

Dear Author

Here are the proofs of your article.

- You can submit your corrections **online**, via **e-mail** or by **fax**.
- For **online** submission please insert your corrections in the online correction form. Always indicate the line number to which the correction refers.
- You can also insert your corrections in the proof PDF and email the annotated PDF.
- For **fax** submission, please ensure that your corrections are clearly legible. Use a fine black pen and write the correction in the margin, not too close to the edge of the page.
- Remember to note the **journal title**, **article number**, and **your name** when sending your response via e-mail or fax.
- **Check** the metadata sheet to make sure that the header information, especially author names and the corresponding affiliations are correctly shown.
- Check the questions that may have arisen during copy editing and insert your answers/corrections.
- Check that the text is complete and that all figures, tables and their legends are included. Also check the accuracy of special characters, equations, and electronic supplementary material if applicable. If necessary refer to the *Edited manuscript*.
- The publication of inaccurate data such as dosages and units can have serious consequences. Please take particular care that all such details are correct.
- Please **do not** make changes that involve only matters of style. We have generally introduced forms that follow the journal's style.
- Substantial changes in content, e.g., new results, corrected values, title and authorship are not allowed without the approval of the responsible editor. In such a case, please contact the Editorial Office and return his/her consent together with the proof.
- If we do not receive your corrections within 48 hours, we will send you a reminder.
- Your article will be published **Online First** approximately one week after receipt of your corrected proofs. This is the **official first publication** citable with the DOI. **Further changes are, therefore, not possible.**
- The **printed version** will follow in a forthcoming issue.

Please note

After online publication, subscribers (personal/institutional) to this journal will have access to the complete article via the DOI using the URL:

http://dx.doi.org/10.1007/s11302-013-9403-2

If you would like to know when your article has been published online, take advantage of our free alert service. For registration and further information, go to: <u>http://www.springerlink.com</u>.

Due to the electronic nature of the procedure, the manuscript and the original figures will only be returned to you on special request. When you return your corrections, please inform us, if you would like to have these documents returned.

Metadata of the article that will be visualized in OnlineFirst

1	Article Title	Investigation of the functional expression of purine and pyrimidine receptors in porcine-isolated pancreatic arteries		
2	Article Sub-Title			
3	Article Copyright - Year	Springer Science+Business Media Dordrecht 2013 (This will be the copyright line in the final PDF)		
4	Journal Name	Purinergic Sign	alling	
5		Family Name	Ralevic	
6		Particle		
7		Given Name	V.	
8	Corresponding	Suffix		
9	Author	Organization	University of Nottingham	
10		Division	School of Biomedical Sciences	
11		Address	Nottingham NG7 2UH, UK	
12		e-mail	vera.ralevic@nottingham.ac.uk	
13		Family Name	Alsaqati	
14		Particle		
15		Given Name	М.	
16	Author	Suffix		
17	Author	Organization	University of Nottingham	
18		Division	School of Biomedical Sciences	
19		Address	Nottingham NG7 2UH, UK	
20		e-mail		
21		Family Name	Chan	
22		Particle		
23		Given Name	S. L. F.	
24	Author	Suffix		
25	Author	Organization	University of Nottingham	
26		Division	School of Biomedical Sciences	
27		Address	Nottingham NG7 2UH, UK	
28		e-mail		
29	Sebadula	Received	2 July 2013	
30	Schedule	Revised		

