaite AJ, Batchelor TP & Hardy ICW (2006) The importance of 11 valuing resources: host weight and contender age as determinants of parasitoid wasp 12 contest outcomes. Animal Behaviour 72: 891-898. 13 Irvin NA & Hoddle MS (2006) The effect of intraspecific competition on progeny sex ratio in 14 Gonatocerus spp. for Homalodisca coagulata egg masses: economic implications for 15 mass rearing and biological control. Biological Control 39: 162-170. 16 Infante F, Mumford J, Baker P, Barrera J & Fowler S (2001) Interspecific competition 17 between Cephalonomia stephanoderis and Prorops nasuta (Hym., Bethylidae), 18 parasitoids of the coffee berry borer, Hypothenemus hampei (Col., Scolytidae). Journal 19 of Applied Entomology 125: 63–70. 20 Jayaratnam TJ (1941) The bethylid parasite (*Perisieriola nephantidis* M.) of the coconut 21 caterpillar (Nephantidis serinopa Meyr.). Tropical Agriculturalist and Magazine of the 22 Ceylon Agricultural Society 97: 115-125. 23 Jervis MA (2005) Insects as Natural Enemies: A Practical Perspective. Springer, Dordrecht, 24 The Netherlands. 25 Jervis MA, Copland MJW & Harvey JA (2005) The life-cycle. Insects as Natural Enemies: A 26 Practical Perspective (ed. by MA Jervis), pp. 73-165. Springer, Dordrecht, The 27 Netherlands. 28 de Jong PW, Hemerik L, Gort G & van Alphen JJM (2011) Rapid establishment of a regular 29 distribution of adult tropical *Drosophila* parasitoids in a multi-patch environment by 30 patch defence behaviour. PLoS ONE 6: e20870. 31 Kapranas A & Hardy ICW (2014) One hundred years of parasitoids. Biocontrol News and 32 Information 35: 1-4. 33 Kapranas A, Hardy ICW, Tang X, Gardner A & Li B (2016) Sex ratios, virginity and local 34 resource enhancement in a quasi-social parasitoid. Entomologia Experimentalis et

1	Applicata, this issue.
2	Karieva P (1989) Renewing the dialogue between theory and experiments in population
3	ecology. Perspectives in Ecological Theory (ed. by J Roughgarden, RM May & SA
4	Levin), pp. 68-88. Princeton University Press, Princeton, NJ, USA.
5	Kidd NAC & Jervis MA (2005) Population dynamics. Insects as Natural Enemies: A
6	Practical Perspective (ed. by MA Jervis), pp. 435-523. Springer, Dordrecht, The
7	Netherlands.
8	Legner EF & Gordh G (1992) Lower navel orangeworm (Lepidoptera: Phycitide) population
9	densities following establishment of Goniozus legneri (Hymenoptera: Bethylidae) in
10	California. Journal of Economic Entomology 85: 2153-2160.
11	Legner EF & Silveira-Guido A (1983) Establishment of Goniozus emigratus and Goniozus
12	legneri (Hym: Bethylidae) on navel orangeworm, Amyelois transitella (Lep:
13	Phycitidae) in California and biological control potential. Entomophaga 28: 97-106.
14	Legner EF & Warkentin RW (1988) Parasitization of Goniozus legneri (Hymenoptera:
15	Bethylidae) at increasing parasite and host, Amyelois transitella (Lep: Phycitidae),
16	densities. Annals of the Entomological Society of America 81: 774-776.
17	Le Lann C, Outreman Y, van Alphen JJM & van Baaren J (2011) First in, last out:
18	asymmetric competition influences patch exploitation of a parasitoid. Behavioral
19	Ecology 22: 101-107.
20	Lewontin RC & Lewins R (1989) On the characteristics of density and resource availability.
21	American Naturalist 134: 513-524.
22	Lizé A, Khidr SK & Hardy ICW (2012) Two components of kin recognition influence
23	parasitoid aggression in resource competition. Animal Behaviour 83: 793-799.
24	Luo S, Li H, Lu Y, Zhang F, Haye T et al. (2014) Functional response and mutual
25	interference of Peristenus spretus (Hymenoptera: Braconidae), a parasitoid of Apolygus
26	lucorum (Heteroptera: Miridae). Biocontrol Science and Technology 24: 247-256.
27	Lyla KR, Beevi SP & Venkatesan T (2006) Field evaluation of Goniozus nephantidis
28	(Muesebeck) against coconut black-headed caterpillar in Kerala using different release
29	techniques. Journal of Biological Control 20: 33-36.
30	Mayhew PJ (1997) Fitness consequences of ovicide in a parasitoid wasp. Entomologial
31	Experimentalis et Applicata 84: 115–126.
32	Mesterton-Gibbons M & Hardy ICW (2004) The influence of contests on optimal clutch size:
33	a game-theoretic model. Proceedings of the Royal Society of London B 271: 971-978.
34	Meunier J & Bernstein C (2002) The influence of local mate competition on host-parasitoid

