

1 Similarity- and Substructure-Based Development of β_2 -Adrenergic 2 Receptor Ligands Based on Unusual Scaffolds

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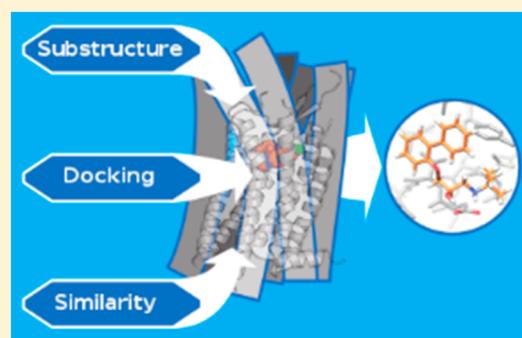
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8 **S** Supporting Information

9 **ABSTRACT:** The β_2 -adrenergic receptor (β_2 AR) is a G protein-coupled
10 receptor (GPCR) and a well-explored target. Here, we report the discovery
11 of 13 ligands, ten of which are novel, of this particular GPCR. They have
12 been identified by similarity- and substructure-based searches using
13 multiple ligands, which were described in an earlier study, as starting
14 points. Of note, two of the molecules used as queries here distinguish
15 themselves from other β_2 AR antagonists by their unique scaffold. The
16 molecules described in this work allow us to explore the ligand space
17 around the previously reported molecules in greater detail, leading to
18 insights into their structure–activity relationship. We also report
19 experimental binding and selectivity data and putative binding modes for
20 the novel molecules.

21 **KEYWORDS:** β_2 -adrenergic receptor, similarity searches, docking, SAR-by-catalog



22 **T**he membrane receptors of the G protein-coupled receptor
23 (GPCR) family are flexible heptahelical bundles trans-
24 ferring signals from the outside to the inside of a cell. This is
25 achieved by a conformational change of the receptor upon
26 binding of a signaling molecule to a cavity located at the
27 extracellular end between the seven helices. GPCRs are
28 expressed in almost all tissues,¹ and it is thus not surprising
29 that approximately 1/3 of present-day drugs interact with a
30 GPCR.² Among these receptors, the β_2 -adrenergic receptor
31 (β_2 AR) is considered a prototypical representative and has been
32 investigated for more than 60 years. It was also the first
33 pharmacologically relevant GPCR to succumb to crystallization
34 in 2007.^{3,4}

35 In a previous work,⁵ we have identified six ligands (originally
36 labeled 1–6, and referred to as Q1–Q6 in this work to avoid
37 confusion, Chart S1) of the β_2 AR through *in silico* docking
38 studies, with affinities ranging from 9 nM to 3.2 μ M. Notably,
39 these included two molecules (5 and 6 in ref 5, denoted as Q5
40 and Q6, respectively, in the following) that did not follow the
41 classical adrenaline-based scaffold.⁶ This was remarkable, as
42 nobody had discovered these scaffolds earlier, despite more
43 than six decades of medicinal chemistry in this area. Building
44 upon the discovery of the six ligands, we wanted to expand
45 chemical space around them. In particular, we wanted to
46 investigate the two ligands with unusual scaffolds by employing
47 *in silico* similarity and substructure searches in the ZINC⁷
48 database. Candidate molecules identified in either way were
49 then docked into the β_2 AR, in order to ascertain that their
50 binding modes were consistent. Here we report the results of

this combined ligand- and structure-based screen, which also
51 provides insights into the structure–activity relationship (SAR)
52 of molecules Q5 and Q6 and their derivatives. 53