31		Accepted 5 November 2013
32	Abstract	Receptors for purines and pyrimidines are expressed throughout the cardiovascular system. This study investigated their functional expression in porcine-isolated pancreatic arteries. Pancreatic arteries (endothelium intact or denuded) were prepared for isometric tension recording and preconstricted with U46619, a thromboxane A ₂ mimetic; adenosine-5'-diphosphate (ADP), uridine-5'-triphosphate (UTP) and MRS2768, a selective P2Y ₂ agonist, were applied cumulatively, while adenosine-5'-triphosphate (ATP) and $\alpha\beta$ -methylene-ATP ($\alpha\beta$ -meATP) response curves were generated from single concentrations per tissue segment. Antagonists/enzyme inhibitors were applied prior to U46619 addition. ATP, $\alpha\beta$ -meATP, UTP and MRS2768 induced vasoconstriction, with a potency order of $\alpha\beta$ -meATP > MRS2768 > ATP ≥ UTP. Contractions to ATP and $\alpha\beta$ -meATP were blocked by NF449, a selective P2X1 receptor antagonist. The contraction induced by ATP, but not UTP, was followed by vasorelaxation. Endothelium removal and DUP 697, a cyclooxygenase-2 inhibitor, had no significant effect on contraction to ATP but attenuated that to UTP, indicating actions at distinct receptors. MRS2578, a selective P2Y ₆ receptor antagonist, had no effect on contractions to UTP. ADP induced endothelium-dependent vasorelaxation which was inhibited by MRS2179, a selective P2Y ₁ receptor antagonist. The contractions to ATP and $\alpha\beta$ -meATP were attributed to actions at P2X1 receptors on the vascular smooth muscle, whereas it was shown for the first time that UTP induced an endothelium-dependent vasoconstriction which may involve P2Y ₂ and/or P2Y ₄ receptors. The relaxation induced by ADP is mediated by P2Y ₁ and A _{2A} adenosine receptors. Porcine pancreatic arteries appear to lack vasorelaxant P2Y ₂ and P2Y ₄ receptors.
33	Keywords separated by ' - '	$\alpha\beta$ -meATP - ATP - UTP - ADP - MRS2578 - P2Y_1 - P2Y_2 - P2X1 - A_{2A} adenosine receptors - Vasoconstriction - Relaxation - Endothelium
34	Foot note information	

Purinergic Signalling DOI 10.1007/s11302-013-9403-2

ORIGINAL ARTICLE

Investigation of the functional expression of purine

and pyrimidine receptors in porcine-isolated pancreatic arteries

8 M. Alsaqati • S. L. F. Chan • V. Ralevic

10

32

4

5

6

Received: 2 July 2013 / Accepted: 5 November 2013
 © Springer Science+Business Media Dordrecht 2013

Abstract Receptors for purines and pyrimidines are 13expressed throughout the cardiovascular system. This study 14investigated their functional expression in porcine-isolated 15pancreatic arteries. Pancreatic arteries (endothelium intact or 16denuded) were prepared for isometric tension recording and 1718 preconstricted with U46619, a thromboxane A₂ mimetic; adenosine-5'-diphosphate (ADP), uridine-5'-triphosphate 19(UTP) and MRS2768, a selective P2Y₂ agonist, were applied 2021cumulatively, while adenosine-5'-triphosphate (ATP) and $\alpha\beta$ methylene-ATP ($\alpha\beta$ -meATP) response curves were generated 22from single concentrations per tissue segment. Antagonists/ 2324enzyme inhibitors were applied prior to U46619 addition. 25ATP, αβ-meATP, UTP and MRS2768 induced vasoconstriction, with a potency order of $\alpha\beta$ -meATP > MRS2768 > ATP > 26UTP. Contractions to ATP and $\alpha\beta$ -meATP were blocked by 2728NF449, a selective P2X1 receptor antagonist. The contraction induced by ATP, but not UTP, was followed by vasorelax-29ation. Endothelium removal and DUP 697, a cyclooxygenase-30 2 inhibitor, had no significant effect on contraction to ATP but 31 32 attenuated that to UTP, indicating actions at distinct receptors. MRS2578, a selective P2Y₆ receptor antagonist, had no effect 33on contractions to UTP. ADP induced endothelium-dependent 34vasorelaxation which was inhibited by MRS2179, a selective 35P2Y₁ receptor antagonist, or SCH58261, a selective adeno-36 37 sine A2A receptor antagonist. The contractions to ATP and $\alpha\beta$ -meATP were attributed to actions at P2X1 receptors on 38 39the vascular smooth muscle, whereas it was shown for the first time that UTP induced an endothelium-dependent vasocon-40striction which may involve P2Y2 and/or P2Y4 receptors. The 41 relaxation induced by ADP is mediated by P2Y₁ and A_{2A} 42

