

PqsBC, a Condensing Enzyme in the Biosynthesis of the *Pseudomonas aeruginosa* Quinolone Signal

CRYSTAL STRUCTURE, INHIBITION, AND REACTION MECHANISM[✉]◆

Received for publication, December 7, 2015, and in revised form, January 11, 2016 Published, JBC Papers in Press, January 25, 2016, DOI 10.1074/jbc.M115.708453

 Steffen Lorenz Drees[‡], Chan Li[§], Fajar Prasetya[§], Muhammad Saleem[§], Ingrid Dreveny[§], Paul Williams[¶], Ulrich Hennecke^{||}, Jonas Emsley^{§1}, and Susanne Fetzner^{‡2}

 From the [‡]Institute for Molecular Microbiology and Biotechnology and ^{||}Organic Chemistry Institute, University of Münster, D-48149 Münster, Germany and the Centre for Biomolecular Sciences, Schools of [§]Pharmacy and [¶]Life Sciences, University of Nottingham, Nottingham NG7 2RD, United Kingdom

Pseudomonas aeruginosa produces a number of alkylquinolone-type secondary metabolites best known for their antimicrobial effects and involvement in cell-cell communication. In the alkylquinolone biosynthetic pathway, the β -ketoacyl-(acyl carrier protein) synthase III (FabH)-like enzyme PqsBC catalyzes the condensation of octanoyl-coenzyme A and 2-aminobenzoylacetate (2-ABA) to form the signal molecule 2-heptyl-4(1H)-quinolone. PqsBC, a potential drug target, is unique for its heterodimeric arrangement and an active site different from that of canonical FabH-like enzymes. Considering the sequence dissimilarity between the subunits, a key question was how the two subunits are organized with respect to the active site. In this study, the PqsBC structure was determined to a 2 Å resolution, revealing that PqsB and PqsC have a pseudo-2-fold symmetry that unexpectedly mimics the FabH homodimer. PqsC has an active site composed of Cys-129 and His-269, and the surrounding active site cleft is hydrophobic in character and approximately twice the volume of related FabH enzymes that may be a requirement to accommodate the aromatic substrate 2-ABA. From physiological and kinetic studies, we identified 2-aminoacetophenone as a pathway-inherent competitive inhibitor of PqsBC, whose fluorescence properties could be used for *in vitro* binding studies. In a time-resolved setup, we demonstrated that the catalytic histidine is not involved in acyl-enzyme formation, but contributes to an acylation-dependent increase in affinity for the second substrate 2-ABA. Introduction of Asn into the PqsC active site led to significant activity toward the desamino substrate analog benzoylacetate, suggesting that the substrate 2-ABA itself supplies the asparagine-equivalent amino function that assists in catalysis.

Pseudomonas aeruginosa is a pathogenic Gram-negative bacterium that inhabits soil and aquatic environments. *P.*

aeruginosa infections are often nosocomial and usually associated with compromised host defenses (1). The production of many *P. aeruginosa* virulence factors is controlled by quorum sensing (QS),³ a mechanism where bacteria communicate by sensing self-generated signaling molecules. QS systems allow bacterial populations to synchronize their behavior and so to act cooperatively (2). Because QS controls the virulence of many pathogenic bacteria, QS receptor function and QS signal generation have been proposed as alternative targets for anti-virulence drug development (3–7).

The QS network of *P. aeruginosa* is highly complex, comprising several signaling circuits that are interconnected (8). The Las and Rhl circuits use specific *N*-acylhomoserine lactones as signal molecules, whereas the Pqs circuit employs the alkylquinolone (AQ) signals 2-heptyl-4(1H)-quinolone (HHQ) and 2-heptyl-3-hydroxy-4(1H)-quinolone (*Pseudomonas* quinolone signal (PQS)). AQ signaling influences biofilm development and the production of a number of virulence factors, such as pyocyanin, siderophores, rhamnolipid biosurfactant, the cytotoxic lectin LecA, and elastase LasB (9, 10). Moreover, PQS induces membrane vesicle formation (11), acts as ferric iron chelator (12, 13) and pro-oxidant (14), and exerts host immune modulatory and pro-apoptotic activities (15–17).

Biosynthesis of HHQ and PQS from anthranilic acid and fatty acid precursors is initiated by the coenzyme A (CoA) ligase PqsA, which catalyzes the activation of anthranilic acid to anthraniloyl-CoA (18). A subsequent condensation reaction with malonyl-CoA, catalyzed by PqsD, yields the highly unstable intermediate 2-aminobenzoylacyl-CoA (2-ABA-CoA); however, PqsD also uses malonyl-acyl carrier protein (ACP) as substrate (19). Although 2-ABA-CoA is highly susceptible to spontaneous cyclization to form 2,4-dihydroxyquinoline (DHQ) (19–21), *in vivo* this is counterbalanced by the activity of PqsE, which acts as a 2-ABA-CoA thioesterase to release 2-aminobenzoylacetate (2-ABA) (21). 2-ABA is another branching point in the pathway and can undergo decarboxylation to 2-aminoacetophenone (2-AA) (20), a secondary metab-

* This work was supported by Deutsche Forschungsgemeinschaft Grant FE 383/23-1 (to S. F.) and by a Ph.D. studentship (to F. P.) from the Directorate General of Indonesia Higher Education. The authors declare that they have no conflicts of interest with the contents of this article.

✂ Author's Choice—Final version free via Creative Commons CC-BY license.

◆ This article was selected as a Paper of the Week.

✉ This article contains supplemental Movie S1.

The atomic coordinates and structure factors (code 5DWZ) have been deposited in the Protein Data Bank (<http://www.pdb.org/>).

¹ To whom correspondence may be addressed. Tel.: 44-1158467092; Fax: 44-1158468002; E-mail: jonas.emsley@nottingham.ac.uk.

² To whom correspondence may be addressed. Tel.: 49-251-8339824; Fax: 49-251-8338388; E-mail: fetzner@uni-muenster.de.

³ The abbreviations used are: QS, quorum sensing; 2-AA, 2-aminoacetophenone; 2-ABA, 2-aminobenzoylacetate; 2-ABA-CoA, 2-aminobenzoylacyl-CoA; AQ, 2-alkyl-4(1H)-quinolone; BA, benzoylacetate; CV, column volume; DHQ, 2,4-dihydroxyquinoline; HHQ, 2-heptyl-4(1H)-quinolone; PQS, *Pseudomonas* quinolone signal; TEV, tobacco etch virus; PDB, Protein Data Bank; ACP, acyl carrier protein.

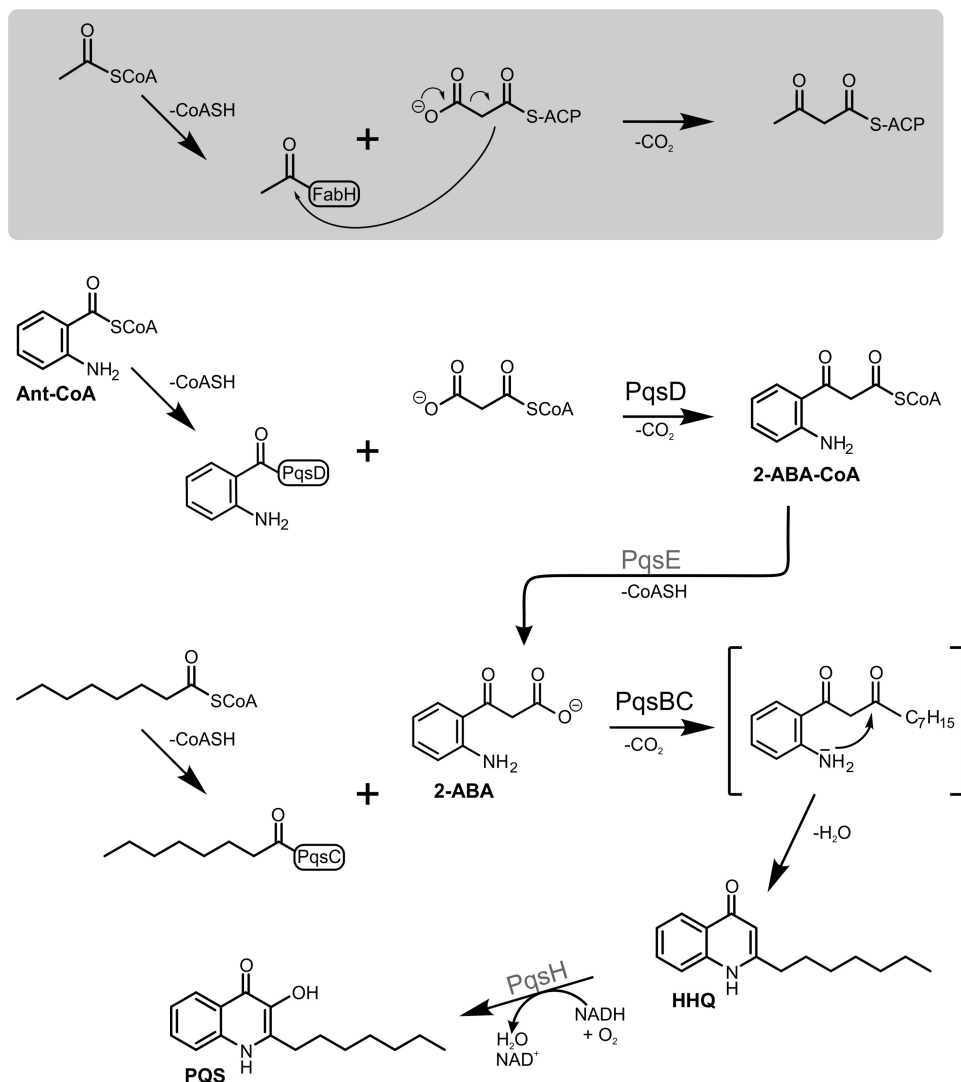


FIGURE 1. Reactions of FabH (shaded box) and the FabH-like proteins PqsD and PqsBC involved in 2-alkyl-4(1H)-quinolone biosynthesis. The PqsD- and PqsBC-catalyzed reactions proceed in a similar manner to that of the model enzyme FabH. In the initial step, the acyl moiety of an activated carboxylic acid is transferred to a strictly conserved active-site cysteine residue. Subsequently, a β -ketoalkanoic acid (malonyl-ACP in case of FabH, malonyl-CoA or malonyl-ACP in PqsD, and 2-ABA in PqsBC) is decarboxylated, and the resulting reactive enolate intermediate (not shown) attacks the thioester bond of the acyl-enzyme to form the reaction product. Intermediates and products of the alkylquinolone biosynthetic pathway: Ant-CoA, anthraniloyl-coenzyme A; 2-ABA-CoA, 2-aminobenzoylacetyl-CoA; 2-ABA, 2-aminobenzoylacetate; HHQ, 2-heptyl-4(1H)-quinolone; PQS, *Pseudomonas* quinolone signal.

olite reported to promote chronic infection phenotypes of *P. aeruginosa* (22, 23) and to modulate the host innate immune response (24, 25). Alternatively, it can be channeled into HHQ biosynthesis by the activity of PqsBC, which catalyzes the condensation of 2-ABA and octanoyl-CoA to form HHQ (Fig. 1) (20, 21). Hydroxylation of HHQ to PQS is catalyzed by the flavin monooxygenase PqsH (26).

PqsD is a member of the FabH family of condensing enzymes (27). PqsC and PqsB, which form a tight complex (20, 21), also have been annotated to belong to this family. FabH (β -ketoacyl-ACP synthase III), a key enzyme in fatty acid biosynthesis, catalyzes the decarboxylating condensation of acetyl-CoA and malonyl-ACP to form acetoacetyl-ACP (Fig. 1). The active site of *Escherichia coli* FabH is composed of residues Cys-112, His-244, and Asn-274, which are required to catalyze the complete condensation reaction. Cys-112 is loaded with the acetyl residue in the initial step of the reaction, whereas His-244 and Asn-274 are involved in the subsequent decarboxylation reac-

tion of malonyl-ACP (28). To identify the determinants of catalysis, to gain insight into the PqsBC reaction mechanism, and to probe possible effects of AQ metabolites on PqsBC activity, we performed the first structural and mechanistic investigation of this unique condensing enzyme. FabH-like enzymes are usually homodimers in solution; however, the PqsBC crystal structure we report here reveals that PqsB and PqsC form a novel heterodimer. PqsC lacks the conserved asparagine, and PqsB lacks all three conserved residues of the FabH catalytic triad, raising the question of how PqsBC mediates the condensation reaction.

Experimental Procedures

Expression and Purification of PqsBC—The *pqsB* and *pqsC* genes of *P. aeruginosa* PAO1 were cloned into plasmid pET28b using restriction-free cloning (29). In the process, *pqsC* was fused with the sequences encoding a short linker (Ser-Ala-Gly), a recognition site for tobacco etch virus (TEV) protease, and an

Structure and Function of the Condensing Enzyme PqsBC

N-terminal octahistidine tag. The vector-internal sequences for affinity tags were not used. The coding sequence of the resulting plasmid pET28b::pqsBC-NHis₈ was verified using commercial sequencing services. Site-specific mutations within the pqsC gene were introduced using restriction-free cloning (29). For protein overproduction, *E. coli* Rosetta2(DE3)[pLysS] carrying the respective expression plasmid was grown in TB medium (Carl Roth, Karlsruhe, Germany) supplemented with 150 $\mu\text{g ml}^{-1}$ kanamycin (Applichem, Darmstadt, Germany). Cells were cultivated in baffled flasks at 37 °C until reaching a cell density (A_{600}) of 0.8 and then cooled to 20 °C before gene expression was induced with 0.2 mM isopropyl β -D-thiogalactopyranoside (Thermo Scientific, Waltham, MA). After 8–12 h, cells were harvested by centrifugation, flash-frozen in liquid N₂, and stored at –80 °C until use.

