| 1 | Development of cordycepin formulations for preclinical and clinical studies |
|----|--|
| 2 | Jong Bong Lee ¹ , Cecilia Adrower ^{1,2} , Chaolong Qin ¹ , Peter M. Fischer ¹ , Cornelia H. de Moor ¹ , |
| 3 | Pavel Gershkovich ^{1,*} |
| 4 | |
| 5 | ¹ School of Pharmacy, University of Nottingham, Nottingham, UK |
| 6 | ² School of Pharmacy, Universita di Roma Tor Vergata, Rome, Italy |
| 7 | |
| 8 | |
| 9 | *Corresponding author: Pavel Gershkovich, PhD |
| 10 | School of Pharmacy, Centre for Biomolecular Sciences |
| 11 | University of Nottingham, University Park |
| 12 | Nottingham, UK |
| 13 | NG7 2RD |
| 14 | Tel: +44 (0) 115 846 8014 |
| 15 | Fax: +44 (0) 115 951 3412 |
| 16 | Email: pavel.gershkovich@nottingham.ac.uk |
| 17 | |
| 18 | SUGGESTED RUNNING TITLE: |
| 19 | Formulation development of cordycepin |
| 20 | |
| 21 | |

23 ABSTRACT

There is extensive literature on *in vivo* studies with cordycepin but these studies were generally 24 conducted without validation of the various formulations, especially in terms of the solubility 25 26 of cordycepin in the dosing vehicles used. Cordycepin is a promising drug candidate in multiple therapeutic areas and there is a growing interest in studies aimed at assessing the 27 pharmacological activity of this compound in relevant animal disease models. It is likely that 28 many reported in vivo studies used formulations in which cordycepin was incompletely soluble. 29 This can potentially confound the interpretation of pharmacokinetics and efficacy results. 30 Furthermore, the presence of particles in intravenously administered suspension can cause 31 adverse effects and should be avoided. Here we present the results from our development of 32 33 simple and readily applicable formulations of cordycepin based on quantitative solubility assessment. Homogeneous solutions of cordycepin were prepared in phosphate-buffered saline 34 (PBS) at different pH levels, suitable as formulations for both intravenously and oral 35 administration. For the purpose of high-dose oral administration we also developed propylene 36 37 glycol (PPG)-based vehicles in which cordycepin is completely soluble. The stability of the newly developed formulations was also assessed, as well the feasibility of their sterilisation by 38 filtration. Additionally, an HPLC-UV method for the determination of cordycepin in the 39 40 formulations, which may also be useful for other purposes, was developed and validated. Our study could provide useful information for improvement of future preclinical and clinical 41 42 studies involving cordycepin.

43

44 KEYWORDS

45 Cordycepin; formulation; solubility; stability; HPLC-UV.

47 INTRODUCTION

Cordycepin (3'-deoxyadenosine) is a nucleoside that differs from adenosine by the absence of 48 the 3'-hydroxyl group. It is the main therapeutically active component in extracts of the insect 49 50 fungus Cordyceps militaris, which is a widely used in traditional medicines of the Far East (1-3). The therapeutic potential of cordycepin has been recognised for a wide range of applications 51 52 and was demonstrated in numerous studies (4). Proposed pharmacological activities of cordycepin include anti-microbial (4), anti-tumour (5-8), anti-mutagenic (9), anti-metastatic (1, 53 10), anti-angiogenesis (11), anti-fungal (12), anti-diabetic (8, 13), anti-inflammatory (8, 13-17), 54 anti-platelet aggregation (18), immunomodulatory (19, 20), hypoglycaemic (21) and anti-55 herpes (22) effects. The molecular mechanisms of action of cordycepin are not completely 56 57 understood. However, like many natural products, cordycepin interacts with multiple 58 biological processes, many of which remain to be investigated in detail (2).

59

Based on diverse *in vitro* studies, *in vivo* preclinical research have been conducted for various
indications including cancer (23, 24), cancer metastasis (25), vascular disorders (26),
neurodegeneration (27, 28), fungal infection (12), osteoporosis (29, 30), osteogenesis (31),
hyperlipidaemia (32), hyperglycaemia (21), viral infections (33), steroidogenesis (34), allergy
(35), central nervous system disorders (36, 37), amongst others (38-40) (Table 1). Moreover,
further studies are anticipated on the pharmacological modes of action and clinical efficacy of
cordycepin.

67

From the many *in vivo* experiments reported that involved cordycepin, it is noted that a substantial range of doses, and therefore a wide range of concentrations of cordycepin in different dosing vehicles, were used in these studies (Table 1). However, the solubility of cordycepin in water or other dosing vehicles used in these studies has not been reported as far as we can ascertain. Moreover, reports of such studies often do not indicate whether cordycepin formulations at the time of administration were actual homogeneous solutions or merely suspensions. If a formulation is administered orally or intraperitoneally as a suspension, factors such as dissolution and solubilisation could complicate interpretation of the results (41). Moreover, if a formulation for intravenous administration is a suspension rather than a solution, acute adverse effects can occur, especially if the suspension contains large particles. Such adverse effects can include embolism of blood vessels and inflammation (42-45).

79

Therefore, in this study, simple and readily applicable formulations of cordycepin were developed based on quantitative solubility assessment. Homogeneous solutions of cordycepin were prepared in phosphate-buffered saline (PBS) at different pH levels, yielding formulations that can be administered both intravenously and orally. A propylene glycol (PPG)-based formulation for higher concentration oral cordycepin administration was also developed. Stability and sterilisation studies for the formulations developed were also carried out.