The similarity screen among the 8.5 million molecules of the
54 ZINC database resulted in 6363 molecules, which were
55 distributed across the six query molecules as shown in Table
56 S1. From the substructure-based screen, approximately 653 000
57 hits emerged. Duplicates were removed from both sets. After
58 docking, 5838 and 587 099 molecules remained, respectively,
59 and the top-scoring 500 of each run were visually inspected.
60 After weeding out molecules with artificially inflated scores due
61 to the absence of corrective terms in present-day scoring
62 functions, e.g., unfavorable desolvation contributions or
63 unsatisfied hydrogen-bond donors, during this inspection, we
64 were left with eight and nine molecules from the similarity and
65 substructure searches, respectively. These were acquired from
66 their respective vendors for further experimental testing (Table
67 S5). Three compounds (1, 2, and 3) contained a biaryl moiety
68 and a charged amine and thus resembled the classical motif of a
69 β_2 binder. Indeed, a thorough literature search revealed that
70 these compounds had been described before (Table 1; by the
71 time of selection, these compounds had not been annotated in
72 ChEMBL⁸). To analyze the selectivity of the compounds, we
73 also evaluated them against the closely related β_1 AR. The 74

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Table 1. Affinity (K_D Values) and β_2 -Selectivity for Compounds as Measured by [^3H](–)CGP 12177 Whole Cell Binding to CHO- β_1 and CHO- β_2 Cells; Values Are Mean \pm SEM of n Separate Experiments

ID	Structure	$\beta_2\text{AR pK}_D$		n	$\beta_1\text{AR pK}_D$		n	β_2
1 ^c		5.42	± 0.14	5	4.34	± 0.07	4	12.0
2 ^c		5.58	± 0.06	6	4.56	± 0.06	6	10.5
3 ^d		10.45	± 0.05	8	9.01	± 0.04	5	27.5
4		4.63	± 0.07	5	4.01 ^b	± 0.05	5	4.2
5		4.41	± 0.08	3	3.59 ^b	± 0.1	3	6.6
6		4.76	± 0.09	5	4.58	± 0.03	5	1.5
7		4.66	± 0.16	5	4.35	± 0.04	4	2
8		4.60 ^b	± 0.11	4	4.33 ^b	± 0.05	4	1.9
9		4.84 ^b	± 0.13	4	4.42 ^b	± 0.11	4	2.6
10		6.05	± 0.11	6	5.51	± 0.07	6	3.5
11		5.31	± 0.12	6	4.86	± 0.05	5	2.8
12		4.75 ^b	± 0.12	5	n.c.		4	
13		5.26	± 0.06	6	4.45	± 0.04	5	6.5
ICI 118551		9.61	± 0.05	5	6.74	± 0.01	5	741
CGP 20712A		5.84	± 0.10	5	8.96	± 0.13	4	0.0008

Table 1. continued

^aSelectivity: $\beta_2/\beta_1 = K_D(\beta_2)/K_D(\beta_1)$ ^b Apparent K_D values: here the maximum concentration of the compound was not sufficient to fully inhibit specific binding; however, the majority of specific binding was inhibited allowing an apparent measure of affinity. For ligands with less than 50% inhibition of specific binding, the IC_{50} value could not be determined and thus a K_D value could not be calculated (n.c.). ^cUS 20090163545. ^dAntiarrhythmic pharmaceutical (Bipranol/Berlafenone), Arzneimittel-Forschung 1992, 42, 289–291.