For the preparation of PqsBC, cells were resuspended in 20 mM Tris-HCl buffer, pH 8.0, containing 150 mM NaCl, 10 mM imidazole, 5 mM EDTA, 0.5 mM β -mercaptoethanol, and 0.1% Nonidet P-40 (all Carl Roth, Karlsruhe, Germany). Cells were disrupted by sonication (Hielscher UP200S Hielscher, Germany), and subsequently, cell debris was removed by centrifugation for 1 h at 50,000 $\times g$ at 4 °C. Purification of PqsBC was performed using immobilized metal affinity chromatography, anion exchange, and gel filtration chromatography. All chromatographic procedures were carried out using either Äkta PrimePlus (GE Healthcare) or BioLogic (Bio-Rad) chromatography systems. To this end, cell extract supernatants were filtered (0.45 μm polyvinylidene fluoride membrane, Carl Roth, Karlsruhe, Germany) and applied onto a 5-ml nickel-nitrilotriacetic acid-Sepharose column (Qiagen, Hilden, Germany) equilibrated with a wash buffer containing 20 mM Tris, pH 8.0, 150 mM NaCl, and 10 mM imidazole. The column was washed with 10 column volumes (CV) of wash buffer before a 10-CV gradient against an elution buffer containing 20 mM Tris, 150 mM NaCl, and 150 mM imidazole was applied. Protein-containing fractions were pooled and dialyzed at room temperature against a buffer containing 10 mM Tris and 0.5 mM DTT (Carl Roth, Karlsruhe, Germany) using a 14-kDa molecular mass cut-off dialysis tube (Servapor, Serva Electrophoresis, Heidelberg, Germany). After 2 h, TEV protease (see below; 1 μg of TEV protease per 20 μg of total protein), arginine, and glutamate (final concentration 50 mM, Sigma) were added directly to the dialysis tube, and the tube was transferred into fresh buffer. Dialysis was continued overnight at 4 °C with gentle stirring. Precipitated material was removed by centrifugation at 50,000 $\times g$ (20 min at 4 °C), and the supernatant was passed through a 1-ml HisTrap column (GE Healthcare) equilibrated with 20 mM Tris-HCl, pH 8, to separate PqsBC from the uncleaved fusion protein, TEV protease, and His tag peptides. The flow-through was collected and subsequently applied to a 6-ml Resource Q column (GE Healthcare) equilibrated with a wash buffer containing 15 mM MOPS, pH 8.1 (Carl Roth, Karlsruhe, Germany), and 1 mM DTT. The column was washed with 20-CV wash buffer before a linear 20-CV gradient against 15 mM MOPS, pH 8.0, 400 mM NaCl, 1 mM DTT was started. PqsBC-containing fractions were pooled and passed through a second 1-ml HisTrap column equilibrated with a buffer containing 15 mM MOPS, pH 8.0, 150 mM NaCl to remove residual

impurities. The flow-through was collected and concentrated to a volume of ~ 1.5 ml using a 10-kDa molecular mass cutoff centrifugal concentrator (Sartorius, Goettingen, Germany). The protein solution was applied onto a HiLoad26/60 Superdex 200 gel filtration column (GE Healthcare) equilibrated with a buffer containing 40 mM MOPS, pH 8.1, and 150 mM NaCl. For molecular weight estimation, a calibration run using a Bio-Rad gel filtration standard was used. Elution fractions containing the PqsBC protein were pooled, and 2 mM DTT was added. The solution was concentrated to >20 mg ml^{-1} and applied onto two serially connected 5-ml HiTrap buffer exchange columns (GE Healthcare) equilibrated with the storage buffer containing 40 mM MOPS, pH 8.1, and 50 mM NaCl. Eluted protein fractions were pooled, aliquoted, flash-frozen, and stored at –80 °C until required for further use. Protein concentration was determined by UV spectroscopy according to Gill and von Hippel (30) using a calculated extinction coefficient of 72,880 $\text{M}^{-1} \text{cm}^{-1}$. Purification of PqsBC protein variants followed the same protocol.

Expression and Purification of Tobacco Etch Virus Protease—The L56V/S135G/S219V variant of TEV protease was overproduced and prepared as described by Cabrita *et al.* (31).

Synthesis of 2-Aminobenzoylacetate (2-ABA)—In a 250-ml flask, 2-nitrobenzoylacetate (32–34) (84 mg, 0.40 mmol) was dissolved in water (10 ml) and aqueous ammonia (0.1 ml, ~ 25 weight % in water). Palladium on charcoal (75 mg, 5% Pd/C, Sigma) was added, and the flask was placed under a hydrogen atmosphere (atmospheric pressure, by three cycles of brief evacuation and backfilling with hydrogen from a balloon). The suspension was rapidly stirred for 40 min under hydrogen and then filtered through a pad of Celite. The solvent was removed under high vacuum to give the product (56 mg, 0.31 mmol, 78%) as a yellow-green gum. ¹H NMR spectra were recorded on an Agilent DD2 600 spectrometer (600 MHz). ¹³C NMR spectra were recorded on an Agilent DD2 600 spectrometer (150 MHz). Chemical shifts δ are given in ppm and are referenced according to IUPAC recommendations on a unified chemical shift scale for all nuclides based on the proton resonance of tetramethylsilane as primary reference. MS-ESI(+) spectra were recorded on a Thermo Scientific™ Orbitrap LTQ XL using nanospray methods. Data were as follows: ¹H NMR (600 MHz, D₂O, 299 K): $\delta = 7.67\text{--}7.62$ (m, 1H), 7.26–7.21 (m, 1H), 6.72–6.68 (m, 1H), 6.65–6.60 (m, 1H), 3.71/3.69 (s, the intensity of these signals, which represents the -CH₂- and -CHD- group depends strongly on the time between dissolution and measurement of the spectrum due to the slow exchange of hydrogen against deuterium). ¹³C NMR (150 MHz, D₂O, 299 K): $\delta = 200.0, 175.9, 150.5, 135.3, 132.1, 118.0, 117.9, 116.7,$ and 49.9–49.4 (m, overlay of signals of CH₂-, -CHD-, and -CD₂- groups). MS-ESI(+): $m/z = 403$ (30) [2M – H + 2Na]⁺, 381 (60) [2M + Na]⁺, 218 (25) [M + K]⁺, 202 (100) [M + Na]⁺, 180 (55) [M + H]⁺, 162 (30) [M – H₂O + H]⁺. MS-ESI(+)-EM: $m/z = 202.04746$ calculated for C₉H₉NO₃Na [M + Na]⁺ and found 202.04705.

Spectrofluorimetric Determination of Substrate and Inhibitor Binding to PqsBC—Binding equilibria as well as octanoylation of PqsBC were measured by spectrofluorimetric titration using Jasco FP-6500 or Jasco FP-8300 instruments (Jasco, Tokyo,

Japan) and 100- μl quartz cuvettes. All measurements were performed in ratio mode with a 5-nm excitation/emission bandwidth, 1-s response, and manually adjusted photomultiplier sensitivity. Titrants were added with an eVol digital analytical syringe (SGE/Trajan Scientific, Melbourne, Australia). For determination of dissociation constants for the PqsBC-2-ABA and PqsBC-2-AA complexes, the fluorescence properties of the substrates were utilized ($\lambda_{\text{Ex}} = 359 \text{ nm}$ and $\lambda_{\text{Em}} = 470 \text{ nm}$). Assays were carried out in 50 mM HEPES, pH 8.2, and 50 mM NaCl at 22 °C, titrating 2-ABA or 2-AA (0–3 mM) into PqsBC (1–10 μM) or vice versa (0–300 μM PqsBC, 1–10 μM 2-ABA or 2-AA). After every titration step, the mixture was equilibrated for 1 min before the measurement was taken. Recorded spectra were corrected for dilution and inner filter effects as described elsewhere (35). For detection of covalent octanoate binding, 2-AA was used as a fluorescence probe. The same conditions as above were used, but 2-AA (0–50 μM) and PqsBC (0–50 μM) were premixed, and octanoyl-CoA (Sigma) was added while fluorescence was monitored over time (0.5 s resolution, 10 s mixing time). All experiments were carried out with at least three technical replicates. To exclude variance between protein preparations, four different batches of PqsBC were assayed. Any influence of the affinity tag on substrate or 2-AA binding could be excluded, because titrations with tagged and protease-processed proteins gave similar binding constants.

Equilibrium kinetic data from spectrofluorimetric titrations were fitted using Matlab R2014b with optimization toolbox (The Mathworks). For titrations of 2-ABA or 2-AA with PqsBC, the concentration of the enzyme in complex with substrate or substrate analog was calculated from the law of mass action as shown in Equation 1,

$$ES = \frac{(E_t + K_D + S_t) - \sqrt{((E_t + K_D + S_t)^2 - 4E_t \cdot S_t)}}{2} \quad (\text{Eq. 1})$$

where ES is the concentration of the enzyme-substrate complex; E_t is the total enzyme concentration; K_D is the dissociation constant, and S_t is the total substrate concentration. Fluorescence data were converted to binding ratios using the expression given in Equation 2,

$$F = F_0 + \frac{ES}{S_t} \cdot F_{\text{amp}} \quad (\text{Eq. 2})$$

when S_t was kept constant, or Equation 3 when E_t was kept constant in the assay.

$$F = F_0 + \frac{ES}{E_t} \cdot F_{\text{amp}} \quad (\text{Eq. 3})$$

F is the measured fluorescence; F_0 is the starting fluorescence value, and F_{amp} is total amplitude of fluorescence change.

Transient Kinetics of PqsBC Octanoylation—Stopped-flow fluorimetry was used to measure the transient kinetics of PqsBC octanoylation. Experiments were conducted using an SFM-3 stopped-flow instrument (Biologic, Claix, France) coupled to a Jasco FP-6500 fluorimeter. 2-AA or 2-ABA (120 μM) was mixed with PqsBC protein (30 μM), octanoyl-CoA (120 μM), and 2-AA or 2-ABA (120 μM) from separate reservoirs.

Fluorescence changes of 2-AA or 2-ABA upon protein binding were used to probe octanoylation. Samples were excited at 360 nm, and emission was measured at 450 nm (bandwidth 10 nm). The minimal detector response time was 10 ms, and the stopped-flow unit was operated at maximum speed (dead time 1.8 ms). 10–20 experiments were averaged for data collection. Data were fitted using a single exponential function, and rate constants were calculated assuming a first order reaction.

Enzyme Activity Assays with 2-ABA and Octanoyl-CoA—Catalytic activity of PqsBC was measured with 2-ABA and octanoyl-CoA. Assays were performed at 25 °C in buffer containing 50 mM HEPES, pH 8.2, 50 mM NaCl, and 100 μM DTT (all from Carl Roth, Karlsruhe, Germany). Discontinuous assays were based on the quantification of HHQ by HPLC formed in the reaction as described previously (21). Continuous spectroscopic assays were based on measuring either 2-ABA consumption or HHQ formation. The extinction coefficient of 2-ABA, quantified by the amount of DHQ formed upon acidification, was 5,025 $\text{M}^{-1} \text{cm}^{-1}$ at 360 nm (in assay buffer, pH 8.2). For setting up a spectroscopic assay detecting HHQ, defined amounts of 2-ABA were fully converted by PqsBC with an excess of octanoyl-CoA, and the resulting HHQ was quantified by HPLC. UV spectra were recorded before and after the reaction (Jasco V-650 spectrophotometer), which, in combination with the known UV-visible spectral properties of HHQ and 2-ABA, allowed the estimation of a differential extinction coefficient for the HHQ formation of $\epsilon_{313 \text{ nm}} = 6,520 \text{ M}^{-1} \text{cm}^{-1}$. However, the value at concentrations of more than 10 μM HHQ is meaningless due to scattering effects caused by precipitating HHQ. The apparent steady-state kinetic constants of PqsBC were estimated by fitting the initial velocities measured at different substrate concentrations with the Michaelis-Menten equation. The assays contained 0.02 μM PqsBC, and 2-ABA was varied from 10 to 500 μM at 100 μM octanoyl-CoA, whereas octanoyl-CoA was varied at concentrations between 1 and 200 μM at 500 μM 2-ABA. For testing the inhibitory potential of 2-AA (Sigma), DHQ (Ferak, Berlin, Germany), and PQS and HHQ (both from Sigma) on 2-ABA conversion by PqsBC, the assays contained 50 μM octanoyl-CoA and varying concentrations of 2-ABA and the potential inhibitor. EC_{50} values were calculated by non-linear fitting using the appropriate logistic function. Inhibition constants were either determined graphically from the Dixon plot (smallest Euclidean distance between regression lines) or calculated by least squares fit over all measured initial velocities using Equation 4,

$$v = \frac{v_{\text{max}} \cdot S}{K_m \cdot \left(1 + \frac{I}{K_i}\right) + S} \quad (\text{Eq. 4})$$

where v is the measured initial reaction velocity; v_{max} is the maximum velocity; S is the initial substrate concentration; K_m is the apparent Michaelis constant without inhibitor added; I is the inhibitor concentration, and K_i is the competitive inhibition constant. All data processing, fitting, and calculations were performed with MATLAB R2014A with curve fitting and optimization toolboxes (The Mathworks, Natick, MA).