86

87 MATERIALS AND METHODS

88 Materials

Cordycepin was purchased from Carbosynth Ltd (Berkshire, UK). Adenosine and Dulbecco's phosphate buffered saline (PBS) were obtained from Sigma (Gillingham, UK). Water for injection was purchased from Gibco (Paisley, UK) and normal saline was from Baxter (Berkshire, UK). Costar Spin-X Centrifuge Tubes and propylene glycol were purchased from Fisher Scientific (Loughborough, UK). Sartorius Minisart syringe filters were obtained from Scientific Laboratory Supplies Ltd (Nottingham, UK). All solvents used in the study were HPLC grade or higher.

97 Solubility assay

To assess the solubility of cordycepin in water for injection, normal saline, pH 4.0 PBS, pH 98 4.5 PBS and pH 5.0 PBS, saturated suspensions of cordycepin were prepared (15 mg of 99 100 cordycepin per mL of solvent). The pH of the formulations in PBS were readjusted using HCl 101 after dissolving cordycepin powder, to readjust the effect of solubilised cordycepin itself on 102 the pH of the formulation. The test suspensions were contained in glass vials and were mixed using a vial roller, overnight at room temperature. The test suspensions were then filtered using 103 Costar Spin-X Centrifuge Tubes (cellulose acetate, 0.22 µm pore size) by centrifugation at 104 2400 g for 5 min at room temperature (Heraeus Fresco 17 Centrifuge, Thermo Electron, MA, 105 106 USA). The filtrate was analysed for the concentration of cordycepin. Solubility of cordycpin 107 in PPG was assessed by preparing saturated suspensions at 20 mg of cordycepin per mL of 108 solvent and sonicating for 15 min. The test suspensions were filtered using the method described above and the filtrates were analysed for cordycepin concentrations. The assay was 109 conducted in quadruplicate. 110

111

112 Formulation preparation

Formulations of cordycepin were prepared based on PBS or PPG. For PBS-based formulations, 113 114 the pH of PBS was adjusted to 4.0, 4.5 and 5.0 using NaOH or HCl prior to addition of cordycepin. PBS-based formulations were prepared by dissolving cordycepin in PBS at pH 4.0, 115 4.5 and 5.0 at concentrations of 5.5, 4.5 and 3.0 mg/mL, respectively. The formulations were 116 117 vortex-mixed and the pH of each formulation was then adjusted to its initial pH using HCl to account for the effect of solubilised cordycepin on the pH of the solution. The vortex-mix and 118 pH re-adjustment steps were repeated until they resulted in clear solutions. For the PPG-based 119 120 formulation, cordycepin was dissolved in PPG at a concentration of 13.0 mg/mL and the formulation was then sonicated in a water bath for 15 min. After confirming that the 121

formulation was a clear solution, deionised-distilled water (DDW) was added to the formulation to give a final concentration of 10.0 mg/mL. This resulted in a PPG-based formulation with a composition of PPG:DDW = 77:23.

125

126 Formulation stability test

Formulations of cordycepin prepared as described above were tested for their stability at four different storage conditions: -80 °C, -20 °C, 4 °C and room temperature. Prepared formulations were aliquoted into 1.5 mL centrifuge tubes and were stored under different conditions. Three samples of each storage condition were withdrawn after 4 and 11 days, or 3, 8 and 12 weeks after preparation to assess the stability of the formulations.

132

133 Filter sterilisation

PBS-based formulations of cordycepin were filter-sterilised using Sartorius Minisart syringe
filters (polyethersulfone, 0.2 µm pore size). Aliquots (700 µL) of the formulations prepared as
described above were filtered through the devices and air was purged using a 1 mL syringe.
Concentrations of cordycepin in the formulations were measured before and after the filter
sterilisation procedure. The evaluation was performed in quadruplicate.

139

140 Analytical method

Analysis of cordycepin concentrations in the samples was performed using a HPLC-UV system consisting of a Waters Alliance 2695 separations module equipped with a Waters 996 photodiode array detector. The autosampler temperature was maintained at 10 °C and the column temperature was set at 40 °C. The stationary phase was a Capcell Pak C18 4.6×150 mm, 3 µm particle size column (Shiseido, Tokyo, Japan), protected by a SecurityGuard 2×4 mm, 3 µm particle size column (Phenomenex, Macclesfield, UK). The mobile phase was an 147 isocratic composition of acetonitrile:DDW at a ratio of 6:94 (v/v). The flow rate of the mobile 148 phase was 0.8 mL/min and the chromatograms were monitored at a wavelength of 259 nm. The 149 injection volume was 40 μ L.

150

151 Sample preparation

For HPLC-UV sample preparation, formulation samples were firstly diluted with DDW to yield 153 1.0 mg/mL of cordycepin (dilution ratios were 18.2:81.8, 22.2:77.8, 33.3:66.7 and 10:90 for 154 formulations of PBS pH 4.0, PBS pH 4.5, PBS pH 5.0 and **PPG-based formulation**, 155 respectively). From these diluted samples, 10 μ L was withdrawn and added to 980 μ L of DDW 156 with 10 μ L of internal standard stock solution (1.0 mg/mL adenosine in DDW). The samples 157 were then vortex-mixed and 100 μ L aliquots were transferred to HPLC vials for analysis.

