Running Title: Whole-exome sequencing of the BDR cohort: Evidence to support the role of the *PILRA* gene in Alzheimer's disease

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Abstract

Aim: Late-onset Alzheimer's disease (LOAD) accounts for 95% of all Alzheimer's cases and is genetically complex in nature. Overlapping clinical and neuropathological features between AD, FTD and Parkinson's disease highlight the potential role of genetic pleiotropy across diseases. Recent GWAS have uncovered 20 new loci for AD risk, however these exhibit small effect sizes. Using NGS, here we perform association analyses using exome-wide and candidate-gene driven approaches.

Methods: Whole-exome sequencing was performed on 132 AD cases and 53 control samples. Exomewide single variant association and gene burden tests were performed for 76,640 non-singleton variants. Samples were also screened for known causative mutations in familial genes in AD and other dementias. Single variant association and burden analysis was also carried out on variants in known AD and other neurologic dementia genes.

Results: Tentative single variant and burden associations were seen in several genes with kinase and protease activity. Exome-wide burden analysis also revealed significant burden of variants in *PILRA* (P=3.4x10⁻⁵), which has previously been linked to AD via GWAS, hit *ZCWPW1*. Screening for causative mutations in familial AD and other dementia genes revealed no pathogenic variants. Variants identified in *ABCA7*, *SLC24A4*, *CD33* and *LRRK2* were nominally associated with disease (P<0.05) but did not withstand correction for multiple testing. *APOE* (P=0.02) and *CLU* (P=0.04) variants showed significant burden on AD.

Conclusions: In addition, polygenic risk scores (PRS) were able to distinguish between cases and controls with 83.8% accuracy using 3,268 variants, sex, age at death and *APOE* ε4 and ε2 status as predictors.

Abbreviations

AD – Alzheimer's disease AUC – area under curve

BDR – Brains for Dementia Research
DLB – Dementia with Lewy bodies

fEOAD – familial early onset Alzheimer's disease

FTD - Frontotemporal dementia

GWAS – genome-wide association study LOAD – late onset Alzheimer's disease

MAF – minor allele frequency NGS – next generation sequencing

OR – odds ratio P – P-value

PCR – polymerase chain reaction
PD – Parkinson's disease
PRS – polygenic risk score

1. Introduction

Alzheimer's disease (AD) is the most common form of dementia, affecting over 850,000 people in the UK alone, a number expected to rise to 1 million by 2025 [1]. There are two forms distinguished by the age when symptoms first appear. In the early-onset familial form (fEOAD), symptoms appear before 65 years of age, however this only accounts for about 5% of cases [2]. Mutations in the familial genes *APP*, *PSEN1* and *PSEN2* are rare but highly penetrant. Individuals with these mutations are almost certain to develop fEOAD [3]. The majority of cases are sporadic in nature and classified as late-onset (LOAD), with symptoms appearing at 65 years or later. This represents the other 95% of all incidences [4]. Presence of the *APOE* ε4 allele is the largest known genetic risk factor for LOAD, with a 2-3 fold increase in risk for carriers and 15 fold for individuals homozygous for the ε4 allele [5]. The rare ε2 allele confers a protective effect and appears to reduce AD risk by up to 40%.

Genome-wide association studies (GWAS) have identified 20 risk variants associated with LOAD [6–9]; this has implicated several new pathways in AD, such as endocytic processing, inflammation and cholesterol transport [8]. Although highly replicable in Caucasian groups, these effects have been difficult to replicate in other populations [10]. These common variants also exert only small effects on disease risk, which does not account for much of the missing heritability in AD. It is likely that low frequency variants, not detected by GWAS, could have greater effect sizes and therefore explain more of the heritable component.

Advances in next-generation sequencing (NGS) in recent years have allowed exomes and entire genomes to be explored at single-base level. The exome accounts for approximately 1% of the human genome, yet it harbours almost 85% of known mutations underlying disease-related traits [11]. Therefore, rare mutations can be identified using this technology. Whole-exome sequencing (WES) has identified a rare variant in the *TREM2* gene, R47H, associated with a 5-fold increase in AD risk [12]. These studies have also found rare causative variants in *CLU* and *SORL1* that were overlooked by GWAS [13]. By identifying the genetic variants of individuals, WES has the potential to uncover more rare variants associated with AD risk.

There are several overlapping clinical and neuropathological features across different dementias. For example, dementia with Lewy bodies (DLB) shares clinical features with AD and Parkinson's disease (PD), often resulting in misdiagnosis. PD and DLB are both synucleinopathies presenting with alphasynuclein deposits in the brain, whereas *APOE* £4 increases risk of disease in AD and DLB [14]. This suggests that genetic risk factors may contribute to more than one disease, known as genetic pleiotropy, whereby a gene or DNA variant can influence multiple phenotypes.

Clinically well-characterised brain tissue samples from healthy individuals remains a limiting factor in the study of neurological disorders [15], Brains for Dementia Research (BDR) (www.brainsfordementiaresearch.org.uk) is a network of six leading UK brain banks (jointly funded by

Alzheimer's Research UK and Alzheimer's Society), specifically created to address the shortages of high-quality brain tissue samples from healthy individuals as well as those with dementia. This project is a planned brain donation programme with over 3000 participants, aged 65 years and above, with and without the diagnosis of dementia. Regular, standardised cognitive and psychiatric assessment of potential brain donors during life is critical in optimising the value of brain tissue for research [16,17].

We performed single variant and burden analysis on coding variants to identify significant associations with LOAD. We also report on screening of 132 LOAD patients from the Brains for Dementia Research (BDR) resource with the aim to identify causative or predicted pathogenic coding variants in 40 selected genes. Of these, 16 are associated with familial forms of neurodegeneration, including fEOAD (*APP*, *PSEN1* and *PSEN2*), frontotemporal dementia (FTD) and Amyotrophic lateral sclerosis (ALS) (*C9or72*, *CHMP2B*, *FUS*, *GRN*, *MAPT*, *TARDBP* and *VCP*), PD (*LRRK2*, *PARK2*, *PARK7*, *PINK1* and *SNCA*) and Prion disease (*PRNP*). The remaining genes were selected from AD GWAS and NGS (20 GWAS, *APOE* and *TREM2*).

Polygenic risk scores (PRS) have been increasingly used to investigate the effect of multiple genetic variants on disease traits. It is based on the notion that many variants with small effects will not be detectable at genome-wide significance, however collectively they may have a strong effect [18]. PRS were generated to examine the association between multiple genetic markers and their collective effects on LOAD.