Structure and Function of the Condensing Enzyme PqsBC

Activities of PqsBC with Substrate Analogs—Because the reactivity of PqsBC with octanoyl-CoA and substrate analogs benzoylacetate (BA) or malonyl-CoA cannot be analyzed by spectral changes upon product formation, a colorimetric assay based on the reaction of Ellman's reagent with free thiols originating from transacylation of octanoyl-CoA was employed (36). For assaying the activity of PqsBC with octanoyl-CoA and BA, 2.5 mM BA and 250 μ M octanoyl-CoA were mixed in a buffer containing 50 mM HEPES, pH 8.2, and 50 mM NaCl before PqsBC (concentration between 0.001 and 1 μ M) was added. The sample volume was 50 μ l. Samples were incubated between 1 and 30 min before reactions were stopped by adding 48 μ l of 200 mM Tris, pH 7.4, 1% SDS. The chromogenic reaction was started by adding 2 μ l of 50 mM 5,5'-dithiobis-(2-nitrobenzoic acid) (DTNB). Sample absorbances were immediately measured at 412 nm ($\epsilon_{\text{DTNB}} = 13,700 \text{ cm}^{-1} \text{ M}^{-1}$ (36)). All sample data were corrected against equivalently treated samples in which the enzyme was inactivated with 0.5% SDS before adding it to the assay mixture. Assays with octanoyl-CoA and malonyl-CoA were conducted analogously, using 100 μ M octanoyl-CoA and 250 μ M malonyl-CoA. Samples were incubated for 1 h before reactions were stopped, and free thiol content was measured.

Analysis of the *in Vivo* Inhibition of HHQ Biosynthesis by 2-AA—*Pseudomonas putida* KT2440 [pBBR-pqsABCD-his], when grown in mineral salts medium supplemented with anthranilic acid, produces substantial amounts of HHQ (37). To test the effect of 2-AA on the *in vivo* HHQ synthesis, the recombinant strain was grown as described previously (37) in the presence of different concentrations of 2-AA. Culture samples were withdrawn in 30–120-min intervals and analyzed by HPLC for HHQ contents. Possible effects of 2-AA on the production of the biosynthetic enzymes were monitored by Western blotting and immunodetection of the His₆-tagged PqsD protein using a pentahistidine-specific monoclonal antibody (Qiagen, Hilden, Germany).

Crystallization and Structure Determination of PqsBC^{C129A}—Protein samples of wild-type PqsBC and the active site protein variant PqsBC^{C129A} in 40 mM MOPS buffer, pH 8.1, 100 mM NaCl, 0.3 mM DTT at protein concentrations of 31.5 and 36.5 mg ml⁻¹, respectively, were diluted with 100 mM NaCl to give 3 mg/ml. These samples were subjected to screens, including Pi-PEG, Procomplex, classics lite, Mbclass, Index, PACT, Morpheus, JCSG+, and MIDAS, using the sitting drop vapor diffusion method (Mosquito robot, TTP Labtech, Melbourne, UK). Crystals of PqsBC^{C129A} resulted from the conditions of 0.2 M NaCl, 0.1 M Tris, pH 8.5, 25% PEG 3350 within 24 h, whereas octanoylated or apo-forms of wild-type PqsBC did not form crystals. Optimization was performed using 24-well sitting drop plates varying PEG in 1% increments, 23–27%, as well as Tris, pH 8.3–8.7. The final optimized crystallization condition was 25% PEG 3350, 0.1 M Tris, pH 8.3, and 0.2 M NaCl with 3 mg ml⁻¹ protein concentration and a total volume of 4 μ l. PqsBC^{C129A} crystals were cryoprotected in a solution of 27% PEG 3350, 0.1 M Tris, pH 8.3, 0.2 M NaCl, 25% glycerol, and flash-cooled in liquid nitrogen prior to data collection at the Diamond Light Source synchrotron facility beamline I02 to 2.0 Å resolution with space group P2₁2₁2₁ and a unit cell of

TABLE 1
Data collection and refinement statistics

Data collection	PqsBC ^{C129A}
Space group	P2 ₁ 2 ₁ 2 ₁
<i>a</i> , <i>b</i> , <i>c</i> (Å)	78.5, 115.0, 289.7
α , β , γ (°)	90, 90, 90
Wavelength	0.97949
Resolution (Å)	2.0
<i>R</i> _{merge} ^{a,b}	0.069 (0.696)
<i>I</i> / σ <i>I</i>	12.5 (1.9)
Completeness (%)	99.2 (96.7)
Redundancy	4.5 (4.6)
CC 1/2 ^c	0.998 (0.661)
Unique reflections	166,566
<i>R</i> _{work} ^d	0.190 (0.291)
<i>R</i> _{free}	0.237 (0.321)
Overall <i>B</i> factor (Å ²)	38.1
Bond lengths (Å)	0.016
Bond angles (°)	1.81
Most favored (%) ^e	97.7
Allowed (%)	2.3
Outliers (%)	0

^a Values in parentheses correspond to the highest resolution shell.

^b $R_{\text{merge}} = \sum_h \sum_i |I_i(h) - \langle I(h) \rangle| / \sum_h \sum_i I_i(h)$, where *I* is the observed intensity, and $\langle I(h) \rangle$ is the average intensity of multiple observations from symmetry-related reflections calculated with XDS.

^c Correlation coefficient value was calculated using XDS to determine the resolution cutoff.

^d All values were calculated using REFMAC. $R_{\text{work}} = \sum_h |F_o(h) - |F_c(h)|| / \sum_h |F_o(h)|$, where *F_o* and *F_c* are the observed and calculated structure factors, respectively. *R*_{free} computed as in *R*_{work}, but only for (5%) randomly selected reflections, which were omitted in refinement.

^e Ramachandran plot is shown.

a = 78.5 Å, *b* = 115.0 Å, and *c* = 289.7 Å, α = 90°, β = 90°, and γ = 90° (Table 1). This unit cell was predicted to have four PqsB and four PqsC polypeptides in the asymmetric unit with ~50% solvent content. Molecular replacement was performed with PHASER using a variety of templates generated using MrBUMP (38) from PDB codes 3h76, 3h77, and 3h78 of the PqsD structure (39). Using different resolution cutoffs, this resulted in an incomplete solution from which a partial model was built using BUCCANEER (40), and electron density maps were subsequently improved using 4-fold non-crystallographic symmetry and solvent flattening with the CCP4 programs DM (41) and PARROT (42). The resulting density-modified 2 Å electron density map was of high quality allowing 95% of the model to be built with BUCCANEER. The model was completed manually using COOT and refined with REFMAC5 (43) giving 2,474 residues in total, and crystallographic statistics are listed in Table 1 (PDB code 5DWZ). Values for surface area buried were calculated using PISA (44).

Results

Synthesis and Stability of 2-ABA and Benzoylacetate—Quantitative investigation of the PqsBC reaction requires highly purified substrates applicable at defined concentrations; however, 2-ABA was previously described as relatively unstable in solution, decomposing to 2-AA and also to DHQ (20, 34). After testing a series of conditions, we discovered that hydrogenation in an alkaline aqueous solution is the key to obtain 2-ABA in high yield and purity. Using dilute ammonia solution as medium for hydrogenation of the 2-nitrobenzoylacetate precursor, we obtained 2-ABA in 78% yield with ~95% purity (as indicated by HPLC and NMR data). In aqueous solution at a pH above 8, it is stable for several days at room temperature (in the

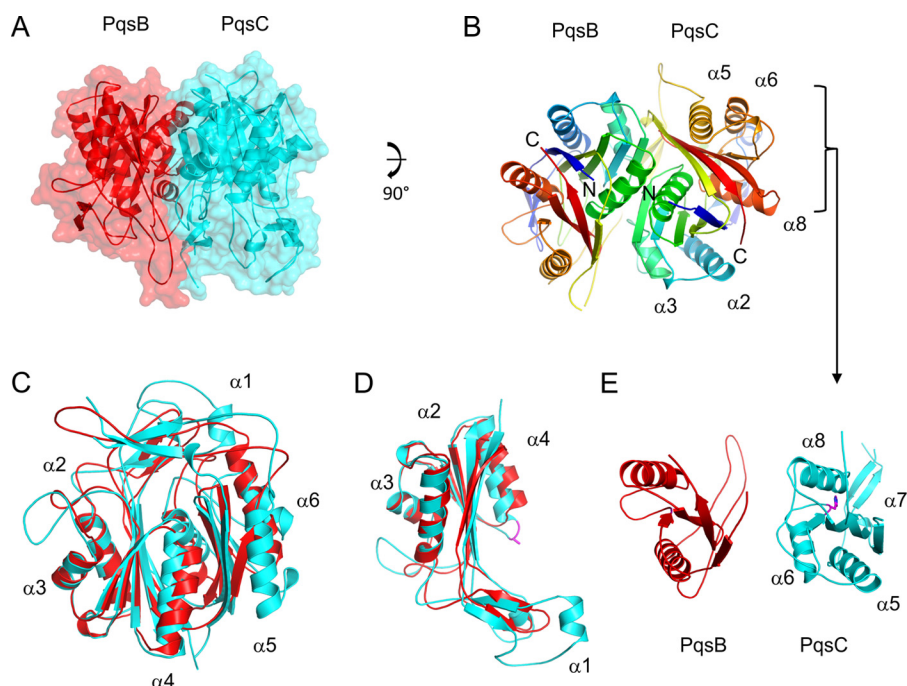


FIGURE 2. **Schematic diagrams of the PqsBC^{C129A} structure.** *A*, PqsBC^{C129A} heterodimer is shown with PqsC (cyan) and PqsB (red) covered with a transparent molecular surface illustrating the interface. *B*, PqsB and PqsC^{C129A} structures are colored in a rainbow from the N (blue) to the C terminus (red) with secondary structure elements labeled for PqsC^{C129A} in all panels. *C*, superposition of PqsB and PqsC^{C129A}. *D*, N-terminal sub-domains of PqsB(1–154) and PqsC(1–184) superposed with the catalytic dyad residue Ala-129 shown as sticks (purple). *E*, C-terminal sub-domains of PqsB(155–279) and PqsC(188–348) arranged side by side viewed with the same orientation showing PqsC^{C129A} helix $\alpha 6$ is missing in PqsB. PqsC residue His-269 is shown as sticks (purple).

dark) and can be stored at -80°C for months. When solutions of 2-ABA were acidified, the compound decomposed to DHQ. However, when exposed to organic solvents such as methanol, acetonitrile, or dimethyl sulfoxide, rapid decarboxylation to 2-AA as the sole decomposition product was observed. BA, synthesized as described previously (21), was used as an analog of 2-ABA in the PqsBC reaction. It turned out to be unstable in aqueous solution at neutral pH and decarboxylated within minutes to hours, yielding acetophenone.

PqsBC^{C129A} Crystal Structure Reveals a Heterodimer—PqsBC^{C129A} crystallized in space group $P2_12_12_1$ and the structure was determined to 2 Å resolution with the final model refined to crystallographic R -factors of $R_{\text{work}} = 0.190$ and $R_{\text{free}} = 0.237$ (Table 1). The crystallographic asymmetric unit contains four copies of PqsB and four copies of PqsC. An analysis of the interfaces reveals a large $\sim 2,238 \text{ \AA}^2$ surface area buried by interactions formed between the PqsB and PqsC subunits as shown in Fig. 2*A*. This is consistent with gel filtration data indicating that PqsB and PqsC form a tight complex with an apparent molecular mass of 51 kDa. The structure is thus referred to as the PqsBC^{C129A} heterodimer. This large interface between PqsB and PqsC is observed to be almost identical in the four copies observed in the asymmetric unit (values of buried surface area are 2,277, 2,240, 2,223, and 2,212 Å² averaged to 2,238 Å²), and the heterodimers superimpose onto each other well with an average root mean square deviation of 0.243 Å (8,182 atoms).

The arrangement of secondary structural elements colored from the N to the C termini shows PqsB and PqsC assemble into a dimer with the N termini (blue) co-localizing close to the pseudo 2-fold axis and the C termini (red) being placed adjacent

directed away from the axis (Fig. 2*B*). The fold of PqsC and PqsB is closely related to the FabH enzymes and PqsD with two subdomains each composed of a four-stranded β -sheet surrounded by α -helices. The structural superposition of PqsB and PqsC shown in Fig. 2*C* is largely driven by similarities in the N-terminal subdomain (Fig. 2*D*). Substantial differences occur in the C-terminal subdomains with PqsB having a shorter sequence (124 amino acids) with only two α -helices and PqsC (160 amino acids) having four α -helices (Fig. 2*E*). The large difference in the amino acid sequence of the PqsB C-terminal region compared with PqsC means that it is difficult to align the primary sequences, and there is no identifiable catalytic triad or dyad in the PqsB structure indicating it likely contributes a supporting structural role to the enzyme functionality.

