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3 ***OpenSimRoot: Widening the scope and application of root architectural models***

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30 Summary

- 31 • **Research Conducted and Rationale:** *OpenSimRoot* is an open sourced, functional-
32 structural plant model and mathematical description of root growth and function. We
33 describe *OpenSimRoot* and its functionality to broaden the benefits of root modeling to the
34 plant science community.
- 35 • **Description:** *OpenSimRoot* is an extended version of *SimRoot*, established to simulate root
36 system architecture, nutrient acquisition, and plant growth. *OpenSimRoot* has a plugin,
37 modular infrastructure, coupling single plant and crop stands to soil nutrient, and water
38 transport models. It estimates the value of root traits for water and nutrient acquisition in
39 environments and plant species.
- 40 • **Key results and unique features:** The flexible *OpenSimRoot* design allows upscaling from
41 root anatomy to plant community to estimate 1) resource costs of developmental and
42 anatomical traits, 2) trait synergisms, 3) (inter species) root competition. *OpenSimRoot* can
43 model 3D images from MRI and X-ray CT of roots in soil. New modules include: 1) soil
44 water dependent water uptake and xylem flow, 2) tiller formation, 3) evapotranspiration, 4)
45 simultaneous simulation of mobile solutes, 5) mesh refinement, and 6) root growth
46 plasticity.
- 47 • **Conclusion:** *OpenSimRoot* integrates plant phenotypic data with environmental metadata to
48 support experimental designs and gain mechanistic understanding at system scales.

49

50 **Keywords:** *Root system architecture, Functional Structural Plant Model, OpenSimRoot, Root*
51 *architectural traits, Simulation, Model driven Phenotyping, Plant nutrition*

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54 Introduction

55 Functional-structural plant models combine a representation of 3D plant structure with
56 physiological functions to advance plant science and its applications (Vos *et al.*, 2010; Dunbabin *et al.*, 2013). Those that incorporate below-ground root parameters (Dunbabin *et al.*, 2002; Pagès *et al.*, 2004; Wu *et al.*, 2007; Pierret *et al.*, 2007; Javaux *et al.*, 2008; Leitner *et al.*, 2010; Lobet *et al.*, 2014; Gérard *et al.*, 2017), require significant time and expertise in biological, mathematical, computational and digital image analyses, and therefore their development benefits greatly from an open and global setting. *SimRoot* is one of the most feature-rich and highly cited functional-structural root architectural models. However the last full description dates back twenty years (Lynch *et al.*, 1997), and subsequent papers report application of the model, with successive changes embedded in methods sections (Postma & Lynch, 2011a,b; Dathe *et al.*, 2013). Here we describe fully a new, open source version, branded *OpenSimRoot*, that is freely available for download (<http://rootmodels.gitlab.io/OpenSimRoot>). New features in this version allow simulation of more growth scenarios and crops, and its application has been widened to support emerging root phenotyping technologies.

69 *SimRoot* was originally designed to reconstruct root system architecture (RSA, see Table 1) from empirical data such as growth rates, angles and branching frequencies of different root classes. A post-simulation analysis of root geometry, nutrient uptake, and carbon costs enabled comparison of different RSAs with respect to their efficiency in taking up phosphorus relative to carbon costs (Nielsen *et al.*, 1994; Lynch & Beebe, 1995; Nielsen *et al.*, 1997; Lynch *et al.*, 1997; Ge *et al.*, 2000; Rubio *et al.*, 2001; Walk *et al.*, 2004, 2006). Later versions coupled physiological mechanisms such as root respiration, nutrient uptake, canopy photosynthesis, and RSA to simulate how the root phenotype dynamically interacts with the soil environment, and how this interaction influences acquisition of soil resources and consequently plant growth (Postma & Lynch, 2011a,b, 2012; Dathe *et al.*, 2013; Postma *et al.*, 2014a; Dathe *et al.*, 2016; York *et al.*, 2016). The initial focus was on phosphorus capture (Lynch & Beebe, 1995; Ge *et al.*, 2000; Ma *et al.*, 2001; Postma & Lynch, 2011b), which was later expanded to include C (photosynthesis), N, K, and water (Postma *et al.*, 2008; Postma & Lynch, 2011a; Dathe *et al.*, 2013). Microeconomic theory, in which resource acquisition is compared to resource investment costs, has guided the interpretation of results (Lynch, 2007; Postma *et al.*, 2014b). Although *SimRoot* was designed as a heuristic model, i.e., a tool for exploring implications of existing knowledge, and gaps in that knowledge, it proved surprisingly accurate for predicting fitness outcomes of root phenotypes (Chen *et al.*, 2011; Saengwilai *et al.*, 2014; Zhan *et al.*, 2015).

87 *SimRoot* is one of several root models that have been developed. Dunbabin et al. (2013) presents an
88 exhaustive review of all root models to date and their capabilities. To our knowledge *OpenSimRoot*
89 is currently the only plant root model that is openly version controlled (GIT) and GPLv3 licensed,
90 allowing community-driven development. We envision that *OpenSimRoot* will be used and
91 expanded by both modelers and non-modelers to simulate RSA and nutrient and water uptake in an
92 ever widening scenario for species, environments and crop management practices to advance root-
93 based opportunities to increase resource-efficient agricultural productivity. A design goal of
94 *OpenSimRoot* is a flexible model structure that can be controlled by the user rather than the
95 programmer. This means that, through a plugin infrastructure, the user can directly vary components
96 of the model and compare the results. Model behavior can further be studied through sensitivity
97 analysis, which has been a major focus in past publications.

98 In this paper we initially provide a short description of the design of the *OpenSimRoot* model and
99 definitions, then present the major submodels in *OpenSimRoot* which simulate RSA, the shoot,
100 carbon, water and nutrient acquisition and utilization, root growth plasticity, and geometric
101 descriptors. After this model description we discuss model implementation, which is designed for
102 flexibility, extensibility, transparency and robust numerics. We conclude with several examples of
103 *OpenSimRoot* usage.

104 **Model description**

105 *OpenSimRoot* has, compared to other root models, a unique design which centers on coupling
106 various mini-models (For definitions see Table 1). The distinction between parameter and algorithm
107 has been, in line with object oriented programming, removed by encapsulating both within classes
108 which share a common interface for coupling and data exchange.

109 *OpenSimRoot* design

110 *OpenSimRoot* contains a command line interface (CLI), a simulation engine, a plugin library, and
111 classes responsible for reading and writing of data (Figure 1, Notes S1,2,3&4). The simulation
112 engine implements an application programming interface (API) through which different modules
113 can request information (See Note S1). The plugin infrastructure allows developers to implement
114 new modules with limited knowledge about the rest of the code. Each plugin establishes
115 dependencies between minimodels through the API and requests data of other minimodels in order
116 to compute the necessary information. At the start of execution the import module reads an XML
117 file (see below) and, based on that file, constructs a tree of minimodels. According to the
118 specification in the XML, the minimodels load (instantiate) appropriate algorithms from a registry

119 which lists all available plugins (Note S3). The plugin infrastructure not only allows the user to
120 implement new processes, but also to implement alternative algorithms and compare model results.
121 The behavior of the modules described below is thus not fixed but can be adapted to hypotheses.
122 Simulation is driven based on data requests that originate from the users request for output. Upon
123 instantiation of the object tree, the modules that write output start requesting information in order to
124 write the output files. The CLI has a small number of options (listed with -h) with the most
125 important one the input file name. Runs are non-interactive such that many runs with different
126 parameter combinations can be fully automated on a computational cluster. This capability is
127 important when large numbers of simulations are required, for example when exploring parameter
128 sensitivity or processing real root structures (see Results) from large numbers of plants.

129 Description of the various modules

130 **Root growth and RSA.** The root system is represented by vertices and edges in *OpenSimRoot*.
131 Every root tip has its own vertex with dynamic coordinates, and all other vertices have stationary
132 coordinates that are placed behind the root tip as it extends. The final discretization of the root
133 system can be coarser than the frequency of each growth point's directional change. A fine scale
134 discretization request can automatically reduce the integration time step. In the case of a coarse
135 discretization, the length of a root segment is not the linear distance between two vertices, but the
136 true distance that the root grew, based on growth rate at that given time. We thus "simplify" the
137 growth trajectory for computational reasons, without losing the true root length.

138 To grow a root system, we need: 1) when and where root tips (primordia) are created, 2) how fast
139 root tips grow, and 3) in what direction. To start, we assume that, at a minimum, one primary root
140 and a hypocotyl are present in the seed embryo. The term hypocotyl is used here freely to include
141 any shoot axiles (stems) that are the origins of adventitious roots, whether simulating dicotyledons
142 or monocotyledons. Branch roots and their own branch roots (classed according to order), are
143 assumed to emerge from the primary root, based on rules that control the timing and placing of the
144 branches. Adventitious roots (crown or nodal roots in grasses) can branch from the stem according
145 to different schemes; the simplest defined by a starting time and position of a single whorl of roots.
146 Formation of branch roots from these axiles is typically based on branching frequencies, which can
147 be expressed in time, or space, or both, where the missing information is computed based on the
148 growth rate of the parent root. Roots can branch from either phloem or xylem poles, depending on
149 species (Casimiro *et al.*, 2003). The number of poles determines number of positions of the radial
150 branching angle, while the axial branching angle (angle between parent root and branch) is given in
151 the parameter section (for detailed explanation see Lynch, 1997, and Figure 2).

152 Elongation rates of individual roots are predefined in the parameter space, but might be scaled
153 according to a “root vigor” scaling factor; for example drawn from a lognormal distribution of
154 elongation rates (scaled to unity), thus creating variation in length. The vigor factor can also scale
155 the root specific root diameter, to allow an allometric relation between elongation rate and root
156 diameter expansion within a root class (Pagès, 2000; Wu *et al.*, 2016). Initial root diameter is
157 otherwise a root class specific input parameter.

158 While initial growth direction is set by specified radial and axial branching angles (Figure 2), the
159 direction can be changed with a tropism vector. The tropism vector is the sum of several vectors
160 representing gravitropism, random impedance, and nutrient tropisms and is added to the normalized
161 growth direction vector, to obtain the new direction.

162 Once the root is growing, its branching rules allow it to branch off new roots of different classes and
163 the whole process is repeated. Although *OpenSimRoot* currently does not simulate shoot
164 architecture, a simple tiller model is included. Tillers can form their own leaf area, and their own
165 root systems. In grasses, tillers produce nodal roots which can form a significant fraction of the total
166 root system, depending on species and environment (Atkinson *et al.*, 2014; Sebastian *et al.*, 2016).
167 Tiller formation is done on the basis of a table that indicates the time dependent delay till the next
168 tiller is formed. Dicotyledonous roots have secondary growth from cambium which thickens the stele
169 and periderm in the root. Secondary growth is simulated using a time dependent radial growth rate,
170 scaled to distance along the root.

171 **Simulation of shoot growth and related processes.** A simple shoot model can be constructed with
172 *OpenSimRoot* plugins. The shoot model is non-geometric and represents the shoot by the state
173 variables leaf area and leaf and stem dry weight. Increase in dry weights is based on carbon
174 allocated to leaves and stems, multiplied by a dry weight to carbon factor. Increase in leaf area is the
175 increase in leaf dry weight multiplied by the specific leaf area (SLA). Carbon partitioning can be
176 based on predefined time dependent values (van Ittersum *et al.*, 2003). Carbon partitioning tables
177 are typically established from dry weight measurements, and thus instead of entering carbon
178 partitioning tables, *OpenSimRoot* can also compute partitioning directly from dry weight
179 measurements. This predefined growth represents “potential” growth under a well-watered and
180 fertilized condition, whereas nutrient or carbon limitations may alter carbon partitioning (see
181 below). Total carbon available for plant growth is computed by subtracting the carbon costs (for
182 example respiration, and root exudates) from the total carbon fixed in the leaves, and or available
183 from seed or non-structural carbon reserves. Carbon costs depend on rates of respiration or carbon
184 expenditure on exudates or nitrate uptake, and these are integrated over the whole plant or root
185 system. Total carbon fixation is based on a radiation use efficiency (RUE) model, whereby

186 intercepted light is converted linearly to carbon fixation. Intercepted light is computed from leaf
187 area index, assuming that the simulated plant is in a homogeneous canopy of equally spaced and
188 identical plants. Tillers are simulated as new plants with their own leaf area, but sharing resources.

189 **Carbon allocation to roots.** The root growth module can compute the carbon for growth for each
190 root segment (edge) using its volumetric increase, and a specific root volume (g cm^{-3}). Volume
191 increases arise from primary and/or secondary growth, and root segments are assumed cylindrical
192 or, in the case of varying diameters, a truncated cone. *OpenSimRoot* compares available to required
193 carbon, and if source strength is greater than sink strength, stores the carbon left over into a labile
194 pool. *OpenSimRoot* thus considers that plant growth may be physiologically, not resource,
195 constrained (Postma *et al.*, 2014b). The labile pool is depleted when sink strength (defined by
196 carbon needed for potential growth) is greater than source strength. Once stored carbon is depleted,
197 growth rates decline. Various rules for carbon allocation under source limiting conditions have been
198 implemented. The most used rule to date prioritizes shoot over roots, and within the root system,
199 secondary growth (root cambial thickening) over elongation, and within the root classes, elongation
200 of major bearing roots over branch roots. Consequently, when plant growth is carbon limited,
201 growth rates of branch roots are reduced more than the growth of the parent roots. These rules do
202 not have a physiological basis, rather a pragmatic basis in which source sink imbalances are seen as
203 errors in the parameterization and estimation of the growth rates, and the assumption is that these
204 errors are more likely in the branch root growth than in the shoot growth. However, other rules,
205 such as equal scaling of all organs have been implemented, and can be used if the user assumes that
206 all sinks compete equally for the available carbon.

207 Although the inputs of the model are absolute growth rates, allometric scaling, based on the ratio
208 between actual and potential leaf area (not mass), can reduce the attainable growth rate of the
209 canopy and the rate of formation of new root branches. This implies that plants can never fully
210 recover from a stress. However, a recovery rate can be defined which allows the plant to grow, for
211 example, 10% faster when resources permit. Allometric scaling can also be used for the formation
212 of branches. For example, the number of nodal roots per whorl in maize is dependent on the size of
213 the shoot.

214 **Hydrology.** *OpenSimRoot* includes a hydrology module (Figure 3). The implementation of the
215 hydrology module involves the coupling of three models that simulate the movement of water
216 through soil, plant and into the atmosphere. *OpenSimRoot* includes a simplified C++
217 implementation of the SWMS model which is used to simulate soil water transport in Hydrus
218 (Diamantopoulos *et al.*, 2013) and RSWMS (Šimunek *et al.*, 1995). Water transport through the
219 xylem is simulated using a hydraulic network model (Alm *et al.*, 1992; Doussan *et al.*, 1998) and

220 evapotranspiration is simulated using the Penman-Monteith equation (Penman, 1948; Monteith,
221 1964). Small adjustments of these models, to achieve good coupling, are described in Note S5.

222 The hydrology module provides 3D water uptake profiles, drives convective nutrient transport, and
223 can simulate compensatory water uptake and hydraulic redistribution, which may occur when the
224 top soil dries out, causing nutrient uptake from dry soil domains to be reduced. It currently does not
225 simulate drought related growth responses.

226 **Nutrients.** *OpenSimRoot* has a nutrient module to simulate the simultaneous uptake of solutes,
227 originally implemented to simulate the impact of RSA on nutrient uptake, and to test tradeoffs for
228 acquisition of nutrients (Postma & Lynch, 2011a; Dathe *et al.*, 2013). Postma *et al.* (2014a) showed
229 how the optimal branching density in maize depends on the relative availability of phosphorus and
230 nitrogen. The module involves three parts: 1) simulation of plant nutrient requirements, 2)
231 simulation of nutrient acquisition, and 3) stressors which define how suboptimal plant nutrient
232 concentrations affect physiology or growth (Figure 4). Nutrients are simulated independently of
233 each other, except that in step (3) the impact of suboptimal nutrient concentrations on a given state
234 variable is aggregated using a maximum or averaging function. For example, nitrogen might affect
235 photosynthesis more than phosphorus, but phosphorus might affect the leaf area expansion rate
236 more strongly (see Dathe *et al.*, 2013).

237 The nutrient requirements of the plant are determined by integrating over the whole plant biomass
238 predefined optimal and minimal nutrient concentrations. The plant acquires nutrients through seed
239 reserves, uptake by the root system, and optional nitrogen fixation. Uptake of nutrients by the root
240 system is simulated by Michaelis-Menten kinetics, where movement of nutrients in the soil towards
241 roots is simulated through convection-dispersion-diffusion equations. *OpenSimRoot* includes two
242 different implementations for solving these equations: 1) The Barber-Cushman model (Itoh &
243 Barber, 1983), which simulates depletion zones around individual root segments at high resolution,
244 and is suitable for immobile nutrients like phosphorus; and 2) a reimplement of the solute
245 model included in SWMS3D (Šimunek *et al.*, 1995), which couples to the soil water model within
246 the hydrology module (above), simulates the whole soil domain and is suitable for mobile nutrients
247 like nitrate. More detailed descriptions of these models are given in Note S5.

248 When acquisition falls short of what is required, plant stress is assumed. Stress impact functions can
249 be defined for components such as leaf expansion rate, photosynthesis rates, respiration rates, and
250 root elongation rates or secondary growth. By making the initial response of the shoot stronger than
251 that of the roots, the plant decreases shoot to root ratios when nutrient deficient (Postma & Lynch,
252 2011a). *OpenSimRoot* will move towards a functional equilibrium, although, due to the inherent
253 slow nature of growth, and the relatively fast dynamics of other processes, this functional

254 equilibrium might not be reached (Postma & Lynch, 2011b; Postma *et al.*, 2014b). The current
255 implementation assumes that, internally, reallocation of nutrients is fast and perfect, such that all
256 organs experience equal stress. This might be true for a nutrient like nitrogen, which typically
257 causes chlorosis everywhere in the shoot, but might not be correct for other nutrients. The
258 importance of simulation of nutrient redistribution in the plant still needs study, and would require
259 implementation of a shoot architectural model in which the age and position of individual leaves or
260 canopy strata are simulated.

261 **Mineralization and rhizosphere processes.** *OpenSimRoot* implements the Yang and Janssen
262 model for mineralization (Yang & Janssen, 2000). This model assumes exponential decline of a
263 carbon pool, via aging and decline in break down rate. Based on C/N ratios of the substrate and C/N
264 ratios of the microbial biomass, the net mineralization or immobilization of N can be computed.
265 *OpenSimRoot* assumes that ammonium is readily converted to nitrate, and soil water content and
266 temperature are currently ignored. The implementation of the Yang and Janssen model in
267 *OpenSimRoot* simulates mineralization for every FEM node independently and thus mineralization
268 rates may vary in space. The user can define a nitrogen fixation rate as a percentage of the nitrogen
269 requirements of the plant. Fixation will not directly reduce nitrogen uptake from soil, but improves
270 plant nitrogen status.

271 Root exudation is not explicitly simulated, but is instead described as a root class- and time-
272 dependent carbon cost. Furthermore, exudation may increase the soluble nutrient concentration in
273 the soil at the cost of the insoluble fraction and thereby increase nutrient availability locally
274 (Barber-Cushman model only).

275 **Root growth plasticity.** *OpenSimRoot* can define reaction curves to local environmental factors, to
276 simulate a localized growth behavior of roots (Figure 5), often termed “plasticity” (Bradshaw, 1965;
277 Palmer *et al.*, 2001). 3D interpolation of available environmental data is used to define values at the
278 root surface. For example, a reaction curve (norm, (Pigliucci *et al.*, 1996)) could describe how
279 gravitropism is scaled according to the local concentration of a nutrient. Similarly, branching
280 frequency or root elongation rates can be scaled according to a local soil variable. For example,
281 static fields for soil compaction can be defined in three dimensions, using lists of coordinates and
282 associated values in conjunction with a spatial interpolation algorithm. Root elongation can then be
283 defined as a function of local soil compaction.