158

Calibration curve samples were prepared with working standard solutions of cordycepin (in 159 DDW) at concentrations of 5, 10, 25, 50, 100, 250, 500 and 1000 µg/mL. DDW-based diluents 160 were prepared to match the composition of each formulation sample medium (0.187% PBS pH 161 4.0, 0.227% PBS pH 4.5, 0.344% PBS pH 5.0 and 0.079% PPG). Samples were prepared by 162 adding 20 µL of the cordycepin working standard solutions to 970 µL of the DDW-based 163 164 diluents with 10 μ L of internal standard stock solution (1.0 mg/mL adenosine in DDW). The resulting calibration curve points were 0.1, 0.2, 0.5, 1, 2, 5, 10 and 20 µg/mL. The samples 165 166 were then vortex-mixed and 100 µL aliquots were transferred to HPLC vials.

167

168 Validation of the analytical method

Validation of the analytical method was performed for accuracy and precision. Accuracy was
expressed as relative error (RE), which represents the bias from nominal concentrations.
Precision was expressed as relative standard deviation (RSD), which represents the coefficient

of variation at each concentration level (46, 47). Both accuracy and precision were assessed by
intra-day (six replicates in one day) and inter-day (replicates on six different days) variability.
The concentration levels of quality control (QC) samples were the lower limit of quantification
(LLOQ, 0.1 µg/mL), low QC (LQC, 0.25 µg/mL), medium QC (MQC, 1.5 µg/mL) and high
QC (HQC, 15 µg/mL). The QC samples were prepared in the same manner as calibration curve
samples in each formulation sample medium.

178

179 Statistical analysis

All data were expressed as mean \pm standard deviation (SD). Statistically significant differences between measurements in different media were assessed using the unpaired two-tailed t-test. The differences between measurements in one sample before and after sterilisation were assessed using the paired two-tailed t-test. Statistical significance was declared when p < 0.05.

185 **RESULTS**

186 Prior to formulation development, the maximal solubility of cordycepin in water for injection, normal saline, PBS at three different pH levels (4.0, 4.5 and 5.5) and PPG was assessed. The 187 results are shown in Figure 1. The use of PBS at pH 4.0 and PPG resulted in significantly higher 188 189 solubility of cordycepin compared to water or normal saline (both p < 0.001). Following the solubility results, formulations were developed in each medium at concentrations lower than 190 the maximal solubility in the respective medium (Table 2). The PBS-based formulations were 191 192 intended for intravenous and oral administration, while the PPG-based formulation was developed for higher dose oral administration. The achievable dose for each route of 193 administration was calculated based on recommendation from a previous report (42, 43). 194

The analytical method used for the stability assessment of the formulations was fully validated by intra-day and inter-day validation in order to evaluate the reliability of the method. The results in Table 3 show that the method has acceptable precision and accuracy at all QC levels tested. Representative chromatograms from the analytical method are shown in Figure 2.

200

201 Stability of the formulations was assessed under four different storage conditions (-80 °C, -20 °C, 4 °C and room temperature) up to 3 months for PBS-based formulations and up to 3 202 weeks for PPG-based formulations. The results in Table 4 show that all PBS-based 203 formulations were stable at -80 °C, -20 °C and at room temperature for up to 3 months. 204 205 Interestingly, crystallisation of cordycepin was observed in samples stored at 4 °C in PBS-206 based formulations. The rate of crystallisation differed with pH of the formulations, as crystallisation at pH 4.0 and at pH 4.5 in PBS was found after 6 days but at pH 5.0 in PBS only 207 after 11 days. The PPG-based formulation was stable under all four storage conditions up to 3 208 months and crystallisation was not observed in any of the samples. 209

210

The feasibility of simple filter sterilisation of the PBS-based formulations was assessed by measuring the concentration of cordycepin before and after filtration. The results in Table 5 show that there was no statistically significant loss during the filter sterilisation (p > 0.05). Therefore, filter sterilisation can be applied without compromising the concentration of the PBS-based formulations.

216

217 **DISCUSSION**

Knowledge of the maximal solubility of cordycepin in various dosing vehicles is essential information for the development of solution phase-based formulations. Some preclinical studies listed in Table 1 used concentrations that in fact exceed the maximal solubility of cordycepin in the respective dosing vehicles. Therefore, it is likely that the formulations used
in these studies were in a suspension state. As previously mentioned, suspension-based
formulations could complicate the interpretation of the results due to dissolution-related factors
involved following oral or intraperitoneal administration. Intravenous administration of
heterogeneous suspensions can also a cause adverse effects.

226

PBS is an isotonic vehicle that causes minimal discomfort when administered intravenously or orally. The solubility of cordycepin in PBS was highest at pH 4.0, probably due to the fact that cordycepin is a weak base ($pK_a = 3.6$, predicted by ACD/Labs, Toronto, Canada). Decreasing the pH further would likely result in even higher solubility. However, the recommended pH range for intravenous formulations is 4-9, and therefore the lowest pH tested in this work was 4.0 (42).

