

1 EVALUATING RESISTANT BRASSICA TRAP CROPS TO MANAGE *HETERODERA*  
2 *SCHACHTII* (SCHMIDT) INFESTATIONS IN EASTERN ENGLAND

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4 *Evaluating brassica trap crops for H. schachtii control in East England*

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29 EVALUATING RESISTANT BRASSICA TRAP CROPS TO MANAGE *HETERODERA*  
30 *SCHACHTII* (SCHMIDT) INFESTATIONS IN EASTERN ENGLAND

31

32 **Abstract**

33 BACKGROUND:

34 The beet cyst nematode (BCN), *Heterodera schachtii* Schmidt, is a plant parasitic nematode  
35 which causes severe losses to yields of sugar beet. Resistant brassicas (radish and  
36 mustard) have been bred to be planted after the harvest of a main crop, such as a cereal,  
37 and encourage hatch of BCN juveniles. The resistant plants stimulate hatch of the juveniles  
38 but are not suitable hosts. Juveniles are unable to complete their lifecycle and thus  
39 populations are lowered. This research aimed to investigate the effectiveness of a range of  
40 these brassicas in terms of BCN control when grown in infested fields in Eastern England.

41 RESULTS:

42 Experiments were sown using four different radish cultivars, which differed in their resistance  
43 to BCN, and one resistant mustard variety. Field experiments were sown in early September  
44 in 2016 and 2017. Significant reductions in BCN populations were only found following the  
45 resistant mustard and the radish with the greatest resistance level.

46 CONCLUSIONS:

47 Further research is needed to understand how best to utilise the brassicas and whether they  
48 are economically viable when alternative management options for BCN are available. Time  
49 of planting may be crucial to fully achieve their BCN reducing potential.

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54 Keywords

55 Beet cyst nematode, radish, mustard, nematode management, IPM

56

## 57 1. Introduction

58 The beet cyst nematode (BCN), *Heterodera schachtii* Schmidt, has been a well-known pest  
59 of sugar beet for over 150 years<sup>1</sup>. It can now be found in all major sugar beet producing  
60 areas of the world<sup>2</sup> and is a major problem for growers with infestations, especially in years  
61 of limited water supply<sup>3</sup>. Yield losses can be severe (30-40% on susceptible cultivars<sup>4</sup>) and  
62 have been calculated to be as high as £3.8 million per year in the UK.<sup>5</sup> However, modern  
63 varieties of sugar beet with BCN tolerance may be able to overcome the majority of the yield  
64 losses<sup>6</sup>. These tolerant varieties still lead to a build-up of BCN populations,<sup>6</sup> which pose a  
65 problem for future sugar beet crops in the infested field and increase the chance of the  
66 transfer of infested soil to other fields. BCN is also harmful to a wide range of crop species,  
67 particularly brassicas such as oilseed rape (*Brassica napus*)<sup>7</sup> and, therefore, alternative  
68 methods of reducing populations are important to investigate, especially since chemical  
69 control options for BCN control are no longer available<sup>8</sup>.

70

71 The use of BCN resistant brassicas has become commonplace in Europe. In countries such  
72 as Germany, where BCN infestations are widespread, and may exceed 50% of the area  
73 cultivated with sugar beet<sup>4</sup>, they are a popular option to cultivate. The use of oilseed rape as  
74 a trap crop was proposed in the late 19<sup>th</sup> century by Prof. Julius Kühn.<sup>1</sup> The oilseed rape,  
75 which is susceptible to BCN, would be planted to stimulate juvenile emergence and root  
76 invasion in the summer prior to sugar beet being planted in the following spring. The  
77 seedlings of the oilseed rape would then need to be destroyed before the nematodes could  
78 reproduce. Therefore, the use of susceptible trap crops is a risk for the grower, as failure to  
79 destroy the crop before a completed lifecycle could exacerbate an infestation and this  
80 method was not implemented in the 1800s.<sup>1</sup> However, there are now tools available to  
81 advise growers when to destroy OSR volunteer seedlings in Germany to maximise BCN  
82 population control.<sup>9</sup> More suitable for BCN control is the use of resistant brassica varieties  
83 which offer an option for BCN control with much less risk to the grower. The resistant  
84 brassicas stimulate the hatch of juveniles from the cysts in the soil, and whilst they allow for

85 root invasion, they are resistant to the nematode, and prevent the completion of its lifecycle,  
86 hence lowering populations within the soil.<sup>10</sup> Resistant varieties of both white mustard  
87 (*Sinapis alba*) and oil radish (*Raphanus sativus*) are now available for cultivation as a trap  
88 crop and could be incorporated as part of an integrated pest management (IPM) strategy for  
89 BCN control.

90

91 The use of trap crops for BCN control has been investigated in many locations worldwide  
92 and mixed results have been obtained. Research in Germany in the 1990s found reductions  
93 in BCN populations using brassica trap crops and improvement to yield of subsequent beet  
94 crops<sup>1</sup> and similar results were found by Hafez<sup>11</sup> in the USA. However, Koch et al. <sup>12</sup> found  
95 significant reductions in BCN populations using oil radish at only one site out of four also in  
96 the USA. Kenter et al.<sup>13</sup> found BCN populations were lowered at twelve out of thirteen sites  
97 in Germany where resistant radish variety 'Adagio' was planted and the best results were  
98 obtained where the cover crop was sown in July rather than August. Hauer et al. <sup>6</sup> found  
99 resistant mustard to reduce populations of BCN at five out of eight environments tested  
100 between 2013 and 2014, also in Germany. Mustard varieties Luna and Accent and oil radish  
101 variety Colonel (the latter two varieties are also evaluated in this paper) have been found to  
102 reduce BCN populations at one site in Iran but did not produce significant reductions at a  
103 second site.<sup>14</sup> It appears, therefore, that the use of such trap crops is highly variable and  
104 further research into their use is necessary, especially as different results are obtained by  
105 different researchers in different countries and in different years. Findings from one country  
106 cannot simply be expected to work in another country and, likewise, the findings from one  
107 variety of trap crop cannot be expected to carry over to another variety.