PqsC^{C129A} Structure Defines a Catalytic Dyad—The active-site residues His-269 and the mutated Ala-129 (replacing Cys-129) are clearly defined in the electron density (Fig. 3*A*). In a noticeable difference with other members of the FabH enzyme family, an asparagine amino acid does not form a catalytic triad with His-269 and Cys-129, and in its place a valine (Val-299) is present (Fig. 3*B*) shows the superposition of PqsC with the PqsD catalytic triad). Fig. 4*A* shows PqsC^{C129A} superposed onto the PqsD structure (PDB code 3h78) viewed in the area of the active site cleft illustrating significant structural differences. The area of the structure known as the flap in the FabH enzymes (45) is conserved as a short α -helix in PqsD, whereas in PqsC it is formed by a short antiparallel $\beta 9$ - $\beta 10$ sheet. Directly C-terminal to this, further differences are apparent as the PqsC helix $\alpha 5$ is shorter than the equivalent α -helix in PqsD/FabH. Pro-242 breaks the helix $\alpha 5$ in PqsC and directs an alternate path for the

Structure and Function of the Condensing Enzyme PqsBC

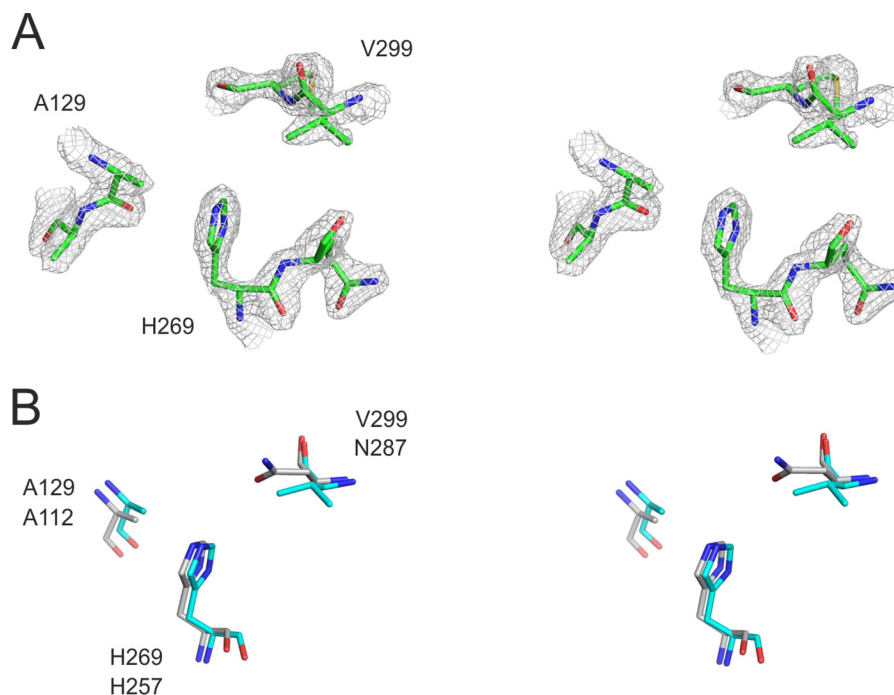


FIGURE 3. **Stereo figures to illustrate the PqsBC^{C129A} catalytic dyad.** *A*, 2.0 Å electron density map is shown (gray) contoured at 2.0 σ level calculated using the final refined weighted $2Fo - Fc$ coefficients (REFMAC) and rendered with PyMOL showing PqsC residues Ala-129, Ala-130, His-269, Gln-270, Val-299, and Met-300 as sticks in green. *B*, PqsBC^{C129A} Ala-129, His-269 catalytic dyad is shown in cyan superposed with the PqsD^{C112A} triad residues (gray).

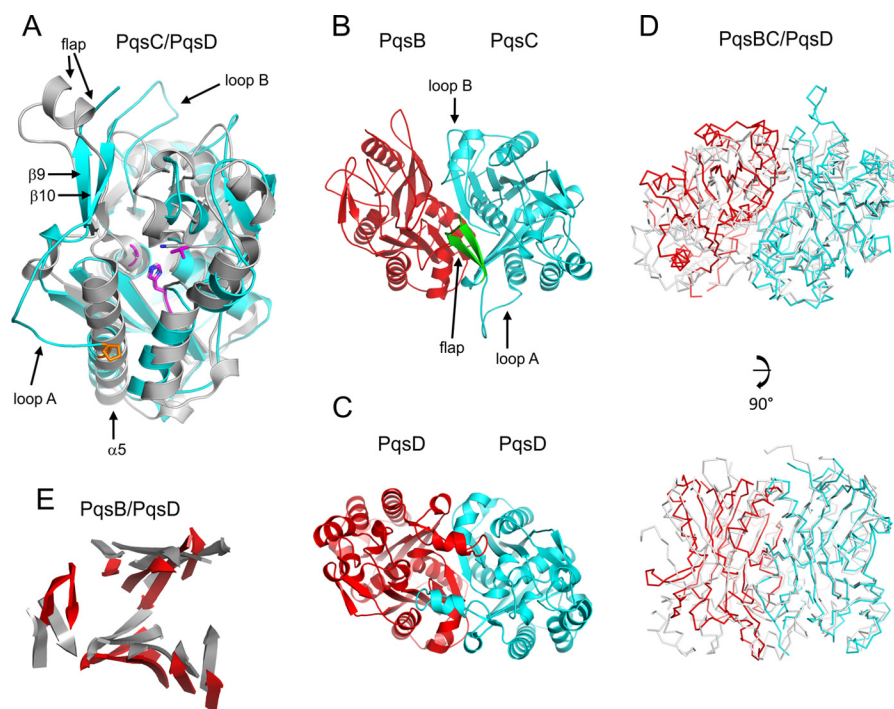


FIGURE 4. **Comparison of PqsD and PqsBC^{C129A}.** *A*, PqsC^{C129A} structure is shown as a schematic (cyan) superposed onto PqsD (gray). Residues from PqsC shown as sticks in purple are Ala-129, His-269, Val-299, and Pro-242 (orange). PqsD residues Ala-112, His-257, and Asn-287 are shown as sticks (gray). *B*, schematic diagram of PqsBC (red, cyan) with the flap (PqsC residues 207–227) in green shown in the same orientation as the PqsD homodimer below (C). *D*, superposition of the PqsBC structure (red, cyan) with the PqsD homodimer (gray) illustrated as C α -backbone representations with two views shown. *E*, same as *D*, but only β -strands for PqsB (red) and PqsD (gray) are shown as schematics.

polypeptide compared with FabH/PqsD forming an irregular loop structure, termed loop A, which effectively opens up the active site cleft. Another difference between PqsC and FabH/PqsD structures is an insertion of 14 amino acids in PqsC between strand β_4 and helix α_3 (Fig. 5; PqsC residues 88–102).

This insertion is termed loop B and forms a network of interactions with the PqsC flap and contributes to the dimer interface with PqsB.

Fig. 4*B* illustrates the structure of the PqsBC^{C129A} heterodimer compared with the PqsD homodimer below viewed

Structure and Function of the Condensing Enzyme PqsBC

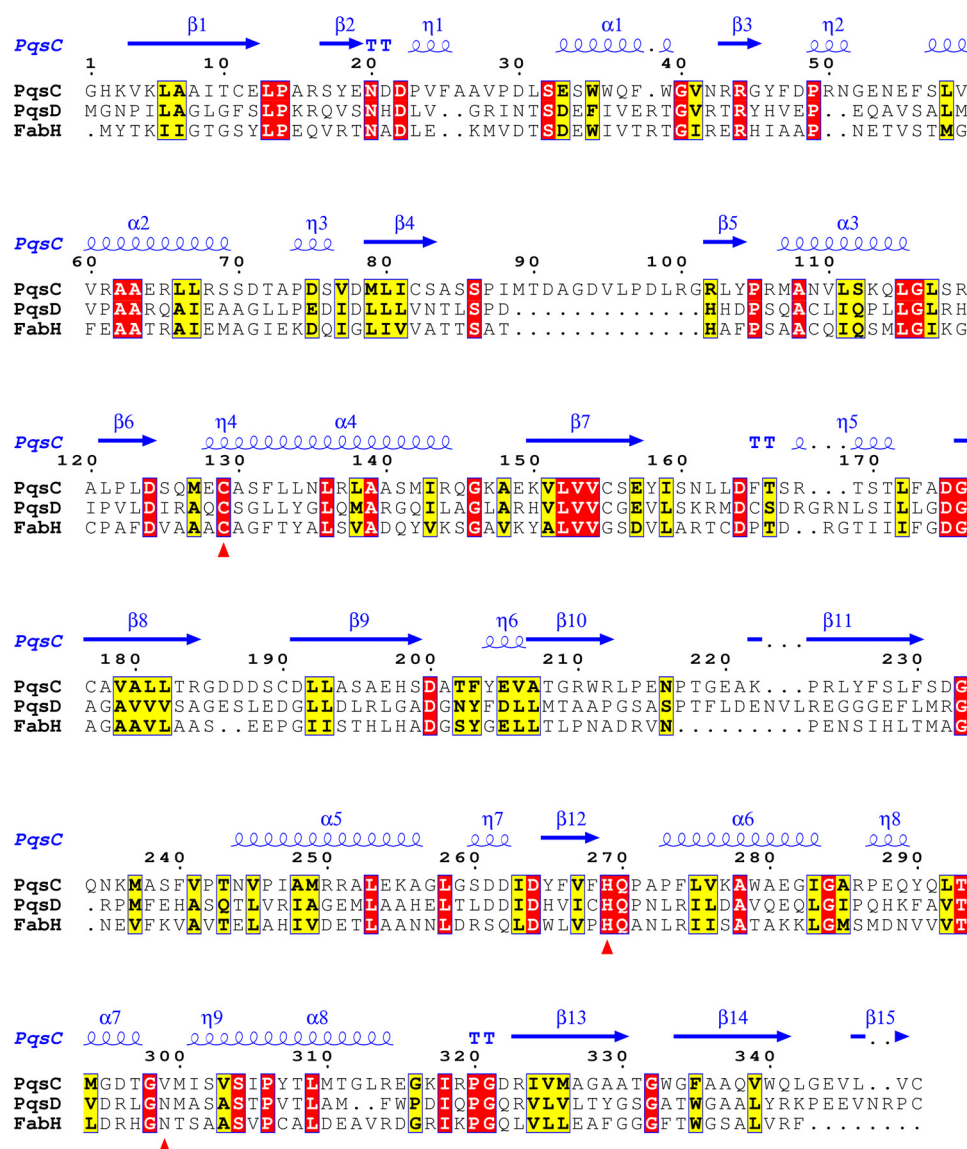


FIGURE 5. Alignment of the PqsC amino acid sequence with related enzymes showing the assignments of the PqsC^{C129A} structure at the top (blue). The catalytic residues are marked by red triangles. The sequences shown are PqsD (*P. aeruginosa*) and FabH (*E. coli*).

with the flap region lying close to the PqsD dimer axis (Fig. 4C). Fig. 4D shows two orientations of a C- α trace of PqsBC polypeptide backbone superposed on the PqsD homodimer. The superposition is driven largely by the similarity between PqsC and PqsD and shows the close relative positioning of the PqsBC subunits compared with the PqsD homodimer. The PqsB β -sheets are displaced by 2–3 Å when overlaid with PqsD (Fig. 4E). The PqsBC active site cavity is elongated in the direction of the heterodimer interface where it narrows and extends into PqsB. The gap in the interface is not observed in FabH/PqsD and in part accounts for the smaller ~2,238 Å² surface area buried by formation of the PqsBC heterodimer compared with the larger value of 3,000 Å² for the FabH/PqsD homodimer.