2. Materials and Methods

2.1 BDR samples

The BDR cohort comprised of 132 clinically diagnosed LOAD (age at onset ≥65 years) cases and 53 cognitively normal controls; all diagnoses were neuropathologically confirmed (Supplementary Table S1). Neuropathological diagnoses were undertaken by experienced neuropathologists within the Brains for Dementia Research network and were based on Thal Aβ phases [19], neurofibrillary tangle Braak stages [20], Consortium to Establish a Registry for Alzheimer's disease (CERAD) criteria for AD [21] which are all combined in the National Institute on Aging − Alzheimer's Association guidelines [22], together with the Newcastle / McKeith criteria for Lewy body disease [23] and for FTLD-TDP as described by [24]. Whilst there are no fully established criteria for vascular pathology the VCING criteria were used [25] along with those proposed by Smallwood et al (2012) [26] and Grinberg and Thal (2010) [27].

Demographics of case and control samples for each centre are shown in Table 1. The average age at death was 82.5 years (range 65-101 years) for LOAD samples. For control individuals, average age at

death was 85.9 years (range 58-104 years). The proportion of females between cases and controls were similar, accounting for around 50% of the total sample size. *APOE* ϵ 4 carriers were 3-fold higher in cases (64.7%) with almost one fifth (17.3%) being homozygous for the ϵ 4 allele. In comparison only 24.5% of controls were carriers and no individuals were ϵ 4 ϵ 4. The ϵ 2 allele was present in 10 control samples (18.9%) and 8 cases (6.2%), with both a case and control sample being homozygous for the allele. The higher 3-fold frequency observed in control samples correlates with the protective effect of the ϵ 2 allele. All samples in the BDR cohort used for analysis were classified as AD or control by neuropathology.

The BDR has a number of neuropathological features available for more definitive analysis of genotype-phenotype correlation [28]. Data includes the CERAD scale, Braak tangle and Lewy body staging, with simplified measures (present/absent; mild/moderate/severe) of small vessel disease, deposition of TDP-43 protein, arteriolar Aβ-CAA and cerebrovascular atherosclerosis. Detailed clinical and cognitive information on the samples is also available upon request, demonstrating the potential value of the BDR cohort for very detailed analyses in future studies as more extensive genetic data is generated.

2.2 DNA extraction

DNA was extracted from brain tissue using standard phenol-chloroform procedures. Samples were analysed on the Agilent TapeStation and quantified using the Nanodrop 3300 spectrometer to ensure high concentration and quality material was obtained. Samples were genotyped for *APOE* ε2, ε3 and ε4 alleles using the TaqMan method (Applied Biosystems) to determine *APOE* status.

2.3 Exome sequencing library prep

DNA libraries were hybridised to exome-capture probes with Agilent SureSelect Human All Exon Kit V4 for Illumina GA (Agilent Technologies) as per manufacturer's protocol. Exome-enriched libraries were sequenced on the Illumina HiSeq 2500 using 2 x 100bp paired end read cycles. The Agilent capture library includes 5' and 3' untranslated regions.

2.4 Bioinformatics

Paired-end sequence reads were aligned to the human reference genome build 19 (UCSC hg19) using Burrows-Wheeler Aligner [29]. Format conversion, indexing and removal of PCR duplicates were performed with Picard (www.picard.sourceforge.net/index.shtml). The Genome Analysis Toolkit was used for recalibration of base quality scores, realignment around indels and variant calling [30]. Variants were annotated using ANNOVAR [31] and Variant Effect Predictor [32] predicted SIFT and Polyphen2 scores of protein coding variants. Consistency between SIFT and Polyphen2 predictions and the databases allowed more reliable classification. Variants were also checked against

established databases (dbSNP v.149, 1000 Genomes Project and Exome Variant Server).

2.5 Filtering

Singleton variants with MAF \leq 0.002 were removed in VCFtools [33]. Coding variants in genes were filtered by annotation with SnpSift [34]. Visualisation of variants was performed, when necessary, using Integrative Genomics Viewer [35]. Individuals with a calculated age at onset below 65 years were removed and samples were screened for causative mutations in fEOAD genes *APP*, *PSEN1* and *PSEN2* to ensure only sporadic cases were used for analyses. Pathogenicity of variants was determined using pathogenic status in AD&FTD and PD mutation databases [36].

2.6 Association analyses

Quality control filtering was performed on the VCF using VCFtools. Individuals were removed if genotyping rate \leq 97%, followed by markers with call rate \leq 98%. Markers with significant deviation (P<0.001) from the Hardy-Weinberg equilibrium in control individuals were removed. After removing samples and markers failing quality control, 290 individuals remained with 76,640 non-singleton variants in coding regions. The average genotyping rate was 99.9%.

Plink files were imported to PLINK-SEQ [37] (https://atgu.mgh.harvard.edu/plinkseq/). Single-variant association was performed on samples using a logistic regression test correcting for the covariates sex, age at death and *APOE* ε4 allele count.

Gene-based association for genes of interest was calculated in R using a SKAT-O [38] burden test. The C-alpha test was used for exome-wide analysis and the SKAT-O test was used for selected genes.

2.7 Polygenic risk scoring

PRS were generated for BDR samples using PRSice [39]. The International Genomics of Alzheimer's Project (IGAP) summary data was used as the base dataset, collated from 17,008 LOAD cases and 37,154 controls. A region of 500kb around the *APOE* locus was excluded from the analysis. The best-fit model with the greatest predictive accuracy was computed using area under the curve (AUC) in SPSS. Additional predictor variables included were the number of *APOE* ε4 and ε2 alleles, age, sex and genotypes for the GWAS SNPs.

3. Results

3.1 Overview of data

Exome-sequencing was performed on a total of 292 individuals. The final cohort consisted of 132

LOAD cases and 53 control samples after quality control filtering. A total of 157,217 non-singleton variants were present in 290 individuals, with a minimum of 2 alleles observed per variant. Filtering to retain only coding mutations resulted in 76,640 variants for exome-wide analysis.

3.2 Exome-wide analyses

Burden analysis using a C-alpha test highlighted some nominally significant gene associations with AD, shown in Table 2. *PILRA* and *PRSS45* are just below the Bonferroni-corrected threshold (P=2x10⁻⁶) at P=3.4x10⁻⁵ and P=5.9x10⁻⁵, respectively. *PILRA* has previously been linked to AD through *ZCWPW1*, which was highlighted by the GWAS meta-analysis [9]. Five variants in *PILRA* contribute to the effect: intronic variants rs7792525, rs190071731 and rs148891131, synonymous mutation rs2405442 and missense variant p.S279L (rs34266222). GWAS SNP rs1476679 is in weak LD with rs2405442 (R²=0.50). This mutation is tolerated as predicted by SIFT and Polyphen2. The other genes have not previously been linked to AD. *PRSS45* contains 5 variants that drive this signal, of which 2 were highly associated with disease (Table 3), therefore it is possible that these SNPs are contributing to the signal.