108

109 It is clear that gaps in our understanding of the use of trap crops remain. As their use  
110 appears variable between years and countries in which the experiments are conducted, due  
111 to different climatic conditions and weather patterns, as well as time of sowing. We therefore  
112 decided to conduct two field trials, over two years, to see how a range of commercially

113 available trap crop varieties could reduce BCN populations on infested fields in Eastern  
114 England. To do this, on farm field experiments were conducted in fields with known  
115 infestations of BCN.

116

117

## 118 **2. Materials and methods**

### 119 Brassica variety selection

120 Following a recommendation from a seed merchant five brassica treatments were selected  
121 to be sown in a field experiments. Four of these treatments reportedly had some resistance  
122 to BCN from bioassays performed on these varieties in Germany<sup>6</sup>. These were one class  
123 one radish (with  $\geq 90\%$  resistance to BCN), two class two radishes (70-90% resistance to  
124 BCN) and one class two mustard. The final brassica treatment was a tillage radish with no  
125 reported resistance to BCN. A fallow treatment was also included in the experimental design.  
126 Details of the brassica treatments are listed in table 1.

127

128 Table 1 – Descriptions of the varieties of radish and mustard used in the field experiments in  
129 Norfolk, England.

130

Species	Cultivar name	BCN resistance Class †	Seed Rate Kg ha <sup>-1</sup>	Treatment name
<i>Raphanus sativus</i>	Colonel	1	22	Class 1 Radish
<i>R. sativus</i>	Defender	2	22	Class 2 Radish A
<i>R. sativus</i>	Bokito	2	20	Class 2 Radish B
<i>R. sativus</i>	Early Mino	-	12	Susceptible Radish
<i>Sinapis alba</i>	Accent	2	18	Class 2 Mustard

131 † Class 1 is stated as having an reproductive factor (*Rf*) of  $\leq 0.1$  and class 2 a reproductive  
132 factor (*Rf*) of between 0.1 and 0.3 when tested in official laboratory bioassays<sup>6</sup>.

133

134 Field experiments

135 Fields with known infestations of BCN were selected in 2016 (Brettenham, Norfolk, 52° 24'  
136 22.6296" N, 0° 51' 1.4688" E) and 2017 (Bridgham, Norfolk, 52° 26' 0.1932" N, 0° 52'  
137 9.5016" E). Both fields used are of a loamy sand soil type over chalk bedrock.<sup>15</sup> Areas of  
138 each field were surveyed for BCN populations on a 50 x 50 m grid spacing comprising 40  
139 soil cores (150 mm deep, 25 mm diameter) and extraction of the cysts. Once areas with the  
140 greatest populations had been identified, the field was lightly cultivated using a disc harrow  
141 (Simba Xpress 5.5, Simba International Limited, Sleaford, UK) and then the plot layout was  
142 marked on the field. Plots were 6x9 m and centred between wheelings in the field. In both  
143 years, 24 plots were sampled over the high population areas. Surveying and sampling took  
144 place immediately after harvest of the previous crop, which was winter wheat (*Triticum*  
145 *aestivum*) in both years. From each plot, 40 soil cores were taken in a zig-zag pattern, using  
146 an AMS EZ-Eject soil probe (AMS Inc. American Falls, ID, USA) (25mm diameter), to a  
147 depth of 150 mm to create a bulk sample of each plot. This bulk sample was then sieved  
148 through a 4 mm sieve to remove large stones. The soil was then thoroughly mixed together  
149 before two 200 ml subsamples were taken. Both were weighed and one was washed using a  
150 Wye washer to extract cysts<sup>16</sup>. The other subsample was dried at 105°C for 24 hours to  
151 determine moisture content of the soil and thus the dry weight of the washed sample was  
152 calculated. Cysts from the sample were counted and then crushed using a Reid aluminium  
153 slide.<sup>17</sup> To determine the mean number of eggs per cyst, the crushed cysts were then diluted  
154 in 50ml of water inside a measuring cylinder and thoroughly mixed using a glass pipette and  
155 pipette controller (Powerpette, VWR International, Radnor, PA, USA). A 1ml subsample was  
156 then dispensed into a Fenwick slide<sup>18</sup> and the number of viable eggs and juveniles counted  
157 using a stereomicroscope (Leica M80, Leica, Wetzlar, Germany) and tally counter. The  
158 population of BCN eggs per gram of dry soil was determined for each plot using the cyst and  
159 egg data. The field experiments were established using randomised block designs with the  
160 field plots grouped into blocks of similar *Pi* prior to sowing of the brassicas. Each block  
161 contained one replicate of each treatment (Table 1). Initial plot populations ranged from 2.1

162 to 14.8 eggs g<sup>-1</sup> dry soil in 2016 (mean 7.8) and 2.1 to 24.9 eggs g<sup>-1</sup> dry soil in 2017 (mean  
163 7.5).