233

234 A PPG-based formulation was developed in order to enable higher doses of cordycepin to be 235 delivered by the oral route. PPG is a widely-used and a well-tolerated dosing vehicle in 236 preclinical studies. A previous study reported that oral administration of as much as 1000 mg/kg/day of PPG for 90 days was tolerable in rats (48). The solubility of cordycepin in PPG 237 238 was 2.7-fold higher than in pH 4.0 PBS (Figure 1). However, pure PPG as a dosing vehicle can present difficulties in practical use during preclinical experiments due to its high viscosity. 239 Therefore PPG was diluted with DDW to produce the final formulation. As a result, our PPG-240 241 based formulation is composed of PPG:DDW = 77:23 at 10 mg/mL cordycepin and addition of water did not cause any precipitation of cordycepin in this formulation. 242

243

The analytical method used in this study was developed based on a previous report where cordycepin was analysed in extracts of *Cordyceps* using HPLC-UV methods (49). However, this previously reported analytical method was limited by the fact that an internal standard was not used and validation results were not provided. Therefore, in the present study, we incorporated an internal standard to improve the precision and accuracy of the method and we also performed a full validation of the method (Table 3). This HPLC-UV method could provide useful information for future studies where concentrations of cordycepin are to be analysed in other formulations.

Formulations intended for intravenous administration should not only be free of particles but should also be aseptic (42). In this study, filter sterilisation was tested for its feasibility, as it is one of the techniques that does not require heating of the formulation. One concern for filter sterilisation can be that the drug might have non-specific binding to the filter or the device. The results in our case showed that the PBS-based cordycepin formulations can be filter-sterilised without affecting the concentration of the formulations (Table 5).

259

Surprisingly, and rather counterintuitively, we observed that a temperature of 4°C is the only 260 261 storage condition to be avoided for PBS-based formulations. This is probably because the slow cooling process provided upon storage at 4°C resulted in crystallisation of cordycepin. It is 262 263 often reasonable to assume that pharmaceutical formulations are more stable at lower temperatures (50). Moreover, when a label instructs to store a drug below a certain temperature 264 (e.g. "store below 25 °C"), this usually means that storage in the fridge would provide stable 265 266 conditions (51). However, the current study emphasises that storage of the formulations or dosage forms at lower temperature might in fact result in poorer stability from a physical point 267 of view. 268

269

²⁵²

271 CONCLUSION

Herein, we report a study where simple and readily applicable formulations of cordycepin were 272 developed and tested for their stability. An HPLC-UV method for quantification of cordycepin 273 274 in the formulations was developed and validated. Additionally, the PBS-based formulations could be sterilised by filtration without loss of material. The PBS-based formulations were 275 stable for up to 3 months at room temperature, -20°C and -80°C but not at 4°C, which suggests, 276 rather counterintuitively, that storage of the formulations or dosage forms at lower temperature 277 might in fact result in poorer **physical** stability. This study could provide useful information 278 for future preclinical and clinical studies with cordycepin. 279 280 281 ACKNOWLEDGEMENT The research was funded by Arthritis Research UK grant 20795 awarded to CHdM. 282 283 REFERENCES 284 Nakamura K, Shinozuka K, Yoshikawa N. Anticancer and antimetastatic effects of 285 1. cordycepin, an active component of Cordyceps sinensis. J Pharmacol Sci. 2015;127(1):53-6. 286 2. Tuli HS, Sharma AK, Sandhu SS, Kashyap D. Cordycepin: a bioactive metabolite 287 288 with therapeutic potential. Life Sci. 2013;93(23):863-9. 3. Kim HG, Shrestha B, Lim SY, Yoon DH, Chang WC, Shin DJ, et al. Cordycepin 289 inhibits lipopolysaccharide-induced inflammation by the suppression of NF-kappaB through 290 291 Akt and p38 inhibition in RAW 264.7 macrophage cells. Eur J Pharmacol. 2006;545(2-3):192-9. 292 Tuli HS, Sandhu SS, Sharma AK. Pharmacological and therapeutic potential of 293 4. 294 Cordyceps with special reference to Cordycepin. 3 Biotech. 2014;4(1):1-12.

295 5. Lee JH, Yoon JY, Myoung H, Hong SM, Kim SM, Kim MJ. Anti-cancer effects of
296 cordycepin on oral squamous cell carcinoma proliferation and apoptosis in vitro. Oral Oncol.
297 2011;47:S75-S.

Jeong JW, Jin CY, Park C, Hong SH, Kim GY, Jeong YK, et al. Induction of
 apoptosis by cordycepin via reactive oxygen species generation in human leukemia cells.
 Toxicol In Vitro. 2011;25(4):817-24.

301 7. Patel S, Goyal A. Recent developments in mushrooms as anti-cancer therapeutics: a
302 review. 3 Biotech. 2012;2(1):1-15.

303 8. Wong YY, Moon A, Duffin R, Barthet-Barateig A, Meijer HA, Clemens MJ, et al.

304 Cordycepin inhibits protein synthesis and cell adhesion through effects on signal

305 transduction. J Biol Chem. 2010;285(4):2610-21.

306 9. Cho MA, Lee DS, Kim MJ, Sung JM, Ham SS. Antimutagenicity and cytotoxicity of
307 cordycepin isolated from Cordyceps militaris. Food Sci Biotechnol. 2003;12(5):472-5.

308 10. Nakamura K, Konoha K, Yoshikawa N, Yamaguchi Y, Kagota S, Shinozuka K, et al.

309 Effect of cordycepin (3'-deoxyadenosine) on hematogenic lung metastatic model mice. In

310 Vivo. 2005;19(1):137-41.

311 11. Yoo HS, Shin JW, Cho JH, Son CG, Lee YW, Park SY, et al. Effects of Cordyceps

militaris extract on angiogenesis and tumor growth. Acta Pharmacol Sin. 2004;25(5):657-65.

313 12. Sugar AM, McCaffrey RP. Antifungal activity of 3'-deoxyadenosine (cordycepin).