Exome-wide association analysis was performed on the non-singleton variants, correcting for age, sex and *APOE* ε4 status of individuals. As expected, *APOE* SNP rs429358 showed the most significant association prior to adjusting for covariates (P=7.2x10⁻⁹, OR=6.5 [3.2-13.1]). There were no significant associations at the genome-wide threshold (P=5x10⁻⁸) or at the suggestive threshold (P=1x10⁻⁵) after correction. However due to low sample numbers, we do not have the power to detect any association at that level. Some tentative associations were observed and are shown in Table 3.

Several of the genes encompassing variants are involved in signaling pathways, including serine proteases *PRSS42* and *PRSS45* and inositol triphosphate receptor *ITPR3*. SIFT and Polyphen2 predictions indicate that most mutations are benign or tolerated. However missense mutations *TMEM260* p.A245S/T (rs17776256) and *AVPR1B* p.K65N (rs35369693) were predicted to be probably damaging by both software. Both variants are more frequent in control samples, signifying a protective effect. *MEP1B* is a metalloprotease recently implicated in *APP* cleavage and has been implicated in inflammation. The synonymous variant p.S537 (rs173032) has an odds ratio of 3.1 (1.6-5.9) and has a significantly greater frequency in cases than controls, inferring an association with AD. Two missense mutations in *PRSS45* are associated with protection against AD, p.I190L (rs58830807) and P130Q (rs58943210) however both are predicted to be benign. None of the genes aside from *MEP1B* have been directly linked to dementia.

3.3 Polygenic risk scoring

A 500kb region around the APOE gene containing 227 variants was excluded from the analysis to

identify effects independent of *APOE*. The predictive accuracy of each tested model is given in Table 4, denoted by area under the curve (AUC), with 95% confidence intervals. The *APOE* ϵ 4 allele alone has 71.8% accuracy in discriminating between cases and controls, however this is a poor fit model. Other covariates ϵ 2, age, sex and GWAS greatly increase the predictive power 83.0%. In total 3,268 variants were utilised to score risk of developing AD using the best fit model, which had a predictive accuracy of 83.8% when combined with all covariates as predictors. The addition of PRS only increased this accuracy by 0.8%, which is similar to improvements seen in other studies. There is overlap between scores for both groups, however on average scores were higher for cases. Mean scores for AD cases were 3.6×10^{-4} compared to 2.7×10^{-4} for control samples.

3.4 Screening for familial mutations

Samples were screened for mutations in fEOAD genes to remove any non-sporadic cases. A total of 6 coding variants were found in *APP* (1), *PSEN1* (1) and *PSEN2* (4), shown in Table 5. All individuals were heterozygous for the variants listed and mutations were synonymous except two identified as missense. *PSEN1* p.E318G (rs17125721) was classified as a risk modifier but not pathogenic, found in 5 cases and 1 control sample. *PSEN2* p.S130L (rs63750197) was previously identified as possibly damaging *in silico* with unclear pathogenicity, present in 1 case and 1 control. Both SNPs were predicted to be deleterious depending on the transcript. However, no causative or fully penetrant pathogenic mutations were observed in these genes, confirming that these samples are representative of sporadic AD.

Other known neurologic genes were also screened for potential pathogenic mutations to identify genetic overlap between sporadic LOAD and other neurodegenerative diseases (Table 6). Mutations in Parkinson's genes *LRRK2*, *PARK2* and *PINK1* appear to have some possibly damaging consequences on the proteins. *PARK2* p.R275W (rs34424986) is very rare and present in 1 AD case and 1 control sample with mild cerebral amyloid angiopathy and presence of an unspecified dementia. SIFT/Polyphen2 predictions both indicate a potentially deleterious effect of this mutation. Variant p.P246L (rs149953814) was also found in 1 case and control, both also presenting with mild non-amyloid small vessel disease. Samples were heterozygous for both variants.

Previously uncharacterised mutations were found in *CHMP2B* and *LRRK2*. The frameshift variant in *CHMP2B* was seen in a healthy control and an individual with AD. The *LRRK2* mutation was heterozygous in a sample of each phenotype and results in a p.L1271P change. *PRNP* p.M129V (rs1799990) has previously been implicated as a risk factor for prion disease, here however it is observed in an equal number of case and controls, suggesting no effect in AD.

3.5 Association analyses of known neurologic genes

Following on from this, genes were selected based on whether they had been linked to AD or other neurological diseases. Direct functionality was inferred from data filtered for coding variants only. A total of 76,640 variants were annotated as coding mutations, with 219 variants in 35 of the selected genes. Association and burden analyses were performed as before on the subset of variants. No coding variants were identified in AD genes *HLA-DRB1*, *HLA-DRB5* and *MEF2C* or the other neurologic genes *MAPT* and *TARDBP*.

We performed single variant association on all 219 variants in the selected genes. No variants reached genome-wide or suggestive levels of significance as identified by a logistic regression test with correction for covariates. The most significant associations (P<0.05) are listed in Table 7. Results for all 219 variants are shown in supplementary Table S2.

A large proportion of the highly associated variants are synonymous mutations. Four *ABCA7* variants are present, with 2 suggesting increased risk (OR>1) and 2 showing a protective effect (OR<1). *ABCA7* rs3752234 and rs3752237 are both synonymous mutations that increase risk more than 2-fold. *SLC24A4* synonymous SNP (rs7144273) also showed strong effects in the risk direction (OR=1.63, P=0.018).

The majority of variants appear to be exhibiting a protective effect as indicated by the odds ratios, as they were observed more frequently in control samples. LRRK2 p.M1646T (rs35303786) missense is predicted as benign, however it is found in a greater frequency in control samples (OR=0.14, P=0.018). We calculated gene-based burden using a SKAT-O test to provide greater statistical power than that of a single-marker test (Table 8). Both sets of familial genes did not appear to exhibit any burden on LOAD. Burden analysis revealed two significant associations; APOE and CLU were the only genes to reach significance (P \leq 0.05). However, they would not pass Bonferroni correction (P=0.0014). Five variants in CLU contributed to the effect seen, which was corrected for age, sex and APOE ϵ 4 status. Two of these variants were significant, synonymous variant rs9331939 and rs149859119 (p.S16R), therefore they could be driving the signal in this gene.

4. Discussion

In this study, we initially investigated genetic association with LOAD using an exome-wide approach. Although the analyses did not find any significant associations when corrected for multiple testing, the sample size only provides enough power to detect common variant (MAF>5%) associations with an effect size above 2.2 with 80% certainty. Nonetheless, single variant analysis highlighted some interesting tentative associations which may merit further exploration.