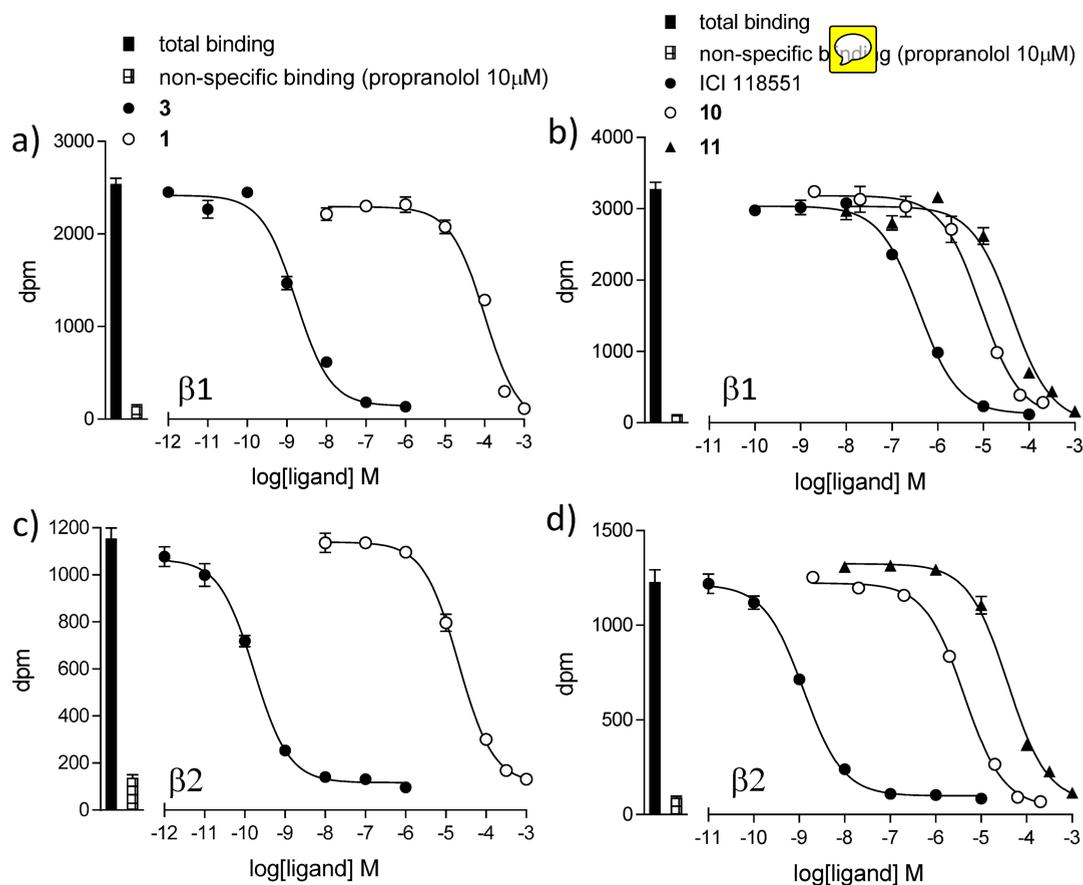


Figure 1. Inhibition of [³H](–)CGP 12177 whole cell binding to (a,b) CHO-β₁ cells and (c,d) CHO-β₂ cells in response to (a,c) 3 and 1 and (b,d) ICI 118551, 10, and 11. Bars represent total and nonspecific binding, and data points are mean ± SEM of triplicate determinations. The concentration of [³H](–)CGP 12177 used in these experiments was (a,c) 0.58 nM and (b,d) 0.44 nM, and they are representative of (a) 4, (b) 5, (c) 5, and (d) 5 separate experiments.

75 efficacy of all compounds was further evaluated in a functional
76 assay.

77 Several of the compounds identified in this work inhibited
78 [³H](–)CGP 12177 whole cell binding (Table 1; see
79 Supporting Information for assay validation and Table S2 for
80 inactive compounds). This assay also demonstrated that
81 compound 3 had very high affinity (pK_D 9.01 at β₁AR and
82 pK_D 10.45 at β₂AR) and was therefore 28-fold β₂-selective
83 (Figure 1a,c, Table 1). While the remaining compounds had
84 relatively poor affinity in comparison to 3, many of them, e.g.,
85 1, 2, 10, 11 and 13, inhibited [³H](–)CGP 12177 binding to
86 yield measurable affinity values (Figure 1b,d, Table 1).

87 Next, characteristics of ligands were examined in a functional
88 assay, namely, CRE-gene transcription. The ability of ligands to
89 stimulate a response (intrinsic efficacy) was assessed, but also,
90 given that the affinity of many of the ligands to inhibit
91 [³H](–)CGP 12177 binding were at the very limit of the
92 binding assay, the ability of ligands to inhibit functional
93 responses was also evaluated, thus giving a totally independent
94 measure of affinity from that achieved in the binding assay.