> M. Alsaqati · S. L. F. Chan · V. Ralevic (⊠) School of Biomedical Sciences, University of Nottingham, Nottingham NG7 2UH, UK e-mail: vera.ralevic@nottingham.ac.uk

adenosme rec	reptors. For the pancieatic arteries appear to fack	45
vasorelaxant	$P2Y_2$ and $P2Y_4$ receptors.	44
Keywords o	$\kappa\beta$ -meATP · ATP · UTP · ADP · MRS2578 ·	45
$P2Y_1 \cdot P2Y_2$	\cdot P2X1 \cdot A _{2A} adenosine receptors \cdot	46
Vasoconstrict	tion · Relaxation · Endothelium	47
Abbreviatio	ns	48
αβ-meATP	$\alpha\beta$ -Methylene-adenosine-5'-triphosphate	50
ADP	Adenosine-5'-diphosphate	53
ATP	Adenosine-5'-triphosphate	54
EDCFs	Endothelium-derived contractile factors	56
ENTPDase	Ecto-nucleotidase 5'-triphosphate	59
	diphosphohydrolase	60
PPADS	Pyridoxal phosphate-6-azo(benzene-2,4-	62
	disulfonic acid)	63
UTP	Uridine-5'-triphosphate	64
VSMCs	Vascular smooth muscle cells	66
XAC	Xanthine amine congener	68
		(1)

Introduction

71

The activities of both exocrine and endocrine cells of the 72pancreas are regulated by autonomic nerves (parasympathetic 73 and sympathetic) as well as by hormones and autocrine and 74paracrine mediators. Although the exact mechanisms remain 75to be established, it is generally agreed that an increase in 76endocrine cell activity during hormone secretion corresponds 77 with an increase in blood flow, to meet the metabolic demand. 78The role of exogenous purine and pyrimidine nucleotides in 79controlling the functions of endocrine and exocrine compo-80 nents of the pancreas are well described [1, 2], but little is 81 known about their effects on pancreatic arterial 82 vasocontractility. 83

There are two main families of P2 purine and pyrimidine 84 receptors, ionotropic P2X and G protein-coupled P2Y recep-85 tors. Molecular cloning has identified seven mammalian P2X 86 87 receptor subunits: P2X1, P2X2, P2X3, P2X4, P2X5, P2X6 88 and P2X7 [3], while eight mammalian P2Y receptors have been identified: P2Y1, P2Y2, P2Y4, P2Y6, P2Y11, P2Y12, 89 90 P2Y₁₃ and P2Y₁₄ [4]. P2X receptors are activated by adenosine-5'-triphosphate (ATP) and its stable, and conse-91quently more potent, analogue $\alpha\beta$ -methylene-ATP ($\alpha\beta$ -9293 meATP) [5, 6]. P2Y receptors can be divided on the basis of 94their endogenous agonists into adenine nucleotide-preferring 95 (P2Y₁, P2Y₁₁, P2Y₁₂ and P2Y₁₃) receptors and uracil nucleotide or UDP-sugar-preferring (P2Y2, P2Y4, P2Y6 and 96 P2Y₁₄) receptors [7]. Among the adenine nucleotide group, 97 the human P2Y₁₁ receptor is selectively activated by ATP and 98 99fails to respond to adenosine-5'-diphosphate (ADP) [8], although the dog orthologue responds to both ADP and ATP 100101 [9]. $P2Y_{1}$, $P2Y_{12}$ and $P2Y_{13}$ receptors are activated by ADP, 102with lower potency by ATP [10-13]. Among the uracil nucleotide or UDP-sugar receptors, P2Y₂ is equally activated by 103104 ATP and uridine-5'-triphosphate (UTP), while P2Y₄ receptor is highly selective for UTP over ATP [14]. The P2Y₆ receptor 105106 is activated by UDP and UTP, while the P2Y₁₄ receptor is activated by UDP and UDP-sugars [6, 15]. 107