Burden analysis revealed a tentative association with PILRA, an inhibitory immunoglobulin receptor

involved in regulating signal transduction in the immune system. This gene has previously been linked to AD via its interaction with paired activation receptor *PILRB* and GWAS hit *ZCWPW1*. It is expressed on myeloid cells and works with *PILRB*, which also associates with *DAP12* and *TREM2* [40]. *PILRA* SNP rs2405442 is in weak LD (r²=0.5) with GWAS SNP rs1476679, suggesting this signal is likely to be independent of the GWAS association. *ZCWPW1* locus SNP rs1476679 was nominally associated with reduced *PILRA* levels [41]. This suggests a potential role for the gene in AD, highlighting the need for further investigation.

Many of the remaining genes on the burden list are enzymes with serine/threonine activity or serine proteases, such as *PRSS45*, *BCR*, *KLK2* and *THNSL2*. Efficient breakdown of proteins is important as impairments in this can lead to the buildup of misfolded proteins. Dysfunction of the amyloid protein degradation pathway has been implicated in AD. None of these genes have been previously linked to AD. However in combination, enzymes regulating protein function and breakdown could play a greater role in disease and this too warrants further exploration.

Multiple *PRSS45* variants were observed, with two missense mutations found to be associated with AD, exerting a protective effect. However, functional predictions indicate that both polymorphisms are benign. This gene encodes a serine protease, part of a group of enzymes that cleave peptide bonds. *PRSS45* SNPs were also highly associated when tested in burden. Missense variants *TMEM260* p.A245S and *AVPR1B* p.K65N were both predicted to be damaging to the protein *in silico* and found more frequently in control samples. The function of *TMEM260* is not clearly understood, whereas *AVPR1B* is a vasopressin receptor located in the anterior pituitary gland that stimulates ACTH release. *AVPR1B* SNP rs35369693 has been linked to mood disorders and found more frequently in affected females [42]. The mutation in *MEP1B*, known as meprin β is synonymous; recent proteomic studies have found that these metalloproteases can cleave APP, affecting Aβ levels [43,44]. While these associations are tentative, examination of other larger datasets could be worthwhile.

PRS generated for individuals showed that, on average, scores were significantly higher in LOAD cases than controls, despite an overlap amongst the cohort. Using sex, age at death, *APOE* £4 and £2 allele counts and GWAS SNP genotypes as variables for prediction, the model was able to distinguish cases and control with 83.8% accuracy. A total of 3,268 variants were used to predict disease risk. The presence of controls with high risk scores suggests that these individuals may have gone on to develop AD had they lived longer. The utility of PRS has already been demonstrated in AD, with individuals' genetic risk profiles able to predict disease susceptibility with more than 80% accuracy [45]. However, there were controls with high PRS and no phenotypic changes indicative of dementia and also cases with low PRS. Although PRS can identify more of the genetic component of AD, this shows that there is still unexplained missing heritability.

Mutations in familial AD genes, APP, PSEN1 and PSEN2 are rare but highly penetrant. Screening of

these genes revealed no pathogenic variants and samples harbouring mutations were heterozygotes, confirming there were no familial EOAD cases amongst the BDR LOAD classified cases. Other neurologic familial genes were also screened for pathogenic mutations linked to related dementias. No known causative mutations were identified, however *PARK2* p.R275W was predicted to be deleterious and has unknown pathogenicity in the PD mutation database. It produces an unusual distribution of parkin with large cytoplasmic and nuclear inclusions [46]. The variant was present in one case and control sample, however, which suggests that it is likely benign and not pathogenic in nature. Previously uncharacterised mutations were identified in *CHMP2B* and *LRRK2*, with a frameshift variant in *CHMP2B* and a missense variant in *LRRK2*. The frameshift variant was only seen in one control sample, suggesting that it could be a sequencing artifact. *LRRK2* p.L1271P is present in a case and control sample so does not appear to segregate with disease. *PRNP* p.M129V has been highlighted as a risk factor for prion disease but appears not to be having any effect in AD.

TREM2 mutation p.R47H was observed in 3 AD case samples which were heterozygotes. This variant can increase risk of developing AD by 2-3 fold [12,47]. DNA was available for these 3 samples and Sanger sequencing confirmed the presence of the variant in these subjects. No control samples harboured this variant. However, given the documented frequency of R47H (MAF=0.002) this cohort appears to have a greater MAF of 0.008. This 4-fold greater frequency will be verified as the BDR sample set increases in size.

Single variant association of all neurologic gene variants revealed several synonymous mutations to be nominally associated with AD at P≤0.05. The majority of variants exerted effects in the protective direction with greater frequency in controls than case samples. Four *ABCA7* variants were significantly associated at P<0.05 with 2 increasing risk and 2 being protective. Synonymous variants rs3752234 and rs3752237 increased AD risk more than 2-fold, which is contradictory to previous findings [48,49], where the effects were protective. Conversely, rs4147915 and missense mutation rs3764645 p.E188G are protective. *ABCA7* p.E188G is predicted as tolerated and previously shown to have no effect on disease risk [48]. These findings need to be validated as the sample size increases.

Missense variant *LRRK2* p.M1646T was associated with protection against AD, but the amino acid substitution is predicted to be tolerated. However the mutation is known to increase risk of developing PD [50]. *LRRK2* mutations have previously been linked to AD with PD risk variant p.R1628P found in greater frequencies in AD cases than controls [51]. The variant increased apoptosis and cell death in transfected human cell lines. Therefore, it is likely that genetic pleiotropy possibly occurs across several neurodegenerative diseases. *LRRK2* is involved in autophagy and recycling proteins in the retrograde trafficking pathway. Mutations in this protein are associated with dendrite shortening in neurons, a possible cause of motor symptoms in PD [46]. With some shared clinical features, mutations in *LRRK2* could also affect the autophagy process in AD.

Gene-based burden analysis in SKAT-O allowed adjustment to correct for the effect of age, sex and number of *APOE* ε4 alleles. Both AD and other neurologic familial genes did not exhibit any burden on LOAD. *APOE* and *CLU* were significant to P<0.05 but did not pass Bonferroni correction. Only SNP p.S16R in *CLU* was significantly associated in single variant testing, indicating that this is driving the signal.

5. Conclusion

Although other familial neurologic genes did not show any burden on LOAD, an individual missense variant in *LRRK2* was tentatively associated; preliminary exploration of the data has indicated that genetic pleiotropy is likely to play a role in diseases with overlapping features. *LRRK2* is involved in PD, yet few studies have investigated its role in other dementias.