314 Antimicrob Agents Chemother. 1998;42(6):1424-7.

315 13. Shin S, Lee S, Kwon J, Moon S, Lee S, Lee CK, et al. Cordycepin Suppresses

316 Expression of Diabetes Regulating Genes by Inhibition of Lipopolysaccharide-induced

317 Inflammation in Macrophages. Immune Netw. 2009;9(3):98-105.

318 14. Jeong JW, Jin CY, Kim GY, Lee JD, Park C, Kim GD, et al. Anti-inflammatory

effects of cordycepin via suppression of inflammatory mediators in BV2 microglial cells. Int
Immunopharmacol. 2010;10(12):1580-6.

321 15. Noh EM, Kim JS, Hur H, Park BH, Song EK, Han MK, et al. Cordycepin inhibits IL-

322 1 beta-induced MMP-1 and MMP-3 expression in rheumatoid arthritis synovial fibroblasts.

323 Rheumatology. 2009;48(1):45-8.

16. Lee YR, Noh EM, Jeong EY, Yun SK, Jeong YJ, Kim JH, et al. Cordycepin inhibits

325 UVB-induced matrix metalloproteinase expression by suppressing the NF-kappa B pathway

in human dermal fibroblasts. Exp Mol Med. 2009;41(8):548-54.

327 17. Kondrashov A, Meijer HA, Barthet-Barateig A, Parker HN, Khurshid A, Tessier S, et
328 al. Inhibition of polyadenylation reduces inflammatory gene induction. RNA.

329 2012;18(12):2236-50.

18. Cho HJ, Cho JY, Rhee MH, Park HJ. Cordycepin (3'-deoxyadenosine) inhibits human
platelet aggregation in a cyclic AMP- and cyclic GMP-dependent manner. Eur J Pharmacol.
2007;558(1-3):43-51.

333 19. Zhou XX, Luo LP, Dressel W, Shadier G, Krumbiegel D, Schmidtke P, et al.

Cordycepin is an Immunoregulatory Active Ingredient of Cordyceps sinensis. Am J Chinese
Med. 2008;36(5):967-80.

336 20. Zhou XX, Meyer CU, Schmidtke P, Zepp F. Effect of cordycepin on interleukin-10

337 production of human peripheral blood mononuclear cells. European Journal of

338 Pharmacology. 2002;453(2-3):309-17.

339 21. Ma L, Zhang S, Du M. Cordycepin from Cordyceps militaris prevents hyperglycemia
340 in alloxan-induced diabetic mice. Nutr Res. 2015;35(5):431-9.

341 22. de Julian-Ortiz JV, Galvez J, Munoz-Collado C, Garcia-Domenech R, Gimeno-

342 Cardona C. Virtual combinatorial syntheses and computational screening of new potential
343 anti-herpes compounds. J Med Chem. 1999;42(17):3308-14.

344 23. Yoshikawa N, Nakamura K, Yamaguchi Y, Kagota S, Shinozuka K, Kunitomo M.
345 Antitumour activity of cordycepin in mice. Clin Exp Pharmacol Physiol. 2004;31 Suppl
346 2:S51-3.

Pan BS, Wang YK, Lai MS, Mu YF, Huang BM. Cordycepin induced MA-10 mouse
Leydig tumor cell apoptosis by regulating p38 MAPKs and PI3K/AKT signaling pathways.
Sci Rep. 2015;5:13372.

350 25. Sato A, Yoshikawa N, Kubo E, Kakuda M, Nishiuchi A, Kimoto Y, et al. Inhibitory
351 effect of cordycepin on experimental hepatic metastasis of B16-F0 mouse melanoma cells. In
352 Vivo. 2013;27(6):729-32.

Won KJ, Lee SC, Lee CK, Lee HM, Lee SH, Fang Z, et al. Cordycepin attenuates
neointimal formation by inhibiting reactive oxygen species-mediated responses in vascular
smooth muscle cells in rats. J Pharmacol Sci. 2009;109(3):403-12.

27. Cheng Z, He W, Zhou X, Lv Q, Xu X, Yang S, et al. Cordycepin protects against
cerebral ischemia/reperfusion injury in vivo and in vitro. Eur J Pharmacol. 2011;664(1-3):208.

359 28. Yuan J, Wang AH, He Y, Si ZH, Xu S, Zhang SC, et al. Cordycepin attenuates
360 traumatic brain injury-induced impairments of blood-brain barrier integrity in rats. Brain Res
361 Bull. 2016;127:171-6.

362 29. Dou C, Cao Z, Ding N, Hou TY, Luo F, Kang F, et al. Cordycepin Prevents Bone
363 Loss through Inhibiting Osteoclastogenesis by Scavenging ROS Generation. Nutrients.
364 2016;8(4).

365 30. Zhang DW, Deng H, Qi W, Zhao GY, Cao XR. Osteoprotective effect of cordycepin
366 on estrogen deficiency-induced osteoporosis in vitro and in vivo. Biomed Res Int.
367 2015;2015:423869.

368 31. Wang F, Yin P, Lu Y, Zhou Z, Jiang C, Liu Y, et al. Cordycepin prevents oxidative
369 stress-induced inhibition of osteogenesis. Oncotarget. 2015;6(34):35496-508.

370 32. Sun Y, Wang YH, Qu K, Zhu HB. Beneficial effects of cordycepin on metabolic

371 profiles of liver and plasma from hyperlipidemic hamsters. J Asian Nat Prod Res.

372 2011;13(6):534-46.