164

165 Field experiments were sown on 2 September 2016 and 1 September 2017 using a Horsch  
166 Pronto 6 metre seed drill (Horsch Maschinen GmbH, Schwandorf, Germany). The selected  
167 varieties of the radish and the mustard were drilled at the seed rates stated in Table 1  
168 following guidance from the seed merchant. There was also a fallow treatment where the  
169 drill and tractor passed over the plot but no seed was sown. Soil temperatures were  
170 monitored throughout the experiments using a temperature logger (Tinytag Plus 2, Gemini  
171 data loggers, Chichester, UK) buried 10 cm into the soil. Accumulated thermal time above  
172 10°C (a temperature which has been reported at which egg hatch activity ceases<sup>19</sup>) was  
173 measured as 278°C days in 2016 and 223°C days in 2017 from sowing to destruction of the  
174 brassicas. All plots received 40kg ha<sup>-1</sup> of nitrogen as ammonium nitrate (NH<sub>4</sub>NO<sub>3</sub>) in liquid  
175 form prior to emergence of the seedlings using a Berthoud raptor FC crop sprayer (Berthoud  
176 Agricole SAS, Belleville, France)

177

178 Canopy spectral reflectance data were recorded during the growth of the brassicas using a  
179 Crop Circle ACS-430 NDVI meter (Holland Scientific, Lincoln, Nebraska, USA) to measure  
180 Normalised Difference Vegetation Index (NDVI).<sup>20</sup> We used NDVI as a measure of growth of  
181 the trap crops as a higher NDVI equates to greater canopy ground cover and therefore  
182 represents greater crop growth, especially as this relationship has been demonstrated in  
183 brassica species previously.<sup>21</sup> The brassicas were destroyed early in the following January  
184 in both years using a 3 m tractor mounted flail (Maschio Gaspardo S.p.A., Campodarsego,  
185 Italy). Prior to flailing a 50x50 cm quadrat was used to take four samples from each plot to  
186 determine shoot biomass. Samples were dried at 70°C until constant weight and weighed.  
187 After flailing, each plot was resampled for BCN cysts as previously described to determine  
188 the *Pf* of each plot and the *Rf* was then calculated as follows:

189 
$$\text{Reproductive factor (Rf)} = \frac{\text{Initial population (Pi)}}{\text{Final population (Pf)}}$$

190

191 The length of the experiments (approximately 16 weeks) followed standard farm practice in  
192 the UK regarding the cultivation of a cover crop and was to allow for any frost susceptible  
193 varieties of the cover crops to die off during the winter rather than result in large amounts of  
194 biomass which would have remained on the surface or require the use of herbicides to kill  
195 the trap crops. Likewise, the fallow treatments allowed weeds to grow to simulate normal  
196 fallow conditions on the fields.

197

#### 198 2.4 Data analysis

199 Data were analysed using Genstat 17<sup>th</sup> edition (VSN International, Hemel Hempstead, UK).  
200 The NDVI data were analysed using a repeated measures ANOVA, and field data analysed  
201 using one way ANOVA. Calculation of the least significant difference (LSD) at 5%  
202 significance was included in the ANOVA analysis. Figures were prepared using Microsoft  
203 Excel (Microsoft Corp, Redmond, Washington, USA) and Graphpad prism version 7  
204 (GraphPad Software Inc. La Jolla, CA, USA)