Exome-wide analysis has revealed a significant burden of *PILRA* variants on AD. Previous studies have identified a possible link with AD via GWAS hit *ZCWPW1* and paired receptor *PILRB* which associates with *DAP12* and *TREM2*. *PILRA* and *PILRB* function may be co-regulated and therefore further investigation should involve looking at both genes in AD.

Limitations in power have made it difficult to find many significant associations, but with on-going data collection, the sample size will increase to address this issue. However, using an exome sequencing approach it has been possible to detect rare variants with greater effect sizes, which previous GWAS did not permit.

Supplementary Information

Supplementary Table S1. Summary data of all case and control samples used for analysis. Supplementary Table S2. Variants found in AD-related genes in 129 LOAD cases and 53 controls.

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Approval

Written informed consent was obtained for all individuals and approved by the appropriate institutional review boards (BDR ethics number 08/H0704/128 + 5).

References

- Alzheimer's Society. Drug treatments for Alzheimer's disease. Natl Inst Heal Care Excell. 2014;1–13.
- 2. Minati L, Edginton T, Bruzzone MG, Giaccone G. American Journal of Alzheimer 's Disease and Other Dementias Current Concepts in Alzheimer 's Disease: A Multidisciplinary Review. 2009;95–121.
- 3. Bertram L, Lill CM, Tanzi RE. The genetics of Alzheimer's disease: Back to the future. Neuron. 2010;68(2):270–81.
- 4. Piaceri I, Nacmias B, Sorbi S. Genetics of familial and sporadic Alzheimer's disease. Front Biosci (Elite Ed). 2013 Jan 1;5:167–77.
- 5. Farrer LA, Cupples LA, Haines JL, Hyman B, Kukull WA, Mayeux R, Myers RH, Pericak-Vance MA, Risch N, Duijn CM van. Effects of Age, Sex, and Ethnicity on the Association Between Apolipoprotein E Genotype and Alzheimer Disease. JAMA. 1997 Oct 22;278(16):1349.
- 6. Harold D, Abraham R, Hollingworth P, Sims R, Gerrish A, Hamshere ML, Pahwa JS, Moskvina V, Dowzell K, Williams A, Jones N, Thomas C, Stretton A, Morgan AR, Lovestone S, Powell J, Proitsi P, Lupton MK, Brayne C, Rubinsztein DC, Gill M, Lawlor B, Lynch A, Morgan K, Brown KS, Passmore PA, Craig D, McGuinness B, Todd S, Holmes C, Mann D, Smith AD, Love S, Kehoe PG, Hardy J, Mead S, Fox N, Rossor M, Collinge J, Maier W, Jessen F, Schürmann B, Heun R, van den Bussche H, Heuser I, Kornhuber J, Wiltfang J, Dichgans M, Frölich L, Hampel H, Hüll M, Rujescu D, Goate AM, Kauwe JSK, Cruchaga C, Nowotny P, Morris JC, Mayo K, Sleegers K, Bettens K, Engelborghs S, De Deyn PP, Van Broeckhoven C, Livingston G, Bass NJ, Gurling H, McQuillin A, Gwilliam R, Deloukas P, Al-Chalabi A, Shaw CE, Tsolaki M, Singleton AB, Guerreiro R, Mühleisen TW, Nöthen MM, Moebus S, Jöckel K-H, Klopp N, Wichmann H-E, Carrasquillo MM, Pankratz VS, Younkin SG, Holmans PA, O'Donovan M, Owen MJ, Williams J. Genome-wide association study identifies variants at CLU and PICALM associated with Alzheimer's disease. Nat Genet. 2009 Oct;41(10):1088–93.
- 7. Lambert J-C, Heath S, Even G, Campion D, Sleegers K, Hiltunen M, Combarros O, Zelenika D, Bullido MJ, Tavernier B, Letenneur L, Bettens K, Berr C, Pasquier F, Fiévet N, Barberger-Gateau P, Engelborghs S, De Deyn P, Mateo I, Franck A, Helisalmi S, Porcellini E, Hanon O, European Alzheimer's Disease Initiative Investigators, de Pancorbo MM, Lendon C, Dufouil C, Jaillard C, Leveillard T, Alvarez V, Bosco P, Mancuso M, Panza F, Nacmias B, Bossù P, Piccardi P, Annoni G, Seripa D, Galimberti D, Hannequin D, Licastro F, Soininen H, Ritchie K, Blanché H, Dartigues J-F, Tzourio C, Gut I, Van Broeckhoven C, Alpérovitch A, Lathrop M, Amouyel P. Genome-wide association study identifies variants at CLU and CR1 associated with Alzheimer's disease. Nat Genet. 2009 Oct;41(10):1094–9.
- 8. Hollingworth P, Harold D, Sims R, Gerrish A, Lambert J-C, Carrasquillo MM, Abraham R, Hamshere ML, Pahwa JS, Moskvina V, Dowzell K, Jones N, Stretton A, Thomas C, Richards A, Ivanov D, Widdowson C, Chapman J, Lovestone S, Powell J, Proitsi P, Lupton MK, Brayne C, Rubinsztein DC, Gill M, Lawlor B, Lynch A, Brown KS, Passmore PA, Craig D, McGuinness B, Todd S, Holmes C, Mann D, Smith AD, Beaumont H, Warden D, Wilcock G, Love S, Kehoe PG, Hooper NM, Vardy ERLC, Hardy J, Mead S, Fox NC, Rossor M, Collinge J, Maier W, Jessen F, Rüther E, Schürmann B, Heun R, Kölsch H, van den Bussche H, Heuser I, Kornhuber J, Wiltfang J, Dichgans M, Frölich L, Hampel H, Gallacher J, Hüll M, Rujescu D, Giegling I, Goate AM, Kauwe JSK, Cruchaga C, Nowotny P, Morris JC, Mayo K, Sleegers K, Bettens K, Engelborghs S, De Deyn PP, Van Broeckhoven C, Livingston G, Bass NJ, Gurling H, McQuillin