The PqsC^{C129A} active site volume is twice as large as those for FabH/PqsD with a calculated volume of 761 Å³ compared with 389 Å³ for PqsD (PDB code 3h76) and 367 Å³ for FabH (PDB code 2QNZ) (calculated using 3vee (46)). Fig. 6A highlights internal cavities in the PqsBC structure calculated using meta-

pocket (47) and rendered with PyMOL. The PqsC pocket surrounding the catalytic dyad extends toward the interface with PqsB and is bifurcated with one branch formed by loop B and residues from PqsB and a second branch remains within PqsC in the area of the flap (Fig. 6B). Fig. 6, B and C, shows the decyl-formate ligand from the FabH-decyl formate complex crystal structure (PDB code 2QNZ (45)), which has been superposed onto the PqsC^{C129A} structure illustrating that the active site pocket extends in two directions from Ala-129 as in FabH (supplemental Movie 1). Fig. 6D schematically shows the residues lining the area of the pocket. The binding site of coenzyme A has been characterized in both FabH (PDB code 1hnj) and PqsD (PDB code 3h77) complex crystal structures (27, 48, 49). Two PqsC residues, Trp-35 and Arg-168, homologous to the PqsD (Trp-32 and Arg-151) coenzyme A-binding site are conserved (27, 50). In PqsD these form interactions on either side of the coenzyme A nucleobase suggesting a similar interaction may occur in PqsC (Fig. 6D). However, no other similar residues are conserved due to the large

Structure and Function of the Condensing Enzyme PqsBC

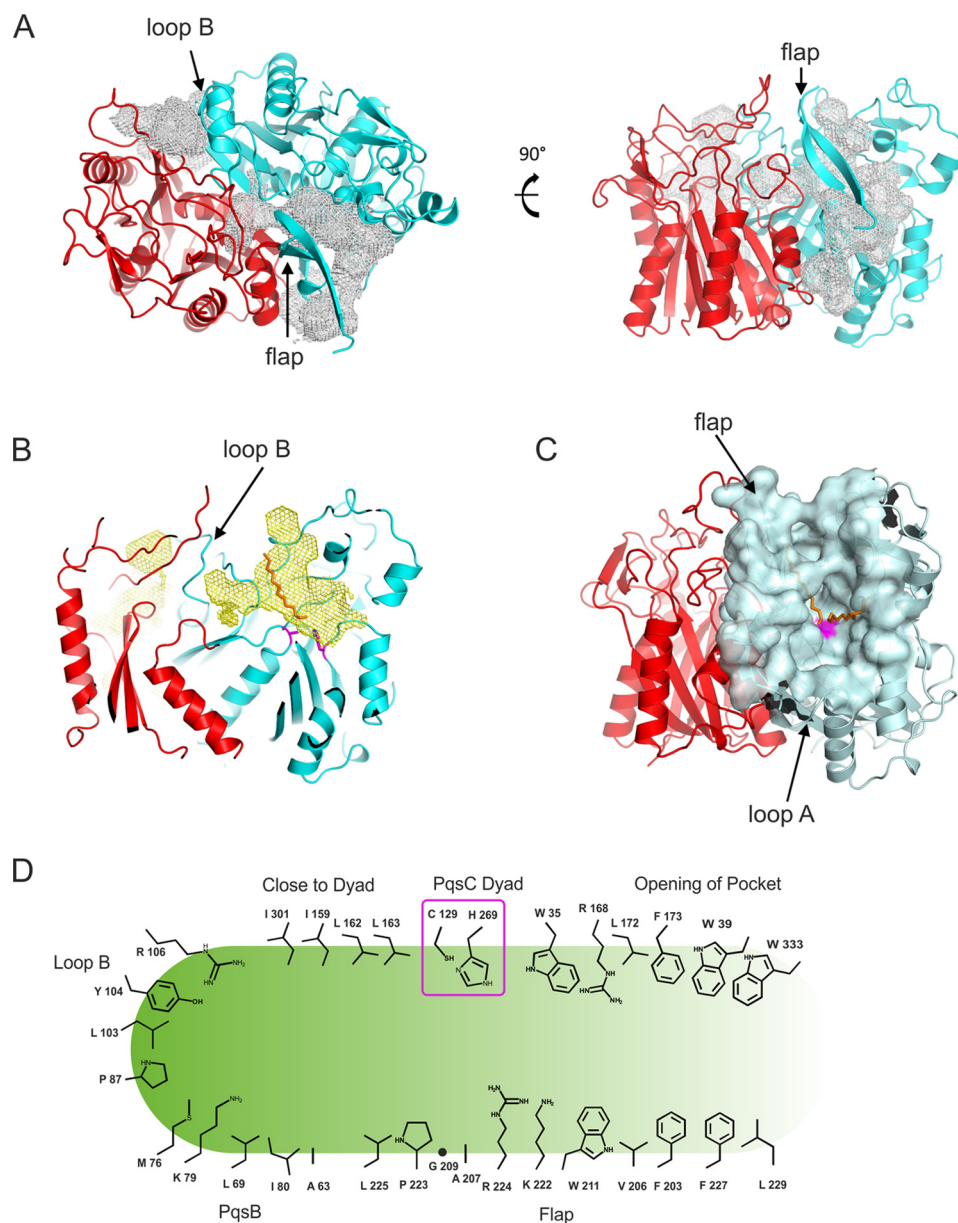


FIGURE 6. **PqsBC^{C129A} active site pocket.** *A*, two views showing the internal pockets calculated using metapocket shown as mesh (gray) together with schematic representations of the PqsBC^{C129A} structure. *B*, PqsBC structure is shown with the view clipped such that the PqsC flap is removed to show the relationship between the PqsC catalytic dyad (purple) and the PqsC cavity (yellow mesh). For perspective, a decyl formate molecule is shown as sticks (orange) derived from a superposition of the Mtb FabH–decyl formate complex structure (PDB code 2QNZ). *C*, transparent surface representation of the PqsBC active site pocket is shown (light cyan) with residue Ala-129 in purple. For perspective, the decyl formate molecules showing the orientation of the proposed entry and exit portals from the FabH–decyl formate complex structure are shown (orange). *D*, graphical representation of the PqsBC active site cleft. Residues conserved with CoA-binding sites of FabH/PqsD enzymes are Trp-35 and Arg-168.

changes associated with loss of the N-terminal region of helix $\alpha 5$ in PqsC (Fig. 4A).

Steady-state Kinetics Suggest Competitive Inhibition of PqsBC by 2-AA—Using a continuous spectrophotometric assay, initial reaction rates were determined for the PqsBC reaction at substrate concentrations of up to 1 mM 2-ABA and 200 μ M octanoyl-CoA. Because a scattering band is emerging at higher concentrations of HHQ (Fig. 7A), initial rates were evaluated before the solubility limit of HHQ was reached. Under the conditions of the assay, a k_{cat} of $6.8 \pm 0.5 \text{ s}^{-1}$ was observed, and apparent K_m values of PqsBC were $6.0 \pm 0.5 \mu\text{M}$ for octanoyl-CoA and $105 \pm 12 \mu\text{M}$ for 2-ABA. The His-tagged and untagged protein showed no difference in their kinetic parameters.

To determine whether the various products of the AQ biosynthetic pathway have an effect on PqsBC activity, enzyme assays were performed in the presence of HHQ, PQS, 2-AA, or DHQ. HHQ and PQS as well as DHQ at concentrations up to 50 μM did not affect PqsBC activity. 200 μM DHQ, a concentration that is not reached in *P. aeruginosa* cultures (51), caused an $\sim 10\%$ drop in activity. In contrast, 2-AA is an efficient inhibitor of PqsBC with an apparent EC_{50} of 46 μM in the *in vitro* assay. It increased the apparent K_m value of the enzyme for 2-ABA, whereas the k_{cat} value remained unaffected (Fig. 7, B–D), indicating a competitive binding mode with an estimated inhibition constant K_i of 54.5 μM (graphic determination from Dixon plot, Fig. 7D) or

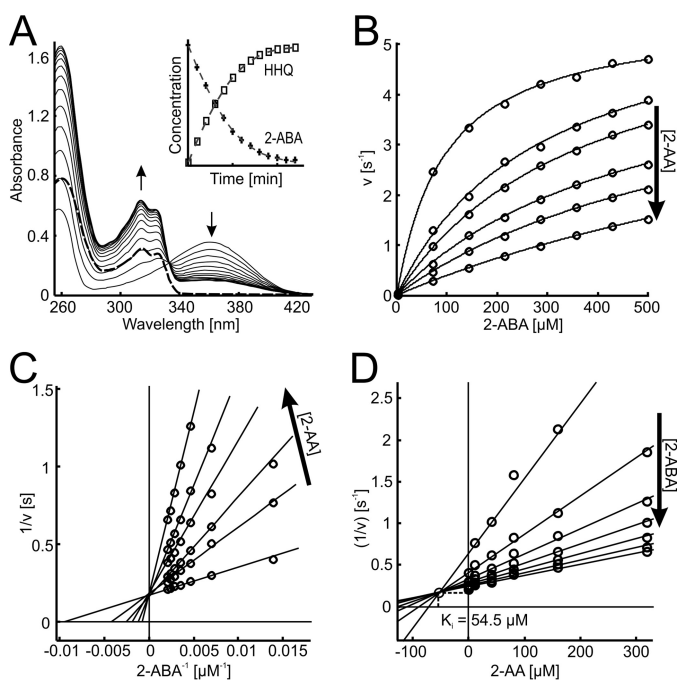


FIGURE 7. Steady-state kinetics and competitive inhibition of PqsBC by 2-AA. *A*, conversion of 2-ABA (λ_{\max} at 360 nm) and octanoyl-CoA (absorbing at 260 nm) to HHQ (λ_{\max} at 313 nm). 20 nM PqsBC was mixed with 50 μM of each substrate. Spectra were measured every 60 s with a bandwidth of 1 nm. HHQ formation could be observed at 313 nm with a differential extinction coefficient of $6,520 \text{ M}^{-1} \text{ cm}^{-1}$. The remaining absorption above 340 nm after full conversion of the substrates (*bold trace*) is due to scattering and can be eliminated by diluting to 50% in 2-propanol/HCl (*dashed trace*). *Inset*, conversion of 10 μM substrates, monitored at 313 nm (HHQ) and 360 nm (2-ABA) with the determined extinction coefficients. Data interval is 1 min. *B–D*, inhibition of PqsBC activity by 2-AA. *B*, nonlinear fits of measured initial rates. The Lineweaver-Burk plot (*C*) shows the characteristic pattern of competitive inhibition. Using the Dixon plot (*D*), the inhibitor constant K_i of the competitive reaction could be determined graphically. Experiments were performed with 50 μM octanoyl-CoA and 20 nM PqsBC in HEPES buffer, pH 8.2, at 25 $^{\circ}\text{C}$; arrows indicate ascending concentrations.

$56 \pm 3.1 \mu\text{M}$ (global fit according to Equation 4, denoted error is the S.E. of cross-validation).

2-AA Inhibits Biosynthesis of HHQ in Vivo—In view of the observed *in vitro* inhibition of PqsBC activity by 2-AA, which is produced by planktonic *P. aeruginosa* cultures in concentrations of up to 200 μM (52), we used a recombinant strain of *P. putida* constitutively expressing the *pqsABCD* genes for testing whether exogenously added 2-AA can inhibit AQ biosynthesis *in vivo*. Any measured inhibitory effect on HHQ biosynthesis in this strain would be caused by inhibition of the biosynthetic proteins. Immunodetection of PqsD, the last protein translated from the polycistronic mRNA, in total cell proteins demonstrated that exposure to 2-AA did not alter the production of Pqs proteins by the cells. 2-AA inhibited HHQ biosynthesis by *P. putida* KT2440 [pBBR-*pqsABCD*], with an EC_{50} value of $319 \pm 38 \mu\text{M}$ (Fig. 8).

Inhibition of HHQ synthesis by 2-AA was observed before for *P. aeruginosa* and was in part attributed to down-regulation of *pqsABCDE* expression by inhibition of the transcriptional regulator PqsR (Mvfr). However, post-transcriptional interference of 2-AA with HHQ biosynthesis was also proposed (22). Our *in vivo* and *in vitro* data suggest that competitive inhibition of PqsBC accounts for the post-transcriptional inhibition.

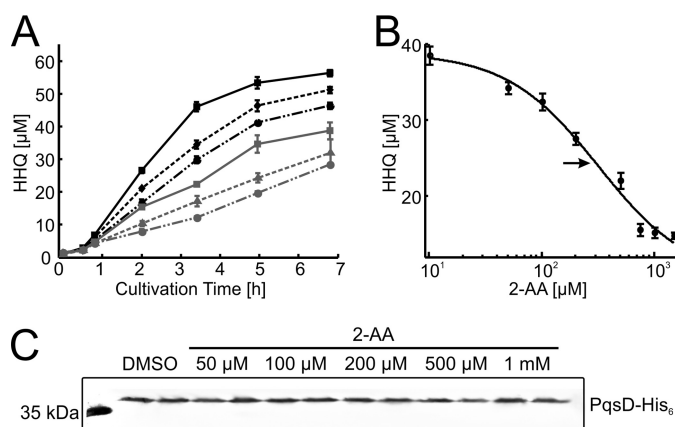


FIGURE 8. *In vivo* inhibition of HHQ biosynthesis by 2-AA. Cultures of *P. putida* KT2440 [pBBR-*pqsABCD*-his] were grown in the presence of 1 mM anthranilic acid (as precursor for HHQ synthesis) and various concentrations of 2-AA. 2-AA does not affect growth of *P. putida* KT2440, and it is not utilized as a carbon source. *A*, HHQ contents of culture samples, as determined by HPLC. *Black solid line*, DMSO control; *black dashed line*, 50 μM ; *black dash-dotted line*, 100 μM ; *gray solid line*, 200 μM ; *gray dashed line*, 500 μM ; *gray dash-dotted line*, 1 mM 2-AA. Errors reflect standard errors (S.E.) of three biological replicates. *B*, dose-response plot, using cultures sampled 3.5 h after inoculation. The arrow indicates the calculated EC_{50} of $319 \pm 38 \mu\text{M}$. *C*, immunodetection of PqsD-His₆ in Western blots of cell extract supernatants confirms synthesis of the Pqs proteins. Cell extracts were from cultures (independent duplicates) harvested after 7 h of incubation. Each lane contained 50 μg of total protein.