284 Currently, only absolute values (scalars) of local environmental variables such as soil compaction or
285 nutrient concentrations can be used to simulate plasticity responses. Gradient sensing (i.e. relative
286 values or tensors) of environmental factors may be important for nutrient- or hydro-tropism, or root
287 proliferation responses into enriched patches. However, the biological mechanism for sensing

288 gradients is unclear, and currently no such mechanism has been implemented. *OpenSimRoot* does,
289 however, include a mechanism to scale the strength of the local plasticity response on the basis of
290 yet another reaction norm which might couple plasticity to whole plant status.

291 **Root length distribution and virtual coring.** *OpenSimRoot* can compute several geometric
292 metrics, specifically root length density profiles, virtual coring, root length below D90 for nitrate,
293 and overlap of depletion zones. Others, like explored soil volume, or fractal dimensions, can be
294 computed by the user on the basis of the geometric model output.

295 **Root anatomy.** Root anatomy is not simulated in 3D explicitly, but *OpenSimRoot* can represent the
296 stele diameter, thickness of the cortex, the degree of cortical senescence, the degree of root cortical
297 aerenchyma formation, and the length, diameter and density of root hairs. These anatomical traits
298 may influence processes at the root segment level, specifically nutrient content, respiration, nutrient
299 uptake and hydraulic conductivity (Fan *et al.*, 2003; Hu *et al.*, 2014).

300 Implementation

301 *OpenSimRoot* is written in C++, an object oriented programming language. *OpenSimRoot* couples
302 minimodels, which encapsulate the simulation of a single state variable. State variables are assumed
303 to be associated with time and space and always have a unit. Minimodels are implemented as single
304 C++ classes which inherit from the same base class (named SimulaBase), such that they all have the
305 same interface (API). This interface allows minimodels to connect to other minimodels and request
306 data. Minimodels might encapsulate a constant, an interpolation table, a random number generator
307 or may make use of helper functions for computation. These helper functions are of the class type
308 IntegrationBase and DerivativeBase and are registered under their specific names, such that, based
309 on the input, the correct helper function can be instantiated. Helper functions compute a variable,
310 and when associated with an integration function, can be integrated over time. The true
311 functionality of *OpenSimRoot* is thus dispatched to the helper functions. Through a plugin
312 framework, developers can add new helper functions and thus extend the functionality of the model.
313 Example code for a plugin is given in Note S4.

314 Thus, coupling of the state variables is done through a simple common interface guaranteeing that
315 minimodels are, from a programmer point of view, standalone objects. Computations are quite
316 indifferent as to how dependent variables are computed. This creates high flexibility in the input
317 files, where the state variables can be defined in a variety of ways, i.e. constant, stochastic,
318 interpolation table, or based on a plugin (Table 2, Note S6).

319 One big challenge in coupling independent (mini)models is the implementation of numerical
320 integration when different models have different time steps, and when implicit coupling is desired.
321 In *OpenSimRoot* we implemented a general framework for predictor corrected methods, by default
322 RungeKutta4, with three components: 1) Interpolation, 2) Prediction, and 3) Dependency tracking.
323 Each minimodel keeps a time table to interpolate between time steps and return historical
324 information. Different minimodels can run at different time steps, which are however synchronized
325 at every globally defined maximum time step. Since all data requests loop through the SimulaBase
326 API, *OpenSimRoot* tracks forward dependencies and predictions, to determine whether to keep the
327 step taken. Interdependent minimodels (For an exemplar graph of dependencies see Note S7) update
328 using a predictor corrector method with interpolation to ensure compatibility of time steps. Whilst
329 the precise order may have some influence on numerical accuracy or efficiency, there is typically no
330 rational basis on which to prefer any one order of evaluation and is therefore simply dependent on
331 the order of information requests (Typically breadth-first search, see hierarchical contextualization).
332 The independent minimodel approach can create a significant computational overhead. However,
333 simulations of RSA are still relatively fast compared to soil and we regard the ease with which new
334 functionality can be added with no or little programming effort or knowledge about the rest of the
335 code as more important than runtime.

336 The current implementation of *OpenSimRoot* only depends on the standard C++ libraries (ISO C++
337 +11, and a few system libraries for the CLI), and on our website (rootmodels.gitlab.io) we provide
338 directions for compilation and running on Linux, Mac and Windows operating systems.

339 **Hierachical contextualization.** Many dynamic models are structured along a sequence of events;
340 the 'time loop'. However, *OpenSimRoot* represents the plant as a hierarchy of interacting
341 components to allow the main purpose of understanding of the function of root traits for the whole
342 plant. Minimodels are placed in a simple hierarchy which provides them context, while the object
343 oriented paradigm “hides” the internal workings of each component.

344 **Dynamic adding of components.** *OpenSimRoot* adds (instantiates) new components during
345 simulation to represent newly grown roots. This contrasts with crop models that represent plant
346 growth by an increase in values of the state variables. Dynamic memory management, connected to
347 an object oriented programming paradigm, is a useful programming feature for adding new
348 components (Dingkuhn *et al.*, 2005). Each minimodel can optionally have a class (inherited from
349 the class ObjectGeneratorBase) attached to it, which, when the children of the minimodel are
350 requested, is run to update the list of children. For example there are classes that will create new
351 branch roots or will insert new vertices (rootNodes) into the hierarchy. Most of these classes do this

352 by copying templates, which contain all the necessary minimodels that are defined in the input files.
353 An example of an ObjectGenerator plugin is given in Note S4.

354 Input files

355 *OpenSimRoot* uses a hierarchical file of parameter values, which not only contains parameter
356 values, but all state variables, and their metadata, such as names and units. Hierarchy provides
357 context, such that parameter lists can be specific for different root classes of different plant species.
358 Input files are implemented in XML, a general language for describing data together with metadata
359 that is also hierarchical, flexible, allows comments, is supported by many software tools, and can be
360 rendered in a browser as a more readable document. Note S6 gives an example of an input file that
361 simulates a simple relative growth model.

362 *OpenSimRoot* allows the user not only to enter initial values, but arrays of initial time series. This
363 way, part of the RSA can be predefined, based on measurements (also see examples in Results).
364 This approach may be different from most models, but creates the opportunity to use the model as
365 an extension to phenotyping as partial information derived from phenotypic measurements can be
366 directly entered into the input files (Fiorani & Schurr, 2013, Figure 6). Parameterizations exist for
367 maize, squash, bean, lupin, *Arabidopsis*, and barley, and are now being developed for wheat and
368 rice (Ma *et al.*, 2001; Chen *et al.*, 2011; Postma & Lynch, 2012). Input files for maize and bean, a
369 predefined root system, a small crop model and other testing scenarios are included in the source
370 code repository (<https://gitlab.com/rootmodels/OpenSimRoot>).

371 Output files

372 *OpenSimRoot* includes export modules that can be enabled or disabled to retrieve specified output
373 forms that include tables in text files, 3D models in various VTK (visual tool kit, www.vtk.org)
374 formats, 3D raster images, and a XML formatted dump of the model in the format of
375 *OpenSimRoot*'s own input files. For example: tables can be further processed with statistical
376 software (like R), VTK files can be opened with 3D data viewers (e.g. Paraview,
377 <http://www.paraview.org/>), and the model dump can be viewed in a web browser (Note S6).

378 License

379 *OpenSimRoot* is available under the GPLv3 Licence (<https://www.gnu.org/licenses/gpl-3.0.en.html>)
380 which is an opensource – copyleft license. The license enables the practice of “good science” by

381 making the model transparent and by facilitating contributions from a wider range of expertise in
382 the community. Access the version controlled code at <https://gitlab.com/rootmodels/OpenSimRoot>.

383 **Application examples for *OpenSimRoot***

384 *SimRoot* has found useful application in several domains, including 1) geometric analysis of root
385 system form and function, 2) simulation of processes that are very difficult to measure empirically,
386 3) simulation of dynamic systems, 4) sensitivity analyses, and 5) simulation of hypothetical
387 systems. In addition, a new capability of *OpenSimRoot* to read in (partially) predefined RSA enables
388 application as an extension to 3D phenotyping techniques such as X-ray CT (Computed
389 tomography) and MRI (Magnetic Resonance Imaging). Examples of all of these applications are
390 provided below.

391 **Studies on the function of RSA traits**

392 A primary output of *OpenSimRoot* is the RSA phenotype emerging from input parameters
393 simulating specific phenes like gravitropic setpoint angle or lateral root initiation interacting with
394 environmental conditions. For example, due to spatio-temporal heterogeneity in soil nutrient
395 availability, growth angles may differentially affect phosphorus and nitrogen uptake but also affect
396 the degree of inter- versus intra-plant root competition (Ge *et al.*, 2000; Rubio *et al.*, 2001; Dathe *et al.*,
397 2013). Results of simulated maize-bean-squash intercropping systems showed that RSA and
398 nitrogen fixation (bean) work towards reduced competition and increased biomass (Postma &
399 Lynch, 2012; Zhang *et al.*, 2014). Competition among branches of the same parent root may
400 become stronger when the root branching density increases, and since this increase results in greater
401 sink strength, but not greater source strength (in carbon available for growth), the individual roots
402 may stay shorter. Simulating these processes, Postma *et al.* (2014a) estimated that the optimal
403 branching density (assuming parent roots have the same root branching density) for maize was
404 lower when nitrogen availability decreased. The benefit of fewer but longer laterals in low nitrogen
405 soils was confirmed in a genotypic contrast study (Zhan *et al.*, 2015). Walk *et al.* (Walk *et al.*, 2006)
406 estimated the tradeoffs between basal root growth and adventitious root growth in bean and
407 concluded that adventitious roots might be of most benefit when phosphorus availability is low.
408 While these RSA traits represent tradeoffs, other traits may work in synergy towards greater
409 productivity on low nutrient soils (Ma *et al.*, 2001; Postma & Lynch, 2011a; Miguel *et al.*, 2015).
410 *OpenSimRoot* has also increased understanding of how integrated phenotypes function. This was
411 demonstrated by York *et al.* (2015) who used *SimRoot* to estimate how changes in maize RSA,
412 introduced by breeding over 100 years, might affect the nutrient uptake efficiency of modern

413 cultivars. New functionality described here will enable new studies of the function of whole plant
414 traits, such as tiller formation and its influence on RSA.

415 Relationships between RSA traits and root system descriptors.

416 Many researchers determine what might be called geometric descriptors of RSA: root length density
417 profiles, fractal geometry, specific root length, total root length, rooting depth and convex-hull
418 (Fitter & Stickland, 1992; Clark *et al.*, 2011). These descriptors can be computed on simulated roots
419 and their relation to architectural, anatomical or functional traits can be inferred. For example,
420 differences in the specific root length of a root system may be related to anatomical changes, or a
421 different ratio of thick to finer roots. Nielsen *et al.* (1997) determined differences in fractal
422 dimensions between phosphorus efficient and inefficient genotypes, and Walk *et al.* (2004) applied
423 *SimRoot* to show how soil exploration for P related to the fractal dimensions of the root system.
424 Miguel *et al.* (2015) applied *SimRoot* to do “virtual coring” in order to support the idea that
425 genotypic differences in rooting depth might best be seen when coring in between rows. These
426 studies show how the geometric aspects of the root system can be related to root traits and function,
427 something not easily derived from empirical measurements of actual root systems.

428 Scaling up from root anatomy to crop

429 At its smallest spatial scale, *OpenSimRoot* represents root anatomy, and at its largest scale it
430 simulates crop measures like biomass, nutrient uptake and root zone depletion and leaching. For
431 example, Ma *et al.*, (2001) focused on root hairs in *Arabidopsis thaliana* and concluded that their
432 length and density contribute synergistically towards greater phosphorus uptake. Chen *et al.*, (2011,
433 2013) used *SimRoot* and lupin phenotypic data to compute that the contribution of root hairs to total
434 phosphorus uptake might vary strongly among genotypes. Postma and Lynch (2011a,b) and
435 Schneider *et al.* (unpublished) simulated the root class- and time-dependent formation of Root
436 Cortical Aerenchyma (RCA) and Root Cortical Senescence (RCS) respectively, and determined that
437 RCA and RCS may be mechanisms underlying greater growth on low nutrient soils in maize, bean
438 and barley, possibly via efficient use and recycling of resources. Genotypic contrast studies on low
439 N soils concur with these simulation results (Saengwilai *et al.*, 2014) which suggests that
440 *OpenSimRoot* can be used for scaling up from anatomy to crop stands.

441 *OpenSimRoot* as an extension to plant phenotyping

442 Technologies like X-ray CT and MRI have been adapted to image root systems non-destructively
443 and provide non-invasive ways to phenotype whole root systems in 3D in soil (Mooney *et al.*, 2012;
444 Mairhofer *et al.*, 2013; van Dusschoten *et al.*, 2016). The utility to feed such data to a model was
445 demonstrated by Stingaciu *et al.* (2013) for a non-growing lupin root system. Using time estimates,
446 *OpenSimRoot* can simulate the growth of a root system such that the RSA is identical to that
447 imaged. Figures 6a,b (for animation see Movie S1) show an MRI image, and the simulated root
448 system. The simulation does not include a small portion (~8%) of the roots visible in the 3D image
449 data because of limitations in image segmentation, rather than in the model. *OpenSimRoot* can add
450 “MRI-non-visible” finer roots to the simulation according to existing model rules, and the
451 simulation can be extended beyond the measured time, to predict continued growth of the root
452 system. Importantly, *OpenSimRoot* modules for nutrient and water uptake can be enabled with the
453 architectural phenotypes derived from measurements and simulation, and functions can be ascribed
454 to the traits. This may help researchers and breeders go from image to functional understanding of
455 the measured root systems, and compare genotypes not only on the basis of geometry, but also on
456 the basis of modeled ability to take up water and nutrients. For example, Figures 6c,d show a CT
457 image, and corresponding *OpenSimRoot* simulation of nitrate depletion zones around the root
458 system. Integration of the model into phenotyping pipelines is also likely to help find deficits of the
459 model, and give modelers a basis for improving parameterization and/or algorithms. This important
460 development considerably widens the scope of application of *OpenSimRoot*.

461 **Discussion & Conclusions**

462 We have described the first open source version of the RSA model *SimRoot*, which is now available
463 for use by biologists and modelers. New features that expand its use include hydrology to simulate
464 and understand root system hydraulic properties. A novel area of application includes simulation of
465 non-invasive 3D phenotypic data of RSA from MRI and X-ray CT, and their putative functions in
466 nutrient and water uptake. To our knowledge, *OpenSimRoot* is currently the most feature rich and
467 widely published multiplatform RSA model (Dunbabin *et al.*, 2013) that is freely available for
468 direct download (<http://rootmodels.gitlab.io/OpenSimRoot>). The new open-source implementation
469 combines features that will enable expansion of use for plant and crop science:

- 470 • a modular, plugin infrastructure for extending the model;
- 471 • a default predictor-corrected numerical scheme for integration and coupling;
- 472 • the ability to predefine any data that was measured, where the model will use the measured
473 data instead of its algorithm for simulation (e.g. the root system, and optionally its history,
474 may be partly pre-defined based on MRI or CT images);

- 475 • integration with a shoot model;
- 476 • ability to simulate competition among plants of different species;
- 477 • maintained by an international community of root researchers.

478 Relationships in crop models that are typically only defined empirically, such as competition among
479 roots for nutrients, or root length density profiles, are actually a result of RSA, and therefore, RSA
480 models provide insight into relations between measurable traits and emerging properties at the crop
481 level. We regard the heuristic value of the model, and its use as a tool for developing and testing of
482 concepts, and prediction of mechanisms and trends, as the more important motivation for model
483 studies with, and continued development of, *OpenSimRoot*. The model may have further utility in
484 extending phenotyping pipelines by estimating genotype performance based on measured root
485 phenotypes.

486 Future development will be community driven, and may include new processes such as root
487 signaling networks, drought responses, soil microbial interactions and soil chemistry. As our
488 mechanistic understanding of different processes increases, *OpenSimRoot*'s hierarchical structure
489 allows new empirical data to be represented by new algorithms. For example, gravitropism may be
490 simulated on the basis of understanding of differential cell elongation rather than on the current
491 empirically derived input. Open sourcing allows other modelers to couple *OpenSimRoot* to their
492 models. For example shoot architectural models might be coupled to *OpenSimRoot*, in order to
493 understand competition for light and shoot architectural traits in relation to RSA traits. Finally,
494 opening up the code enables developers of other RSA models to compare the results of
495 *OpenSimRoot* to those of their models, which may lead to constructive critique and improvements
496 of all RSA models, and by extension, discoveries for improvements in understanding of plant and
497 crop resource efficiencies.

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509 **Author contributions**

510 J.A.P. and M.W. planned the manuscript. J.P.L. conceived of SimRoot and led its development
511 through 2011, J.A.P. rewrote the code, expanded its capabilities, and has led its development since
512 2011, with mathematical support from C.K. since 2013. J.A.P., C.K., N.M. and M.R.O. programmed
513 various parts of the model code. All authors were involved in open sourcing of the code and
514 forming a development team. M.J.B., N.M., M.G. and M.R.O. contributed the CT image data and
515 the simulation output based upon that data. J.A.P., C.K., M.W., J.P.L. M.R.O. and M.J.B. wrote
516 various parts of the manuscript, with input from all authors.

517 **Supplemental files**

518 Note S1: Description of the SimulaBase API

519 Note S2: How to run *OpenSimRoot*: description of the command line interface CLI.

520 Note S3: Overview of all classes that form *OpenSimRoot*, including list of plugins

521 Note S4: Example C++ code for a plugin

522 Note S5: Technical description of water and nutrient modules

523 Note S6: Example input file

524 Note S7: Example graph of state variables and their dependencies

525 Movie S1: Animation of Figure 6.

- Alm DM, Cavalier J, Nobel PS. 1992.** A finite-element model of radial and axial conductivities for individual roots: development and validation for two desert succulents. *Annals of Botany* **69**: 87–92.
- Atkinson JA, Rasmussen A, Traini R, Voß U, Sturrock C, Mooney SJ, Wells DM, Bennett MJ. 2014.** Branching Out in Roots: Uncovering Form, Function, and Regulation. *Plant Physiology* **166**: 538–550.
- Bradshaw AD. 1965.** Evolutionary significance of phenotypic plasticity in plants (EWC and JM Thoday, Ed.). *Advances in Genetics* **13**: 115–155.
- Casimiro I, Beeckman T, Graham N, Bhalerao R, Zhang H, Casero P, Sandberg G, Bennett MJ. 2003.** Dissecting Arabidopsis lateral root development. *Trends in Plant Science* **8**: 165–171.
- Chen YL, Dunbabin VM, Postma JA, Diggle AJ, Kadambot H. M. Siddique, Rengel Z. 2013.** Modelling root plasticity and response of narrow-leaved lupin to heterogeneous phosphorus supply. *Plant and Soil* **372**: 319–337.
- Chen Y, Dunbabin V, Postma J, Diggle A, Palta J, Lynch J, Siddique K, Rengel Z. 2011.** Phenotypic variability and modelling of root structure of wild *Lupinus angustifolius* genotypes. *Plant and Soil* **348**: 345–364.
- Clark RT, MacCurdy RB, Jung JK, Shaff JE, McCouch SR, Aneshansley DJ, Kochian LV. 2011.** Three-dimensional root phenotyping with a novel imaging and software platform. *Plant Physiology* **156**: 455–465.
- Dathe A, Postma JA, Lynch JP. 2013.** Modeling resource interactions under multiple edaphic stresses. In: Timlin D, Ahuja LR, eds. *Advances in Agricultural Systems Modeling. Enhancing Understanding and Quantification of Soil–Root Growth Interactions*. Madison, Wis., USA: American Society of Agronomy, Crop Science Society of America, Soil Science Society of America. 273–294.
- Dathe A, Postma JA, Postma-Blaauw MB, Lynch JP. 2016.** Impact of axial root growth angles on nitrogen acquisition in maize depends on environmental conditions. *Annals of Botany* **118**: 401–414.
- Diamantopoulos E, Iden SC, Durner W. 2013.** Modeling non-equilibrium water flow in multistep outflow and multistep flux experiments. *HYDRUS Software Applications to Subsurface Flow and Contaminant Transport Problems*: 69–76.
- Dingkuhn M, Luquet D, Quilot B, de Reffye P. 2005.** Environmental and genetic control of morphogenesis in crops: towards models simulating phenotypic plasticity. *Australian Journal of Agricultural Research* **56**: 1289–1302.
- Doussan C, Pagès L, Vercambre G. 1998.** Modelling of the hydraulic architecture of root systems: An integrated approach to water absorption - Model description. *Annals of Botany* **81**: 213–223.
- Dunbabin VM, Diggle AJ, Rengel Z, van Hugten R. 2002.** Modelling the interactions between water and nutrient uptake and root growth. *Plant and Soil* **239**: 19–38.