205

206

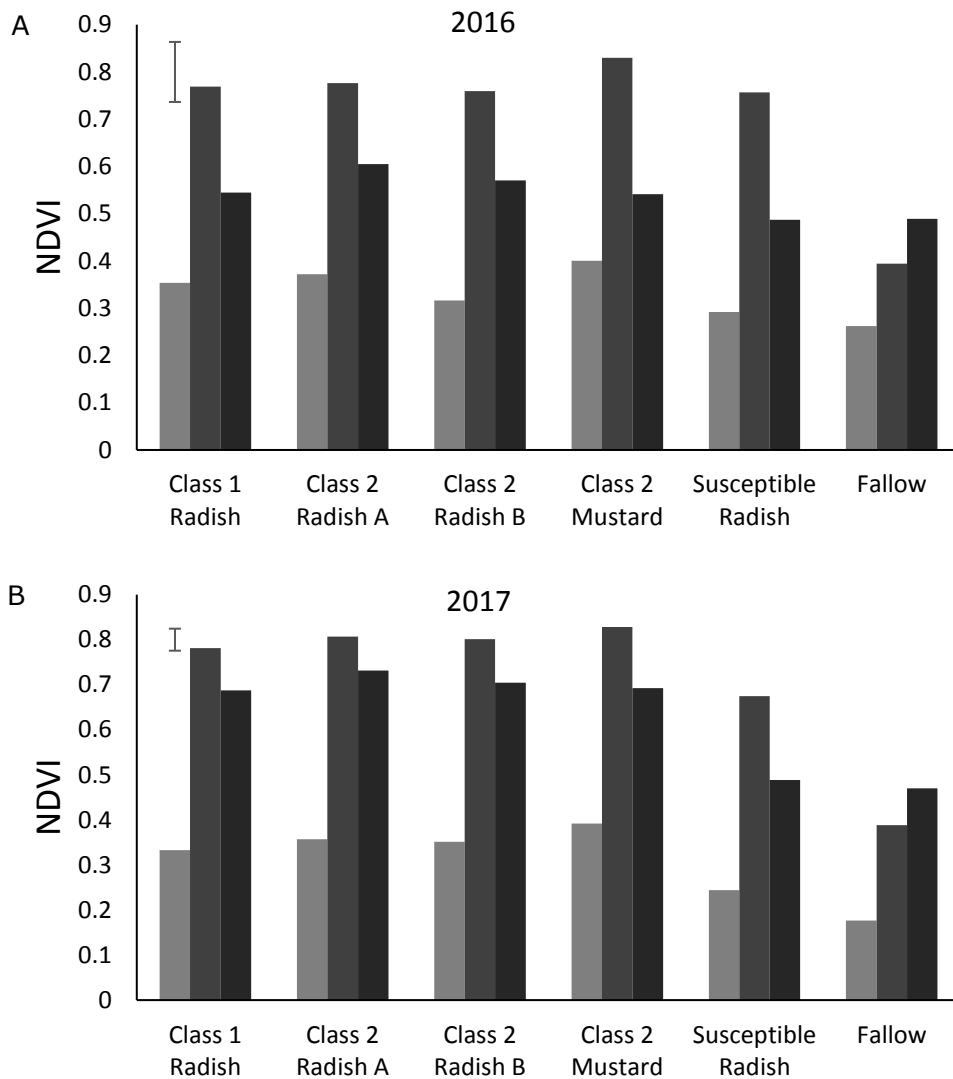


207 **3. Results**

208 In both years of the field experiments similar trends were observed in the growth of the trap  
209 crops as measured using NDVI (Fig1). NDVI increased between the first and second  
210 measurements, as the radishes or mustard plants were rapidly growing, and then declined  
211 by the third measurement, in early January. Only the fallow treatment shows a continual  
212 increase in NDVI values, due to the development of weeds on the fallow plots. In 2016 there  
213 were no significant differences in NDVI between trap crops but they had a greater NDVI at  
214 58 DAS than the fallow treatment. In 2017, the susceptible radish had a significantly lower  
215 NDVI than the other trap crops at every measurement and resulted in an NDVI at 128 DAS  
216 similar to the fallow treatment.

217 In 2016, the mustard had a significantly greater biomass than all of the other trap crops (Fig  
218 2,  $P < 0.001$ ). In 2017 the mustard again shows the greatest development of biomass and  
219 was significantly greater than the class 1 radish and class 2 radish B. Class 2 radish A also  
220 shows significantly greater growth than class 2 radish B. However, all resistant treatments  
221 resulted in greater growth than the susceptible radish in 2017 ( $P < 0.001$ ).

222 When analysing the two years separately, populations of BCN were not significantly reduced  
223 by the class one radish or class two mustard compared with the fallow control (2016  
224  $P = 0.085$ , 2017  $P = 0.125$ ). Equally, the susceptible radish and the two class 2 radish varieties  
225 did not cause any clear reductions in BCN populations. However, when data from the two  
226 years were combined into a multi-year analysis, significant differences were found between  
227 the treatments ( $P = 0.01$ ) (Fig 3). The class one radish and the class two mustard showed  
228 significant mean population reductions compared to the fallow treatment but differences  
229 were not found from using the two class two radish varieties or the susceptible radish.



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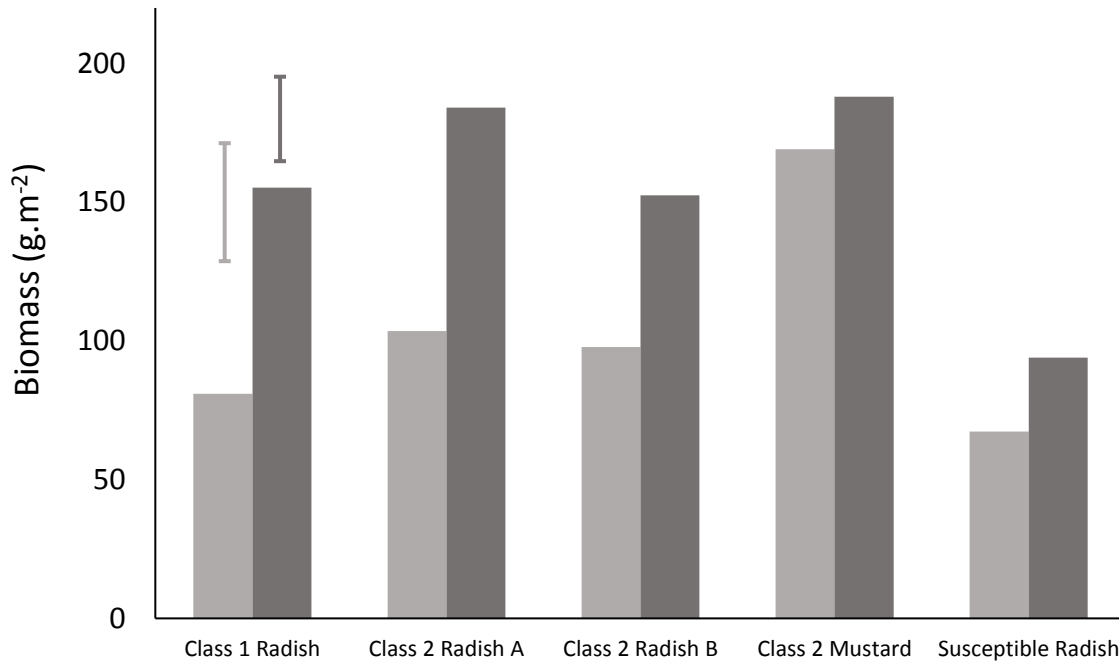
Fig 1. NDVI (Normalised Difference Vegetative Index) of brassica crops grown in fields infested with *H. schachtii* in Norfolk. NDVI was measured three times in each season (2016/17 ■ = 20 DAS [days after sowing], ■ = 58 DAS, ■ = 126 DAS & 2017/18 ■ = 23 DAS, ■ = 56 DAS & ■ = 128 DAS) Differences between treatments were found in both years ( $P < 0.001$  for both years). Error Bars shows LSD at 5% time x treatment interaction. Fallow treatments were not sterile so NDVI values for these plots shows development of weed plants.