Except for compound 3, no other compound stimulated a 95
measurable response ($n = 4–5$ for each compound) in this 96
assay (see Supporting Information for more details and assay 97
validation). However, several compounds antagonized the 98
cimaterol response to give a parallel shift of the cimaterol 99
concentration response curve and thus yield measurable K_D 100
values (Figure S1, Table S3). For some compounds, e.g., 1, 2, 101
and 13, this gave selectivity values similar to those obtained in 102
the binding assay. For other compounds, e.g., 16 and 17, no 103
rightward shift of the cimaterol response was observed, 104
suggesting no inhibition at the maximum concentration 105
possible (100 μM in each case). For few of the ligands, the 106
highest concentration possible caused a marked fall in CRE- 107
SPAP production to below basal in a manner more consistent 108
with toxicity, cell death, or assay interference, rather than 109
receptor-mediated inverse agonism (see Supporting Informa- 110
tion for full details). In these instances, compound concentra- 111
tions used to inhibit cimaterol responses were reduced until 112
such a time as the reduction in basal was minimal. An example 113
of this was compound 10, which reduced basal at the maximum 114
concentration of 20 μM but not at 2 μM (see Supporting 115

Information). At 2 μM , **10** was still able to cause a rightward shift of the cimaterol concentration response curve at the $\beta_2\text{AR}$, but not the $\beta_1\text{AR}$, consistent with its β_2 -selectivity. The fall from maximum of the concentration response to cimaterol (most likely because the assay is at the limit of its capability) means that an apparent K_D is reported (calculated from the shift of the lower part of the curve where the lines are parallel), this apparent K_D is however similar to the K_D values obtained from the binding assay, confirming that this is receptor-mediated and β_2 -selective.

Compound **3** on its own stimulated a partial agonist response at both the β_1 - and $\beta_2\text{AR}$. This response was inhibited by CGP 20712A in the CHO- β_1 -cells with high affinity and by ICI 118551 in the CHO- β_2 -cells (Figure S4, Table S3). Furthermore, **3** was able to inhibit the cimaterol responses in both cell lines in a manner consistent with that of a partial agonist (Figure S2, Table S3). Finally, **3** inhibited the response to fixed concentrations of cimaterol in both cell lines in a manner consistent with competition at a single receptor conformation⁹ (Figure S4 and Supplementary Procedures for full details).

Altogether, the high affinity of CGP 20712A and ICI 118551 for the CHO- β_1 and CHO- β_2 cells confirm the presence of the β_1 - and $\beta_2\text{AR}$ in the respective cell lines. Several of the compounds (e.g., **16** and **17**) did not interact with the receptors in either the binding assay or functional assay up to the maximum concentration possible for the compounds (20–100 μM). Of the molecules with novel scaffolds, **10** and **11** show the highest affinities at $\text{p}K_D$ values of 6.05 and 5.31, respectively, for the $\beta_2\text{AR}$ and are thus in a range comparable to those of the established compounds **1** and **2**. These compounds did not induce a functional response in the receptor and are therefore neutral antagonists. However, we emphasize that the outcome of a virtual screening campaign in the manner conducted here is the prediction of binding, not efficacy. Of the novel compounds, **13** exhibited affinity in the binding as well as in the functional assay with low micromolar activity.

The more traditional biaryl compounds **1**, **2**, and **3** display the highest affinities at the $\beta_2\text{AR}$, as was to be expected. In particular, compound **3** was confirmed as a very high affinity partial agonist at both receptors, but with some $\beta_2\text{AR}$ selectivity. At the $\beta_2\text{AR}$, the affinity measured by binding ($\text{p}K_D$ 10.45) and the affinity measured as antagonism of the cimaterol response ($\text{p}K_D$ 10.74) are very similar, confirming the very high affinity ligand–receptor interaction. The partial agonist was itself antagonized by ICI 118551 (yielding a similar $\text{p}K_D$ for ICI 118551 as that for antagonism of the cimaterol response), confirming that signaling is indeed occurring via the $\beta_2\text{AR}$. Compound **3** is therefore a very high affinity, weak partial agonist of the human $\beta_2\text{AR}$. Moreover, **3** was found to be a partial agonist of the $\beta_1\text{AR}$, with the agonist response occurring through the primary catecholamine conformation of the receptor (see Supplementary Results).