- A, Gwilliam R, Deloukas P, Al-Chalabi A, Shaw CE, Tsolaki M, Singleton AB, Guerreiro R, Mühleisen TW, Nöthen MM, Moebus S, Jöckel K-H, Klopp N, Wichmann H-E, Pankratz VS, Sando SB, Aasly JO, Barcikowska M, Wszolek ZK, Dickson DW, Graff-Radford NR, Petersen RC, Alzheimer's Disease Neuroimaging Initiative, van Duijn CM, Breteler MMB, Ikram MA, DeStefano AL, Fitzpatrick AL, Lopez O, Launer LJ, Seshadri S, CHARGE consortium, Berr C, Campion D, Epelbaum J, Dartigues J-F, Tzourio C, Alpérovitch A, Lathrop M, EADI1 consortium, Feulner TM, Friedrich P, Riehle C, Krawczak M, Schreiber S, Mavhaus M, Nicolhaus S, Wagenpfeil S, Steinberg S, Stefansson H, Stefansson K, Snaedal J, Björnsson S, Jonsson P V, Chouraki V, Genier-Boley B, Hiltunen M, Soininen H, Combarros O, Zelenika D, Delepine M, Bullido MJ, Pasquier F, Mateo I, Frank-Garcia A, Porcellini E, Hanon O, Coto E, Alvarez V, Bosco P, Siciliano G, Mancuso M, Panza F, Solfrizzi V, Nacmias B, Sorbi S, Bossù P, Piccardi P, Arosio B, Annoni G, Seripa D, Pilotto A, Scarpini E, Galimberti D, Brice A, Hannequin D, Licastro F, Jones L, Holmans PA, Jonsson T, Riemenschneider M, Morgan K, Younkin SG, Owen MJ, O'Donovan M, Amouyel P, Williams J. Common variants at ABCA7, MS4A6A/MS4A4E, EPHA1, CD33 and CD2AP are associated with Alzheimer's disease. Nat Genet. 2011 May;43(5):429-35.
- 9. Lambert J, Ibrahim-Verbaas C, Harold D, Naj A. Meta-analysis of 74,046 individuals identifies 11 new susceptibility loci for Alzheimer's disease. Nature. 2013;
- 10. Jun G, Naj AC, Beecham GW, Wang L-S, Buros J, Gallins PJ, Buxbaum JD, Ertekin-Taner N, Fallin MD, Friedland R, Inzelberg R, Kramer P, Rogaeva E, St George-Hyslop P, Alzheimer's Disease Genetics Consortium, Cantwell LB, Dombroski BA, Saykin AJ, Reiman EM, Bennett DA, Morris JC, Lunetta KL, Martin ER, Montine TJ, Goate AM, Blacker D, Tsuang DW, Beekly D, Cupples LA, Hakonarson H, Kukull W, Foroud TM, Haines J, Mayeux R, Farrer LA, Pericak-Vance MA, Schellenberg GD. Meta-analysis confirms CR1, CLU, and PICALM as alzheimer disease risk loci and reveals interactions with APOE genotypes. Arch Neurol. 2010 Dec;67(12):1473–84.
- 11. Choi M, Scholl UI, Ji W, Liu T, Tikhonova IR, Zumbo P, Nayir A, Bakkaloğlu A, Ozen S, Sanjad S, Nelson-Williams C, Farhi A, Mane S, Lifton RP. Genetic diagnosis by whole exome capture and massively parallel DNA sequencing. Proc Natl Acad Sci U S A. 2009 Nov 10;106(45):19096–101.
- Guerreiro R, Wojtas A, Bras J, Carrasquillo M, Rogaeva E, Majounie E, Cruchaga C, Sassi C, Kauwe JSK, Younkin S, Hazrati L, Collinge J, Pocock J, Lashley T, Williams J, Lambert J-C, Amouyel P, Goate A, Rademakers R, Morgan K, Powell J, St. George-Hyslop P, Singleton A, Hardy J. TREM2 Variants in Alzheimer's Disease. N Engl J Med. 2013;368(2):117–27.
- 13. Tosto G, Reitz C. Genome-wide Association Studies in Alzheimer's Disease: A Review. Curr Neurol Neurosci Rep. 2013 Oct 17;13(10):381.
- 14. Bras J, Guerreiro R, Darwent L, Parkkinen L, Ansorge O, Escott-Price V, Hernandez DG, Nalls MA, Clark LN, Honig LS, Marder K, Van Der Flier WM, Lemstra A, Scheltens P, Rogaeva E, St George-Hyslop P, Londos E, Zetterberg H, Ortega-Cubero S, Pastor P, Ferman TJ, Graff-Radford NR, Ross OA, Barber I, Braae A, Brown K, Morgan K, Maetzler W, Berg D, Troakes C, Al-Sarraj S, Lashley T, Compta Y, Revesz T, Lees A, Cairns N, Halliday GM, Mann D, Pickering-Brown S, Dickson DW, Singleton A, Hardy J. Genetic analysis implicates APOE, SNCA and suggests lysosomal dysfunction in the etiology of dementia with Lewy bodies. Hum Mol Genet. 2014 Dec 1;23(23):6139–46.
- Samarasekera N, Salman RA-S, Huitinga I, Klioueva N, McLean CA, Kretzschmar H, Smith C, Ironside JW. Brain banking for neurological disorders. Lancet Neurol. 2013 Nov;12(11):1096– 105.