232

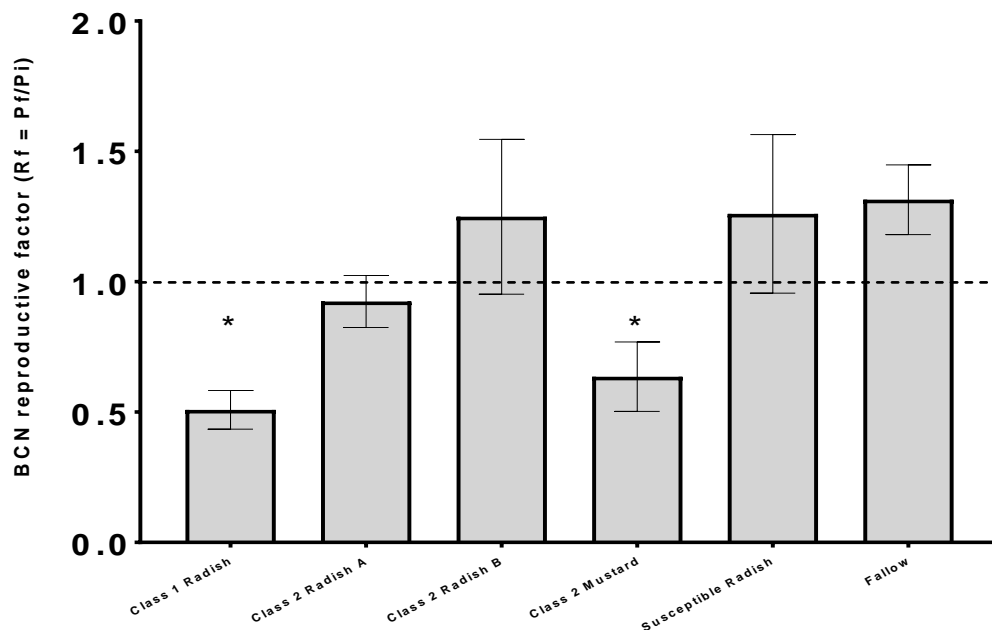
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235



236 Fig 2. Above ground biomass of the brassica trap crops in 2016 ■ and 2017 ■ in the  
 237 field experiments in Norfolk. Significant differences were found in both years ( $P < 0.001$   
 238 in both years) Error bars show LSD at 5% for each respective year. N.B. no samples  
 were taken from the fallow treatments.



249 Fig 3. Combined *H. schachtii* reproduction factor data from the brassica trap crops grown  
 250 in field trials in 2016 and 2017. Significant differences were found between the  
 251 treatments ( $P = 0.01$ ) and significantly reduced populations from the fallow control  
 252 treatment are denoted by \*. The dashed line shows where  $P_f/P_i = 1$ , i.e. where no net  
 253 BCN population change was measured. LSD at 5% significance = 0.53. Error bars show  
 254  $\pm$ SEM.

256 **4. Discussion**

257 The interest in the use of resistant brassicas for control of BCN has increased in recent  
258 years but limited data has been available on their effectiveness in UK field conditions. We  
259 have investigated their use over two field seasons to understand how effective they are at  
260 reducing nematode populations and which types are most suited to climatic conditions in  
261 Britain.

262

263 The class 1 radish and class 2 mustard were the only two treatments to significantly reduce  
264 the population of the nematodes when compared to the fallow control and we hypothesise  
265 that these varieties stimulate significantly greater levels of BCN hatch when compared to the  
266 other varieties. To have found 30 to 40% population reductions from these two varieties  
267 when grown for such a short amount of time, and when temperatures have begun to decline,  
268 will be important to those dealing with BCN infestations, especially in the UK. Growing class  
269 1 radish and class 2 mustard ahead of sugar beet should reduce the population of BCN,  
270 which could infect seedlings and limit growth.

271

272 The differences in *Rf* caused by the brassicas are likely explained by a combination of their  
273 growth habit and differences in their ability to stimulate BCN juvenile hatch. We found that all  
274 of the brassica treatments achieved good levels of ground cover when measured by NDVI,  
275 and when the biomass was measured the resistant radishes all seemed to grow equally well  
276 in both years, and the mustard also grew well. Therefore, none of the treatments were  
277 limited in their growth. As root growth is highly related to shoot growth<sup>22</sup> we can expect that  
278 rooting was similar between the treatments and therefore differences likely exist in how  
279 stimulating the different varieties are in stimulating BCN J2 hatch. Hatching stimulation must  
280 be considered when choosing which variety of trap crop to grow, as this trait might be more  
281 important than the resistance rating. Current assessments for resistance to BCN using a  
282 bioassay and hatched juveniles might produce misleading results as the ability to stimulate  
283 hatch is not assessed. Alternative methods may be needed, such as the use of laboratory

284 hatching assays<sup>23,24</sup> to screen future varieties for their hatching ability combined with  
285 confirming resistance to advise farmers with infested fields on which varieties to choose for  
286 maximum population control.