1	dynamics. Ecological Modelling 152: //-88.
2	Mills NJ (2000) Biological control: the need for realistic models and experimental
3	approaches to parasitoid introductions. Parasitoid Population Biology (ed. by ME
4	Hochberg & AR Ives), pp. 218-233. Princeton University Press, Princeton, NJ, USA.
5	Mohamad R, Wajnberg E, Monge J-P & Goubault M (2015) The effect of direct interspecific
6	competition on patch exploitation strategies in parasitoid wasps. Oecologia 177: 305-
7	315.
8	Mohan C, Nair CPR, Rajan O & Bindhumol PN (2004) Influence of temperature on
9	biological parameters of Goniozus nephantidis Mues. and Elasmus nephantidis Rohw.,
10	two promising parasitoids of the coconut black headed caterpillar, Opisina arenosella
11	Wlk. Journal of Planation Crops 32: 306-308.
12	Mohan C, Nair CPR, Nampoothiri CK & Rajan P (2010) Leaf-eating caterpillar (Opisina
13	arenosella) - induced yield loss in coconut palm. International Journal of Tropical
14	Insect Science 30: 132-137.
15	Mohan C & Shameer KS (2003) Galleria mellonella L. (Lepidoptera: Galleridae) as a new
16	host for Goniozus nephantidis Mues. (Hymenoptera: Bethylidae). Entomon 28: 169-
17	172.
18	Moore JC & Greeff JM (2003) Resource defence in female pollinating fig wasps: two's a
19	contest, three's a crowd. Animal Behaviour 66: 1101-1107.
20	Nakamatsu Y, Harvey JA & Tanaka T (2009) Intraspecific competition between adult
21	females of the hyperparasitoid Trichomalopsis apanteloctena (Hymenoptera:
22	Chelonidae), for domination of Cotesia kariyai (Hymenoptera: Braconidae) cocoons.
23	Annals of the Entomological Society of America 102: 172-180.
24	Ode PJ & Hardy ICW (2008) Parasitoid sex ratios and biological control. Behavioral Ecology
25	of Insect Parasitoids: From Theoretical Approaches to Field Applications (ed. by E
26	Wajnberg, C Bernstein & JJM van Alphen), pp. 253-291. Blackwell Publishing,
27	Oxford, UK.
28	Pérez-Lachaud G, Hardy ICW & Lachaud J-P (2002) Insect gladiators: competitive
29	interactions between three species of bethylid wasps attacking the coffee berry borer,
30	Hypothenemus hampei (Coleoptera: Scolytidae). Biological Control 25: 231-238.
31	Petersen G & Hardy ICW (1996) The importance of being larger: parasitoid intruder-owner
32	contests and their implications for clutch size. Animal Behaviour 51: 1363-1373.
33	Rogers DJ & Hassell MP (1974) General models for insect parasite and predator searching
34	behaviour: interference. Journal of Animal Ecology 43: 239-253.