- 16. Vonsattel JPG, del Amaya MP, Keller CE. Twenty-first century brain banking. Processing brains for research: The Columbia University methods. Acta Neuropathol. 2008 May 6;115(5):509–32.
- 17. Kretzschmar H. Brain banking: Opportunities, challenges and meaning for the future. Nat Rev Neurosci. 2009 Jan 3;10(1):70–8.
- 18. Wray NR, Goddard ME, Visscher PM. Prediction of individual genetic risk of complex disease. Curr Opin Genet Dev. 2008;18(3):257–63.
- 19. Thal DR, Rüb U, Orantes M, Braak H. Phases of A beta-deposition in the human brain and its relevance for the development of AD. Neurology. 2002 Jun 25;58(12):1791–800.
- 20. Braak H, Alafuzoff I, Arzberger T, Kretzschmar H, Del Tredici K. Staging of Alzheimer disease-associated neurofibrillary pathology using paraffin sections and immunocytochemistry. Acta Neuropathol. 2006 Oct 12;112(4):389–404.
- 21. Mirra SS, Heyman A, McKeel D, Sumi SM, Crain BJ, Brownlee LM, Vogel FS, Hughes JP, van Belle G, Berg L. The Consortium to Establish a Registry for Alzheimer's Disease (CERAD). Part II. Standardization of the neuropathologic assessment of Alzheimer's disease. Neurology. 1991 Apr;41(4):479–86.
- 22. Montine TJ, Phelps CH, Beach TG, Bigio EH, Cairns NJ, Dickson DW, Duyckaerts C, Frosch MP, Masliah E, Mirra SS, Nelson PT, Schneider JA, Thal DR, Trojanowski JQ, Vinters H V., Hyman BT, National Institute on Aging, Alzheimer's Association. National Institute on Aging—Alzheimer's Association guidelines for the neuropathologic assessment of Alzheimer's disease: a practical approach. Acta Neuropathol. 2012 Jan 20;123(1):1–11.
- 23. McKeith IG, Dickson DW, Lowe J, Emre M, O'Brien JT, Feldman H, Cummings J, Duda JE, Lippa C, Perry EK, Aarsland D, Arai H, Ballard CG, Boeve B, Burn DJ, Costa D, Del Ser T, Dubois B, Galasko D, Gauthier S, Goetz CG, Gomez-Tortosa E, Halliday G, Hansen LA, Hardy J, Iwatsubo T, Kalaria RN, Kaufer D, Kenny RA, Korczyn A, Kosaka K, Lee VMY, Lees A, Litvan I, Londos E, Lopez OL, Minoshima S, Mizuno Y, Molina JA, Mukaetova-Ladinska EB, Pasquier F, Perry RH, Schulz JB, Trojanowski JQ, Yamada M, Consortium on DLB. Diagnosis and management of dementia with Lewy bodies: Third report of the DLB consortium. Neurology. 2005 Dec 27;65(12):1863–72.
- 24. Mackenzie IRA, Neumann M, Baborie A, Sampathu DM, Du Plessis D, Jaros E, Perry RH, Trojanowski JQ, Mann DMA, Lee VMY. A harmonized classification system for FTLD-TDP pathology. Vol. 122, Acta Neuropathologica. 2011. p. 111–3.
- 25. Skrobot OA, Attems J, Esiri M, Hortobágyi T, Ironside JW, Kalaria RN, King A, Lammie GA, Mann D, Neal J, Ben-Shlomo Y, Kehoe PG, Love S. Vascular cognitive impairment neuropathology guidelines (VCING): the contribution of cerebrovascular pathology to cognitive impairment. Brain. 2016 Nov 2;139(11):2957–69.
- 26. Smallwood A, Oulhaj A, Joachim C, Christie S, Sloan C, Smith AD, Esiri M. Cerebral subcortical small vessel disease and its relation to cognition in elderly subjects: a pathological study in the Oxford Project to Investigate Memory and Ageing (OPTIMA) cohort. Neuropathol Appl Neurobiol. 2012 Jun;38(4):337–43.
- 27. Grinberg LT, Thal DR. Vascular pathology in the aged human brain. Acta Neuropathol. 2010 Mar 14;119(3):277–90.
- 28. Costello H, Hayes GM, Highton-Williamson E, Nurock S, Hanbury D, Francis PT. A pilot study

- of potential brain donor satisfaction and attitudes towards telephone assessment. Int J Geriatr Psychiatry. 2016;
- 29. Li H, Durbin R. Fast and accurate long-read alignment with Burrows–Wheeler transform. Bioinformatics. 2010 Mar 1;26(5):589–95.
- 30. McKenna A, Hanna M, Banks E, Sivachenko A, Cibulskis K, Kernytsky A, Garimella K, Altshuler D, Gabriel S, Daly M, DePristo MA. The Genome Analysis Toolkit: a MapReduce framework for analyzing next-generation DNA sequencing data. Genome Res. 2010 Sep 1;20(9):1297–303.
- 31. Wang K, Li M, Hakonarson H. ANNOVAR: functional annotation of genetic variants from high-throughput sequencing data. Nucleic Acids Res. 2010 Sep 1;38(16):e164–e164.
- 32. McLaren W, Gil L, Hunt SE, Riat HS, Ritchie GRS, Thormann A, Flicek P, Cunningham F. The Ensembl Variant Effect Predictor. bioRxiv. 2016;42374.
- 33. Danecek P, Auton A, Abecasis G, Albers CA, Banks E, DePristo MA, Handsaker RE, Lunter G, Marth GT, Sherry ST, McVean G, Durbin R. The variant call format and VCFtools. Bioinformatics. 2011 Aug 1;27(15):2156–8.
- 34. Cingolani P, Platts A, Wang LLL, Coon M, Nguyen T, Wang LLL, Land SJ, Lu X, Ruden DM. A program for annotating and predicting the effects of single nucleotide polymorphisms, SnpEff. Fly (Austin). 2012 Apr 27;6(2):80–92.
- 35. Thorvaldsdottir H, Robinson JT, Mesirov JP. Integrative Genomics Viewer (IGV): high-performance genomics data visualization and exploration. Brief Bioinform. 2013 Mar 1;14(2):178–92.
- 36. Cruts M, Theuns J, Van Broeckhoven C. Locus-specific mutation databases for neurodegenerative brain diseases. Hum Mutat. 2012 Sep 1;33(9):1340–4.
- 37. Purcell S. A whole-genome association toolset. 2008;
- 38. Lee S, Miropolsky L, Wu M. SKAT: SNP-set (Sequence) kernel association test. 082, UR L http://CRAN R-project org/package= SKAT. 2013;
- 39. Euesden J, Lewis C, O'Reilly P. PRSice: polygenic risk score software. Bioinformatics. 2014;
- 40. Karch CM, Ezerskiy LA, Bertelsen S, Goate AM, Albert MS, Albin RL, Apostolova LG, Arnold SE, Baldwin CT, Barber R, Barmada MM, Barnes LL, Beach TG, Beecham GW, Beekly D, Bennett DA, Bigio EH, Bird TD, Blacker D, Boeve BF, Bowen JD, Boxer A, Burke JR, Buxbaum JD, Cairns NJ, Cantwell LB, Cao C, Carlson CS, Carney RM, Carrasquillo MM, Carroll SL, Chui HC, Clark DG, Corneveaux J, Crane PK, Cribbs DH, Crocco EA, Cruchaga C, De Jager PL, DeCarli C, DeKosky ST, Demirci FY, Dick M, Dickson DW, Duara R, Ertekin-Taner N, Evans D, Faber KM, Fallon KB, Farlow MR, Ferris S, Foroud TM, Frosch MP, Galasko DR, Ganguli M. Gearing M, Geschwind DH, Ghetti B, Gilbert JR, Gilman S, Glass JD, Graff-Radford NR, Green RC, Growdon JH, Hakonarson H, Hamilton-Nelson KL, Hamilton RL, Hardy J, Harrell LE, Head E, Honig LS, Huentelman MJ, Hulette CM, Hyman BT, Jarvik GP, Jicha GA, Jin LW, Kamboh MI, Karydas A, Kauwe JSK, Kaye JA, Kim R, Koo EH, Kowall NW, Kramer JH, Kramer P, Kukull WA, LaFerla FM, Lah JJ, Larson EB, Leverenz JB, Levey AI, Li G, Lin CF, Lieberman AP, Lopez OL, Lunetta KL, Lyketsos CG, Mack WJ, Marson DC, Martin ER, Martiniuk F, Mash DC, Masliah E, McCormick WC, McCurry SM, McDavid AN, McKee AC, Mesulam M, Miller BL, Miller CA. Miller JW. Montine TJ. Morris JC. Murrell JR. Myers AJ. Nai AC. Olichney JM. Pankratz VS, Parisi JE, Peskind E, Petersen RC, Pierce A, Poon WW, Potter H, Quinn JF, Raj