373 33. Du Y, Yu J, Du L, Tang J, Feng WH. Cordycepin enhances Epstein-Barr virus lytic

374 infection and Epstein-Barr virus-positive tumor treatment efficacy by doxorubicin. Cancer

375 Lett. 2016;376(2):240-8.

376 34. Leu SF, Poon SL, Pao HY, Huang BM. The in vivo and in vitro stimulatory effects of
377 cordycepin on mouse leydig cell steroidogenesis. Biosci Biotechnol Biochem.

378 2011;75(4):723-31.

379 35. Tianzhu Z, Shihai Y, Juan D. The effects of cordycepin on ovalbumin-induced

allergic inflammation by strengthening Treg response and suppressing Th17 responses in

ovalbumin-sensitized mice. Inflammation. 2015;38(3):1036-43.

382 36. Cai ZL, Wang CY, Jiang ZJ, Li HH, Liu WX, Gong LW, et al. Effects of cordycepin
383 on Y-maze learning task in mice. Eur J Pharmacol. 2013;714(1-3):249-53.

384 37. Hu Z, Lee CI, Shah VK, Oh EH, Han JY, Bae JR, et al. Cordycepin Increases

385 Nonrapid Eye Movement Sleep via Adenosine Receptors in Rats. Evid Based Complement

386 Alternat Med. 2013;2013:840134.

387 38. Rottenberg ME, Masocha W, Ferella M, Petitto-Assis F, Goto H, Kristensson K, et al.

388 Treatment of African trypanosomiasis with cordycepin and adenosine deaminase inhibitors in

a mouse model. J Infect Dis. 2005;192(9):1658-65.

- 390 39. Wei HP, Ye XL, Chen Z, Zhong YJ, Li PM, Pu SC, et al. Synthesis and
- 391 pharmacokinetic evaluation of novel N-acyl-cordycepin derivatives with a normal alkyl
- 392 chain. Eur J Med Chem. 2009;44(2):665-9.
- 393 40. Tsai YJ, Lin LC, Tsai TH. Pharmacokinetics of adenosine and cordycepin, a bioactive
 394 constituent of Cordyceps sinensis in rat. J Agric Food Chem. 2010;58(8):4638-43.
- 395 41. Nickerson DF, Weaver ML, Tse FL. The effect of oral dose volume on the absorption
- of a highly and a poorly water-soluble drug in the rat. Biopharm Drug Dispos.
- 397 1994;15(5):419-29.
- 398 42. Turner PV, Pekow C, Vasbinder MA, Brabb T. Administration of substances to
- 399 laboratory animals: equipment considerations, vehicle selection, and solute preparation. J Am
- 400 Assoc Lab Anim Sci. 2011;50(5):614-27.
- 401 43. Turner PV, Brabb T, Pekow C, Vasbinder MA. Administration of substances to
- 402 laboratory animals: routes of administration and factors to consider. J Am Assoc Lab Anim
 403 Sci. 2011;50(5):600-13.
- 404 44. Hind CR. Pulmonary complications of intravenous drug misuse. 1. Epidemiology and
 405 non-infective complications. Thorax. 1990;45(11):891-8.
- 406 45. Wong J, Brugger A, Khare A, Chaubal M, Papadopoulos P, Rabinow B, et al.
- 407 Suspensions for intravenous (IV) injection: a review of development, preclinical and clinical
- 408 aspects. Adv Drug Deliv Rev. 2008;60(8):939-54.
- 409 46. U.S. Department of Health and Human Servies, Food and Drug Administration,
- 410 Guidance for Industry, Analytical Procedures and Methods Validation for Drugs and
- 411 Biologics. 2015.
- 412 47. U.S. Department of Health and Human Servies, Food and Drug Administration,
- 413 Guidance for Industry, Bioanalytical Method Validation. 2001.

| 414 | 48. | Thackaberry EA, Kopytek S, Sherratt P, Trouba K, McIntyre B. Comprehensive |
|-----|---------|--|
| 415 | investi | gation of hydroxypropyl methylcellulose, propylene glycol, polysorbate 80, and |
| 416 | hydrox | cypropyl-beta-cyclodextrin for use in general toxicology studies. Toxicol Sci. |
| 417 | 2010;1 | 17(2):485-92. |
| 418 | 49. | Ikeda R, Nishimura M, Sun Y, Wada M, Nakashima K. Simple HPLC-UV |
| 419 | determ | nination of nucleosides and its application to the authentication of Cordyceps and its |
| 420 | allies. | Biomed Chromatogr. 2008;22(6):630-6. |
| 421 | 50. | Skeist R, Carlson G. Storing medications safely. Geriatr Nurs. 1981;2(6):429-32, 41. |
| 422 | 51. | World Health Organization. Guide to good storage practices for pharmaceuticals. |
| 423 | WHO | Technical Report Series. 2003;908(Annex 9). |