These three molecules, **1**, **2**, and **3**, were selected by similarity to compounds **Q2**, **Q3**, and **Q4**, all of which contain a biaryl moiety. Not unexpectedly, these hits not only show high affinities but also highest similarities to known (again exclusively biaryl-containing) compounds that are annotated in the ChEMBL database (Table S6). This is encouraging with respect to the performance of similarity screening methods and the value of docking in identifying such compounds. However, it also strongly emphasizes the need for methods that allow for scaffold-hopping to fully explore the ligand space of a target.

By reducing the biaryl scaffold to a 2-ethoxy-ethylamine (**S6** in Chart S2) for the substructure search, two more substances, **4** and **14**, were identified. Compound **4** showed two-digit micromolar affinity, whereas the inhibition by **14** was so weak that no reliable affinity value could be calculated. Interestingly, in **14** the nitrogen matched in the substructure search is the one in the benzoxazine portion, not the exocyclic amine.

Turning to the hits derived from reference molecules **Q5** and **Q6**, we note that they show a much lower Tanimoto similarity of approximately 0.3 and below (when compared to molecules from the ChEMBL database using ECFP4 fingerprints) than the other hits reported in ref 5 (Table S6). This is in line with the fact that these compounds are not based on the classical propanolamine scaffold and underlines the structural novelty of these two scaffolds.

Starting from the benzothiazole-based compound **Q5**, six molecules were identified with benzothiazole (**5**, **10**, **11**, **15**) and benzimidazole (**16**, **17**) motifs. Of these, all benzothiazole-containing molecules except **15** show affinity toward the $\beta_2\text{AR}$ in the micromolar range. Docking poses indicate that the orientation of the benzothiazole ring is comparable to the one of **Q5**, with a polarized methyl group interacting with Asp113^{3,32} (Figures S5 and S6). The benzimidazole compounds **16** and **17** show no activity in our assay. These compounds might be more sterically hindered in the vicinity of the positively charged nitrogen atom, in particular compound **16**. Furthermore, the different polarity of the ring system, owing to the variation of the heteroatoms, might render the predicted interaction with Asp113^{3,32} less likely.

Six additional compounds could be identified on the basis of the parent molecule **Q6**. All these molecules (**6**, **7**, **8**, **9**, **12**, and **13**) share a benzofuran-based moiety, independent of whether they originated from the substructure or the similarity search. This moiety, namely, a 3-oxo-4-methyl-6-hydroxy-benzofuran, is present in the parent molecule **Q6**, too, and can thus be considered a “stable scaffold” in terms of SAR. All molecules display affinity, with $\text{p}K_D$ values varying between 5.26 and 4.6. Interestingly, **8**, which is the substance with the weakest affinity in this set, differs from **7** only by a methoxy group, which is absent in **8**. This methoxy group could act as an acceptor, which is also present in all remaining molecules of this series as (benzo-)furan or methoxy group. The role of this group is not clearly evident from the docking predictions, but an interaction with Thr195^{ECL2} seems to be the most likely explanation (Figures S5 and S6). Furthermore, the docking poses indicate a binding mode of this scaffold, which resembles the key interactions seen in biaryl-based compounds. The benzofuran scaffold forms interactions with Phe193^{45,52}, Phe289^{6,51}, Phe290^{6,52}, and Val114^{3,33}. The hydroxy group at position 6 forms an additional hydrogen bond to Asp113^{3,32}, while the ketone serves as acceptor for a hydrogen bond from Ser203^{5,42}. A second aromatic moiety is attached at position 2, interacting with Tyr199^{5,38}, Tyr308^{7,35}, and, presumably, Thr195^{ECL2}. An increased size of the aromatic system appears to be detrimental for affinity (methoxyphenyl in **13** vs benzofuran in **9**). The charged amine in the pyrrolidine moiety is expected to form a salt bridge with Asp113^{3,32}.