- Dunbabin VM, Postma JA, Schnepf A, Loïc Pagès, Mathieu Javaux, Lianhai Wu, Daniel Leitner, Ying L. Chen, Zed Rengel, Art J. Diggle. 2013.** Modelling root–soil interactions using three–dimensional models of root growth, architecture and function. *Plant and Soil* **372**: 93–124.
- van Dusschoten D, Metzner R, Kochs J, Postma JA, Pflugfelder D, Buehler J, Schurr U, Jahnke S. 2016.** Quantitative 3D analysis of plant roots growing in soil using magnetic resonance imaging. *Plant Physiology* **170**: 1176–1188.
- Fan M, Zhu J, Richards C, Brown KM, Lynch JP. 2003.** Physiological roles for aerenchyma in phosphorus-stressed roots. *Functional Plant Biology* **30**: 493–506.
- Fiorani F, Schurr U. 2013.** Future scenarios for plant phenotyping. *Annual Review of Plant Biology* **64**: 267–291.
- Fitter AH, Stickland TR. 1992.** Fractal characterization of root system architecture. *Functional Ecology* **6**: 632–635.
- Ge ZY, Rubio G, Lynch JP. 2000.** The importance of root gravitropism for inter-root competition and phosphorus acquisition efficiency: results from a geometric simulation model. *Plant and Soil* **218**: 159–171.
- Gérard F, Blitz-Frayret C, Hinsinger P, Pagès L. 2017.** Modelling the interactions between root system architecture, root functions and reactive transport processes in soil. *Plant and Soil* **413**: 161–180
- Hu B, Henry A, Brown KM, Lynch JP. 2014.** Root cortical aerenchyma inhibits radial nutrient transport in maize (*Zea mays*). *Annals of Botany* **113**: 181–189.
- Itoh S, Barber SA. 1983.** A numerical solution of whole plant nutrient uptake for soil-root systems with root hairs. *Plant and Soil* **70**: 403–413.
- van Ittersum MK, Leffelaar PA, van Keulen H, Kropff MJ, Bastiaans L, Goudriaan J. 2003.** On approaches and applications of the Wageningen crop models. *European Journal of Agronomy* **18**: 201–234.
- Javaux M, Schroeder T, Vanderborght J, Vereecken H. 2008.** Use of a three-dimensional detailed modeling approach for predicting root water uptake. *Vadose Zone Journal* **7**: 1079–1088.
- Leitner D, Klepsch S, Bodner G, Schnepf A. 2010.** A dynamic root system growth model based on L-Systems. *Plant and Soil* **332**: 177–192.
- Lobet G, Pagès L, Draye X. 2014.** A modeling approach to determine the importance of dynamic regulation of plant hydraulic conductivities on the water uptake dynamics in the soil-plant-atmosphere system. *Ecological Modelling* **290**: 65–75.
- Lynch J. 1995.** Root Architecture and Plant Productivity. *Plant Physiology* **109**: 7–13
- Lynch JP. 2007.** Rhizoeconomics: The roots of shoot growth limitations. *HortScience* **42**: 1107–1109.
- Lynch JP, Beebe SE. 1995.** Adaptation of beans (*Phaseolus vulgaris* L.) to low phosphorus availability. *HortScience* **30**: 1165–1171.

- Lynch JP, Nielsen KL, Davis RD, Jablokow AG. 1997.** *SimRoot*: Modelling and visualization of root systems. *Plant and Soil* **188**: 139–151.
- Ma Z, Walk TC, Marcus A, Lynch JP. 2001.** Morphological synergism in root hair length, density, initiation and geometry for phosphorus acquisition in *Arabidopsis thaliana*: A modeling approach. *Plant and Soil* **236**: 221–235.
- Mairhofer S, Zappala S, Tracy S, Sturrock C, Bennett MJ, Mooney SJ, Pridmore TP. 2013.** Recovering complete plant root system architectures from soil via X-ray μ -Computed Tomography. *Plant Methods* **9**: 8.
- Miguel MA, Postma JA, Lynch JP. 2015.** Phenological synergism between root hair length and basal root growth angle for phosphorus acquisition. *Plant Physiology* **167**: 1430–1439.
- Monteith JL. 1964.** Evaporation and environment. *Symposia of the society for experimental biology* **19**: 205–234.
- Mooney SJ, Pridmore TP, Helliwell J, Bennett MJ. 2012.** Developing X-ray computed tomography to non-invasively image 3-D root systems architecture in soil. *Plant and soil* **352**: 1–22.
- Nielsen KL, Lynch JP, Jablokow AG, Curtis PS. 1994.** Carbon cost of root systems: an architectural approach. *Plant and Soil* **165**: 161–169.
- Nielsen KL, Lynch JP, Weiss HN. 1997.** Fractal geometry of bean root systems: correlations between spatial and fractal dimension. *American Journal of Botany* **84**: 26–33.
- Pagès L. 2000.** How to include organ interactions in models of the root system architecture? The concept of endogenous environment. *Annals of Forest Science* **57**: 535–541.
- Pagès L, Vercambre G, Drouet JL, Lecompte F, Collet C, Le Bot J. 2004.** RootTyp: A generic model to depict and analyse the root system architecture. *Plant and Soil* **258**: 103–119.
- Palmer CM, Bush SM, Maloof JN. 2001.** Phenotypic and developmental plasticity in plants. eLS. John Wiley & Sons, Ltd.
- Penman HL. 1948.** Natural evaporation from open water, bare soil and grass. *Proceedings of the Royal Society of London. Series A. Mathematical and Physical Sciences* **193**: 120–145.
- Pierret A, Doussan C, Capowicz Y, Bastardie F, Pagès L. 2007.** Root functional architecture: A framework for modeling the interplay between roots and soil. *Vadose Zone Journal* **6**: 269–281.
- Pigliucci M, Schlichting CD, Jones CS, Schwenk K. 1996.** Developmental reaction norms: the interactions among allometry, ontogeny and plasticity. *Plant Species Biology* **11**: 69–85.
- Postma JA, Dathe A, Lynch JP. 2014a.** The optimal lateral root branching density for maize depends on nitrogen and phosphorus availability. *Plant Physiology* **166**: 590–602.
- Postma JA, Jaramillo RE, Lynch JP. 2008.** Towards modeling the function of root traits for enhancing water acquisition by crops. In: Ahuja LR, Reddy VR, Saseendran SA, Yu Q, eds. *Advances in Agricultural Systems Modeling. Response of Crops to Limited Water: Understanding and Modeling Water Stress Effects on Plant Growth Processes*. Madison, Wis., USA: ASA-CSSA-SSSA, 251–276.

- Postma JA, Lynch JP. 2011a.** Root cortical aerenchyma enhances the growth of maize on soils with suboptimal availability of nitrogen, phosphorus, and potassium. *Plant Physiology* **156**: 1190–1201.
- Postma JA, Lynch JP. 2011b.** Theoretical evidence for the functional benefit of root cortical aerenchyma in soils with low phosphorus availability. *Annals of Botany* **107**: 829–841.
- Postma JA, Lynch JP. 2012.** Complementarity in root architecture for nutrient uptake in ancient maize/bean and maize/bean/squash polycultures. *Annals of Botany* **110**: 521–534.
- Postma JA, Schurr U, Fiorani F. 2014b.** Dynamic root growth and architecture responses to limiting nutrient availability: linking physiological models and experimentation. *Biotechnology Advances* **32**: 53–65.
- Rubio G, Walk T, Ge Z, Yan X, Liao H, Lynch JP. 2001.** Root gravitropism and below-ground competition among neighbouring plants: A modelling approach. *Annals of Botany* **88**: 929–940.
- Saengwilai P, Nord E, Chimungu J, Brown K, Lynch J. 2014.** Root cortical aerenchyma enhances nitrogen acquisition from low nitrogen soils in maize (*Zea mays* L.). *Plant Physiology* **166**: 726–735.
- Sebastian J, Yee M-C, Viana WG, Rellán-Álvarez R, Feldman M, Priest HD, Trontin C, Lee T, Jiang H, Baxter I, et al. 2016.** Grasses suppress shoot-borne roots to conserve water during drought. *Proceedings of the National Academy of Sciences* **113**: 8861–8866.
- Šimuněk J, Huang K, van Genuchten MT. 1995.** *The SWMS 3D code for simulating water flow and solute transport in three-dimensional variably-saturated media*. California: U. S. Salinity laboratory, USDA.
- Stingaciu L, Schulz H, Pohlmeier A, Behnke S, Zilken H, Javaux M, Vereecken H. 2013.** In Situ Root System Architecture Extraction from Magnetic Resonance Imaging for Water Uptake Modeling. *Vadose Zone Journal* **12**: 9.
- Vos J, Evers JB, Buck-Sorlin GH, Andrieu B, Chelle M, Visser PHB de. 2010.** Functional–structural plant modelling: A new versatile tool in crop science. *Journal of Experimental Botany* **61**: 2101–2115.
- Walk TC, Jaramillo R, Lynch JP. 2006.** Architectural tradeoffs between adventitious and basal roots for phosphorus acquisition. *Plant and Soil* **279**: 347–366.
- Walk TC, vanErp E, Lynch JP. 2004.** Modelling applicability of fractal analysis to efficiency of soil exploration by roots. *Annals of Botany* **94**: 119–128.
- Wu L, McGechan MB, McRoberts N, Baddeley JA, Watson CA. 2007.** SPACSYS: Integration of a 3D root architecture component to carbon, nitrogen and water cycling--Model description. *Ecological Modelling* **200**: 343–359.
- Wu Q, Pagès L, Wu J. 2016.** Relationships between root diameter, root length and root branching along lateral roots in adult, field-grown maize. *Annals of Botany* **117**:379–390.
- Yang HS, Janssen BH. 2000.** A mono-component model of carbon mineralization with a dynamic rate constant. *European Journal of Soil Science* **51**: 517–529.

York LM, Galindo-Castañeda T, Schussler JR, Lynch JP. 2015. Evolution of US maize (*Zea mays* L.) root architectural and anatomical phenes over the past 100 years corresponds to increased tolerance of nitrogen stress. *Journal of Experimental Botany* **66**: 2347–2358.

York LM, Silberbush M, Lynch JP. 2016. Spatiotemporal variation of nitrate uptake kinetics within the maize (*Zea mays* L.) root system is associated with greater nitrate uptake and interactions with architectural phenes. *Journal of Experimental Botany* **67**: 3763–3775.

Zhan A, Schneider H, Lynch JP. 2015. Reduced lateral root branching density improves drought tolerance in maize. *Plant Physiology* **168**: 1603–1615.

Zhang C, Postma JA, York LM, Lynch JP. 2014. Root foraging elicits niche complementarity-dependent yield advantage in the ancient ‘three sisters’ (maize/bean/squash) polyculture. *Annals of Botany* **114**: 1719–1733.

Term	Definition
State variable	A quantity that has a unit and may depend on time and or space.
Minimodel	An object that encapsulates a state variable and is of a type derived from SimulaBase (Note S3). Minimodels place state variables in a context, give them a lifetime, a name, a unit and provide a general API for coupling of minimodels.
Module	A set of minimodels that form together a major component, like the carbon, nutrient or water modules.
Plugin	A class which adds functionality to the model without changing the main code (For example see Note S4). Plugins can be of derived type ObjectGenerator, DerivativeBase, or IntegrationBase
ObjectGenerator	plugin which instantiates new minimodels
DerivativeBase	Base classes for plugins that add new computational ability and/or new dependencies among minimodels
IntegrationBase	Base classes for plugins that add new integration procedures.
CLI	Command line interface, as opposed to a graphical user interface.
Root segment, root, root system, root system architecture (RSA)	Root segment is a short piece of root that can be represented by two coordinates, root is a single root axis, without branches, unless it stands in contrast to shoot, whereby it represents the whole root system (as in “root to shoot ratios”). Root system is a system of connected roots. Root system architecture is the spatio-temporal arrangement of the root system (Lynch, 1995) and is characterized by RSA traits such as branching frequencies or root gravitropism. RSA is often described by its geometric attributes, such as depth, width, specific root length, etc.

529

530 Table 1: Definition of terms.

531

532

533

Declaration of minimodel	Explanation
<code><SimulaDerivative name="rootGrowthRate" function="usePath" unit="cm/day"></code>	Declaration of a minimodel named <code>rootGrowthRate</code> which uses the plugin “ <code>usePath</code> ” to simulate a growth rate with the unit <code>cm/day</code> .
<code><SimulaConstant name="path" type="string"> rootGrowth </SimulaConstant></code>	Declaration of a minimodel named “ <code>path</code> ” which contains a string of the path to which “ <code>rootGrowthRate</code> ” needs to be coupled
<code><SimulaConstant name="multiplier"> 0.1 </SimulaConstant></code>	Declaration of a minimodel named “ <code>multiplier</code> ” which is a simple constant with which the result of minimodel named by “ <code>path</code> ” should be multiplied with.
<code></SimulaDerivative></code>	Closing of the declaration of minimodel “ <code>rootGrowthRate</code> ”, so it is clear that “ <code>path</code> ” and “ <code>multiplier</code> ” are owned by it.
<code><SimulaVariable name="rootGrowth" function="useName+Rate" integrationFunction="RungeKutta4" unit="cm" > 1. </SimulaVariable></code>	Declaration of minimodel named <code>rootGrowth</code> , which will use function “ <code>useName+Rate</code> ” to retrieve data and will integrate that data with the default integration function, <code>RungeKutta4</code> . Start value is 1.

534

535 Table 2: A simple example of how a simple relative growth rate model can be constructed with
536 *OpenSimRoot* by coupling two minimodels, one simulating the rate of growth ($rgr=0.1 \cdot \text{length}$), and
537 one that integrates that rate (analytical result would be $\text{length}=\exp(0.1 \cdot t)$). The rate calculation is
538 done using the plugin “`usePath`” which simply retrieves the length using the declared path and uses
539 the multiplier to calculate the fraction (0.1). The integration is done by the default integration
540 method, `RungeKutta4`, which integrates the result computed by the plugin “`useName+Rate`”. This
541 plugin simply retrieves the values of minimodel “`rootGrowthRate`”. If the user would like the
542 relative growth rate to be time dependent, the minimodel “`multiplier`” can be declared as an
543 interpolation table, i.e. `<SimulaTable name="multiplier" ...> 0 0.1 10 0.05 </SimulaTable>`.
544 Alternatively, stochasticity could be introduced by declaring the multiplier as of class
545 `SimulaStochastic`. This model is obviously superfluous, and most plugins will implement more
546 complex computations, with more dependencies (see also Note S6).

547

548 Figure legends

549 Figure 1: Schematic representation of the *OpenSimRoot* code. Code encompasses three major
550 components, the command line interface (CLI), different types of minimodels and a library of
551 plugins. The class hierarchy for each component is given in Note S3.

552 Figure 2: Simulated root system of bean (left) and maize (right) as rendered with ParaView. Root
553 systems are made up of different root classes, each with their own root diameter, branching rules,
554 growth direction and growth rates. Root cross-sections are not simulated but illustrate root segment
555 traits that are represented in *OpenSimRoot*.

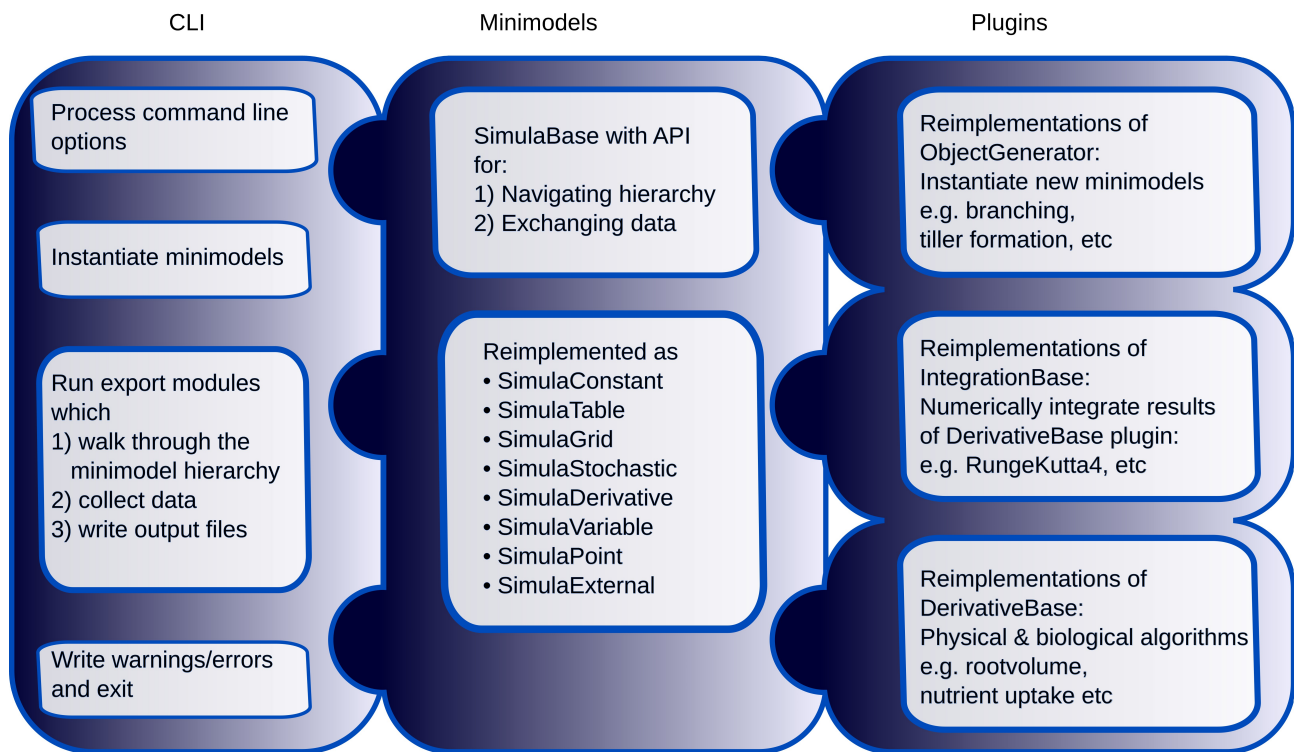
556 Figure 3: Schematic representation of the coupling of the Evapotranspiration, xylem transport and
557 soil water modules. a) Soil pedon with the hydraulic head indicated in pseudo color (left) and three
558 barley root systems (right) taking up water from that column. At the dry top water uptake is
559 negative, meaning that some hydraulic lift occurred in this scenario. b) The Penman-Monteith
560 equation for simulating transpiration and evaporation. c) Zoomed version of roots, showing the
561 edges and vertices. d) Network model for simulating water flow through the roots. e) Water
562 transport in three dimensions in the soil is simulated by solving the Richards equation, which
563 combines Darcy's law with mass conservation, using the Finite Element Method.