Because the IC_{50} of 2-AA of $\sim 370 \mu\text{M}$ in *P. aeruginosa* PA14 (indicated by the relative activity of the *pqsA* promoter (22)) is in a similar range as the EC_{50} value determined for the recombinant *P. putida* strain used in this study, the effect of 2-AA on PqsBC likely dominates over its effects on *pqsABCDE* transcription.

Octanoylation of PqsBC Has a Major Effect on Binding of the Second Substrate—The apparent Michaelis constant of PqsBC for 2-ABA is rather high, at least when compared with the K_m values of PqsA, PqsD, and PqsH (18, 19, 26). This led us to analyze the binding equilibria of PqsBC with 2-ABA, as well as with its competitive inhibitor 2-AA. Usefully, it turns out that the fluorescence characteristics of 2-AA and 2-ABA, both containing a 2-aminobenzoyl moiety, are strongly influenced by their molecular environment such that the increase in fluorescence upon protein binding could be used for determination of the binding affinities to PqsBC.

The K_D value of about 370 μM determined for the complex of PqsBC with 2-ABA (Table 2) suggests that the affinity of non-octanoylated PqsBC for 2-ABA is even lower, more than 3-fold, than expected from the corresponding apparent K_m value. Exchange of Cys-129 by Ala or Ser had only a minor effect on the K_D value, indicating that this position does not play a crucial role in the binding of 2-ABA. K_D values of the PqsBC·2-AA complex were roughly 2-fold lower, which matches the observed ratio of $K_m(2\text{-ABA})/K_i(2\text{-AA})$ of 2:1.

The exceedingly high K_D value of the PqsBC·2-ABA complex and the discrepancy between equilibrium binding and kinetic data led us to hypothesize that octanoylation at Cys-129 of PqsC (20), which, based on the current understanding of FabH-like reactions is proposed to be the initial step of the reaction, modulates the affinity of the enzyme for 2-ABA. Assuming that 2-AA binds to PqsBC and to octanoyl-PqsBC in the same way as

Structure and Function of the Condensing Enzyme PqsBC

TABLE 2

Dissociation constants of PqsBC in complex with 2-ABA or 2-AA

K_D values were deduced from fluorimetric titrations (1–10 μM PqsBC and 0–3 mM 2-AA or 2-ABA). Errors reflect standard errors of mean (S.E., $n \geq 3$). Titrations were carried out in the absence (–) or presence (+Oct-CoA) of a 10-fold excess of octanoyl-CoA over PqsBC. The values of the H269A and C129S proteins are possibly affected by minor residual activity (k_{cat} (H269A) = 0.0005 s^{-1} and k_{cat} (C129S), below detection limit of the spectrophotometric assay).

Protein	K_D (2-ABA)		K_D (2-AA)	
	–	+Oct-CoA	–	+Oct-CoA
	μM		μM	
PqsBC	368 ± 19	– ^a	161 ± 8	5 ± 1
PqsBC ^{H269A}	198 ± 11	166 ± 12	117 ± 6	88 ± 5
PqsBC ^{C129A}	330 ± 30	312 ± 38	158 ± 12	141 ± 19
PqsBC ^{C129S}	411 ± 43	420 ± 45	170 ± 10	177 ± 8
PqsBC ^{V299N}	NB ^b	– ^a	NB ^b	89 ± 4

^a Experiment was not possible due to turnover.

^b No binding or K_D values above detection limit (estimated to >1 mM).

2-ABA, we used 2-AA for probing the effect of PqsBC acylation on the affinity for the second substrate. As illustrated in Fig. 9B, octanoyl-PqsBC has an ~35-fold higher affinity for 2-AA than the unloaded enzyme. However, such a change in affinity was not observed for the PqsBC^{C129S} and PqsBC^{C129A} proteins incubated with octanoyl-CoA, supporting the notion that covalent binding of the octanoyl moiety to Cys-129 increases the binding affinity of PqsBC for 2-AA and, most probably, also for 2-ABA. Octanoylation of PqsBC^{C129S} was not observed.

His-269 of PqsC Modulates 2ABA/2-AA Binding—Interestingly, replacement of His-269 in PqsC by alanine resulted in significantly altered binding properties of the PqsBC^{H269A} protein for both 2-ABA and 2-AA. The affinity of the (non-acylated) mutant protein was 1.5–2-fold higher than that of wild-type PqsBC for both compounds, although octanoylated PqsBC^{H269A} did not show the increase in affinity for 2-AA that was observed for the octanoylated wild-type enzyme (Fig. 9B and Table 2). Again, assuming that the interaction with 2-AA reflects 2-ABA binding, these findings suggest that His-269 acts as modulator, which, depending on the absence or presence of an octanoyl moiety in the active site, has restricting or promoting effects, respectively, on binding of the second substrate. However, when comparing different structures of cysteine-loaded and unloaded FabH-like enzymes, no distinct structural change of the catalytic histidine is observable, suggesting that electronic rather than spatial changes drive the observed affinity shift.

Octanoylation of Cys-129 of PqsC Is Independent of His-269—In previous studies of FabH and FabH-like proteins, a possible involvement of the active-site histidine in deprotonation of the cysteine to act as a nucleophile in the initial acylation step has been discussed (28, 49). In the case of PqsBC, the observed competitive inhibition of the reaction by 2-AA and the high affinity of 2-AA for the octanoylated protein suggested that this compound could be used as a fluorescence probe to monitor the acylation step in a time-resolved setup (Fig. 10A). The binding velocity of 2-AA to PqsBC was faster than resolvable by the stopped-flow instrument (~5 ms detection limit), which was a prerequisite for using 2-AA as a probe for determination of the octanoylation rate. From single turnover stopped-flow fluorimetric measurements (Fig. 10, B–D), rate constants of $k_{\text{WT}} = 86.9 \pm 2.6 \text{ s}^{-1}$, $k_{\text{H269A}} = 76.6 \pm 1.9 \text{ s}^{-1}$, and $k_{\text{H269A}}(2\text{-ABA}) = 59.4 \pm 5.1 \text{ s}^{-1}$ were deduced for the octanoy-

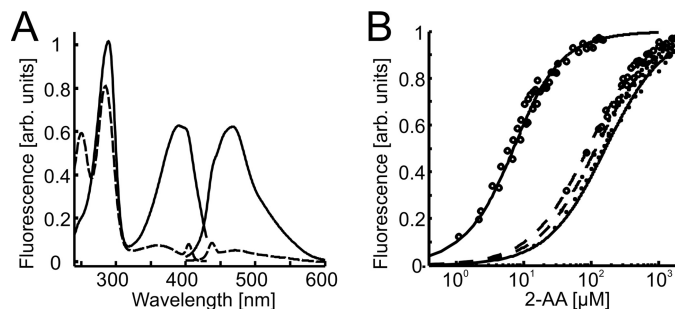


FIGURE 9. Equilibrium binding of 2-AA to PqsBC proteins in apo- and octanoylated forms. A, fluorescence excitation and emission spectra of 2-AA (1 μM) in unbound (dashed line) and protein-bound (20 μM PqsBC, continuous line) form. B, titration data and curve fittings of 5 μM PqsBC (continuous lines) or PqsBC^{H269A} variant (dashed lines) with 2-AA in the presence (circles) or absence (dots) of 10 μM octanoyl-CoA. Data suggest an increase of binding affinity to 2-AA of the acyl-protein in the wild-type but not in the H269A variant (numerical data provided in Table 2).

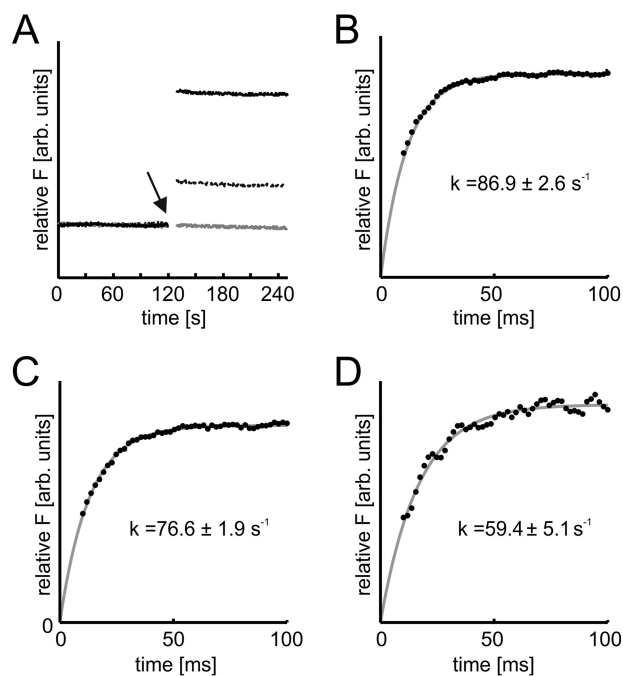


FIGURE 10. Role of His-269 in octanoylation of PqsBC, monitored by transient kinetics. A, time-dependent measurement of 2-AA fluorescence ($\lambda_{\text{ex}} = 400 \text{ nm}$, $\lambda_{\text{em}} = 460 \text{ nm}$, 0.5-s time resolution) in complexes with PqsBC proteins. Octanoylation (addition of 10-fold molar excess of octanoyl-CoA indicated by arrow) shifts the binding equilibrium of 2-AA and PqsBC (solid black line). The effect was reduced in PqsBC^{H269A} (dashed black line), and addition of octanoyl-CoA to PqsBC^{C129A} did not change 2-AA fluorescence (gray line). Enzyme and 2-AA concentrations were varied between 1–20 and 1–100 μM respectively. B–D, octanoylation of PqsBC proteins, analyzed with stopped-flow fluorimetry. B, PqsBC octanoylation probed with 2-AA. C, PqsBC^{H269A}, probed with 2-AA. D, PqsBC^{H269A}, probed with 2-ABA. Assays were conducted at 6 °C, detector response time was 10 ms; enzyme concentration was varied to achieve sufficient signal to noise ratio. Rate constants were calculated assuming a first order reaction characteristic. Denoted errors are standard errors of cross-validation.

lation reaction in wild-type PqsBC, the PqsBC^{H269A} variant, and PqsBC^{H269A} when probed with 2-ABA instead of 2-AA, respectively. If the His-269 residue were important for activation of the catalytic Cys-129, more drastic differences in the acylation rates would have been expected.

Amino Group of 2-ABA Functionally Replaces the Canonical Catalytic Asparagine—The catalytic cysteine accepts the acyl moiety, while the histidine and asparagine residues in the active

site of FabH-like enzymes have been shown to interactively mediate the decarboxylation of the second reactant (malonyl-ACP in FabH and PqsD) (19, 48). In *E. coli* FabH, both residues are required for decarboxylation activity (48). Interestingly, exchange of His-269 to alanine in PqsBC nearly abolished catalytic activity ($k_{\text{cat}} = 0.0005 \text{ s}^{-1}$, corresponding to a more than 10,000-fold decrease in activity). This is in striking contrast to FabH, where replacement of the catalytic histidine resulted in only a 10-fold decrease in activity (48). Because the PqsBC^{H269A} protein still undergoes the acylation step (see above), the histidine clearly is essential for a later step of the PqsBC-catalyzed reaction, which apparently does not depend on participation of an Asn residue.

2-ABA, the substrate of PqsBC, in contrast to malonyl-CoA/ACP, features an amino group in close proximity to the β -keto carbonyl, which, as suggested by the increased stability of 2-ABA over its desamino analog BA in aqueous solutions, influences the chemical properties of the β -keto acid moiety. We hypothesized that in the PqsBC catalytic cycle, the amino group of 2-ABA might functionally replace the amide group of the missing active-site asparagine. To test this hypothesis, we analyzed the conversion of BA by PqsBC. Using octanoyl-CoA consumption as a readout for the reaction, a greatly diminished reaction velocity of 0.02 s^{-1} was observed.

As a complementary approach, we reintroduced the asparagine into the active site of PqsBC. The crystal structure reveals Val-299 occupies an identical position to the corresponding asparagine residue in PqsD (see Fig. 3B), and no steric hindrance could be predicted for the amide group extension. PqsBC^{V299N} has a very low activity toward 2-ABA ($k_{\text{cat}} = 0.013 \text{ s}^{-1}$). Because the observed K_D value of the complex with 2-AA was 20-fold higher compared with the wild-type enzyme (Table 2), binding of the 2-aminophenyl moiety possibly is spatially impaired, leading to a substrate orientation unfavorable for catalysis. However, BA was converted with a 14-fold increased k_{cat} of 0.18 s^{-1} by PqsBC^{V299N}. Taken together, the data strongly support a model in which 2-ABA itself supplies the asparagine-equivalent amino function that assists in catalysis of the decarboxylation step.