287

288 The BCN population data show an increase in populations even under the fallow treatment.  
289 This was not expected, as we expected populations to remain at levels very close to the *Pi*  
290 or to slightly reduce following the fallow treatment, which has been seen in other  
291 investigations<sup>14</sup>, although Hauer et al.<sup>6</sup> did also find some plots to show large increases in  
292 BCN populations under fallow conditions treated with straw mulch. Our increases under  
293 fallow could be due to a number of factors. Firstly, BCN susceptible weed species may have  
294 been growing and allowed for the population to build up. Weed growth was observed and is  
295 reflected in the NDVI measurements on the fallow plots. Fat hen (*Chenopodium album*) is  
296 commonly found across the host farm, which is a known host of BCN.<sup>25,26</sup> However, fat hen  
297 is classed as a relatively poor host of BCN<sup>27</sup> and Meinecke et al.<sup>25</sup> concluded that common  
298 arable weeds do not require control for nematological reasons. Therefore, it seems unlikely  
299 that weeds are responsible for our observed population increases although we cannot be  
300 certain. Investigations into UK weed populations and their ability to host BCN would be  
301 useful to expand on our findings and possibly confirm if the increased populations are due to  
302 weeds. A clean fallow treatment was not used in our experiments for two reasons, firstly, we  
303 wanted to investigate how a fallow field may cause changes to BCN populations, and  
304 secondly, it would not have been viable for the host farmer to regularly spray or cultivate the  
305 fallow plots. However, should similar experiments be conducted in the future it would be a  
306 good additional treatment and could provide further explanations to our observations.  
307 We hypothesise that consolidation of the soil between the two sampling dates, might have  
308 caused the population increases observed. The samples for *Pi* were taken immediately after  
309 cultivation of the field in the summer. This created an unconsolidated surface to the soil  
310 throughout the area sampled. When the *Pf* samples were taken four months later,  
311 considerable weathering due to rainfall had occurred to the soil which led to observed

312 consolidation, i.e. the same mass of soil now filled a smaller volume and lead to soil layers  
313 below the 150 mm sample depth in the summer (and therefore not sampled for *Pf*) being  
314 included in the winter samples collected to determine *Pf*. This then might have skewed our  
315 population counts, especially as BCN populations at deeper layers may be greater than at  
316 shallower soil layers<sup>28</sup>. If the findings of other investigations, showing no population build up  
317 occurs under fallow<sup>14</sup> and our theory of soil consolidation is correct, it may be the case that  
318 both class two radish treatments, and the susceptible radish, did also result in population  
319 declines, just that they were not significant enough to detect. However, the results could also  
320 be due to sampling error, which is inherently large in field experiments with nematodes<sup>29,30</sup>  
321 although this error should have been equal between the plots as the same method was used  
322 throughout.

323

324

325 The need to survey the field for BCN populations and then sample each plot before planting  
326 the trap crops was very time consuming and also delayed the establishment of the trap  
327 crops by several weeks in which the sampling, cyst extraction and nematode counts took  
328 place. The effectiveness of the trap crops may be further improved by earlier establishment  
329 which would benefit from warmer weather conditions and longer day lengths, thereby  
330 promoting more growth of the trap crops and because of these factors earlier sowing has  
331 been shown to increase the effectiveness of trap crops and the growth of cover crops in  
332 Germany.<sup>6,13,31</sup> However, labour and equipment availability in July and August, typically  
333 when harvest of combinable crops is underway, may not allow for earlier sowing. The  
334 extension of growing the cover crops through the winter is unlikely to have affected BCN  
335 populations due to the temperature of the soil dropping below 10°C, the temperature at  
336 which hatching of BCN juveniles ceases<sup>8</sup>, during early November in both years.

337

338 Population reductions, as a results of growing trap crops may only be useful if they lead to  
339 yield increases in following crops and can be accommodated in the farming rotation. Cooke<sup>32</sup>

340 stated clearly that a trap crop should meet three criteria before being suitable for commercial  
341 use, and these still stand true today. Firstly it should stimulate egg hatch, which we have  
342 shown to be the case with at least two of the varieties used. Secondly, reproduction of the  
343 nematode should be limited, which appeared to be the case as no varieties consistently  
344 showed major increases in populations. Finally, the trap crop should be agriculturally and  
345 economically acceptable. In our case, planting at the beginning of September fitted the  
346 farming system on the host farm as cereal harvest was completed and labour and machinery  
347 was available for drilling the brassicas. However, when the costs of seed are factored in,  
348 along with fertiliser application, and the destruction of the trap crops in the winter, the costs  
349 to the grower may be prohibitive and would require the trap crops to boost yields of the  
350 following crop of sugar beet by at least 5 tonnes ha<sup>-1</sup><sup>i</sup>.

351 The use of BCN tolerant varieties of sugar beet is now common place for growers with  
352 infestations.<sup>6,33</sup> These varieties overcome the majority of yield losses, so returns to the  
353 grower are significantly greater than using a susceptible variety, but losses are not entirely  
354 prevented and a negative relationship between *Pi* and sugar yield has been found by Hauer  
355 et al.<sup>6</sup> Therefore, the addition of a trap crop to the rotation may be useful in these  
356 circumstances, but only if the additional yield benefit can meet the costs to cultivate the trap  
357 crop. Other factors relating to the planting of a trap crop must also be considered and these  
358 may be positive (such as improvements to soil structure, weed control, nutrient retention and  
359 prevention of soil erosion) or negative, such as providing a habitat overwinter for other  
360 agriculturally important pests. Further research is required to consider the use of trap crops  
361 within arable farming systems.