We have elaborated on six previously identified novel binders of the $\beta_2\text{AR}$ through SAR-by-catalog. Using similarity and substructure searches followed by a docking assessment of the interactions of each compound and the receptor, 13 ligands of the $\beta_2\text{AR}$ were verified experimentally. Ten of these molecules are indeed novel ligands for the receptor, while the remaining 241

242 three turned out to have been described before. Based on this
243 data, several conclusions can be drawn.

244 First, the benzofuran scaffold of compound **Q5** and the
245 benzothiazole scaffold of compound **Q6** in ref **5** indeed
246 constitute novel chemotypes with derivatization potential for
247 this receptor. Especially the benzofuran series showed a
248 consistent SAR that is in agreement with the predicted binding
249 modes. This study can thus also provide retrospective evidence
250 that the predicted binding modes are indeed very likely correct.
251 The affinities of the novel compounds are not comparable with
252 those of highly optimized adrenaline- or biaryl-based scaffolds.
253 The latter are exemplified by **Q1** with an affinity of 9 nM and **3**
254 with its pK_D of 10.74. However, the novel compounds can serve
255 as unprecedented starting points for further optimization.

256 Second, that the combination of similarity- and substructure-
257 based searches with protein-structure-based docking constitutes
258 a powerful combination. This is manifest in the quite high hit
259 rate (more than 75% of the molecules bind with an affinity
260 below 100 μ M) and the fact that we (re)discovered a molecule
261 with an affinity of only 35 pM. This compound is also known as
262 *bipranolol* or *berlafenone*, an antiarrhythmia drug.

263 In terms of selectivity, most of the compounds displaying an
264 affinity are mildly selective toward the β_2 AR. Again, **3** takes the
265 lead here at 28-fold selectivity for the β_2 AR. While other
266 compounds such as **1** and **2** still have at least 10-fold preference
267 toward the β_2 AR, all values are far below 100-fold, which for
268 some receptors is considered a ratio that is significant enough
269 to call a compound "selective". Moreover, highly optimized
270 compounds such as ICI 118551 show affinity ratios that are closer
271 to 1000-fold. Interestingly, the top three compounds in terms
272 of selectivity all belong to the biaryl cluster of molecules.

273 Not unexpectedly, most of the compounds with measurable
274 affinity (with the exception of **3**), turned out to be neutral
275 antagonists in the functional assay. This is consistent with what
276 we have seen in our previous study⁵ and the fact that we have
277 been docking to an inactive conformation of the receptor.^{3,4}

278 Future studies will show to which affinities the novel
279 scaffolds can be optimized. It is also encouraging to have
280 confirmed that unbiased computational methods can present us
281 with novel molecules, even for target proteins as well-
282 investigated as the β_2 AR.

283 ■ EXPERIMENTAL PROCEDURES

284 Substructure queries (Chart S2) were manually derived from the
285 original hits. Substructure and similarity searches were run on the
286 ZINC database⁷ and docked to the β_2 AR (PDB 2RH1), as previously
287 described.⁵ [³H](−)CGP 12177 whole cell binding and CRE-SPAP
288 production assays were run using CHO-K1 cells expressing either the
289 human β_1 AR or the human β_2 AR as previously described.^{10,11} See
290 Supporting Information for detailed descriptions of experimental
291 procedures.

292 ■ ASSOCIATED CONTENT

293 ● Supporting Information

294 The Supporting Information is available free of charge on the
295 ACS Publications website at DOI: 10.1021/acsmchem-
296 lett.6b00363.

297 Tables of similar compounds, SMILES codes for all
298 compounds, detailed experimental methods, Supplemen-
299 tary Figures and Charts (PDF)

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Author Contributions

P.K. did the original similarity and substructure searches and
docking calculations. D.S. and J.G. acquired compounds,
prepared assay-ready formats, and supervised initial affinity
measurements. J.G.B. performed pharmacological experiments
and data analysis. P.K., D.S., and J.G. discussed SAR, and D.S.,
J.G., J.G.B., and P.K. wrote the manuscript.

Notes

The authors declare no competing financial interest.

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