Substitution of Val-299 by Asparagine Impacts Substrate Specificity—Besides condensing anthraniloyl-CoA and malonyl-CoA or malonyl-ACP, PqsD can also mediate the condensation of anthraniloyl-CoA and 3-ketodecanoic acid to form HHQ (53). If PqsBC catalyzed the decarboxylative condensation of octanoyl-CoA and malonyl-CoA, 3-ketodecanoyl-CoA would be formed, so PqsBC might provide a substrate for PqsD-mediated HHQ synthesis. We therefore analyzed PqsBC and PqsBC^{V299N} with respect to their activity toward malonyl-CoA. Although the turnover number of wild-type PqsBC with malonyl-CoA was only 0.0005 s^{-1} and the PqsBC^{H269A} protein was even less active, the V299N variant showed a 5-fold higher activity. Thus, substitution of the active-site Asn by Val, among other adaptations, not only sets PqsBC apart from the canonical FabH-like enzymes, but also reduces the cross-reactivity with malonyl-CoA. With respect to AQ production by *P. aeruginosa*, however, the activity of both PqsBC and PqsBC^{V299N} with malonyl-CoA is far too low to be considered an efficient alter-

native path for providing a 3-ketodecanoyl moiety for PqsD-supported synthesis of HHQ.

Discussion

PqsBC, a key enzyme in the biosynthesis of the quorum-sensing signal molecules HHQ and PQS and a member of the β -ketoacyl-ACP-synthase III family of condensing enzymes, has unique features of a heterodimeric subunit composition and the lack of the canonical active-site asparagine. The data presented provide the first crystal structure of PqsBC, and by combining structural data with kinetic and spectroscopic experiments, we were able to characterize individual reaction steps.

Although catalytic involvement of the PqsC subunit has been demonstrated in a previous study by identifying the acylation of Cys-129 (20), the role of PqsB was enigmatic. The crystal structure of PqsBC^{C129A} shows that PqsB has no catalytic residues, and large differences occur in the overall structure compared with PqsC and FabH. The orientation of PqsC and PqsB subunits forms a novel dimer with a pseudo-2-fold symmetry that is unexpectedly similar to the homodimer in FabH-like proteins, and it is likely that PqsB has a scaffolding role in formation of the heterodimer.

The structure of PqsC reveals major structural alterations in the area of the entry to the active site. The shortened PqsC helix α_5 , a prominent difference to PqsD and FabH, widens the entry to the catalytic site and provides an extended cavity that could accommodate a bicyclic molecule such as the reaction product HHQ.

Within the catalytic cavity of PqsC, the distance and the conformation of Cys-129 and His-269, the two remaining canonical catalytic residues of FabH-like enzymes, are conserved, pointing toward conserved functions in the catalytic mechanism. Compared with malonyl-ACP, the second substrate of FabH, the equivalent substrate 2-ABA of PqsBC (Fig. 1) has the advantage of UVA-absorbing and distinct fluorescent properties that allowed a series of experiments that provided insight into the reaction mechanism. Moreover, the spectroscopic properties of 2-AA, which we identified as a potent competitive inhibitor of PqsBC, enabled us to separately address the initial acylation step. Octanoylation of PqsBC induced a significant increase in affinity for 2-AA, which, albeit less pronounced, could also be measured for 2-ABA binding by the octanoyl-PqsBC^{H269A} protein. Even though similar effects have not been demonstrated for other FabH-like enzymes, such an affinity shift caused by the loading step could be a common trait of FabH-like enzymes to prevent premature binding of the second substrate. Equilibrium titrations suggested that His-269 is a major factor for the acylation-dependent affinity shift.

With respect to the role of the catalytic histidine in the acylation reaction, contradictory models on its influence on the catalytic cysteine in FabH (48, 49) and chalcone synthase (54, 55), a closely related polyketide synthase, have been discussed. In PqsBC, the velocity of octanoylation was virtually the same for the wild-type protein and the PqsBC^{H269A} variant, indicating that the histidine is not required for activation of the acyl-accepting cysteine. Given the similar conformations and distances between the catalytic histidines and cysteines in PqsC and FabH and the fact that Cys-129 of PqsC, like Cys-112 in

Structure and Function of the Condensing Enzyme PqsBC

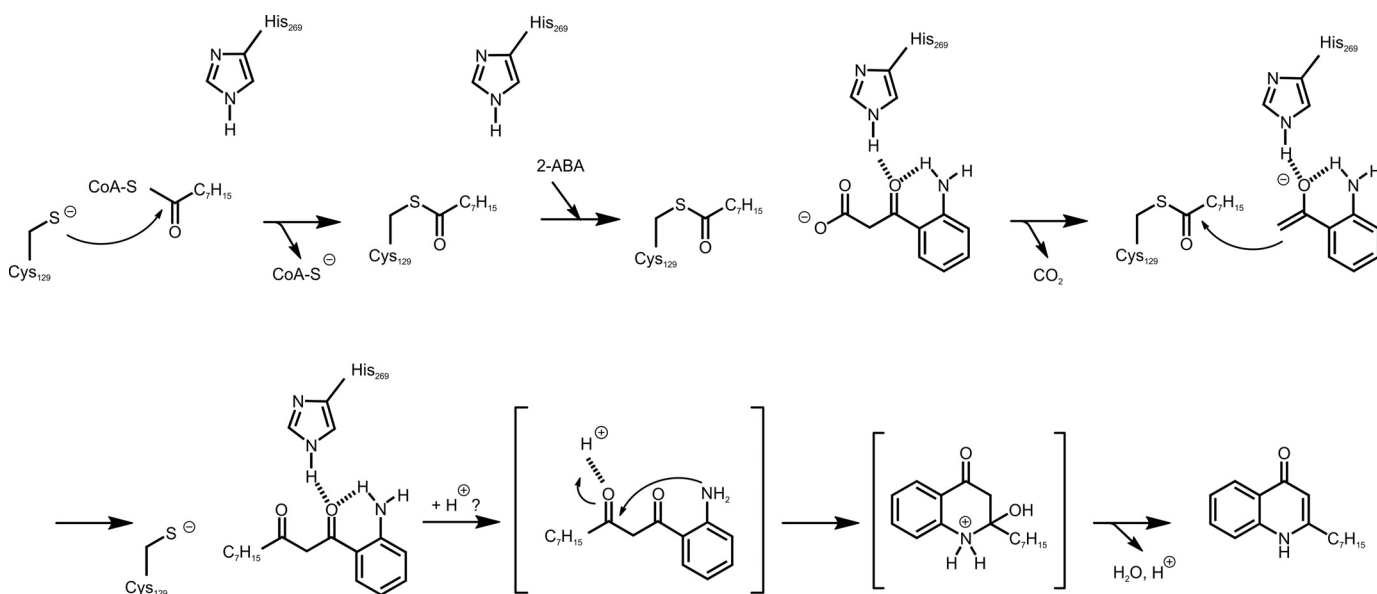


FIGURE 11. Proposed catalytic mechanism of PqsBC toward the formation of the 1-(2-aminophenyl)decane-1,3-dione intermediate and its reaction to HHQ. For details, see text.

FabH, is located at the N terminus of an α -helix (Fig. 5), it is likely that the α -helix dipole effect reduces the pK_a value of Cys to facilitate formation of the thiolate ion necessary for transacylase activity, as suggested by Davies *et al.* (48) for FabH. Also, a proton relay mechanism, as hypothesized by Qiu *et al.* (49), seems unlikely for PqsC.

The major question of how PqsBC is functional without the catalytic asparagine was addressed with the desamino-substrate analog BA and by reintroducing the asparagine into the protein. Whereas for the wild-type protein the activity toward BA was about 300-fold lower than toward 2-ABA, the PqsBC^{V299N} variant performed significantly better with BA than with 2-ABA. As demonstrated previously for 2-AA (56), the amino group of 2-ABA is hydrogen bonded to its β -keto group (Fig. 11). This intramolecular hydrogen bond replaces the hydrogen bond formed by the active-site Asn of FabH with the thioester carbonyl of malonyl-ACP. Taken together with the observed His-269-dependent binding effect, which (as suggested by equilibrium binding experiments) does not seem to distinguish between 2-ABA and 2-AA, we propose that the amine of the substrate and the histidine's imidazole activate the β -keto oxygen by hydrogen bonding, which promotes decarboxylation and stabilizes the enolate required for nucleophilic attack at the acyl-cysteine (Fig. 11). The reaction product of the subsequent decarboxylative Claisen condensation would be 1-(2-aminophenyl)decane-1,3-dione, which continues to react to form HHQ via intramolecular enamine formation. Even though the active site offers enough space for this consecutive step, the way in which the putative intermediate probably is bound (Fig. 11) renders an on-site ring closing reaction unlikely, because activation of the carbonyl for the subsequent cyclization and elimination step should rather involve a general acid that provides a proton to the keto group that is distal to the aromatic ring. Moreover, the cyclization would require a conformational change within the substrate to bring the amino group in proximity to the distal carbonyl carbon.

Given a spatial proximity of the amine and the 3-carbonyl, ring closure could also occur non-enzymatically. Taken together, this work significantly increases our knowledge of the structure, enzyme mechanism, and inhibition of PqsBC, and in a broader dimension, the work has implications for the development of novel therapeutics that control *P. aeruginosa* infection by attenuating virulence gene expression.

Author Contributions—S. F. and P. W. conceived the study. S. F., J. E., and P. W. coordinated the study. S. L. D., S. F., and J. E. wrote the paper. S. L. D. optimized the synthesis of 2-ABA, prepared PqsBC, and designed, performed, and analyzed all spectroscopic, kinetic, and physiological experiments (Figs. 7–11 and Table 2). F. P. and M. S. performed crystallization, and I. D. and C. L. performed data collection. J. E. performed molecular replacement calculations and density modification. C. L. performed data processing and structure refinement. U. H. prepared 2-aminobenzoylacetate and its precursor, performed all MS and NMR analyses, and contributed to the mechanistic discussion. All authors reviewed the results and approved the final version of the manuscript.

Acknowledgment—We acknowledge the Diamond Light Source for provision of the synchrotron radiation source in using the beamline I04.

References

- Lyczak, J. B., Cannon, C. L., and Pier, G. B. (2000) Establishment of *Pseudomonas aeruginosa* infection: lessons from a versatile opportunist. *Microbes Infect.* **2**, 1051–1060
- Williams, P. (2007) Quorum sensing, communication and cross-kingdom signalling in the bacterial world. *Microbiology* **153**, 3923–3938
- Storz, M. P., Maurer, C. K., Zimmer, C., Wagner, N., Brenzel, C., de Jong, J. C., Lucas, S., Müsken, M., Häußler, S., Steinbach, A., and Hartmann, R. W. (2012) Validation of PqsD as an anti-biofilm target in *Pseudomonas aeruginosa* by development of small-molecule inhibitors. *J. Am. Chem. Soc.* **134**, 16143–16146
- Starkey, M., Lepine, F., Maura, D., Bandyopadhyaya, A., Lesic, B., He, J., Kitao, T., Righi, V., Milot, S., Tzika, A., and Rahme, L. (2014) Identification