564 Figure 4: Schematic representation of the nutrient uptake, nutrient requirements and growth
565 regulation modules. a) Root nutrient uptake coupled to model for solute transport in the soil. b)
566 Schematic representation of the radial 1D Barber-Cushman model used for simulating P uptake. (c)
567 summary of how the ratio between nutrient requirements and nutrient uptake determines plant
568 physiology and/or growth.

569 Figure 5: Simulation results for plastic and non-plastic root systems. Root plasticity was defined as
570 increasing lateral branching density with increasing nutrient availability. Phosphorus availability
571 (left two root systems) was high in the top soil, causing branching density to be high in the top as
572 well. At the same time, the reduced branching density deeper down, due to plasticity, allows the
573 plant to grow the individual laterals longer. Pseudo colors show the local phosphorus availability.
574 Nitrate moves throughout the soil, and thereby the plasticity effect is less pronounced and difficult
575 to trace (right two root systems).

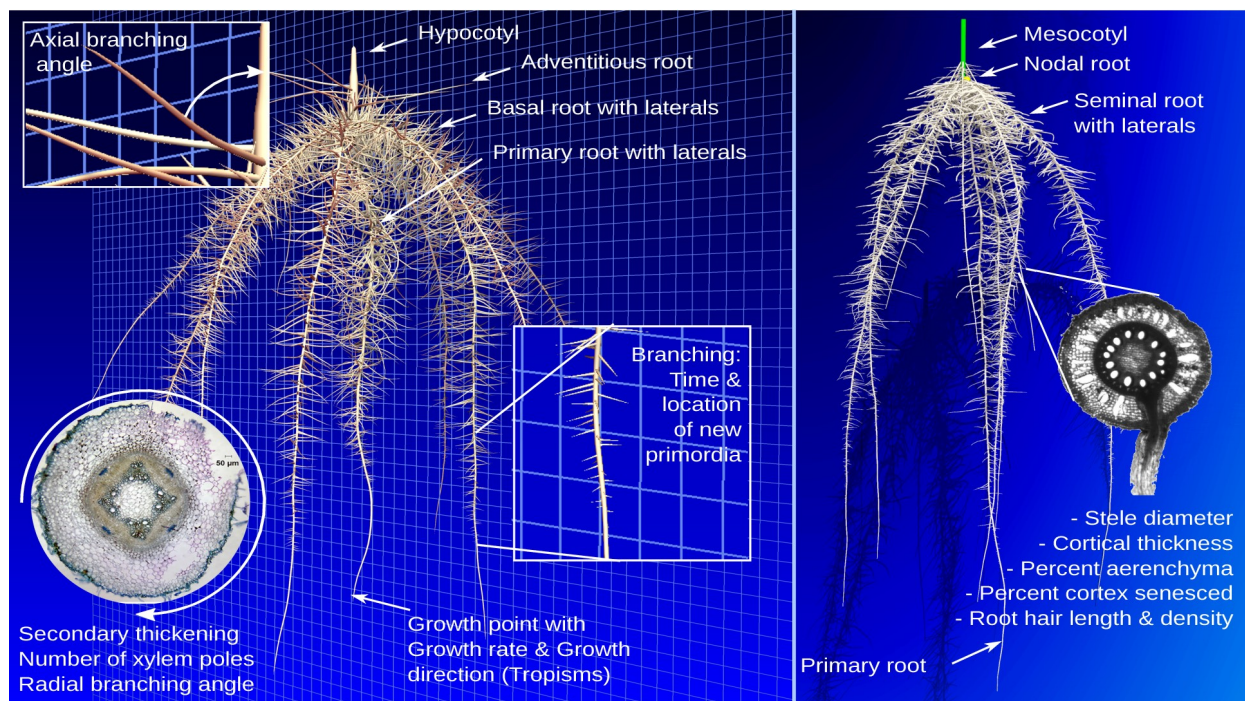
576 Figure 6: Simulation of imaged root phenotypes. a) Rendering of an MRI image of a two week old
577 maize root system and b) the simulation of that root system by *OpenSimRoot* (right). Pseudo colors
578 in 6b show the root segment age as estimated based on root topology, linear interpolation and the
579 assumption that emergence of laterals takes two days. C) Rendering of segmented X-ray CT image

580 of a 10 day old wheat root system. Soil has been sliced to make roots visible. D) *OpenSimRoot*
581 simulation of the predicted nitrate depletion zone of in C imaged root phenotype. We assumed an
582 initially homogeneous distribution of Nitrate within the simulated soil domain. Pseudo colors show
583 the nitrate concentration on a plane cut approximately through the center of the root system.



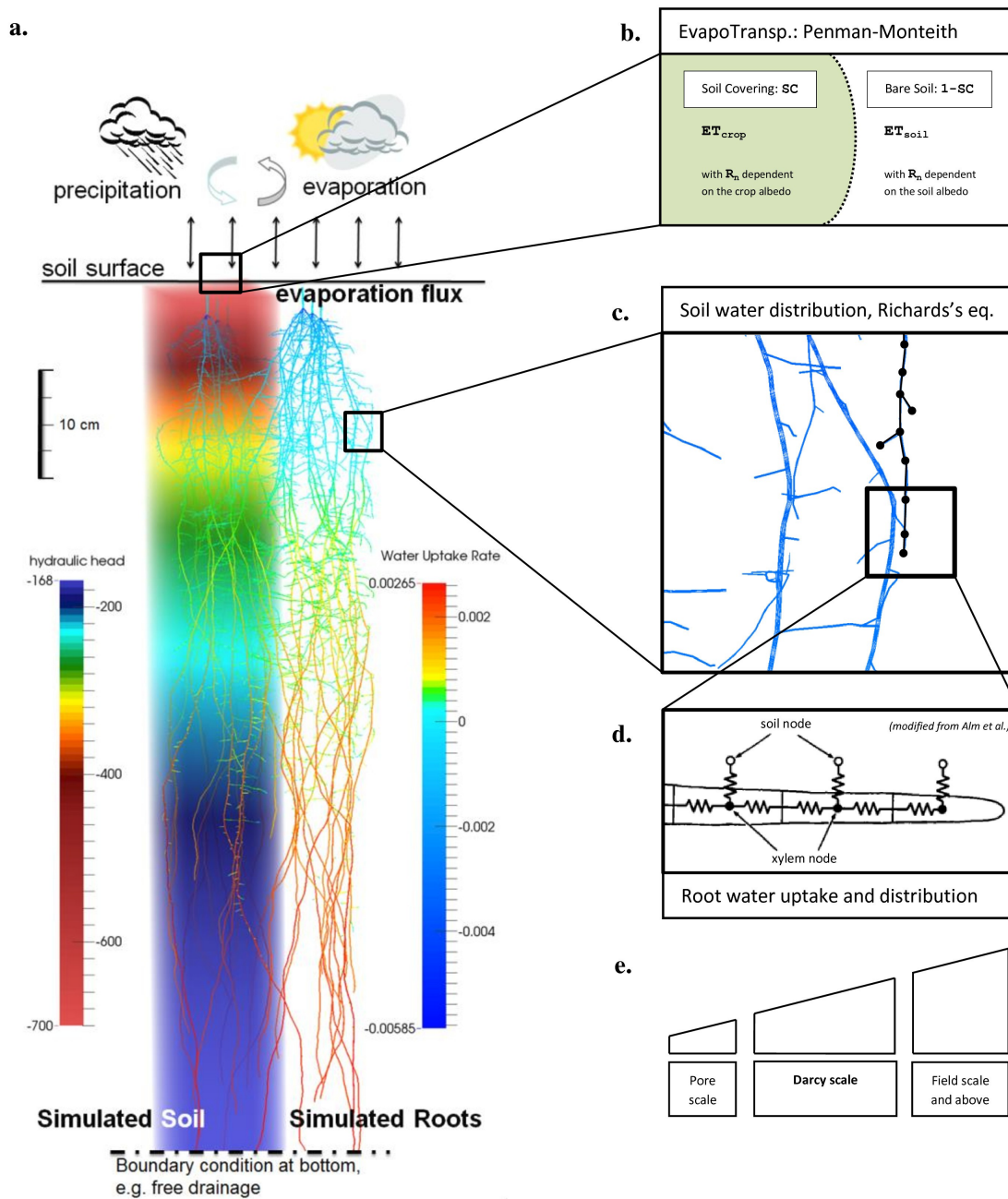
584

585 Figure 1: Schematic representation of the *OpenSimRoot* code. Code encompasses three major
 586 components, the command line interface (CLI), different types of minimodels and a library of
 587 plugins. The class hierarchy for each component is given in Note S3.



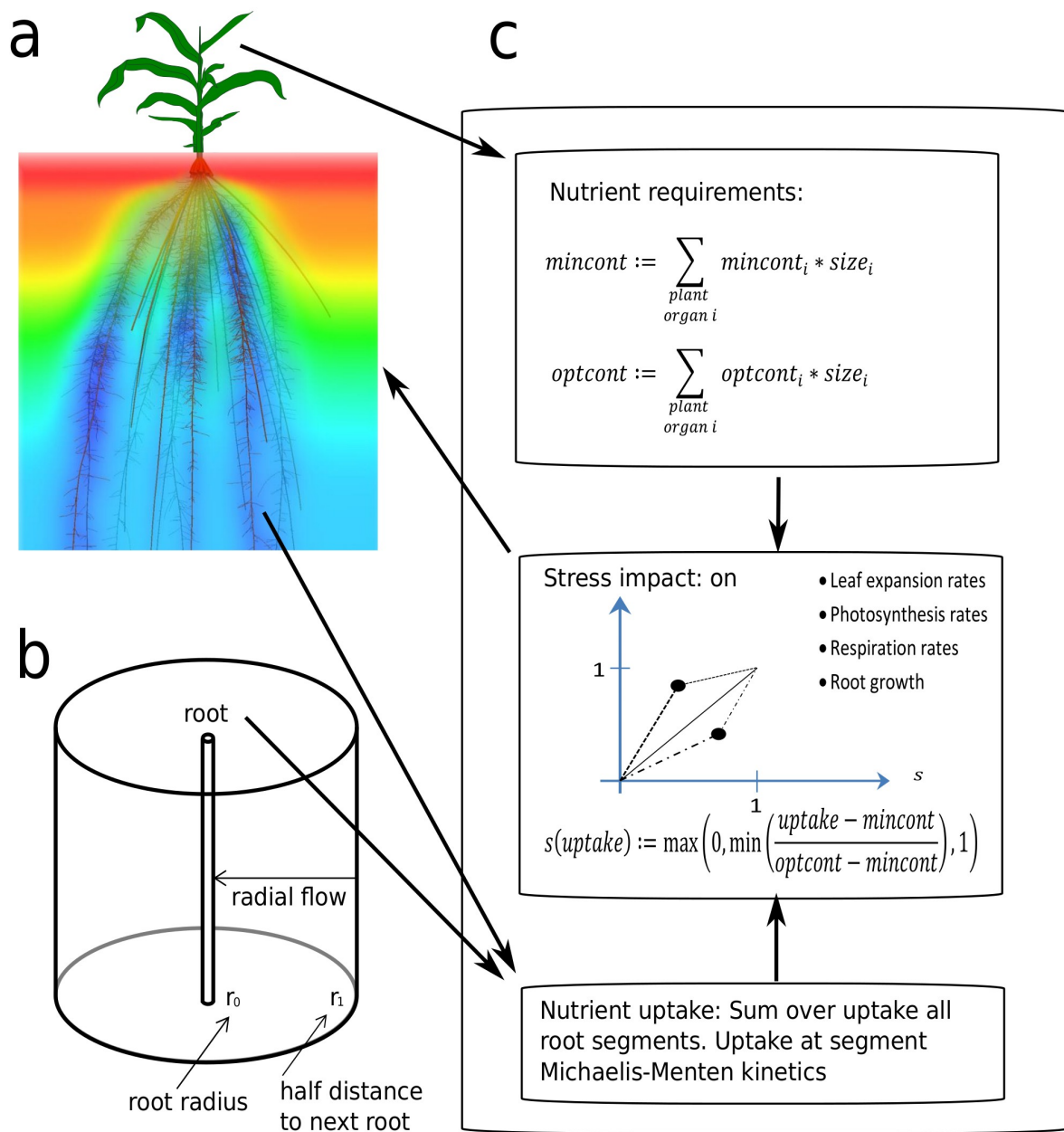
588

589 Figure 2: Simulated root system of bean (left) and maize (right) as rendered with ParaView. Root
 590 systems are made up of different root classes, each with their own root diameter, branching rules,
 591 growth direction and growth rates. Root cross-sections are not simulated but illustrate root segment
 592 traits that are represented in *OpenSimRoot*.



594

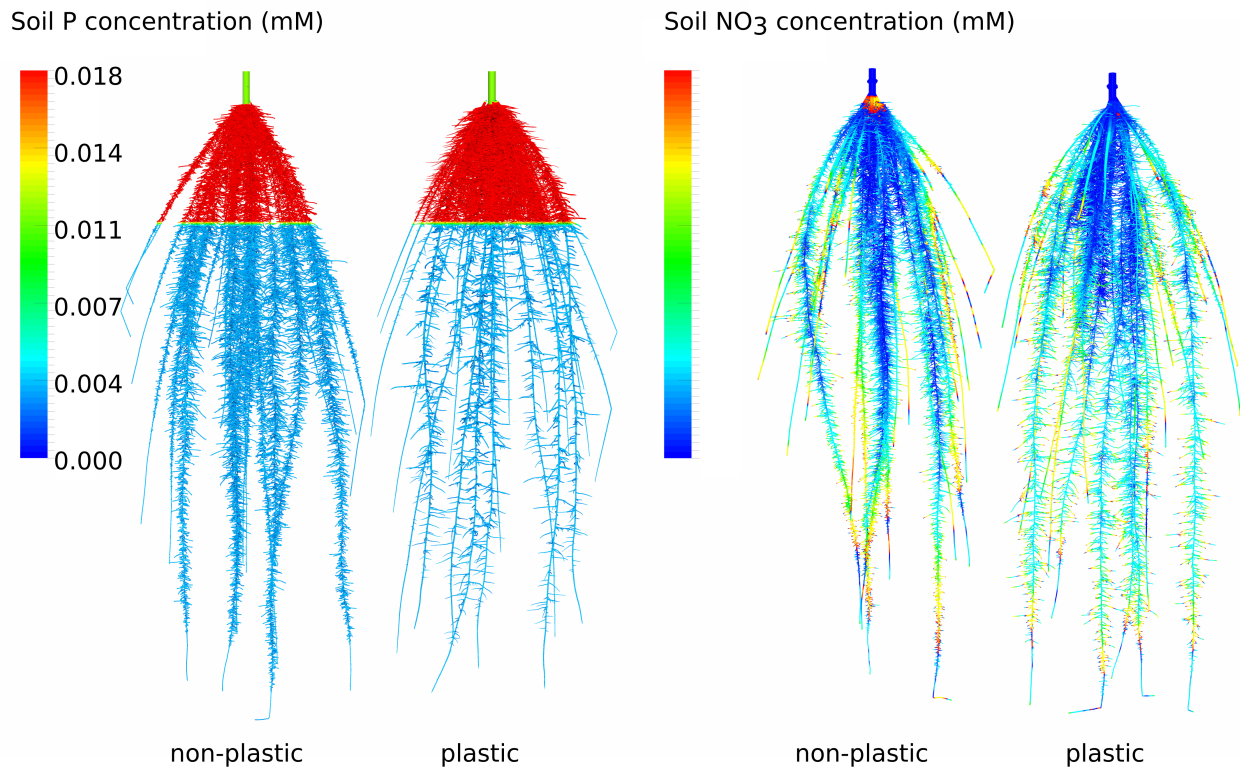
595 Figure 3: Schematic representation of the coupling of the Evapotranspiration, xylem transport and
 596 soil water modules. a) Soil pedon with the hydraulic head indicated in pseudo color (left) and three
 597 barley root systems (right) taking up water from that column. At the dry top water uptake is
 598 negative, meaning that some hydraulic lift occurred in this scenario. b) The Penman-Monteith
 599 equation for simulating transpiration and evaporation. c) Zoomed version of roots, showing the
 600 edges and vertices. d) Network model for simulating water flow through the roots. e) Water
 601 transport in three dimensions in the soil is simulated by solving the Richards equation, which
 602 combines Darcy's law with mass conservation, using the Finite Element Method.



604

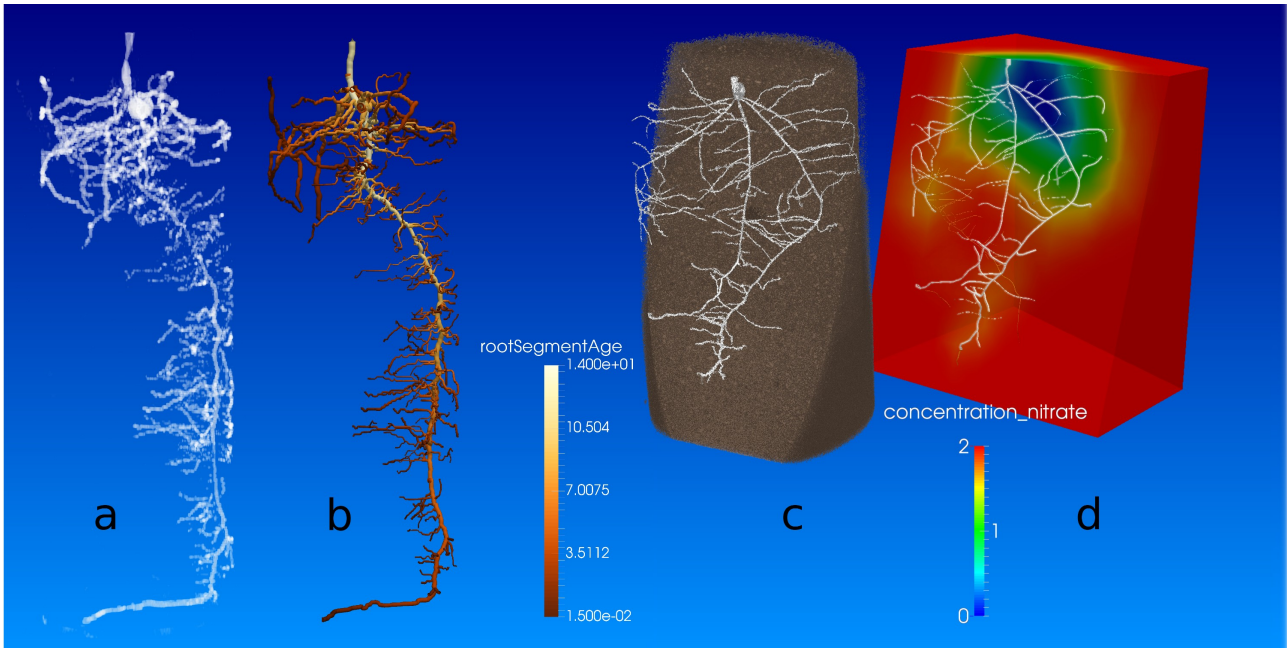
605 Figure 4: Schematic representation of the nutrient uptake, nutrient requirements and growth
 606 regulation modules. a) Root nutrient uptake coupled to model for solute transport in the soil. b)
 607 Schematic representation of the radial 1D Barber-Cushman model used for simulating P uptake. (c)
 608 summary of how the ratio between nutrient requirements and nutrient uptake determines plant
 609 physiology and/or growth.