1	Sadeghi E, Baniameri V & Marouf A (2012) Oviposition behaviour of Goniozus swirskiana				
2	(Hymenoptera: Bethylidae: Bethylinae) a parasitoid of Batrachedra amydraula				
3	Meyrick from the warmest desert of Iran. World Applied Sciences Journal 20: 1493-				
4	1498.				
5	Shameer KS, Mohan C & Nair CPR (2002) Optimum weight of host larvae for the mass				
6	multiplication of Goniozus nephantidis Mues., the larval parasitoid of Opisina				
7	arenosella Walker. Planation Crops Research and Development in the New Millennium				
8	- Proceedings of the International Conference on Planation Crops, Placrosym XIV, 12-				
9	15 December 2000, pp. 452-455. Coconut Development Board, Kochi, India.				
10	Siegel S & Castellan NJ (1988) Nonparametric Statistics for the Behavioural Sciences.				
11	McGraw-Hill, New York, NY, USA.				
12	Stokkebo S & Hardy ICW (2000) The importance of being gravid: egg load and contest				
13	outcome in a parasitoid wasp. Animal Behaviour 59: 1111-1118.				
14	Tang X, Meng L, Kapranas A, Xu F, Hardy ICW & Li B (2014) Mutually beneficial host				
15	exploitation and ultra-biased sex ratios in quasisocial parasitoids. Nature				
16	Communications 5: 4942.				
17	Yazdani M & Keller MA (2015) Mutual interference in Dolichogenidea tasmanica				
18	(Cameron) (Hymenoptera: Braconidae) when foraging for patchily-distributed light				
19	brown apple moth. Biological Control 86: 1-6.				
20	Venkatesan T, Jalali SK, Srinivasa Murthy K & Rao NS (2003) Galleria mellonella				
21	(Linnaeus), an alternate host for the rearing of Goniozus nephantidis (Muesebeck)				
22	(Hymenoptera: Bethylidae), a larval parasitoid of Opisina arenosella Walker.				
23	Biological Control of Lepidopteran Pests (ed. by PL Tandon, CR Ballal, SK Jalali &				
24	RJ Rabindra), pp. 79–82. PDBC, Bangalore, India.				
25	Venkatesan T, Jalali SK, Srinivasa Murthy K, Rabindra RJ & Bharathi Dasan C (2007)				
26	Economics of production of Goniozus nephantidis (Muesebeck), an important				
27	parasitoid of coconut black-headed caterpillar, Opisina arenosella (Walker) for bio-				
28	factories. Journal of Biological Control 21: 53-58.				
29	Venkatesan T, Jalali SK & Srinivasa Murthy K (2009a) Competitive interactions between				
30	Goniozus nephantidis and Bracon brevicornis, parasitoids of coconut pest Opisina				
31	arenosella. International Journal of Pest Management 55: 257-263.				
32	Venkatesan T, Srinivasa Murthy K, Rabindra RJ & Baskaran TV (2009b) Influence of				
33	parasitoid-host density on the behaviour ecology of Goniozus nephantidis				
34	(Muesebeck) (Hymenoptera: Bethylidae), a parasitoid of Opisina arenosella Walker.				

1	Journal of Biological Control 23: 255-264.
2	Venkatraman TV & Chacko MJ (1961) Some factors influencing the efficiency of Goniozus
3	marasmi Kurian, a parasite of maize and jowar leaf roller. Proceedings of the Indian
4	Academy Sciences 53: 275–283.
5	Visser ME (1996) The influence of competition between foragers on clutch size decisions in
6	an insect parasitoid with scramble larval competition. Behavioral Ecology 7: 109-114.
7	Visser ME, van Alphen JJM & Nell HW (1992) Adaptive superparasitism and patch time
8	allocation in solitary parasitoids: The influence of pre-patch experience. Behavioral
9	Ecology and Sociobiology 31: 163–171.
10	Visser ME & Driessen G (1991) Indirect mutual interference in parasitoids. Netherlands
11	Journal of Zoology 41: 214-227.
12	Visser ME, Jones TH & Driessen G (1999) Interference among insect parasitoids: a multi-
13	patch experiment. Journal of Animal Ecology 68: 108-120.
14	Waage JK (1982) Sib-mating and sex ratio strategies in scelionid wasps. Ecological
15	Entomology 7: 103–112.
16	Wajnberg E, Curty C & Colazza S (2004) Genetic variation in the mechanisms of direct
17	mutual interference in a parasitic wasp: consequences in terms of patch-time allocation.
18	Journal of Animal Ecology 73: 1179–1189.
19	Warton DI & Hui FKC (2011) The arcsine is asinine: the analysis of proportions in ecology.
20	Ecology 92: 3-10.
21	Wilson K & Hardy ICW (2002) Statistical analysis of sex ratios: an introduction. Sex Ratios:
22	Concepts and Research Methods (ed. by ICW Hardy), pp. 48-92. Cambridge University
23	Press, Cambridge, UK.
24	Witethom B & Gordh G (1994) Development and life table of <i>Goniozus thailandensis</i> Gordh
25	& Witethom (Hymenoptera: Bethylidae), a gregarious ectoparasitoid of a phycitine fruit
26	borer (Lepidoptera: Pyralidae). Journal of the Scientific Society of Thailand 20: 101-
27	114.
28	Yang Z-Q, Wang X-Y & Zhang Y-N (2014) Recent advances in biological control of
29	important native and invasive forest pests in China. Biological Control 68: 117–218.
30	Zuur AF, Ieno EN, Walker NJ, Saveliev AA & Smith GM (2009) Mixed Effects Models and
31	Extensions in Ecology with R. Springer, New York, NY, USA.
32	
33	