- of anti-virulence compounds that disrupt quorum-sensing regulated acute and persistent pathogenicity. *PLoS Pathog.* **10**, e1004321
5. Lu, C., Maurer, C. K., Kirsch, B., Steinbach, A., and Hartmann, R. W. (2014) Overcoming the unexpected functional inversion of a PqsR antagonist in *Pseudomonas aeruginosa*: an *in vivo* potent antivirulence agent targeting pqs quorum sensing. *Angew. Chem. Int. Ed. Engl.* **53**, 1109–1112
 6. Ilangovan, A., Fletcher, M., Rampioni, G., Pustelny, C., Rumbaugh, K., Heeb, S., Cámara, M., Truman, A., Chhabra, S. R., Emsley, J., and Williams, P. (2013) Structural basis for native agonist and synthetic inhibitor recognition by the *Pseudomonas aeruginosa* quorum sensing regulator PqsR (MvfR). *PLoS Pathog.* **9**, e1003508
 7. Rampioni, G., Leoni, L., and Williams, P. (2014) The art of antibacterial warfare: Deception through interference with quorum sensing-mediated communication. *Bioorg. Chem.* **55**, 60–68
 8. Jimenez, P. N., Koch, G., Thompson, J. A., Xavier, K. B., Cool, R. H., and Quax, W. J. (2012) The multiple signaling systems regulating virulence in *Pseudomonas aeruginosa*. *Microbiol. Mol. Biol. Rev.* **76**, 46–65
 9. Heeb, S., Fletcher, M. P., Chhabra, S. R., Diggle, S. P., Williams, P., and Cámara, M. (2011) Quinolones: from antibiotics to autoinducers. *FEMS Microbiol. Rev.* **35**, 247–274
 10. Huse, H., and Whiteley, M. (2011) 4-Quinolones: smart phones of the microbial world. *Chem. Rev.* **111**, 152–159
 11. Mashburn, L. M., and Whiteley, M. (2005) Membrane vesicles traffic signals and facilitate group activities in a prokaryote. *Nature* **437**, 422–425
 12. Diggle, S. P., Matthijs, S., Wright, V. J., Fletcher, M. P., Chhabra, S. R., Lamont, I. L., Kong, X., Hider, R. C., Cornelis, P., Cámara, M., and Williams, P. (2007) The *Pseudomonas aeruginosa* 4-quinolone signal molecules HHQ and PQS play multifunctional roles in quorum sensing and iron entrapment. *Chem. Biol.* **14**, 87–96
 13. Bredenbruch, F., Geffers, R., Nimtz, M., Buer, J., and Häussler, S. (2006) The *Pseudomonas aeruginosa* quinolone signal (PQS) has an iron-chelating activity. *Environ. Microbiol.* **8**, 1318–1329
 14. Häussler, S., and Becker, T. (2008) The *Pseudomonas* quinolone signal (PQS) balances life and death in *Pseudomonas aeruginosa* populations. *PLoS Pathog.* **4**, e1000166
 15. Hooi, D. S., Bycroft, B. W., Chhabra, S. R., Williams, P., and Pritchard, D. I. (2004) Differential immune modulatory activity of *Pseudomonas aeruginosa* quorum-sensing signal molecules. *Infect. Immun.* **72**, 6463–6470
 16. Skindersoe, M. E., Zeuthen, L. H., Brix, S., Fink, L. N., Lazenby, J., Whittall, C., Williams, P., Diggle, S. P., Froekiaer, H., Cooley, M., and Givskov, M. (2009) *Pseudomonas aeruginosa* quorum-sensing signal molecules interfere with dendritic cell-induced T-cell proliferation. *FEMS Immunol. Med. Microbiol.* **55**, 335–345
 17. Hänsch, G. M., Prior, B., Brenner-Weiss, G., Obst, U., and Overhage, J. (2014) The *Pseudomonas* quinolone signal (PQS) stimulates chemotaxis of polymorphonuclear neutrophils. *J. Appl. Biomater. Funct. Mater.* **12**, 21–26
 18. Coleman, J. P., Hudson, L. L., McKnight, S. L., Farrow, J. M., 3rd., Calfee, M. W., Lindsey, C. A., and Pesci, E. C. (2008) *Pseudomonas aeruginosa* PqsA is an anthranilate-coenzyme A ligase. *J. Bacteriol.* **190**, 1247–1255
 19. Zhang, Y.-M., Frank, M. W., Zhu, K., Mayasundari, A., and Rock, C. O. (2008) PqsD is responsible for the synthesis of 2,4-dihydroxyquinoline, an extracellular metabolite produced by *Pseudomonas aeruginosa*. *J. Biol. Chem.* **283**, 28788–28794
 20. Dulcey, C. E., Dekimpe, V., Fauvelle, D. A., Milot, S., Groleau, M. C., Doucet, N., Rahme, L. G., Lépine, F., and Déziel, E. (2013) The end of an old hypothesis: the *Pseudomonas* signaling molecules 4-hydroxy-2-alkylquinolines derive from fatty acids, not 3-ketofatty acids. *Chem. Biol.* **20**, 1481–1491
 21. Drees, S. L., and Fetzner, S. (2015) PqsE of *Pseudomonas aeruginosa* acts as pathway-specific thioesterase in the biosynthesis of alkylquinolone signaling molecules. *Chem. Biol.* **22**, 611–618
 22. Kesarwani, M., Hazan, R., He, J., Que, Y.-A., Que, Y., Apidianakis, Y., Lesic, B., Xiao, G., Dekimpe, V., Milot, S., Déziel, E., Lépine, F., and Rahme, L. G. (2011) A quorum sensing regulated small volatile molecule reduces acute virulence and promotes chronic infection phenotypes. *PLoS Pathog.* **7**, e1002192
 23. Que, Y.-A., Hazan, R., Strobel, B., Maura, D., He, J., Kesarwani, M., Panooulos, P., Tsurumi, A., Giddey, M., Wilhelmy, J., Mindrinos, M. N., and Rahme, L. G. (2013) A quorum sensing small volatile molecule promotes antibiotic tolerance in bacteria. *PLoS ONE* **8**, e80140
 24. Bandyopadhyaya, A., Kesarwani, M., Que, Y.-A., He, J., Padfield, K., Tompkins, R., and Rahme, L. G. (2012) The quorum sensing volatile molecule 2-amino acetophenon modulates host immune responses in a manner that promotes life with unwanted guests. *PLoS Pathog.* **8**, e1003024
 25. Tzika, A. A., Constantinou, C., Bandyopadhyaya, A., Psychogios, N., Lee, S., Mindrinos, M., Martyn, J. A., Tompkins, R. G., and Rahme, L. G. (2013) A small volatile bacterial molecule triggers mitochondrial dysfunction in murine skeletal muscle. *PLoS ONE* **8**, e74528
 26. Schertzner, J. W., Brown, S. A., and Whiteley, M. (2010) Oxygen levels rapidly modulate *Pseudomonas aeruginosa* social behaviours via substrate limitation of PqsH. *Mol. Microbiol.* **77**, 1527–1538
 27. Bera, A. K., Atanasova, V., Robinson, H., Eisenstein, E., Coleman, J. P., Pesci, E. C., and Parsons, J. F. (2009) Structure of PqsD, a *Pseudomonas* quinolone signal biosynthetic enzyme, in complex with anthranilate. *Biochemistry* **48**, 8644–86455
 28. Heath, R. J., and Rock, C. O. (2002) The Claisen condensation in biology. *Nat. Prod. Rep.* **19**, 581–596
 29. van den Ent, F., and Löwe, J. (2006) RF cloning: a restriction-free method for inserting target genes into plasmids. *J. Biochem. Biophys. Methods* **67**, 67–74
 30. Gill, S. C., and von Hippel, P. H. (1989) Calculation of protein extinction coefficients from amino acid sequence data. *Anal. Biochem.* **182**, 319–326
 31. Cabrita, L. D., Gilis, D., Robertson, A. L., Dehouck, Y., Rooman, M., and Bottomley, S. P. (2007) Enhancing the stability and solubility of TEV protease using *in silico* design. *Protein Sci.* **16**, 2360–2367
 32. Needham, E. R., and Perkin, W. H. (1904) XVI. *o*-Nitrobenzoylacetate. *J. Chem. Soc. Trans.* **85**, 148–155
 33. Sicker, D., and Mann, G. (1988) Synthesis of ethyl *ortho*-substituted benzoylacetates and investigation of the influence of *ortho*-substituents on keto-enol tautomerism and MS fragmentation behaviour. *Collect. Czechoslov. Chem. Commun.* **53**, 839–850
 34. Schöpf, C., Koepke, G., Kowald, B., Schülde, F., and Wunderlich, D. (1956) Darstellung und eigenschaften der *o*-formamino- und der *o*-amino-benzoylessigsäure, zweier zellmöglicher abbauprodukte des tryptophans und heteroauxins. *Chem. Ber.* **89**, 2877–2886
 35. Birdsall, B., King, R. W., Wheeler, M. R., Lewis, C. A., Jr., Goode, S. R., Dunlap, R. B., and Roberts, G. C. (1983) Correction for light absorption in fluorescence studies of protein-ligand interactions. *Anal. Biochem.* **132**, 353–361
 36. Eyer, P., Worek, F., Kiderlen, D., Sinko, G., Stuglin, A., Simeon-Rudolf, V., and Reiner, E. (2003) Molar absorption coefficients for the reduced Ellman reagent: reassessment. *Anal. Biochem.* **312**, 224–227
 37. Niewerth, H., Bergander, K., Chhabra, S. R., Williams, P., and Fetzner, S. (2011) Synthesis and biotransformation of 2-alkyl-4(1*H*)-quinolones by recombinant *Pseudomonas putida* KT2440. *Appl. Microbiol. Biotechnol.* **91**, 1399–1408
 38. Keegan, R. M., and Winn, M. D. (2007) Automated search-model discovery and preparation for structure solution by molecular replacement. *Acta Crystallogr. D Biol. Crystallogr.* **63**, 447–457
 39. McCoy, A. J., Grosse-Kunstleve, R. W., Adams, P. D., Winn, M. D., Storoni, L. C., and Read, R. J. (2007) Phaser crystallographic software. *J. Appl. Crystallogr.* **40**, 658–674
 40. Cowtan, K. (2006) The Buccaneer software for automated model building. 1. Tracing protein chains. *Acta Crystallogr. D Biol. Crystallogr.* **62**, 1002–1011
 41. Winn, M. D., Ballard, C. C., Cowtan, K. D., Dodson, E. J., Emsley, P., Evans, P. R., Keegan, R. M., Krissinel, E. B., Leslie, A. G., McCoy, A., McNicholas, S. J., Murshudov, G. N., Pannu, N. S., Potterton, E. A., Powell, H. R., et al. (2011) Overview of the CCP4 suite and current developments. *Acta Crystallogr. D Biol. Crystallogr.* **67**, 235–242
 42. Zhang, K. Y., Cowtan, K., and Main, P. (1997) Macromolecular crystallography Part B. *Methods Enzymol.* **277**, 53–64
 43. Vagin, A. A., Steiner, R. A., Lebedev, A. A., Potterton, L., McNicholas, S., Long, F., and Murshudov, G. N. (2004) REFMAC5 dictionary: organization of prior chemical knowledge and guidelines for its use. *Acta Crystal-*

Structure and Function of the Condensing Enzyme PqsBC

- logr. D Biol. Crystallogr.* **60**, 2184–2195
44. Krissinel, E., and Henrick, K. (2007) Inference of macromolecular assemblies from crystalline state. *J. Mol. Biol.* **372**, 774–797
 45. Sachdeva, S., Musayev, F. N., Alhamadsheh, M. M., Scarsdale, J. N., Wright, H. T., and Reynolds, K. A. (2008) Separate entrance and exit portals for ligand traffic in *Mycobacterium tuberculosis* FabH. *Chem. Biol.* **15**, 402–412
 46. Voss, N. R., and Gerstein, M. (2010) 3V: cavity, channel and cleft volume calculator and extractor. *Nucleic Acids Res.* **38**, W555–W562
 47. Zhang, Z., Li, Y., Lin, B., Schroeder, M., and Huang, B. (2011) Identification of cavities on protein surface using multiple computational approaches for drug binding site prediction. *Bioinformatics* **27**, 2083–2088
 48. Davies, C., Heath, R. J., White, S. W., and Rock, C. O. (2000) The 1.8 Å crystal structure and active-site architecture of β -ketoacyl-acyl carrier protein synthase III (FabH) from *Escherichia coli*. *Structure* **8**, 185–195
 49. Qiu, X., Janson, C. A., Smith, W. W., Head, M., Lonsdale, J., and Konstantinidis, A. K. (2001) Refined structures of β -ketoacyl-acyl carrier protein synthase III. *J. Mol. Biol.* **307**, 341–356
 50. Pan, H., Tsai Sc., Meadows, E. S., Miercke, L. J., Keatinge-Clay, A. T., O'Connell, J., Khosla, C., and Stroud, R. M. (2002) Crystal structure of the priming β -ketosynthase from the R1128 polyketide biosynthetic pathway. *Structure* **10**, 1559–1568
 51. Lépine, F., Dekimpe, V., Lesic, B., Milot, S., Lesimple, A., Mamer, O. A., Rahme, L. G., and Déziel, E. (2007) PqsA is required for the biosynthesis of 2,4-dihydroxyquinoline (DHQ), a newly identified metabolite produced by *Pseudomonas aeruginosa* and *Burkholderia thailandensis*. *Biol. Chem.* **388**, 839–845
 52. Cox, C. D., and Parker, J. (1979) Use of 2-aminoacetophenone production in identification of *Pseudomonas aeruginosa*. *J. Clin. Microbiol.* **9**, 479–484
 53. Steinbach, A., Maurer, C. K., Weidel, E., Henn, C., Brengel, C., Hartmann, R. W., and Negri, M. (2013) Molecular basis of HHQ biosynthesis: molecular dynamics simulations, enzyme kinetic and surface plasmon resonance studies. *BMC Biophys.* **6**, 10
 54. Jez, J. M., and Noel, J. P. (2000) Mechanism of chalcone synthase. pK_a of the catalytic cysteine and the role of the conserved histidine in a plant polyketide synthase. *J. Biol. Chem.* **275**, 39640–39646
 55. Suh, D. Y., Kagami, J., Fukuma, K., and Sankawa, U. (2000) Evidence for catalytic cysteine-histidine dyad in chalcone synthase. *Biochem. Biophys. Res. Commun.* **275**, 725–730
 56. Shimada, H., Nakamura, A., Yoshihara, T., and Tobita, S. (2005) Intramolecular and intermolecular hydrogen-bonding effects on photophysical properties of 2'-aminoacetophenone and its derivatives in solution. *Photochem. Photobiol. Sci.* **4**, 367–375

PqsBC, a Condensing Enzyme in the Biosynthesis of the *Pseudomonas aeruginosa* Quinolone Signal: CRYSTAL STRUCTURE, INHIBITION, AND REACTION MECHANISM

Steffen Lorenz Drees, Chan Li, Fajar Prasetya, Muhammad Saleem, Ingrid Dreveny, Paul Williams, Ulrich Hennecke, Jonas Emsley and Susanne Fetzner

J. Biol. Chem. 2016, 291:6610-6624.

doi: 10.1074/jbc.M115.708453 originally published online January 25, 2016

Access the most updated version of this article at doi: [10.1074/jbc.M115.708453](https://doi.org/10.1074/jbc.M115.708453)

Alerts:

- [When this article is cited](#)
- [When a correction for this article is posted](#)

[Click here](#) to choose from all of JBC's e-mail alerts

Supplemental material:

<http://www.jbc.org/content/suppl/2016/01/25/M115.708453.DC1>

This article cites 56 references, 6 of which can be accessed free at

<http://www.jbc.org/content/291/13/6610.full.html#ref-list-1>