610



611

612 Figure 5: Simulation results for plastic and non-plastic root systems. Root plasticity was defined as
 613 increasing lateral branching density with increasing nutrient availability. Phosphorus availability
 614 (left two root systems) was high in the top soil, causing branching density to be high in the top as
 615 well. At the same time, the reduced branching density deeper down, due to plasticity, allows the
 616 plant to grow the individual laterals longer. Pseudo colors show the local phosphorus availability.
 617 Nitrate moves throughout the soil, and thereby the plasticity effect is less pronounced and difficult
 618 to trace (right two root systems).



620

621 Figure 6: Simulation of imaged root phenotypes. a) Rendering of an MRI image of a two week old
 622 maize root system and b) the simulation of that root system by *OpenSimRoot* (right). Pseudo colors
 623 in 6b show the root segment age as estimated based on root topology, linear interpolation and the
 624 assumption that emergence of laterals takes two days. C) Rendering of segmented X-ray CT image
 625 of a 10 day old wheat root system. Soil has been sliced to make roots visible. D) *OpenSimRoot*
 626 simulation of the predicted nitrate depletion zone of in C imaged root phenotype. We assumed an
 627 initially homogeneous distribution of Nitrate within the simulated soil domain. Pseudo colors show
 628 the nitrate concentration on a plane cut approximately through the center of the root system.

New Phytologist Supporting Information

Article title: *OpenSimRoot*: Widening the scope and application of root architectural models

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Article acceptance date:

The following Supporting Information is available for this article:

Supplement 1 Description of the SimulaBase API

Supplement 2 How to run *OpenSimRoot*: description of CLI

Supplement 3 Example C++ code for a plugin

Supplement 4 Example C++ code for a plugin

Supplement 5 Technical description of water and nutrient modules

Supplement 6 Example input file

Supplement 7 Example graph of state variables and their dependencies

Supplemental movie 1 Animation of Figure 6b.

Supplement 1: Application programming interface (API) of the SimulaBase class

This interface is used by the plugins to navigate the hierarchy and retrieve necessary data. For an example see, Supplement 4. Developers that would like to develop a new plugin, will need this interface in order to retrieve data from other minimodels. These minimodels are in a hierarchy. The methods listed here can be used to find those minimodels in the hierarchy, and to request data from them. Minimodels are instantiations (objects) of class (type) SimulaBase.

//Method to retrieve meta data on a given minimodel such as its name, path in the hierarchy, lifetime of the object, and its units.

```
std::string getName() const; //name of object
std::string getPrettyName() const; //some what more humen readable name
std::string getPath() const; //path to the object
virtual std::string getType() const; //What type this object has
bool evaluateTime(const Time &t) const; //check if t is within lifetime
Time getEndTime() const; //get the end time of object
Time getStartTime() const; //get the start time of object
virtual Unit getUnit(); //get the unit
void checkUnit(const Unit& unit) const; //check if unit equals given unit
void setUnit(const Unit &newUnit); //change unit
virtual void getXMLtag(Tag &tag); //get the object as tag (xml output)
```

//Methods to navigate the minimodel hierarchy

The difference between the get() and existing() methods is that when the object does not exist get() will throw an error and terminate the simulation, whereas existing() will return a NULL pointer. The getPath() methods will navigate a symbolic path just as a path in a filesystem is navigated. For example getPath("../mySib") translates to getSibling("mysib"), where the later is more efficient.

```
SimulaBase* getParent() const;
SimulaBase* getParent(const unsigned int i) const;
int getNumberOfChildren() const; //does not update!
int getNumberOfChildren(const Time &t); //does update
SimulaBase* getChild(const std::string & name, const Time &t);
SimulaBase* existingChild(const std::string & name, const Time &t);
SimulaBase* getChild(const std::string & name);
SimulaBase* existingChild(const std::string & name);
SimulaBase* getChild(const std::string & name, const Unit & u);
SimulaBase* existingChild(const std::string & name, const Unit & u);
SimulaBase* getSibling(const std::string & name, const Time &t);
SimulaBase* existingSibling(const std::string & name, const Time &t);
SimulaBase* getSibling(const std::string & name);
SimulaBase* existingSibling(const std::string & name);
SimulaBase* getSibling(const std::string & name, const Unit & u);
SimulaBase* existingSibling(const std::string & name, const Unit & u);
//Sibling can be retrieved in alphabetic order.
SimulaBase* getNextSibling(const Time &t);
SimulaBase* getNextSibling() const;
```



```

SimulaBase* getPreviousSibling(const Time &t);
SimulaBase* getPreviousSibling()const;
SimulaBase* getFirstChild(const Time &t);
SimulaBase* getFirstChild()const;
SimulaBase* getLastChild()const;

SimulaBase* getPath(const std::string &name);
SimulaBase* getPath(const std::string &name, const Time &t);
SimulaBase* getPath(const std::string &name, const Unit &u);
SimulaBase* existingPath(const std::string &name);
SimulaBase* existingPath(const std::string &name, const Time &t);
SimulaBase* existingPath(const std::string &name, const Unit &u);

typedef std::vector<SimulaBase*> List;
void getAllChildren(List&, const Time &t);
void getAllChildren(List&)const;

```

//Method for walking along a root axis. Retrieves the minimodel with the same

```

name associated with the next vertex.
virtual SimulaBase* followChain(const Time &t);

```

//Methods to retrieve specific subsets of minimodels based on position

```

typedef std::multimap<Coordinate,SimulaBase*> Positions;
static void getAllPositions(const Time &t, Positions& list);
static void getAllPositions(Positions& list);
void getYSlice(const Time &, const double, const double, Positions&);
void getPositionsWithinRadius(const Time &, const Coordinate& c, const
double &r, Positions&);
void getPositionsInsideBox(const Time &, const Coordinate&, const
Coordinate &, Positions&);

```

//Methods for retrieving data

```

virtual void get(const Time &t, int &returnConstant);
virtual void get(const Time &t, std::string &returnConstant);
virtual void getRate(const Time &t, Time &var);
virtual void get(const Time &t, Coordinate &point);
virtual void get(const Time &t, MovingCoordinate &point);
virtual void getAbsolute(const Time &t, Coordinate &point);
virtual void getBase(const Time &t, Coordinate &point);
virtual void getRate(const Time &t, Coordinate &point);
virtual void getAbsolute(const Time &t, MovingCoordinate &point);
virtual void get(int &returnConstant);
virtual void get(std::string &returnConstant);
virtual void get(bool &returnConstant);
virtual void get(const Time &x, Time &y);
virtual void get(Time &x);
virtual void get(const Time &t, const Coordinate &pos, double &y);
virtual void get(const Time &t, const Coordinate &pos, Coordinate &y);
virtual void getRate(const Time &t, const Coordinate &pos, double &y);
virtual void get(Coordinate &point);

```

```
virtual void getAverageRate(const Time &t1, const Time &t2, double &var);
virtual void getAverageRate(const Time &t1, const Time &t2, Coordinate
&var);
```

```
//reverse data look up: returns time that object was nearest to given
value or position. Only works if the object is not garbage collected
```

```
virtual void getTime(const Coordinate &p, Time &t, Time tmin=-1, Time
tmax=0);
virtual void getTime(const double &p, Time &t, Time tmin=-1, Time tmax=0);
```

//Method for setting data, probably only implemented for timetables.

```
virtual void set(const double &x, const double &y);
```

//Methods to retrieve info on timestepping of a minimodel

```
virtual Time &minTimeStep();
virtual Time &maxTimeStep();
virtual Time &preferredTimeStep();
virtual Time lastTimeStep();
```

//Methods to control garbage collection, which will basically clean up the simulation history

```
virtual void collectGarbage(const Time&); //clean up history
virtual void garbageCollectionOff(); //keep history of this object always
```

//Other methods

```
void stopUpdatefunction(); //When implementing an objectgenerator signal
it has
finished creating all objects for all times, and can be deleted.
static void updateAll(const Time &); //update whole tree
void updateRecursively(const Time &); //update subtree
static void signalMeAboutNewObjects(SimulaBase* me); //if plugin has the
addObject() implemented, it will be signaled when new objects are being
instantiated by any of the object generators.
```

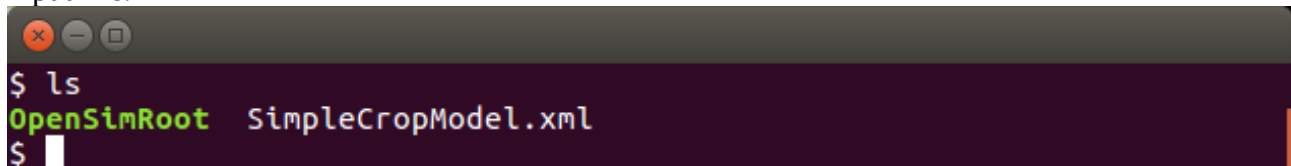

Supplement 2: Command line interface (CLI) of OpenSimRoot: How to run and use the model

OpenSimRoot has a command line interface, which means that you operate the model from a terminal using commands, not with a graphical interface and the mouse.

Step 1: Open a terminal (under windows 10 you may use the program named CMD)

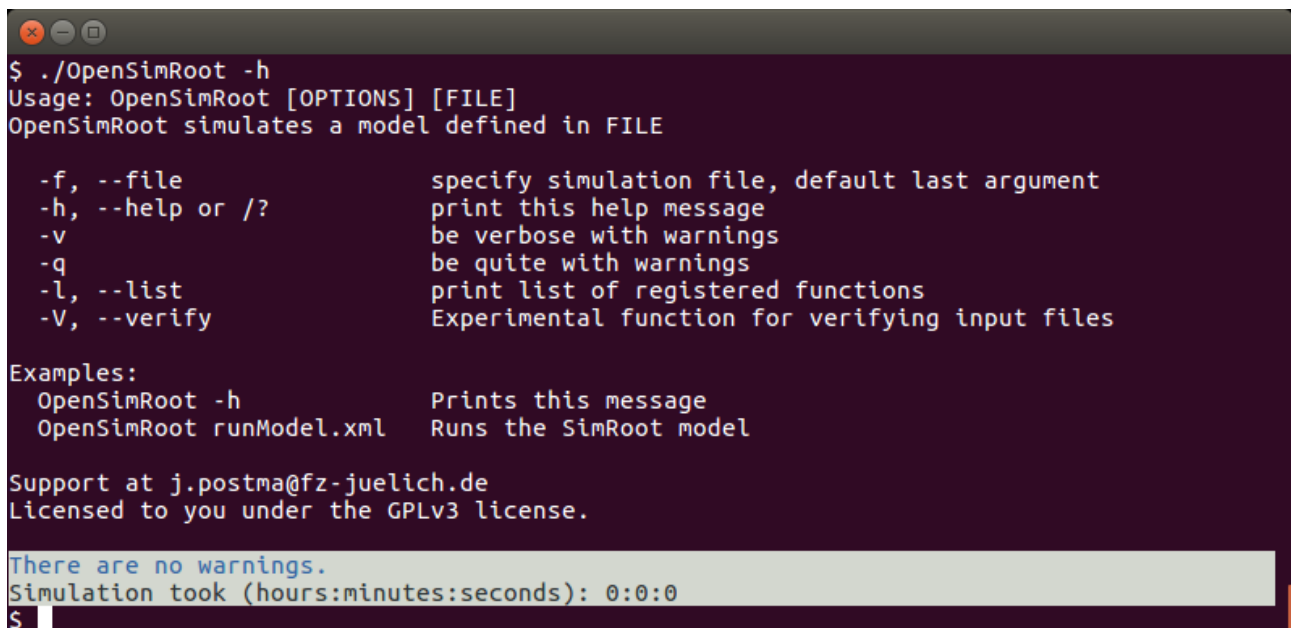
Step 2: Go to the folder where you want to run the model, use the command `cd` to navigate, for example: `cd MyRunFolder`

Step 3: We assume that the folder contains the OpenSimRoot executable. With the “`ls`” command you can list all folders (or on windows the command is “`dir`”). Here we see that my folder contains the executable OpenSimRoot (conveniently made green, as it is executable) and a XML input file.



```
$ ls
OpenSimRoot  SimpleCropModel.xml
$
```

Step 4: OpenSimRoot has a small build in help which we we can run by typing `./OpenSimRoot -h` (on windows you do not type the path “`./`” in front of the executable).



```
$ ./OpenSimRoot -h
Usage: OpenSimRoot [OPTIONS] [FILE]
OpenSimRoot simulates a model defined in FILE

  -f, --file           specify simulation file, default last argument
  -h, --help or /?    print this help message
  -v                   be verbose with warnings
  -q                   be quite with warnings
  -l, --list           print list of registered functions
  -V, --verify        Experimental function for verifying input files

Examples:
  OpenSimRoot -h      Prints this message
  OpenSimRoot runModel.xml  Runs the SimRoot model

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There are no warnings.
Simulation took (hours:minutes:seconds): 0:0:0
$
```

The help shows how to run OpenSimRoot, and gives you some options and their explanation.

Step 5: Like the help shows, running the model is done by appending the input file:
`./OpenSimRoot SimpleCropModel.xml`

```

$ ./OpenSimRoot SimpleCropModel.xml
Trying to load model from file:
Running modules: Running modules: 80.0/80.0 days. Mem 5 mB. #obj.=47 x64b/obj.=15414 OK
Finalizing output:
THE MODEL PUT OUT WARNINGS:
PhotosynthesisLintulV2: no stress impact factor found
Simulation took (hours:minutes:seconds): 0:0:0
$
$ ls
OpenSimRoot SimpleCropModel.xml tabled_output.tab warnings.txt
$

```

Again with ls (dir) you can list the files, the model created two new files, one containing warnings, one containing the simulation results.

Step 6: The results of the simulation are in the tabled_output.tab file which can be viewed with any program that opens text files. Here we simply show the first lines with the command head:

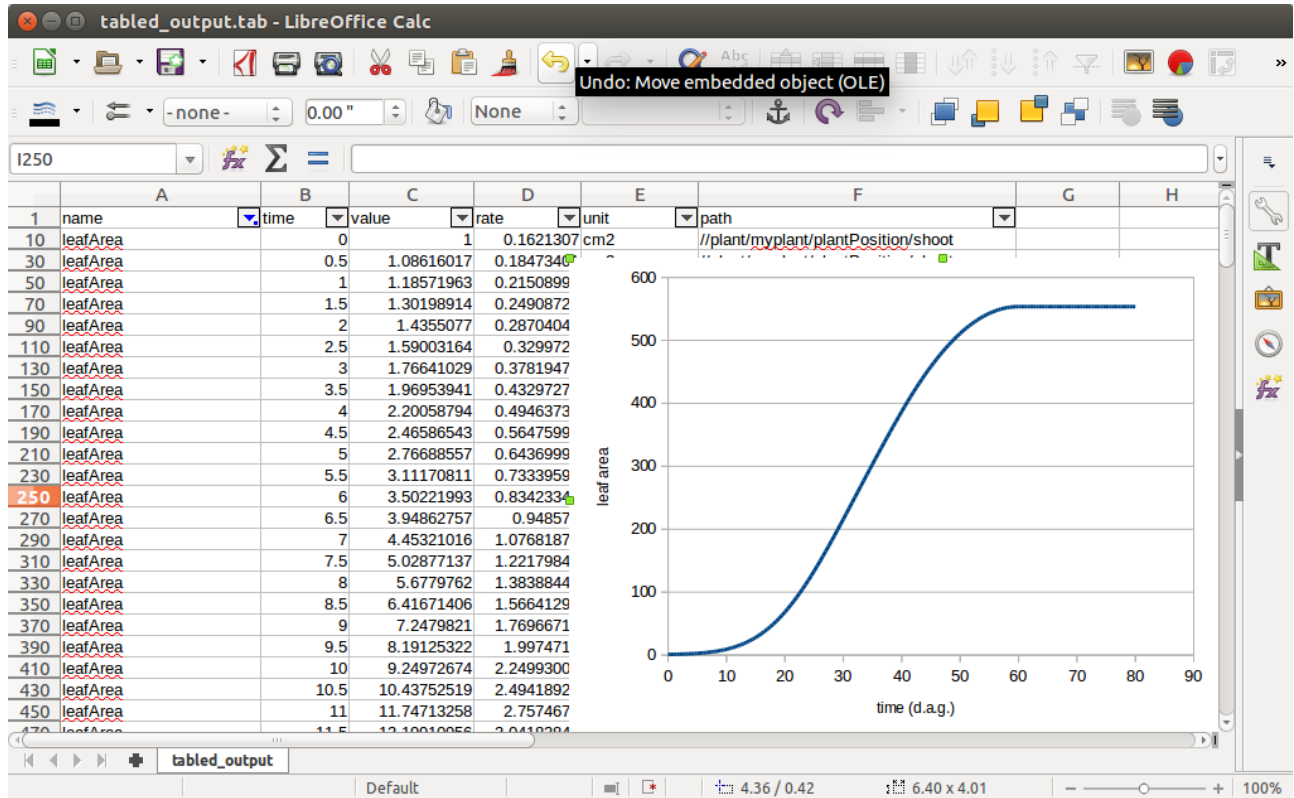
```

$ head tabled_output.tab
name                time  value  rate      unit      path
"irradiation"       0.000 3000.00000000  NA      "umol/cm2/day"  "//environment/atmosphere"
"carbonAllocation2Roots" 0.000 0.00000000  0.00029184  "g"        "//plant/myplant"
"carbonAllocation2Shoot" 0.000 0.00000000  0.00007296  "g"        "//plant/myplant"
"carbonAvailableForGrowth" 0.000 0.00000000  NA         "g"        "//plant/myplant"
"multiplier"        0.000 0.50000000  NA         "noUnit"    "//plant/myplant/carbonAvailableForGrowth"
"carbonToDryWeightRatio" 0.000 0.45000000  NA         "100%"      "//plant/myplant"
"carbonAllocation2Leaves" 0.000 0.00000000  0.00007296  "g"        "//plant/myplant/plantPosition/shoot"
"carbonAllocation2Stems" 0.000 0.00000000  0.00000000  "g"        "//plant/myplant/plantPosition/shoot"
"leafArea"          0.000 1.00000000  0.16213070  "cm2"      "//plant/myplant/plantPosition/shoot"

```

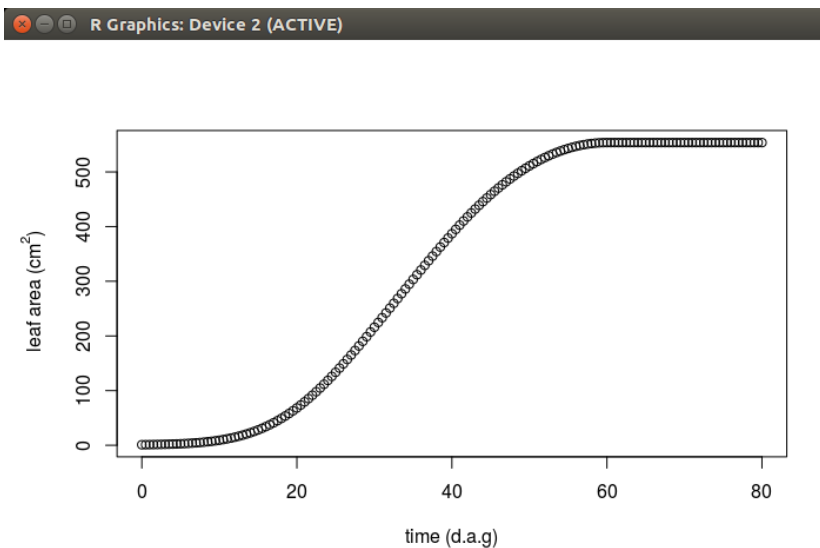
The file contains a header in the first row, and 6 columns listing the name of the state variable, the time, the value, the rate of change of that state variable (if simulated), the unit of the state variable, and the path in the hierarchy to this state variable.