| Literature (Reference) | Dose | Dosing volume | Concentration | Vehicle | Test species | ROA | Indication |
|---------------------------------------|---------------------------|------------------|------------------|--|-----------------|------|----------------------------|
| Won et al., 2009 (26) | 10 mg/kg | 2 mL/kg | 5 mg/mL | Water | Rat | p.o. | Vascular disorder |
| Yoshikawa <i>et al.,</i> 2004 (23) | 5, 15 mg/kg | N.S. | N.S. | Water | Mouse | p.o. | Cancer |
| Cheng <i>et al.</i> , 2011 (27) | 10, 20 mg/kg | N.S. | N.S. | Water | Mouse | p.o. | Neuroprotective activity |
| Rottenberg <i>et al.,</i> 2005 (38) | 2 mg/kg | N.S. | N.S. | N.S. | Mouse | i.p. | African Trypanosomiasis |
| Sugar <i>et al.</i> , 1998 (12) | 1.5 mg/kg | N.S. | N.S. | N.S. | Mouse | i.p. | Antifungal activity |
| Wei et al., 2009 (39) | 500 μmol/kg | N.S. | N.S. | 2% Tween 80 | Mouse | p.o. | Prodrug studies |
| Leu et al., 2011 (34) | 20, 40 mg/kg | 1 mL/mouse | 0.6, 1.2 mg/mL | Saline | Mouse | i.p. | Steroidogenesis |
| Tsai et al., 2010 (40) | 10 mg/kg | 1 mL/kg | 10 mg/mL | Saline | Rat | i.v. | Pharmacokinetic studies |
| Sato et al., 2013 (25) | 0.5, 5 mg/kg | N.S. | N.S. | DPBS | Mouse | i.p. | Cancer metastasis |
| Zhang <i>et al.</i> , 2015 (30) | 22.2, 44.4, 88.9 mg/kg | 10 mL | 0.5, 1, 2 mg/mL | Water | Rat | p.o. | Osteoporosis |
| Sun et al., 2011 (32) | 140 mg/kg | N.S. | N.S. | 2.5% carboxymethyl cellulose sodium | Hamster | p.o. | Hyperlipidaemia |
| Du et al., 2016 (33) | 50 mg/kg | N.S. | N.S. | N.S. | Mouse | i.p. | Epstein-Barr virus |
| Tianzhu <i>et al.</i> , 2015 (35) | 20, 40 mg/kg | N.S. | N.S. | N.S. | Mouse | p.o. | Allergic asthma |
| Ma et al., 2015 (21) | 8, 24, 72 mg/kg | N.S. | N.S. | Saline | Mouse | i.p. | Hyperglycaemia |
| Cai et al., 2013 (36) | 5, 10, 20 mg/kg | N.S. | N.S. | Water | Mouse | p.o. | Cognitive function |
| Hu et al., 2013 (37) | 2, 4 mg/kg | N.S. | N.S. | Saline | Rat | N.S. | Sleep regulation |
| Pan et al., 2015 (24) | 20 mg/kg | N.S. | 0.1 % cordycepin | PBS | Mouse | N.S. | Cancer |
| Wang <i>et al.</i> , 2015 (31) | 1, 5, 10, 20 mg/kg | N.S. | N.S. | N.S. | Mouse | i.p. | Osteogenesis |
| Dou et al., 2016 (29) | 10 mg/kg | N.S. | N.S. | Water | Mouse | p.o. | Osteoporosis |
| Yuan <i>et al.</i> , 2016 (28) | 1, 5, 10, 20 mg/kg | N.S. | N.S. | Saline | Rat | i.v. | Traumatic brain injury |

Table 1. List of preclinical studies conducted with cordycepin

N.S., not specified; p.o., oral; i.p., intraperitoneal; i.v., intravenous.

Table 2. Composition, concentration of developed formulations of cordycepin and their achievable dose in various routes of administration

| | | Cordycepin | Achievable dose (mg/kg) ^a | | |
|-----------|-----------------|----------------------------|--------------------------------------|--------------------|--|
| | Composition | concentration – (mg/mL) | Intravenous (5 mL/kg) | Oral (20 mL/kg) | |
| | pH 4.0 PBS | 5.5 | 27.5 | 110.0 | |
| PBS-based | pH 4.5 PBS | 4.5 | 22.5 | 90.0 | |
| | pH 5.0 PBS | 3.0 | 15.0 | 60.0 | |
| PPG-based | PPG:DDW = 77:23 | 10.0 | - | 200.0 | |

^a Achievable dose in each route of administration calculated according to recommendation from reference (42, 43).

| | Intra | a-day | Inte | r-day |
|----------------------------|---------------------|-----------------------|---------------------|-----------------------|
| - Concentration levels | Accuracy (RE, %) | Precision (RSD, %) | Accuracy (RE, %) | Precision (RSD, %) |
| Samples of pH 4.0 PBS | | | | |
| LLOQ (0.1 µg/mL) | 10.45 | 3.45 | 1.76 | 6.89 |
| LQC (0.25 µg/mL) | 1.98 | 2.77 | -2.29 | 4.40 |
| MQC (1.5 µg/mL) | 4.00 | 0.71 | -1.11 | 6.15 |
| HQC (15 μg/mL) | 0.80 | 0.76 | 1.44 | 3.64 |
| Samples of pH 4.5 PBS | | | | |
| LLOQ (0.1 µg/mL) | 8.95 | 2.85 | 6.54 | 4.01 |
| LQC (0.25 µg/mL) | 2.83 | 0.76 | -0.53 | 3.48 |
| MQC (1.5 µg/mL) | 2.56 | 1.50 | -2.69 | 3.43 |
| HQC (15 μg/mL) | 2.28 | 0.57 | -1.46 | 2.44 |
| Samples of pH 5.0 PBS | | | | |
| LLOQ (0.1 µg/mL) | 7.37 | 6.26 | 4.76 | 9.16 |
| LQC (0.25 µg/mL) | 4.36 | 2.49 | 0.55 | 5.65 |
| MQC (1.5 µg/mL) | 6.73 | 5.56 | -1.13 | 6.34 |
| HQC (15 μg/mL) | 1.08 | 0.93 | -2.42 | 4.70 |
| Samples of PPG:DDW = 77:23 | | | | |
| LLOQ (0.1 µg/mL) | 1.48 | 2.96 | 1.07 | 4.87 |
| LQC (0.25 µg/mL) | 2.43 | 2.98 | 2.22 | 3.14 |
| MQC (1.5 µg/mL) | -2.63 | 2.05 | -1.81 | 2.03 |
| HQC (15 µg/mL) | 2.70 | 2.66 | 4.06 | 1.99 |