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<sup>i</sup> Class 1 radish costs £2.40 per Kg (P. Brown Pers. Comm.) at a seed rate of 22kg.ha<sup>-1</sup> = £52.80 ha<sup>-1</sup>. Costs of sowing = 24.37 ha<sup>-1</sup>, Fertilising 11.16 ha<sup>-1</sup> (plus fertiliser £21.33) and flailing £19.50 ha<sup>-1</sup> = Total costs of 129.16 ha<sup>-1</sup>. With current beet price of £22.50 per tonne, yields need to increase by more than 5.74 tonnes.ha<sup>-1</sup> following the trap crop to be economically viable. Costs are even higher with other treatments due to greater seed costs. [Costs calculated using standard figures<sup>34</sup>]

364 **5. Conclusions**

365 Our experiments have shown that brassica trap crops may form part of an integrated pest  
366 management strategy to manage BCN populations in Eastern England. The class 1 radish  
367 and the class 2 mustard treatments tested produced significant population reductions  
368 whereas the two class 2 radish treatments did not reduce the populations significantly.  
369 Understanding the costs of growing a trap crop and how these might be repaid in the  
370 following crop is needed to provide an agronomic benefit.

371  
372 Further studies are needed to evaluate more of these resistant brassica trap crops over a  
373 wider range of seasons and soil types but our results indicate that two varieties tested offer  
374 some form of BCN population management. However, these population reductions may  
375 soon be undone when a susceptible host species for BCN is planted. In addition, a yield  
376 benefit from the use of the trap crop needs to be identified prior to their widespread adoption  
377 in Britain for BCN control.

378

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386 assistance.

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392 **7. References**

- 393 1 Müller J, The economic importance of *Heterodera schachtii* in Europe, *Helminthologia*  
394 **36**:205–213, Biol Bundesanstalt Land & Forstwirtschaft, Inst Nematol &  
395 Wirbeltierkunde, D-48161 Munster, Germany. Muller, J (reprint author), Biol  
396 Bundesanstalt Land & Forstwirtschaft, Inst Nematol & Wirbeltierkunde, Toppheideweg  
397 88, D-48161 Munster, Germany. (1999).
- 398 2 Moens M, Perry RN, and Jones JT, *Cyst Nematodes - Life Cycle and Economic*  
399 *Importance*, ed. by Moens M, Perry RN, and Jones JT, *Cyst Nematodes*, CAB  
400 International, Wallingford, Oxfordshire, UK, pp. 1–26 (2018).
- 401 3 Cooke DA, Beet Cyst Nematode (*Heterodera Schachtii* Schmidt) and its Control on  
402 Sugar Beet, *Agric Zool Rev* **2**:135–183 (1987).
- 403 4 Blok VC, Tylka GL, Smiley RW, de Jong WS, and Daub M, Resistance Breeding, ed.  
404 by Moens M, Perry RN, and Jones JT, *Cyst Nematodes*, CAB International,  
405 Wallingford, Oxfordshire, UK, pp. 174–214 (2018).
- 406 5 Wright A, Stevens M, Back M, and Sparkes D, Interactions between *Heterodera*  
407 *schachtii* and Sugar Beet, *Asp Appl Biol* **134**:81–88 (2017).
- 408 6 Hauer M, Koch HJ, Krüssel S, Mittler S, and Märländer B, Integrated control of  
409 *Heterodera schachtii* Schmidt in Central Europe by trap crop cultivation, sugar beet  
410 variety choice and nematicide application, *Appl Soil Ecol* **99**:62–69 (2016).
- 411 7 Kakaire S, Grove IG, and Haydock PPJ, Effect of temperature on the life cycle of  
412 *Heterodera schachtii* infecting oilseed rape (*Brassica napus* L.), *Nematology* **14**:855–  
413 867, [Kakaire, Stephen; Grove, Ivan G.; Haydock, Patrick P. J.] Harper Adams Univ  
414 Coll, Crop & Environm Res Ctr, Nematol & Entomol Grp, Newport TF10 8NB, Shrops,  
415 England. Kakaire, S (reprint author), Harper Adams Univ Coll, Crop & Environm Res  
416 Ctr, *Nematol & E* (2012).
- 417 8 Dewar AM and Cooke DA, *Pests*, ed. by Draycott AP, Sugar Beet, Blackwell  
418 Publishing Limited, Oxford, pp. 316–350 (2006).
- 419 9 LIZ, LIZ - Ausfallraps-Manager, 2016.