Step 7: The `tabled_output.tab` file is also easily imported into a spreadsheet program. By enabling the auto filter and selecting `leafArea`, we can easily create a plot.



8. The same can be achieved in R using this script:

```
d<-read.table("tabled_output.tab",header=T)
f=d$name=="leafArea"
plot(value~time,data=d[f,],ylab=~"leaf area (cm^2)", xlab="time (d.a.g)")
```



step 9: Editing the input file can be done with any text file editor. Here I opened the file with the command nano tabled_output.tab and the result is an xml formatted file in which we can change the numbers, save and rerun. In white you see the numbers, and scrolling to the bottom you would see more.

```

GNU nano 2.5.3 File: SimpleCropModel.xml Modified
<SimulationModel>
  <SimulaBase name="plant">
    <SimulaBase name="myplant">
      <SimulaConstant name="plantType" type="string">
        mySpecies
      </SimulaConstant>
      <SimulaConstant name="plantingTime" unit="day" type="Time">
        0
      </SimulaConstant>
      <SimulaConstant name="plantPosition" type="Coordinate">
        0 -2 0
      </SimulaConstant>
      <SimulaBase name="shoot">
        <SimulaDerivative name="lightInterception" unit="umol/cm2/day">
          function="lightInterception" />
        <SimulaVariable name="photosynthesis" unit="g">
          function="photosynthesisSLintulV2" />
        <SimulaDerivative name="leafAreaIndex" unit="cm2/cm2">
          function="leafAreaIndex" />
        <SimulaVariable name="leafArea" unit="cm2" function="leafArea">
          1. </SimulaVariable>
        <SimulaVariable name="leafDryWeight" unit="g">
          function="leafDryWeight.v2" />
        <SimulaDerivative name="relativeCarbonAllocation2Leafs">
          unit="100%" function="relativeCarbonAllocation2LeafsFromInputFile" />
        <SimulaVariable name="carbonAllocation2Leafs" unit="g">
          function="carbonAllocation2Leafs" />
      </SimulaBase>
    </SimulaBase>
  </SimulaBase>
</SimulationModel>
^C Get Help      ^O Write Out    ^W Where Is     ^K Cut Text     ^J Justify     ^C Cur Pos     ^Y Prev Page   ^_ First Line
^X Exit          ^R Read File    ^A Replace      ^U Uncut Text  ^T To Spell    ^G Go To Line  ^V Next Page   ^M Last Line

```

step 10: You see several functions listed that are used to simulate a state variable. To get a list of all functions that are included in your OpenSimRoot version use the command OpenSimRoot -l. This will list all plugins that are included with OpenSimRoot.

```

$
$ ./OpenSimRoot -l
Registered object generator functions:
copyDefaults fieldPlanting generateRoot insertRootBranchContainers rootBranches rootBranchesOfTillers rootDataPoints seedling tillerDevelopment ti
llerFormation

Registered integration functions:
BackwardEuler BarberCushmanSolver Euler ForwardEuler Heuns HeunsII RungeKutta4 convergenceSolver default explicitConvergence iterativeSolver singl
eStepSolver

Registered functions:
BFMemory D95 Grass_reference evapotranspiration Interception InterceptionV2 PriestleyTaylor Radiation SimpleSoilTemperature Simunek Stanghellini
Swms3d Tall_reference Crop ThermalConductivity VolumetricHeatCapacity actualTranspiration aerodynamicResistance atrDensity atrPressure barber_cush
man_1981_nutrient_uptake barber_cushman_1981_nutrient_uptake explicit bnf.V1 carbonAllocation2Leafs carbonAllocation2Roots carbonAllocation2Shoot
carbonAllocation2Stems carbonAvailableForGrowth carbonCostOfBiologicalNitrogenFixation carbonCostOfExudates carbonCostOfNutrientUptake carbonReser
ves carbonToDryWeightRatio constantLeafGrowthRate delta_e_s_e_a_e_s getValuesFromPlantWaterUptake getValuesFromSWMS integrateOverSegment kineticPa
rameters leafArea leafAreaIndex leafAreaReductionCoefficient leafDryWeight leafDryWeight.v2 leafMinimalNutrientContent leafOptimalNutrientContent
leafPotentialCarbonSinkForGrowth leafRespirationRate lightInterception localNutrientResponse meanLeafAreaIndex meanOverAllPlantShoots meanOverAllP
lants meanOverAllPlantsNutrients michaelis_menten nutrient_uptake monteithEQ numberOfRoots numberOfTillers nutrientStressFactor nutrientStressFact
or.V2 penmanEQ photosynthesisLintul photosynthesisSLintulV2 plantCarbonBalance plantCarbonIncomeRate plantTotal plantTotalForNutrients plantTotalRa
tes plantTotalRatesRootFraction plantTotalRatesShootFraction plantTotalRootFraction plantTotalShootFraction pointSensor potentialLeafArea potentia
lLeafGrowthRate potentialRootGrowthRate potentialSecondaryGrowth potentialTranspirationCrop proximity radiusDepletionZoneBarberCushman radiusDeple
tionZoneSimRoot4 randomGravitropism randomImpedance relativeCarbonAllocation2LeafsFromInputFile relativeCarbonAllocation2RootsFromInputFile relati
veCarbonAllocation2RootsOneMinusShoot relativeCarbonAllocation2RootsPotentialGrowth relativeCarbonAllocation2RootsScaledGrowth relativeCarbonAlloc
ation2ShootFromInputFile relativeCarbonAllocation2ShootPotentialGrowth relativeCarbonAllocation2ShootScaledGrowth relativeCarbonAllocation2ShootSw
itch relativeCarbonAllocation2StemsOneMinusLeafs remainingProportion reserves reservesSinkBased rootCircumference rootClassID rootDiameter rootDia
meter.v2 rootDiameterCortex rootDryWeight rootGrowthDirection rootGrowthScalingFactor rootLength2Base rootLengthDensity rootLengthProfile rootNode
PotentialCarbonSinkForGrowth rootNutrientTotal rootPotentialCarbonSinkForGrowth rootSegmentAge rootSegmentDryWeight rootSegmentLength rootSegmentM
inimalNutrientContent rootSegmentNutrientDepletionVolume rootSegmentOptimalNutrientContent rootSegmentRespirationRate rootSegmentRootHairSurfaceAr
ea rootSegmentSpecificWeight rootSegmentSurfaceArea rootSegmentVolume rootSegmentVolumeCortex rootSegmentVolumeSteel rootSystemNutrientTotal roots
ystemTotal rootSystemTotalRates rootTotal rootTotal.v2 rootTotalRates rootsBelowD95Solute scaledRootGrowthRate scaledWaterUptake secondaryGrowth s
egmentMaxNutrientUptakeRate shootDryWeight shootMinimalNutrientContent shootOptimalNutrientContent simplePotentialTranspiration soluteMassBalanceT
est specificHeatCapacityOfAir stemDryWeight stemMinimalNutrientContent stemOptimalNutrientContent stemPotentialCarbonSinkForGrowth stemRespiration
Rate stomatalResistance stressAdjustedPotentialLeafGrowthRate stressFactor sum sumOverAllPlantShoots sumOverAllPlants sumOverAllPlantsNutrients su
mSteelCortex tropisms useDerivative useName+Rate useParameterFromParameterSection usePath useRootClassAndNutrientSpecificTable useRootClassSpecifi
cTable virtualCoring waterMassBalanceTest waterUptakeFromHopmans

There are no warnings.
Simulation took (hours:minutes:seconds): 0:0:0
$

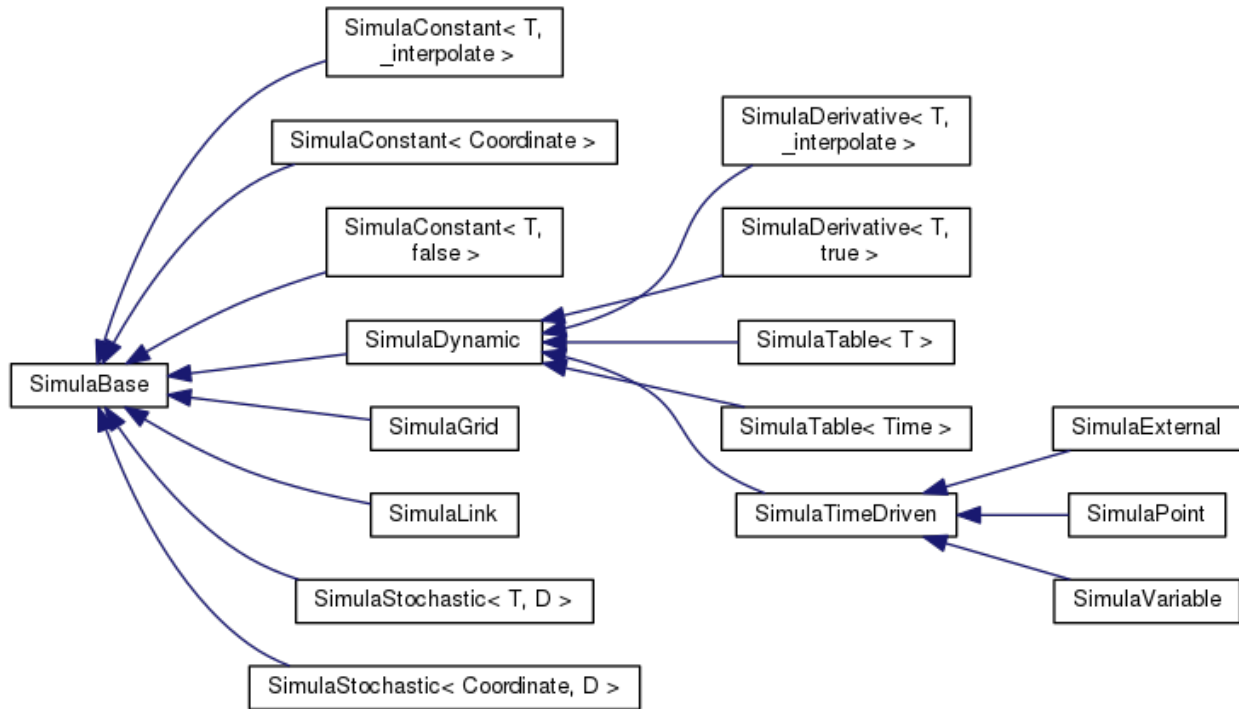
```

Supplement 3: Class hierarchy of OpenSimRoot code

This document lists the class hierarchy for the most important classes in OpenSimRoot.

Minimodels

Minimodels are of type SimulaBase and encapsulate one time and location dependent state variable. The inheritance diagram for all SimulaX classes is given in Supplemental Figure 3.1.



Supplemental Figure 3.1: Inheritance diagram for all SimulaBase classes.

- SimulaConstant encapsulates a constant of various types.
- SimulaDerivative encapsulates an algorithm. Available algorithms are all the DerivativeBase derived plugins.
- SimulaTable encapsulate an array of time,value combinations. Values are interpolated.
- SimulaExternal provides a mechanism for encapsulating other dynamic simulation models.
- SimulaPoint simulates a point and its movement through space.
- SimulaVariable simulates a value and change over time using numerical integration.
- SimulaGrid simulates a static, 3D field using a list of Coordinates with values and a 3D interpolation algorithm
- SimulaLink simply bridges to another minimodel in the hierarchy of minimodels.
- SimulaStochastic draws numbers from a random number generator.

Inherited from DerivativeBase

Below is a list of all the plugins that directly, or indirectly, inherit from DerivativeBase and can be used by SimulaVariable, SimulaPoint or SimulaDerivative for computation.

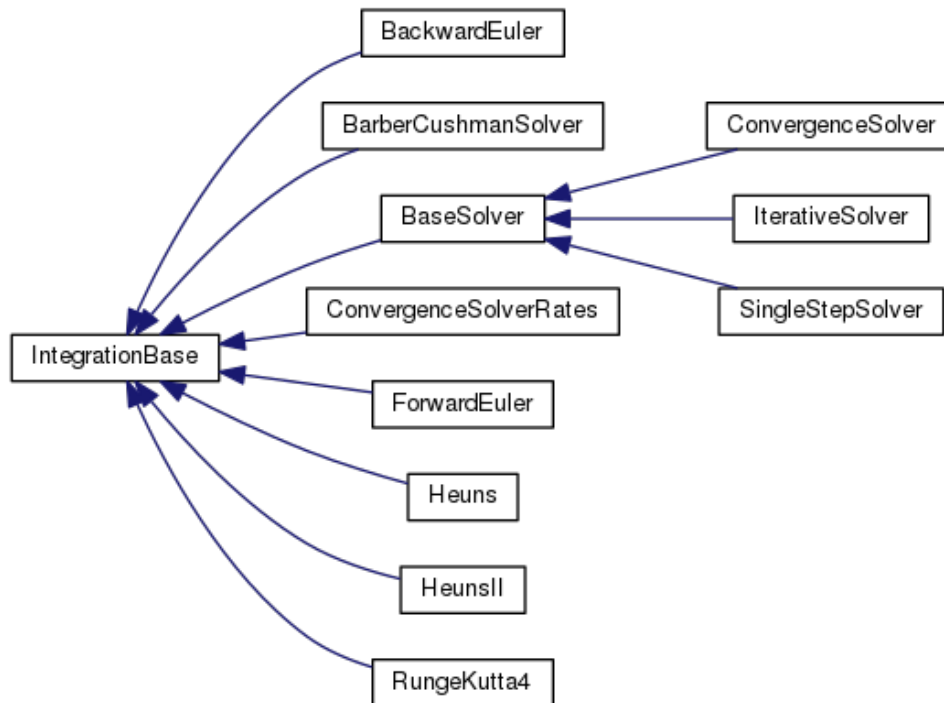
ActualTranspiration	PointSensor	SimplePotentialTranspiration
ActualVaporPressure	PotentialLeafArea	SimpleSoilTemperature
AerodynamicResistance	PotentialTranspirationCrop	SlopeVaporPressure
AirDensity	Proximity	SoluteMassBalanceTest
AirPressure	Radiation	SpecificHeatCapacityOfAir
BFMemory	RadiusDepletionZoneBarberCushman	StemDryWeight
BiologicalNitrogenFixation	RadiusDepletionZoneSimRoot4	StemPotentialCarbonSinkForGrowth
CarbonAllocation2Leafs	RandomGravitropism	StemRespirationRate
CarbonAllocation2Roots	RandomImpedence	StomatalResistance
CarbonAllocation2Shoot	RelativeCarbonAllocation2LeafsFromInputFile	StressAdjustedPotentialLeafArea
CarbonAllocation2Stems	RelativeCarbonAllocation2RootsFromInputFile	StressFactor
CarbonAvailableForGrowth	RelativeCarbonAllocation2RootsOneMinusShoot	SumCarbonCosts
CarbonCostOfBiologicalNitrogenFixation	RelativeCarbonAllocation2RootsPotentialGrowth	SumOverPlants
CarbonCostOfNutrientUptake	RelativeCarbonAllocation2RootsScaledGrowth	SumOverPlantsShoot
CarbonReserves	RelativeCarbonAllocation2ShootFromInputFile	SuperCoring
CinDryWeight	RelativeCarbonAllocation2ShootPotentialGrowth	Swms3d
ConstantRootGrowthRate	RelativeCarbonAllocation2ShootScaledGrowth	ThermalConductivity
D95	RelativeCarbonAllocation2ShootSwitch	TotalBase
ETbaseclass	RelativeCarbonAllocation2StemsOneMinusLeafs	CarbonCostOfExudates
Grass_reference_evapotranspiration	RemainingProportion	CortexDiameter
Penman	Reserves	IntegrateOverSegment
PenmanMonteith	ReservesSinkBased	PotentialSecondaryGrowth
PriestleyTaylor	RootCircumference	RootDiameter
Stanghellini	RootClassID	RootSegmentDryWeight
Tall_reference_Crop	RootDryWeight	RootSegmentLength
GetValuesFromPlantWaterUptake	RootGrowthDirection	RootSegmentSurfaceArea
GetValuesFromSWMS	RootGrowthScalingFactor	RootSegmentVolume
Imax	RootLength2Base	RootSegmentVolumeCortex
Interception	RootLengthDensity	SecondaryGrowth
InterceptionV2	RootLengthProfile	SumSteelCortex
LeafArea	RootNodePotentialCarbonSinkForGrowth	TotalBaseLabeled
LeafAreaIndex	RootPotentialCarbonSinkForGrowth	Barber_cushman_1981_nutrient_uptake
LeafAreaReductionCoefficient	RootsBelowD95Solute	Barber_cushman_1981_nutrient_uptake_explicit
LeafDryWeight	RootSegmentAge	MichaelisMenten
LeafDryWeight2	RootSegmentRespirationRate	OptimalNutrientContent
LeafPotentialCarbonSinkForGrowth	RootSegmentRootHairSurfaceArea	RootSegmentNutrientDepletionVolume
LeafRespirationRate	RootSegmentSpecificWeight	SegmentMaxNutrientUptakeRate
LightInterception	RootSystemTotal	Tropisms
LocalNutrientResponse	RootTotal	UseDerivative
MeanLeafAreaIndex	RootTotal2	UseParameterFromParameterSection
NumberOfRoots	SaturatedVaporPressure	UseRootClassAndNutrientSpecificTable
NumberOfTillers	ScaledRootGrowthRate	VolumetricHeatCapacity
NutrientStressFactor	ScaledWaterUptake	WaterMassBalanceTest
NutrientStressFactorV2	ShootDryWeight	WaterUptakeFromHopmans
PhotosynthesisLintul	ShootOptimalNutrientContent	
PhotosynthesisLintulV2		
PlantCarbonBalance		
PlantCarbonIncomeRate		
PlantTotal		

List of plugins for simulating various processes

Note that these are a list of classes, as they appear in the code. Registration of the plugins may occur under different names. Inputfiles use the registered names, not the class names. Use `OpenSimRoot -l` to get that list. See also operation manual in Supplement 2.

Integration functions

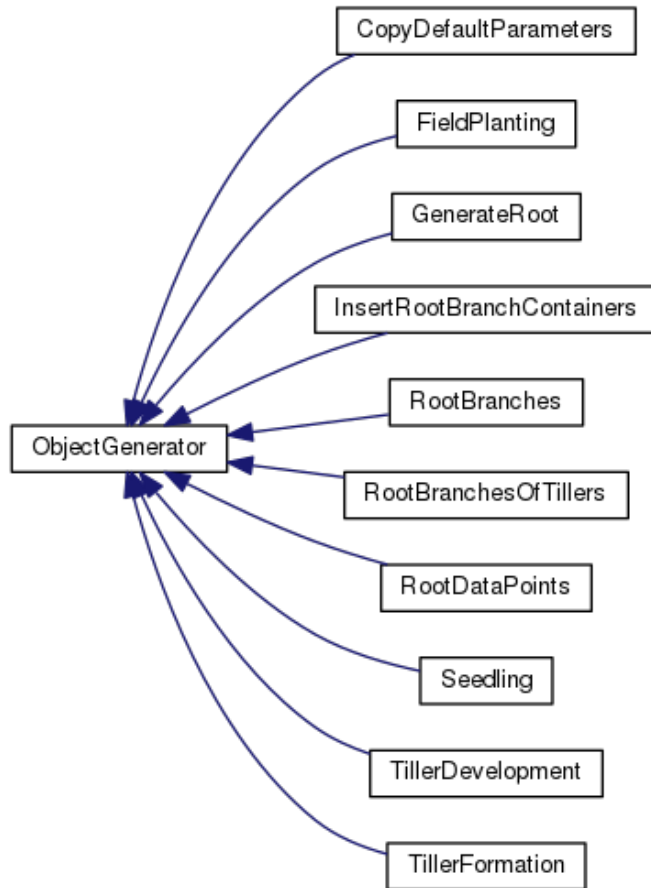
The SimulaVariable and SimulaPoint classes use helper functions for integrating the result. Several integration methods have been implemented (Supplemental figure 3.2). New integration functions can be added and registered, using the plugin framework, similar to the classes that inherit from DerivativeBase.