Table 3. Intra-day and inter-day validation of the analytical method in each formulation sample medium (n = 6)

| Formulation | Storage condition | 4 days | 11 days | 3 weeks | 2 months | 3 months |
|-----------------|-------------------|-----------------|-----------------|-----------------|------------------|-----------------|
| | RT | 98.8 ± 0.2 | 99.9 ± 1.8 | 97.7 ± 2.5 | 102.8 ± 3.7 | 104.8 ± 2.7 |
| pH 4.0 PBS | 4°C | 97.2 ± 2.0 | 88.8 ± 2.2 | 67.3 ± 3.8 | 72.2 ± 9.8 | 62.0 ± 8.0 |
| (5.5 mg/mL) | -20°C | 98.0 ± 0.4 | 97.5 ± 1.7 | 100.5 ± 4.7 | 99.4 ± 1.2 | 101.3 ± 3.4 |
| | -80°C | 98.4 ± 0.8 | 98.2 ± 0.5 | 99.1 ± 9.2 | 99.6 ± 1.4 | 98.8 ± 8.4 |
| | RT | 93.9 ± 0.9 | 99.7 ± 1.2 | 101.9 ± 1.2 | 101.3 ± 0.4 | 97.4 ± 5.2 |
| pH 4.5 PBS | 4°C | 96.0 ± 3.8 | 72.0 ± 9.5 | 76.8 ± 11.1 | 41.7 ± 26.0 | 70.4 ± 18.4 |
| (4.5 mg/mL) | -20°C | 101.3 ± 4.8 | 98.7 ± 1.1 | 99.6 ± 1.2 | 94.0 ± 3.1 | 96.3 ± 4.5 |
| | -80°C | 100.2 ± 7.8 | 99.9 ± 2.9 | 100.8 ± 1.1 | 109.8 ± 11.5 | 99.0 ± 6.2 |
| | RT | 96.9 ± 0.7 | 100.5 ± 2.7 | 102.8 ± 3.8 | 106.2 ± 2.5 | 106.8 ± 1.2 |
| pH 5.0 PBS | 4°C | 96.6 ± 1.4 | 87.4 ± 10.6 | 97.2 ± 11.8 | 85.2 ± 17.7 | 66.5 ± 4.2 |
| (3.0 mg/mL) | -20°C | 98.0 ± 2.8 | 99.2 ± 3.0 | 102.7 ± 5.3 | 110.4 ± 10.5 | 99.7 ± 2.2 |
| | -80°C | 97.4 ± 0.9 | 99.5 ± 1.1 | 104.6 ± 0.3 | 108.3 ± 10.1 | 99.4 ± 7.0 |
| | RT | 102.1 ± 2.6 | 104.7 ± 1.6 | 106.0 ± 3.7 | 109.9 ± 1.0 | 104.0 ± 2.4 |
| PPG:DDW = 77:23 | 4°C | 100.6 ± 0.2 | 104.1 ± 3.4 | 106.2 ± 0.5 | 105.5 ± 3.9 | 107.3 ± 0.8 |
| (10.0 mg/mL) | -20°C | 99.9 ± 1.5 | 104.9 ± 1.0 | 106.3 ± 0.7 | 102.4 ± 3.6 | 106.8 ± 2.0 |
| | -80°C | 98.8 ± 1.3 | 103.8 ± 1.0 | 105.4 ± 2.4 | 104.9 ± 3.5 | 105.3 ± 0.6 |

Table 4. Stability of cordycepin formulation samples (mean \pm SD, n = 3)

RT, room temperature.

Table 5. Effects of filter sterilisation on concentrations of PBS-based cordycepin formulations (mean \pm SD,n = 4)

| | Cordycepin conce | | |
|-------------|----------------------|---------------------|------------------------------------|
| Formulation | Before sterilisation | After sterilisation | % remaining after sterilisation |
| pH 4.0 PBS | 5.52 ± 0.03 | 5.41 ± 0.20 | 98.09 ± 3.35 |
| pH 4.5 PBS | 4.45 ± 0.02 | 4.35 ± 0.11 | 97.91 ± 2.96 |
| pH 5.0 PBS | 3.02 ± 0.05 | 2.95 ± 0.06 | 97.77 ± 1.85 |

LEGENDS TO FIGURES

Figure 1. Solubility of cordycepin in different media (mean \pm SD, n = 4). ***, *p*<0.001 compared to water and saline by unpaired two-tailed t-test.

Figure 2. Representative chromatograms of cordycepin and internal standard (IS, adenosine) from HPLC-UV analysis. A, baseline chromatogram observed after injection of 100% DDW;
B, calibration curve sample spiked with 10 ug/mL cordycepin and IS into pH 4.5 PBS-based medium; C, MQC sample in PPG-based medium; D, stability test sample of pH 4.0 PBS formulation after 3 weeks of storage at 4°C.