- A, Rajbhandary RA, Raskind M, Reiman EM, Reisberg B, Reitz C, Ringman JM, Roberson ED, Rogaeva E, Rosen HJ, Rosenberg RN, Sano M, Saykin AJ, Schneider JA, Schneider LS, Seeley WW, Smith AG, Sonnen JA, Spina S, Stern RA, Tanzi RE, Trojanowski JQ, Troncoso JC, Tsuang DW, Valladares O, Van Deerlin VM, Van Eldik LJ, Vardarajan BN, Vinters H V., Vonsattel JP, Weintraub S, Welsh-Bohmer KA, Williamson J, Woltjer RL, Wright CB, Younkin SG, Yu CE, Yu L. Alzheimer's disease risk polymorphisms regulate gene expression in the ZCWPW1 and the CELF1 loci. PLoS One. 2016;11(2):1–22.
- 41. Allen M, Kachadoorian M, Carrasquillo MMM, Karhade A, Manly L, Burgess JD, Wang C, Serie D, Wang X, Siuda J, Zou F, Chai HS, Younkin C, Crook J, Medway C, Nguyen T, Ma L, Malphrus K, Lincoln S, Petersen RC, Graff-Radford NR, Asmann YW, Dickson DW, Younkin SG, Ertekin-Taner N. Late-onset Alzheimer disease risk variants mark brain regulatory loci. Neurol Genet. 2015;1(2):e15.
- 42. Dempster E, Burcescu I, Wigg K, Kiss E. Evidence of an association between the vasopressin V1b receptor gene (AVPR1B) and childhood-onset mood disorders. Arch Gen. 2007;
- 43. Becker-Pauly C, Pietrzik C. The Metalloprotease meprin β is an alternative β -secretase of APP. Front Mol Neurosci. 2016;
- 44. Schönherr C, Bien J, Isbert S. Generation of aggregation prone N-terminally truncated amyloid β peptides by meprin β depends on the sequence specificity at the cleavage site. Molecular. 2016;
- 45. Escott-Price V, Shoai M, Pither R, Williams J, Hardy J. Polygenic score prediction captures nearly all common genetic risk for Alzheimer's disease. Neurobiol Aging. 2017;49:214.
- 46. Cookson MR, Lockhart PJ, McLendon C, O'Farrell C, Schlossmacher M, Farrer MJ. RING finger 1 mutations in Parkin produce altered localization of the protein. Hum Mol Genet. 2003 Sep 18;12(22):2957–65.
- 47. TREM2 and Neurodegenerative Disease. N Engl J Med. 2013 Oct 17;369(16):1564–70.
- 48. Sassi C, Nalls MA, Ridge PG, Gibbs JR, Ding J, Lupton MK, Troakes C, Lunnon K, Al-Sarraj S, Brown KS, Medway C, Clement N, Lord J, Turton J, Bras J, Almeida MR, Passmore P, Craig D, Johnston J, McGuinness B, Todd S, Heun R, Kölsch H, Kehoe PG, Vardy ERLC, Hooper NM, Mann DM, Pickering-Brown S, Brown K, Lowe J, Morgan K, Smith AD, Wilcock G, Warden D, Holmes C, Holstege H, Louwersheimer E, van der Flier WM, Scheltens P, Van Swieten JC, Santana I, Oliveira C, Morgan K, Powell JF, Kauwe JS, Cruchaga C, Goate AM, Singleton AB, Guerreiro R, Hardy J. ABCA7 p.G215S as potential protective factor for Alzheimer's disease. Neurobiol Aging. 2016;46:235.e1-235.e9.
- 49. De Roeck A, Van den Bossche T, van der Zee J, Verheijen J, De Coster W, Van Dongen J, Dillen L, Baradaran-Heravi Y, Heeman B, Sanchez-Valle R, Lladó A, Nacmias B, Sorbi S, Gelpi E, Grau-Rivera O, Gómez-Tortosa E, Pastor P, Ortega-Cubero S, Pastor MA, Graff C, Thonberg H, Benussi L, Ghidoni R, Binetti G, de Mendonça A, Martins M, Borroni B, Padovani A, Almeida MR, Santana I, Diehl-Schmid J, Alexopoulos P, Clarimon J, Lleó A, Fortea J, Tsolaki M, Koutroumani M, Matěj R, Rohan Z, De Deyn P, Engelborghs S, Cras P, Van Broeckhoven C, Sleegers K, Bessi V, Bagnoli S, do Couto FS, Verdelho A, Fratiglioni L, Padovani A, Rohan Z, Razquin C, Lorenzo E, Iglesias E, Seijo-Martínez M, Rene R, Gascon J, Campdelacreu J, Blesa R. Deleterious ABCA7 mutations and transcript rescue mechanisms in early onset Alzheimer's disease. Acta Neuropathol. 2017 Apr 27;134(3):475–87.
- 50. Ross OOA, Soto-Ortolaza AAI, Heckman MMG, Aasly JOJ, Abahuni N, Annesi G, Bacon JA,

Bardien S, Bozi M, Brice A, Brighina L, Van Broeckhoven C, Carr J, Chartier-Harlin M-C, Dardiotis E, Dickson DW, Diehl NN, Elbaz A, Ferrarese C, Ferraris A, Fiske B, Gibson JM, Gibson R, Hadjigeorgiou GM, Hattori N, Ioannidis JP, Jasinska-Myga B, Jeon BS, Kim YJ, Klein C, Kruger R, Kyratzi E, Lesage S, Lin C-H, Lynch T, Maraganore DM, Mellick GD, Mutez E, Nilsson C, Opala G, Park SS, Puschmann A, Quattrone A, Sharma M, Silburn PA, Sohn YH, Stefanis L, Tadic V, Theuns J, Tomiyama H, Uitti RJ, Valente EM, van de Loo S, Vassilatis DK, Vilariño-Güell C, White LR, Wirdefeldt K, Wszolek ZK, Wu R-M, Farrer MJ. Association of LRRK2 exonic variants with susceptibility to Parkinson's disease: a case—control study. 2011 Oct;10(10):898–908.

51. Zhao Y, Ho P, Yih Y, Chen C, Lee WWL, Tan EEK. LRRK2 variant associated with Alzheimer's disease. 2011 Nov;32(11):1990–3.