- 420 <http://raps.rheinmedia.de/raps/index.php?region=17&datum=2017-07->  
421 [08&enddatum=2017-10-12](http://raps.rheinmedia.de/raps/index.php?region=17&datum=2017-07-08&enddatum=2017-10-12) [accessed 9 May 2018].
- 422 10 Back MA, Cortada L, Grove IG, and Taylor V, Field Management and Control  
423 Strategies, ed. by Perry RN, Moens M, and Jones JT, Cyst Nematodes, CABI,  
424 Wallingford, Oxfordshire, UK, pp. 305–336 (2018).
- 425 11 Hafez SL, The use of green manure crops in a sugarbeet rotation for sugarbeet cyst  
426 nematode management, *J Nematol* **26**:548 (Abstr.) (1994).
- 427 12 Koch DW, Gray FA, and Krall JM, Nematode-Resistant Oil Radish for *Heterodera*  
428 *schachtii* control II. Sugarbeet-Dry Bean-Corn Rotations, *J Sugar Beet Res* **35**:63–75  
429 (1998).
- 430 13 Kenter C, Lukashyk P, Lukashyk P, Daub M, and Ladewig E, Population dynamics of  
431 *heterodera schachtii* schm. and yield response of susceptible and resistant sugar beet  
432 (*beta vulgaris* L.) after cultivation of susceptible and resistant oilseed radish (*raphanus*  
433 *sativus* L.), *J Cultiv Plants* **66**:289–299 (2014).
- 434 14 Hemayati SS, Jahad-e Akbar M-R, Ghaemi A-R, and Fasahat P, Efficiency of white  
435 mustard and oilseed radish trap plants against sugar beet cyst nematode, *Appl Soil*  
436 *Ecol* **119**:192–196, Elsevier (2017).
- 437 15 Hallett SH, Sakrabani R, Keay CA, and Hannam JA, Developments in land  
438 information systems: examples demonstrating land resource management capabilities  
439 and options, *Soil Use Manag* **33**, 514–529 (2017).
- 440 16 Winfield AL, Enfield MA, and Foreman JH, A column elutriator for extracting cyst  
441 nematodes and other small invertebrates from soil samples, *Ann Appl Biol* **111**:223–  
442 231 (1987).
- 443 17 Reid E, A rolling method for opening cysts of potato root eelworm, *Plant Pathol* **4**:28–  
444 29 (1955).
- 445 18 Fenwick DW, A New Modification of the McMaster Slide for Use in Potato-root  
446 Eelworm Investigations, *J Helminthol* **25**:173–176 (1951).
- 447 19 Cooke DA, The effect of resistant cultivars of catch crops on the hatching of

- 448 Heterodera schachtii, Ann Appl Biol **106**:111–120 (1985).
- 449 20 Sharma LK, Bu H, Denton A, and Franzen DW, Active-optical sensors using red NDVI  
450 compared to red edge NDVI for prediction of corn grain yield in north Dakota, U.S.A,  
451 Sensors (Switzerland) **15**:27832–27853 (2015).
- 452 21 Cowley RB, Luckett DJ, Moroni JS, and Diffey S, Use of remote sensing to determine  
453 the relationship of early vigour to grain yield in canola (*Brassica napus* L.) germplasm,  
454 Crop Pasture Sci **65**:1288–1299 (2014).
- 455 22 Vos J, Van Der Putten PEL, Hussein MH, Van Dam AM, and Leffelaar PA, Field  
456 observations on nitrogen catch crops II. Root length and root length distribution in  
457 relation to species and nitrogen supply, Plant Soil **201**:149–155 (1998).
- 458 23 Fatemy S and Abootorabi E, Hatching activity, invasion rate and reproduction of  
459 Heterodera schachtii on oilseed rape cultivars, Nematol Mediterr **30**:163–166 (2002).
- 460 24 Vandenbossche BA, Effect of temperature on the interactions between beet cyst  
461 nematodes (*Heterodera schachtii* and *Heterodera Betae*) and sugar beet, Georg-  
462 August-University Göttingen (2016).
- 463 25 Meinecke A, Ziegler K, Bürcky K, and Westphal A, Composition of the stubble weed  
464 flora and its role for *Heterodera schachtii* in the year preceding sugar beet production,  
465 Weed Res **54**:614–623, Wiley/Blackwell (10.1111) (2014).
- 466 26 BBRO, Beet Cyst Nematode Technical Guide, ed. by British Beet Research  
467 Organisation, Brooms Barn (2009).
- 468 27 Ahmad M, Sedaghatjoo S, and Westphal A, Reproductive capacity of *Heterodera*  
469 *schachtii* on *Thlaspi arvense*, *Capsella bursa-pastoris* and varying populations of  
470 *Chenopodium album*, J Plant Dis Prot **123**:37–42 (2016).
- 471 28 G. WA, Vertical Distribution of Potato, Beet and Pea Cyst Nematodes in some Heavily  
472 Infested Soils, Plant Pathol **26**:85–90.
- 473 29 Southey JF, Principles of sampling for nematodes, Laboratory Methods for work with  
474 Plant and Soil Nematodes, HMSO, London, United Kingdom, pp. 1–4 (1986).
- 475 30 Been TH and Schomaker CH, Distribution Patterns and Sampling, ed. by Perry RN

476 and Moens M, Plant Nematology, CAB International, Wallingford, Oxfordshire, UK, pp.  
477 331–358 (2013).

478 31 Koch HJ, Windt A, Mittler S, and Hauer M, Effect of weather variables on biomass  
479 production and N uptake of catch crops, and their influence on soil water and Nmin  
480 content in Northern Germany, J Cultiv Plants **69**:361–372 (2017).

481 32 Cooke DA, The relationship between numbers of *Heterodera schachtii* and sugar beet  
482 yields on a mineral soil, 1978-81, Ann Appl Biol **104**:121–129 (1984).

483 33 BBRO, British Beet Research Organisation Reference Book, Norwich, Norfolk, United  
484 Kingdom (2017).

485 34 Redman G, The John Nix Pocketbook for Farm management 2018, 48th ed., Agro  
486 Business Consultants, Melton Mowbray (2017).

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