Supplemental figure 3.2: Inheritance diagram for the integration classes

Object generators

Object generators are plugins that can be associated with any SimulaX object and update the list of children when a child is requested.



Supplemental figure 3.3: Inheritance diagram for the object generators

Supplement 4: Plugin example code

Here we give example code for a simple plugin and the code needed to register this plugin with OpenSimRoot. Once the code has been put into a text file, it can be compiled and linked to OpenSimRoot.

1) For new algorithms

```
//Class declaration. Class should inherit from DerivativeBase, have a constructor, and
implements two virtual methods, getName() and calculate(). The example class presented here has
two SimulaBase pointers as private members, which will be used to connect to the minimodels that
simulate length and diameter of a root segment and to retrieve their values.

class RootSegmentSurfaceArea:public DerivativeBase{
public:
    RootSegmentSurfaceArea(SimulaDynamic* pSD);
    std::string getName() const;
protected:
    void calculate(const Time &t,double &var);
private:
    SimulaBase *diameter,*length;
};

//the constructor of our class. pSD is the pointer to the minimodel that uses the plugin for
computation
RootSegmentSurfaceArea::RootSegmentSurfaceArea(SimulaDynamic* pSD):DerivativeBase(pSD)
{
//We check that the user set the unit right
    pSD->checkUnit("cm2");
//We retrieve the pointers
    length=pSD->getSibling("rootSegmentLength","cm");
    diameter=pSD->getSibling("rootDiameter","cm");
}

//the computation
void RootSegmentSurfaceArea::calculate(const Time &t,double &area){
//first we retrieve data
    double d,l;
    diameter->get(t,d);
    length->get(t,l);
//second we compute
    area=l*d*PI;
}

//the name under which the plugin will be registered, make sure it is unique, use OpenSimRoot
-l to see what names are already taken
std::string RootSegmentSurfaceArea::getName() const{
    return "rootSegmentSurfaceArea.v3";
}

//Now we create a function for instantiating our class
DerivativeBase * newInstantiationRootSegmentSurfaceArea(SimulaDynamic* const pSD){
    return new RootSegmentSurfaceArea(pSD);
}

//And we register this plugin using a static instantiation of a class which guarantees that the
constructor is when OpenSimRoot is started
static class AutoRegisterMyNewPlugin {
public:
    AutoRegisterMyNewPlugin() {
//this line does the registration. Make sure you register under the same name as the getName()
method returns. This important for the model dump being loadable again.
        BaseClassesMap::getDerivativeBaseClasses()["rootSegmentSurfaceArea.v3"] =
newInstantiationRootSegmentSurfaceArea;
    } rf9843hh923h; //the one static instance of this class
}
```

2) For new integration functions

```
//class declaration, must inherit from IntegrationBase, has a constructor,  
// a getName() method and at least one integrate method  
class BackwardEuler:public IntegrationBase{  
public:  
    BackwardEuler();  
    std::string getName()const;  
protected:  
    virtual void integrate(SimulaVariable::Table & data, DerivativeBase & rateCalculator);  
    virtual void integrate(SimulaPoint::Table & data, DerivativeBase & movementCalculator);  
};  
  
BackwardEuler::BackwardEuler():IntegrationBase(){}  
  
void BackwardEuler::integrate(SimulaVariable::Table & data, DerivativeBase &rateCalculator){  
    //...Your new algorithm here which should extend the data table, the derivative (rates) that  
    //should be used are retrieved from the rateCalculator. For examples see code.  
}  
  
void BackwardEuler::integrate(SimulaPoint::Table & data, DerivativeBase & movementCalculator){  
    //...Your new algorithm here, but then suitable for Coordinates, not doubles. Intended to  
    //allow the simulation of a point moving through space. Mostly used to simulate the growth  
    //trajectory of the root tip  
};  
  
std::string BackwardEuler::getName()const{  
    return "BackwardEuler";  
}  
  
//function for instantiating the class  
IntegrationBase * newInstantiationBackwardEuler(){  
    return new BackwardEuler();  
}  
  
//Register the instantiation function  
static class AutoRegisterIntegrationFunctions {  
public:  
    AutoRegisterIntegrationFunctions() {  
        BaseClassesMap::getIntegrationClasses()["BackwardEuler"] = newInstantiationBackwardEuler;  
    }  
}  
}p44608510843540385;//the one static instance of this class
```

3) For object generators

```
//class declaration for an object generator
class MyGenerator: public ObjectGenerator {
public:
    void initialize(const Time &t);
    void generate(const Time &t);
    MyGenerator(SimulaBase* const pSB);
};

//construction is delayed. Code is in the initialize method
MyGenerator::MyGenerator(SimulaBase* const pSB) :
    ObjectGenerator(pSB) {
}

//collecting of info, and or construction of minimodels at the start of the simulation
void MyGenerator::initialize(const Time &t) {
    //collect some info about planting time
    Time plantingTime;
    SimulaBase *pt=pSB->existingChild("plantingTime");
    if (pt) {
        //read planting time from file
        pt->get(t, plantingTime);
    } else {
        //copy from parent
        plantingTime = pSB->getStartTime();
    }

    //generate new plant by copying the template
    pSB->copyAttributes(plantingTime, ORIGIN->getChild("plantTemplate"));

    //we are done
    pSB->stopUpdatefunction();
}

void MyGenerator::generate(const Time &t) {
    //add code if there is time dependent generation of objects, not just at the start
}

//the function for instantiation of the class
ObjectGenerator * newInstantiationMyGenerator(SimulaBase* const pSB) {
    return new MyGenerator(pSB);
}

//register the instantiation function
static class AutoRegisterMyGeneratorInstantiationFunctions {
public:
    AutoRegisterMyGeneratorInstantiationFunctions() {
        BaseClassesMap::getObjectGeneratorClasses()["MyGenerator"] =
            newInstantiationMyGenerator;
    }
} p4595582386;
```

Supplement 5: Detailed description of the water and nutrient submodules

Watermodule

Plant transpiration is simulated by OpenSimRoot, assuming that water availability is not limiting and stomatal conductance is constant. Transpiration and evaporation need to be separated within OpenSimRoot. Transpiration can be estimated from a fixed water use efficiency parameter (which simply links carbon fixation linearly to transpiration), or from the Penman-Monteith model, which computes evapotranspiration based on weather conditions (Penman, 1948; Monteith, 1964). When transpiration is calculated based on a water use efficiency parameter, the user needs to provide evaporation values; when the Penman-Monteith model is used, transpiration and evaporation are separated by OpenSimRoot solving the Penman-Monteith model twice, once for full crop cover, and once for a bare soil. Based on the percent light capture by the crop OpenSimRoot scales evaporation and the transpiration terms assuming evaporation is negligible and small under full crop cover (Leaf Area Index ~ 3).

To simulate the soil hydrology, OpenSimRoot has a submodule that solves the Richards equation in three dimensions using finite element method (FEM) on a Cartesian grid. The soil water submodule is a simplified and modified C++ rewrite of the SWMS3D model, which is the basis of Hydrus and R-SWMS (Šimuněk *et al.*, 1995; Diamantopoulos *et al.*, 2013).

Certain exceptional circumstances such as drainage or water ponding at top soil, are excluded. The top boundary condition is a water flux that is the difference between precipitation and evaporation. Evaporation, as computed by the Penman-Monteith equation, is assumed to be potential evaporation (i.e. appropriate for wet soils), and assumed to be equal across the soil surface, shoot geometry is not simulated. Potential evaporation is scaled back to an actual evaporation by including a smooth scaling function which causes evaporation to decrease smoothly from potential, when the top soil is wet, to equal the soil conductivity when the soil is not able to sustain higher evaporation rates. If the top soil is not necessarily uniformly wet, actual evaporation will be non-uniform across the soil surface in OpenSimRoot. The water retention curve and soil hydraulic conductivity are computed using the van Genuchten and Mualem equations.

The Richards equation can include a sink term, which in OpenSimRoot represents water uptake by roots (as described evaporation sink is handled as dynamic boundary condition). To do so we need to know 1) how much water is taken up by each root segment at a given moment in time, and 2) how that uptake is coupled to the FEM nodes of the grid on which the Richards equation is solved. Assuming that root uptake equals transpiration, i.e. we ignore temporal water storage in the plant, OpenSimRoot can either divide the water uptake of the whole root system by assuming each root segment contributes equally to uptake relative to its length (as in Hopmans, (Hopmans & Bristow, 2002)) or by solving the hydraulic architecture represented by a network model and using a circuit analogy likewise motivated by finite element theory (Alm *et al.*, 1992; Doussan *et al.*, 1998). The network model is novel in OpenSimRoot implemented to work with a growing root and used in the study of Schneider (Unpublished). This model requires axial and radial hydraulic conductivities for each root segment, which can be defined in the input files as a function of root age and class, and are scaled (i.e. normalized) with the inverse of the root segment length (axial), or the root segment surface area (radial). The coupling of the root model to the FEM model enables each root segment to have a soil water content at the root surface. The next step is to make sure that water uptake by the root system equals the transpiration which is

achieved by changing the water potential at the root collar (top of the hypocotyl). Getting the root collar potential is a parabolic optimization function which is solved with a Newton solver, typically in three steps. The water potential at the top of the hypocotyl is not allowed to drop below a given threshold. If the threshold is reached, OpenSimRoot assumes that water uptake is less than potential transpiration and will write a warning. Further simulation results might not be correct as currently no effects of drought on photosynthesis, leaf expansion etc have been implemented. However, the model should correctly deal with compensatory uptake of water when soil water distribution is heterogeneous. And this model can show water loss of roots while the same conductivity from xylem to soil is assumed.

Mapping the root model to the FEM model is done based on a neighborhood search. All FEM nodes surrounding the root segment are considered. Sink terms, and local environment are computed based on inverse distance weighted average of the FEM nodes surrounding the root node. An alternative mapping algorithm, by which every FEM node is assigned with every root node has been implemented, in order to ignore root architecture completely in the water and nutrient uptake simulations. This was for example used in Postma and Lynch (2012) where it was concluded that the positioning of the root, that is root architecture, is necessary for simulating niche differentiation for nitrate uptake among maize, bean and squash plants, whereas if roots would be able to take up nutrients from everywhere in the soil, there would be no niche differentiation.

Nutrient module

OpenSimRoot has a nutrient module to simulate the uptake solutes, and in the new version theoretically simultaneously for various nutrients. This module was implemented to simulate the function of root architectural traits for nutrient uptake, and test tradeoffs for acquisition of different nutrients. Time dependent optimal and minimal nutrient content ($\mu\text{mol/g}$) have to be defined for leaves, stems and all root classes, for to be simulated solutes. These amounts are used to compute nutrient requirements of the plant, and compared to total uptake amounts, including initial seed reserves (for uptake see below). When uptake is less than demand, plant stress is assumed, with maximum stress being defined as uptake equal to minimal nutrient content ($\text{stress}(\text{uptake}) = \max(0, \min((\text{uptake}-\text{minimal})/(\text{optimal}-\text{minimal}), 1))$). Stress modifying impact functions can be defined for components such as leaf expansion rate, photosynthesis rates, respiration rates, and root elongation rates or secondary growth. Typically, they should be defined such that, when stress=0, growth ceases altogether. For example, by making the initial response of the shoot stronger than that of the root, the plant will decrease shoot to root ratios when nutrient deficient. Thus OpenSimRoot will move towards a functional equilibrium, although due to the inherent slow nature of growth, and the relative fast dynamics of other processes, this functional equilibrium might not be reached, and oscillatory behavior might occur (Postma & Lynch, 2011; Postma *et al.*, 2014b). The current implementation assumes that internally, reallocation of nutrients is fast and perfect, such that all organs experience equal stress. This might be true for a nutrient like nitrogen, which typically causes chlorosis everywhere in the shoot, but might not be correct for other nutrients. The importance of simulation of nutrient redistribution in the plant still needs study, and would require implementation of a shoot architectural model in which the age and position of individual leaves is tracked.

Nutrient uptake from soil to root is simulated independently of utilization of nutrients within the plant. Two options for simulation are provided: 1) The Barber-Cushman model and 2) a 3D FEM

model. One is a C++ implementation of the original Barber-Cushman model with root hairs. The model is described as radial 1D PDE (Partial Differential Equation) which corresponds to the rhizosphere around the root. It assumes nutrient uptake to be described by a Michaelis-Menten term, and the nutrient transport in the soil to be driven by convection (water flow) and diffusion. A buffer constant replaces a reaction term. The Barber-Cushman model is suitable for immobile nutrients like phosphorus. Phosphorus uptake causes steep gradients in concentrations around the root. These depletion zones are typically only 2-4 mm in diameter, and thereby would require a computationally unacceptably high resolution of the 3D finite element model (~0.1 mm resolution of a 1 m³ soil pedon would result in 1e¹² elements or 8 petabytes to hold a single double precision array).

Competition between roots is computed based on a local average root density which determines the outer boundary of the Barber-Cushman model. OpenSimRoot updates this boundary when new roots grow in the vicinity of other roots and corrects the initial nutrient concentration for new roots with the uptake of nutrients of older roots. Nevertheless, this handling of root competition is only acceptable when the overlap of depletion zones, which can be computed based on raster images of the root system, is relatively small. For crops, overlap in phosphorus depletion zones is typically below 20% because of its low mobility. Inter and intra root competition plays a much more important role in the uptake of mobile nutrients such as nitrate. Nitrate might form diffuse or no depletion zones around the root and for this reason is better simulated using a 3D FEM. SimRoot solved the convection-dispersion equation on the same FEM grid as the water transport is solved which can be restricting, OpenSimRoot alternatively can solve it on a refined grid, where the refinement factor is yet fixed to 2nd, 4th, 8th or the 16th of a reference grid. For each solute a new FEM model is instantiated and linked to the water model. The 3D FEM model for solute transport is coupled to the root systems using the same method as used for the hydraulic model, where the uptake of solutes by the root segments is based on Michaelis-Menten kinetics, as in the Barber-Cushman model. Buffering and diffusion coefficients are dependent on the soil water content, and might thereby deviate from the constant coefficients used in the Barber-Cushman model. The effects must be considered when comparing the output of both models (Postma and Lynch, 2011).

When simulating more than one solute, solutes do not influence each other directly in OpenSimRoot. Indirect effects occur through the influence of nutrient uptake on root growth. Each solute has a stress function to determine how each impacts, for example, photosynthesis. A user specified aggregation function determines the aggregate impact (Dathe *et al.*, 2012). For example, Postma *et al.*, (2014a) showed how the optimal lateral branching density in maize depends on the relative availability of phosphorus and nitrogen.

References

Alm DM, Cavalier J, Nobel PS. 1992. A finite-element model of radial and axial conductivities for individual roots: development and validation for two desert succulents. *Annals of Botany* **69**: 87–92.

Dathe A, Postma JA, Lynch JP. 2012. Modeling resource interactions under multiple edaphic stresses. In: Ahuja LR, In: Reddy VR, In: Saseendran SA, In: Yu Q, eds. *Advances in Agricultural Systems Modeling*. Accepted for publication in. Madison, Wis., USA: ASA-CSSA-SSSA, .

Diamantopoulos E, Iden SC, Durner W. 2013. Modeling non-equilibrium water flow in multistep outflow and multistep flux experiments. *HYDRUS Software Applications to Subsurface Flow and Contaminant Transport Problems*: 69.

Doussan C, Pagès L, Vercambre G. 1998. Modelling of the hydraulic architecture of root systems: An integrated approach to water absorption - Model description. *Annals of Botany* **81**: 213–223.

Hopmans JW, Bristow KL. 2002. Current capabilities and future needs of root water and nutrient uptake modeling. *Advances in Agronomy* **77**: 103–183.

Monteith JL. 1964. Evaporation and environment. *Symposia of the society for experimental biology* **19**: 205–234.

Penman HL. 1948. Natural evaporation from open water, bare soil and grass. Proceedings of the Royal Society of London A: Mathematical, Physical and Engineering Sciences. The Royal Society, 120–145.

Postma JA, Dathe A, Lynch JP. 2014a. The optimal lateral root branching density for maize depends on nitrogen and phosphorus availability. *Plant Physiology* **166**: 590–602.

Postma JA, Lynch JP. 2011. Theoretical evidence for the functional benefit of root cortical aerenchyma in soils with low phosphorus availability. *Annals of Botany* **107**: 829–841.

Postma JA, Lynch JP. 2012. Complementarity in root architecture for nutrient uptake in ancient maize/bean and maize/bean/squash polycultures. *Annals of Botany* **110**: 521–534.

Postma JA, Schurr U, Fiorani F. 2014b. Dynamic root growth and architecture responses to limiting nutrient availability: linking physiological models and experimentation. *Biotechnology Advances* **32**: 53–65.

Šimuněk J, Huang K, van Genuchten MT. 1995. *The SWMS 3D code for simulating water flow and solute transport in three-dimensional variably-saturated media*. California: U. S. Salinity laboratory, USDA.

Supplement 6: Example of a simple OpenSimRoot input file

The XML below is an example of an OpenSimRoot input file that constructs a simple crop model, without any roots. All the SimulaX tags will instantiate a minimodel of the corresponding type, for example a constant (time independent parameter) is declared as `<SimulaConstant ...>`. Metadata for the minimodels, such as name and unit, are given in the attributes lists.

General rules for XML documents

- 1) The document has tags which are between brackets like `<>`
- 2) Tags correspond to minimodels in OpenSimRoot and therefore carry different names, such as SimulaBase, SimulaConstant, etc.
- 3) Tags need to be closed either by putting a `/` before the closing bracket, or if data is nested inside the tag with a corresponding closing tag which is recognized by `</>`. For example `<SimulaConstant></SimulaConstant>`
- 4) Between opening and closing tags you will find data, and or declarations of minimodels which are at the next level in the hierarchy
- 5) Tags carry attributes which describe metadata. Attributes are always listed as `attribute="something"`. In OpenSimRoot all tags have at least a name attribute.
- 6) An XML document is plain text and recognized by a special declaration at the top of the document. `<?xml version="1.0" encoding="UTF-8"?>`
- 7) XML documents can have stylesheets associated with them so the the browser knows how to render the document. Here we have `<?xml-stylesheet type="text/xsl" href="tree-view.xsl"?>`
- 8) Comments are between `<!--` and `-->`.
- 9) All XML documents of a document type tag. For OpenSimRoot the document type is declared as `<SimulationModel></SimulationModel>`. All other tags must be in between these tags.

Here follows an example input file. The comments in black give more explanation as to how a simple crop model is being constructed by this input file. Input files for full root architectural models can be found in the software repository on gitlab:
<https://gitlab.com/rootmodels/OpenSimRoot>

```
<?xml version="1.0" encoding="UTF-8"?>
<?xml-stylesheet type="text/xsl" href="tree-view.xsl"?>
<!--
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CONTRIBUTORS BE LIABLE FOR ANY DIRECT, INDIRECT, INCIDENTAL, SPECIAL, EXEMPLARY, OR CONSEQUENTIAL
DAMAGES (INCLUDING, BUT NOT LIMITED TO, PROCUREMENT OF SUBSTITUTE GOODS OR SERVICES; LOSS OF USE,
```

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<!--This XML constructs a simple, radiation use efficiency based crop model.

Roots and stems are only presented as Carbon (dry weight) pools
Leaf dry weight is converted to leaf area based on specific leaf area (SLA)
Leaf area is converted to light interception using an extinction coefficient.
Light interception is converted to photosynthesis using radiation use efficiency (RUE).
Photosynthesis is converted to structural carbon using a conversion factor (multiplier) which represents relative losses due to respiration
Fixed allocation causes structural carbon to be divided over root, stem and leaves.