Table 1 Effects of experimental treatment on total offspring production at progressive life-

2 history stages

Stage	Log-linear ANOVA			Model simplification ¹	
	F _{3,44}	P	% deviance explained	F _{2,46}	P
Eggs	13.65	< 0.001	48.21	2.27	0.12
Larvae	16.65	< 0.001	49.97	1.30	0.28
Pupae	13.63	< 0.001	48.16	1.35	0.27
Adult females	9.45	< 0.001	39.18	0.62	0.55
Adult males	13.75	< 0.001	48.40	2.80	0.072
Adults (total)	12.28	< 0.001	45.56	1.25	0.30

³ At every life-history stage, the overall significant difference in offspring production was due

- 5 simplification by aggregation of factor levels showed that there were no significant
- 6 differences between block types B, C, and D.

Figure captions

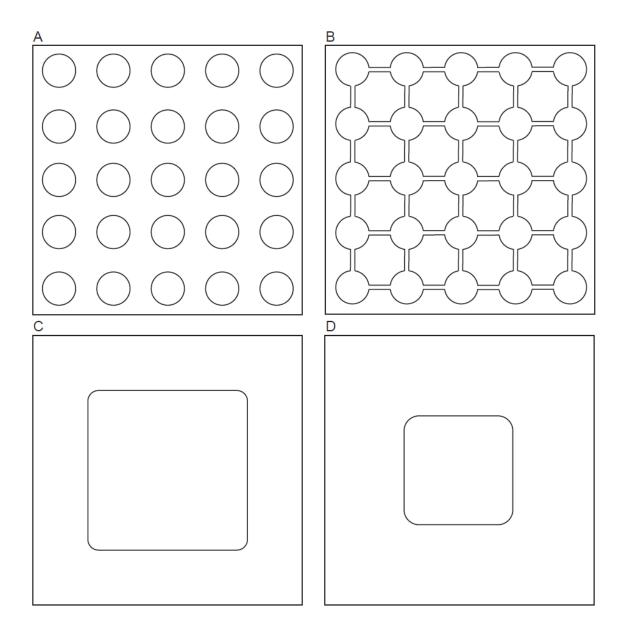
Figure 1 Illustration of treatment blocks to vary sub-division and parasitoid density.

Figure 2 The effect of treatment on the proportion of hosts within a replicate that had eggs laid onto them. Bars represent standard errors of the means and are asymmetrical due to back transformation from the logit scale. Differences between block types B, C and D were not significant.

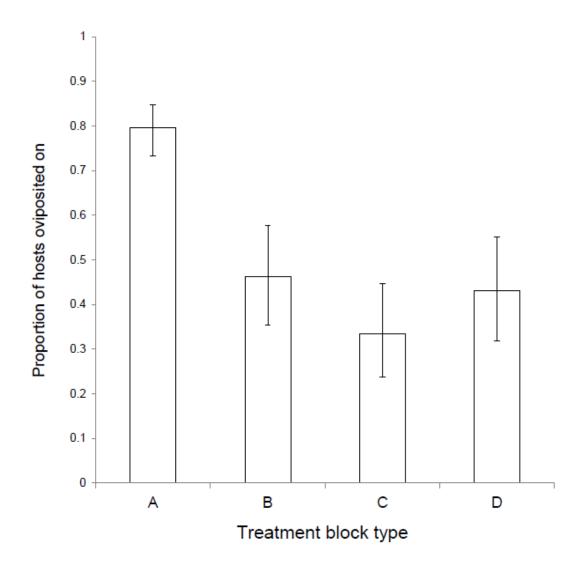
Figure 3 The effect of treatment on the number of adult progeny produced per replicate. Bars represent standard errors of the means and are asymmetrical due to back transformation from the log scale. Differences between block types B, C and D were not significant.

⁴ to greater numbers in block type A than in the other block types (see Figure 1): model

1 Figure 1.



6 Figure 2.



23 Figure 3.