Behavior, is simple exponential growth for which
 $RGR = SLA * C2Leafs * photosynthesis * multiplier$
However, as the light interception with increasing leaf area reaches an asymptote, the model will move towards linear growth.-->

<SimulationModel>

<!-- SimulaBase is a simple container, that holds other SimulaX objects. SimulaBase is thus a minimodel that does not hold or simulate data. It should, like all mini models, have a name. So here we declare a container in which we are going to put all our plants. Inside it we put a container for our plant, named arbitrarily "myPlant". -->

```
<SimulaBase name="plants">
  <SimulaBase name="myplant">
```

<!-- Here follow three SimulaConstant declarations. SimulaConstant is a minimodel that holds time and space independent data of different types. Possible types are double, int, string, Coordinate. Besides the name attribute they must have a unit, and if the data is not a double, a type declaration.

A plant should be of a given species/genotype. The model will look for a parameter set in roottypeParameters with the corresponding type. Here we declare that we want to simulate a plant of type mySpecies -->

```
<SimulaConstant name="plantType" type="string">
  mySpecies
</SimulaConstant>
```

<!-- The time that the plant is planted. 0. is at the start of the simulation. -->

```
<SimulaConstant name="plantingTime" unit="day">
  0.
</SimulaConstant>
```

<!-- Location in space where the seed is planted. -->

```
<SimulaConstant name="plantPosition" type="Coordinate">
  0 -2 0
```

<!-- Container that hold all the minimodels that will simulate shoot related parameters. The shoot and root are inside plantPosition, as OpenSimRoot works with a relative Coordinate system. We achieve that all coordinates that belong to our plant are relative to plantPosition. -->

```
<SimulaBase name="shoot">
```

<!-- Licht interception is simulated by the light interception module. SimulaDerivative declares a minimodel that will use the lightInterception plugin to compute light interception. Attributes are name of what is being computed (name="lightInterception"), the unit of what is being computed (unit="umol/cm2/day"), and the plugin that should be used to compute it (function="lightInterception"). The plugin lightInterception requires leafAreaIndex and from the parameter section and extinctionCoefficient (kdf). Further it needs irradiation levels from the environmental section. All have been declared further down. -->

```

        <SimulaDerivative name="lightInterception" unit="umol/cm2/day"
            function="lightInterception" />
<!--Simulation of photosynthesis rates can be done by the plugin registered as
photosynthesisLintulV2. However, since we want to know the total photosynthesis, the rates need
to be integrated over time. SimulaVariable does this. Thus unit is not g/day, but g. Attributes
are otherwise same as for a SimulaDerivative tag. Optional attributes that control the method of
integration and the timestep can be given. For example integrationFunction="ForwardEuler" will
use the forward euler plugin for integrating. List of all integration methods can be obtained by
running OpenSimRoot -L. maximumTimeStep="0.1" would reduce the maximum timestep from the default
0.2 to 0.1.
-->
        <SimulaVariable name="photosynthesis" unit="g"
            function="photosynthesisLintulV2" />
<!--Declaration of how leafAreaIndex should be simulated, as it is needed by the
lightInterception plugin. -->
        <SimulaDerivative name="leafAreaIndex" unit="cm2/cm2"
            function="leafAreaIndex" />
<!--Declaration of how leafArea should be simulated, as it is needed by the leafAreaIndex plugin.
Here the initial leaf area is given. More time value pairs can be entered in order to specify a
predefined initial leaf area. The leafArea plugin will simulate increases in leaf area on the
basis of carbon allocation to the leaves, the specificLeafArea and the carbonToDryweight ratio,
all declared later on.---->
        <SimulaVariable name="leafArea" unit="cm2" function="leafArea">
            0. 1. </SimulaVariable>
<!--Same as leafArea, but then for leafDryWeight. -->
        <SimulaVariable name="leafDryWeight" unit="g"
            function="leafDryWeight.v2"> 0. 0.001 </SimulaVariable>
<!--Here follow more minimodels, all with their respective plugins declared -->
        <SimulaDerivative name="relativeCarbonAllocation2Leafs"
            unit="100%"
            function="relativeCarbonAllocation2LeafsFromInputFile" />
        <SimulaVariable name="carbonAllocation2Leafs" unit="g"
            function="carbonAllocation2Leafs" />
        <!-- optional to have stem weight -->
        <SimulaDerivative name="relativeCarbonAllocation2Stems"
            unit="100%"
            function="relativeCarbonAllocation2StemsOneMinusLeafs" />
        <SimulaVariable name="carbonAllocation2Stems" unit="g"
            function="carbonAllocation2Stems" />
        <SimulaVariable name="stemDryWeight" unit="g"
            function="stemDryWeight" />
    </SimulaBase>
</SimulaConstant>
<!--In this simulation it was decided to declare the carbonToDryWeight ratio as a simple
constant. -->
        <SimulaConstant name="carbonToDryWeightRatio" unit="100%">
            0.45
        </SimulaConstant>
<!--Carbon allocation -->
        <SimulaDerivative name="relativeCarbonAllocation2Shoot"
            unit="100%"
            function="relativeCarbonAllocation2ShootFromInputFile" />
        <SimulaVariable name="carbonAllocation2Shoot" unit="g"
            function="carbonAllocation2Shoot" />

```

```
<!--Instead of using a process specific plugin to simulate the carbon available for growth, here we use a general plugin named usePath which simply couples the carbon available for growth to photosynthesis. Since this declaration as a child called "multiplier" the photosynthesis rates is halved, so it is assumed that half of all carbon fixed by photosynthesis is converted to plant dry mass, the rest is respired. -->
```

```
    <SimulaDerivative name="carbonAvailableForGrowth"  
      unit="g" function="usePath">  
      <SimulaConstant name="path" type="string">  
        plantPosition/shoot/photosynthesis  
      </SimulaConstant>  
      <!-- half of carbon assumed to be respired -->  
      <SimulaConstant name="multiplier">0.5</SimulaConstant>  
    </SimulaDerivative>
```

```
<!--Some declarations related to roots -->
```

```
    <SimulaDerivative name="relativeCarbonAllocation2Roots" unit="100%"  
      function="relativeCarbonAllocation2RootsOneMinusShoot" />  
    <SimulaVariable name="carbonAllocation2Roots" unit="g"  
      function="carbonAllocation2Roots" />  
    <SimulaVariable name="rootDryWeight" unit="g" function="rootDryWeight" />
```

```
<!--The closing tags for the myPlant and Plants containers. -->
```

```
  </SimulaBase>  
</SimulaBase>
```

```
<!-- Environmental data needs to be declared, here all we need is irradiation in order to know how much light is being captured for photosynthesis -->
```

```
  <SimulaBase name="environment">  
    <SimulaBase name="atmosphere">  
      <SimulaTable name_column1="time" name_column2="irradiation"  
        unit_column1="day" unit_column2="umol/cm2/day">  
        0 3000  
        100 3000  
      </SimulaTable>  
    </SimulaBase>  
  </SimulaBase>
```

```
<!-- here a parameter section for our plant is specified. -->
```

```
  <SimulaBase name="rootTypeParameters">  
    <SimulaBase name="mySpecies">  
      <SimulaBase name="resources">
```

```
<!--relativeCarbonAllocation to leaves (see above) uses a plugin in that simply looks up data from a table. The table is declared here. SimulaTables have two columns. Each column has a name and a unit declared in the attribute list. Here, as will be often the case, the first column is time. This is time since the plant started growing, not since the start of the simulation. First all carbon that is going to the shoot is allocated to leaves, later on more carbon is going to the stems. Values in the table are interpolated linearly, unless a different interpolation method is declared. Currently, only interpolation="step" is implemented as alternative method. -->
```

```
    <SimulaTable name_colum1="time" unit_colum1="day"  
      name_colum2="carbonAllocation2LeafsFactor" unit_colum2="100%">  
      0 1  
      10 0.8  
      40 0.5  
      60 0.  
      80 0.  
    </SimulaTable>
```

```
<!--How much carbon should go to the root. The rest goes to the shoot. -->
```

```
    <SimulaTable name_colum1="time" unit_colum1="day"  
      name_colum2="carbonAllocation2RootsFactor" unit_colum2="100%">
```

```

    0 0.8
    10 0.2
    40 0.2
    80 0.2
  </SimulaTable>
</SimulaBase>

<!--Declaration of several well known shoot related parameters. -->

  <SimulaBase name="shoot">
    <SimulaConstant name="areaPerPlant" unit="cm2">
      100
    </SimulaConstant>
    <SimulaConstant name="extinctionCoefficient" unit="noUnit">
      0.6
    </SimulaConstant>
    <SimulaConstant name="lightUseEfficiency" unit="g/umol">
      0.4E-6
    </SimulaConstant>
    <SimulaTable name_colum1="time" name_colum2="specificLeafArea"
      unit_colum1="day" unit_colum2="g/cm2" note="SLA in lintul">
      0 0.001
      10 0.002
      40 0.003
      80 0.003
    </SimulaTable>
  </SimulaBase>
</SimulaBase>
</SimulaBase>

<!--This section gives the user some control over the output.-->

  <SimulaBase name="simulationControls">
    <SimulaBase name="outputParameters">
      <SimulaBase name="table">

<!--A table should be written containing values for each minimodel, for every half day from day 0
to 80. Hierarchy will be traversed up to depth 10 -->

      <SimulaConstant name="run" type="bool"> 1 </SimulaConstant>
      <SimulaConstant name="searchingDepth" type="int"> 10
      </SimulaConstant>
      <SimulaConstant name="startTime" type="time"> 0.
      </SimulaConstant>
      <SimulaConstant name="endTime" type="time"> 80.
      </SimulaConstant>
      <SimulaConstant name="timeInterval" type="time"> 0.5
      </SimulaConstant>
    </SimulaBase>
  </SimulaBase>
</SimulaBase>

<!--We are done -->
</SimulationModel>

```

User friendly viewing of XML input files

A webbrowser can transform this into more human friendly presentation using the attached tree-view.xml transformation style sheet (available for download at the gitlab repository <https://gitlab.com/rootmodels/OpenSimRoot>). The result when you open this file in a browser is given below.

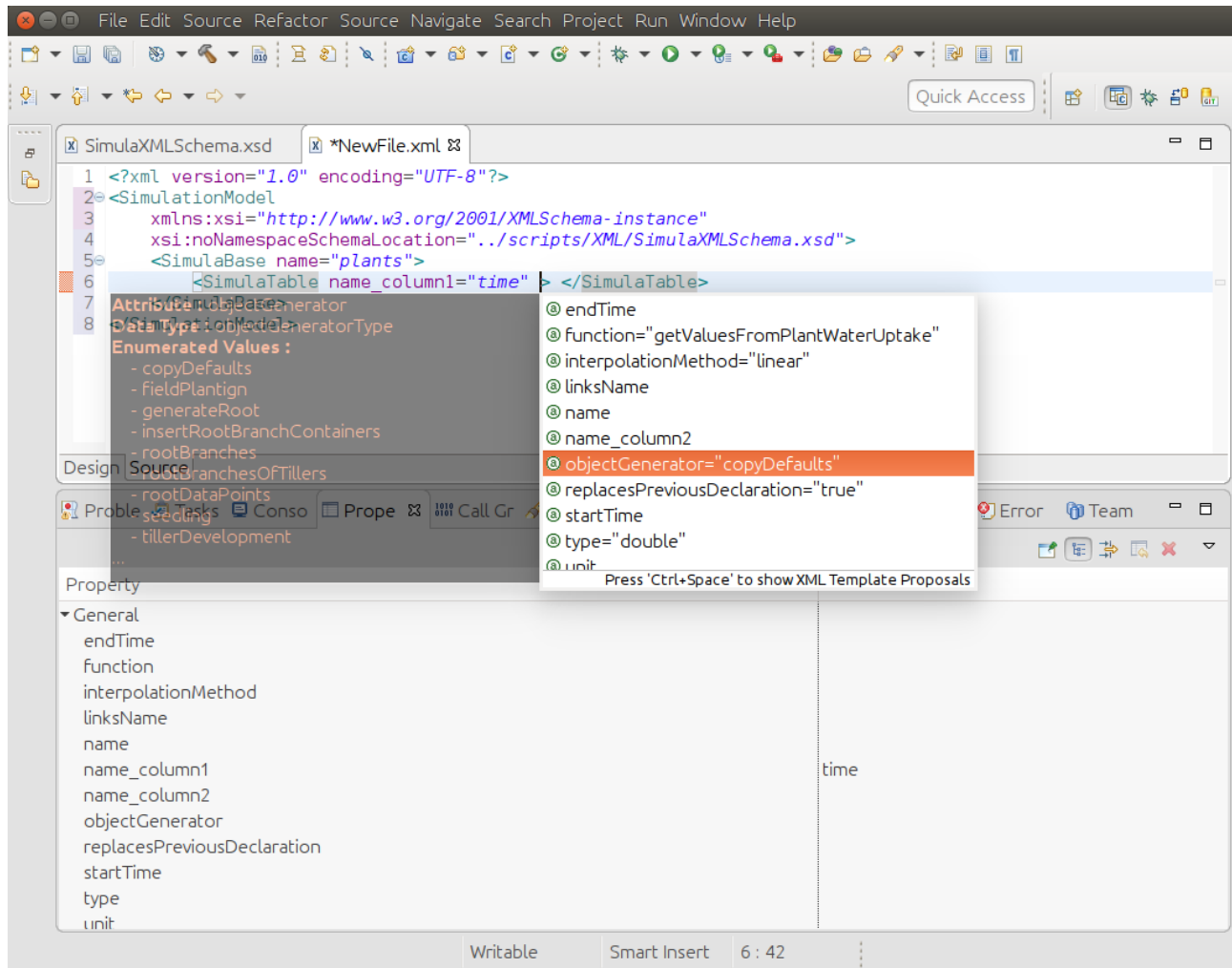
```

|__ Origin
|__ 'plant'
|__ 'myplant'
|__ 'plantType' = mySpecies
|__ 'plantingTime' = 0 (day)
|__ 'plantPosition' = 0 -2 0
|__ 'shoot'
|__ 'lightInterception' (umol/cm2/day)
|__ 'photosynthesis' (g)
|__ 'leafAreaIndex' (cm2/cm2)
|__ 'leafArea' (cm2) initial value = 1.
|__ 'leafDryWeight' (g)
|__ 'relativeCarbonAllocation2Leafs' (100%)
|__ 'carbonAllocation2Leafs' (g)
|__ 'relativeCarbonAllocation2Stems' (100%)
|__ 'carbonAllocation2Stems' (g)
|__ 'stemDryWeight' (g)
|__ 'carbonToDryWeightRatio' = 0.45 (100%)
|__ 'relativeCarbonAllocation2Shoot' (100%)
|__ 'carbonAllocation2Shoot' (g)
|__ 'carbonAvailableForGrowth' (g)
|__ 'path' = plantPosition/shoot/photosynthesis
|__ 'multiplier' = 0.5
|__ 'relativeCarbonAllocation2Roots' (100%)
|__ 'carbonAllocation2Roots' (g)
|__ 'rootDryWeight' (g)
|__ 'environment'
|__ 'atmosphere'
|__ x,y pairs :{ 0 3000 100 3000 }
|__ 'rootTypeParameters'
|__ 'mySpecies'
|__ 'resources'
|__ 'carbonAllocation2LeafsFactor' (100%)=f{'time'} (day) x,y pairs :{ 0 1 10 0.8 40 0.5 60 0. 80 0. }
|__ 'carbonAllocation2RootsFactor' (100%)=f{'time'} (day) x,y pairs :{ 0 0.8 10 0.2 40 0.2 80 0.2 }
|__ 'shoot'
|__ 'areaPerPlant' = 100 (cm2)
|__ 'extinctionCoefficient' = 0.6 (noUnit)
|__ 'lightUseEfficiency' = 0.4E-6 (g/umol)
|__ 'specificLeafArea' (g/cm2)=f{'time'} (day) x,y pairs :{ 0 0.001 10 0.002 40 0.003 80 0.003 }
|__ 'simulationControls'
|__ 'outputParameters'
|__ 'table'
|__ 'run' = 1
|__ 'searchingDepth' = 10
|__ 'startTime' = 0.
|__ 'endTime' = 80.
|__ 'timeInterval' = 0.5

```

User friendly editing of XML input files

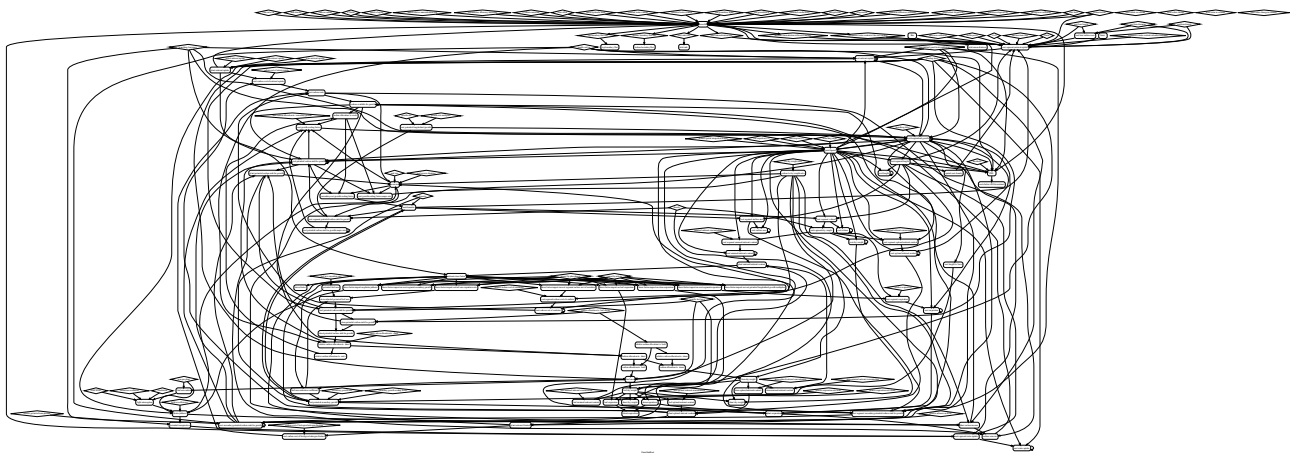
Besides attaching a transformation sheet for transforming xml to html and view it in a webbrowser, a XML schema (xsd) is available, which allows schema aware editors to provide auto completion and validation of the input file. Below is a screenshot from an XML editor (plugin in www.eclipse.org) which shows the declaration of the schema, and a pop down menu for the available arguments for SimulaTable, and the different values that the objectGenerator argument can have.



Supplemental figure 6.1: Screenshot of XML editor in eclipse in which a new file was created, using the new file wizard. The schema is declared with `“xsi:noNamespaceSchemaLocation=“../scripts/XML/SimulaXMLSchema.xsd””` and the black and the black and white pop up boxes show suggestions, as defined in the schema.

Supplement 7: Diagram of all state variables and their dependencies in an exemplar bean simulation

We drew a graph which contains the various state variables in an example simulation and the dependencies among them. Each state variable is simulated by a SimulaObject, here we depicted SimulaConstants, SimulaTables and SimulaStochastic as wedges, whereas all others are depicted as a rounded boxes. The arrows indicate information flow, that is the result of one minimodel goes into the computation of another. The network is strongly dependent on the input file, and somewhat dependent on time, given that computations might switch on given conditions and should thereby be regarded as exemplar. To properly view the graph, enlarge the pdf strongly.



Supplemental figure 7.1: Graph representing all the state variables in a bean simulation, and their connections at day 12. For better viewing, enlarge by about 1200%.