Within the pancreatic vasculature, P2X1, P2X2, P2Y₁ and 108109P2Y₂ receptors were detected by immunohistochemistry [16]. More than two decades ago, it was shown that P2X receptors 110mediate pancreatic vasoconstriction, and P2Y receptors me-111112diate vasodilatation in response to ATP [17], and subsequent studies showed an additional involvement of contractile re-113ceptors sensitive to UTP (named P2U receptors) [18]. Purine 114115receptor subclassification has advanced significantly since that time. A re-evaluation of purine receptors in the pancreatic 116117 vasculature is clearly warranted. In the current study, we 118 describe the pharmacological characterisation of P2Y₁ and 119 A_{2A} receptor-mediated relaxatory responses, in addition to 120 P2X1, P2Y₂ and P2Y₄ receptor-mediated contractile re-121sponses of porcine-isolated pancreatic artery preparations. P2Y₂ and/or P2Y₄ receptors appear to be expressed mainly 122123in endothelial cells, while P2X1 and A2A receptors appear to 124be expressed in smooth muscle cells of the pancreatic arteries. 125A preliminary account of some of these data has previously been presented to the British Pharmacological Society [19]. 126

127 Materials and methods

Pancreases from pigs (either sex, age less than 6 months, wt
~50 kg) were obtained on ice from a local abattoir (G Wood &
Sons Ltd., Mansfield). A crude dissection was conducted to
isolate the porcine pancreatic arteries (greater pancreatic

artery) which were located in the body of the pancreas. The 133vessels were dissected out and placed in Krebs-Henseleit 134buffer containing 2 % (w/v) Ficoll (hydrophilic polysaccha-135ride, type 70) and were refrigerated overnight at 4 °C. The 136next day, a fine dissection was performed on arteries, and the 137 artery segments were cut into rings of about 0.5 cm in length 138and suspended in Krebs-Henseleit buffer (gassed, 95 % O₂, 1395 % CO₂). 140

The endothelium of some arteries was removed by gently 141 rubbing the innermost surface of the artery with forceps on a 142paper tissue before attaching it to the set-up [20]. Successful 143removal of the endothelium was tested using substance P 144 (10 nM). Endothelium-denuded arteries relaxed in response 145to substance P to less than 10 % of the U46619-induced 146contraction, while in endothelium-intact arteries, the relaxa-147tion to substance P was 36 $\% \pm 8$ (*n*=7, data not shown). 148

Responses in the porcine-isolated pancreatic artery 149

Arterial rings were mounted onto wires in tissue baths con-150taining warm (37 °C), oxygenated Krebs-Henseleit solution 151and were connected via isometric force transducers 152(mechanotransducer MLT 050/D; ADInstruments, Sydney, 153Australia) to a PC running the computer programme, 154LabChart (ADInstruments, Sydney, Australia). Rings were 155put under tension (15 g) and allowed to equilibrate for 15660 min before assessing the viability with two challenges of 15775 mM potassium chloride (KCl). The tissues were then 158allowed to equilibrate for 60 min, after which U46619 (10-159100 nM), a thromboxane A2 mimetic, was used to contract the 160tissues to between 40 and 80 % of the second KCl response. 161This ensured that if there was a vasodilator component to the 162response, this could be detected. Once an appropriate level of 163U46619 response had been achieved, ATP, $\alpha\beta$ -meATP, UTP, 164ADP or MRS2768 were added. Antagonists or enzyme inhib-165itors were applied 10 min prior to the addition of U46619, 166allowing them to be incubated with the tissues for a minimal 167 contact time of 30 min prior to the application of agonists. 168Some arteries were incubated with 0.1 % (v/v) DMSO (vehi-169cle control). 170

Reagents and	drugs
--------------	-------

171

Krebs-Henseleit buffer was composed of the following (mM): 172NaCl, 118; KCl, 4.8; CaCl₂·H₂O, 1.3; NaHCO₃, 25.0; 173KH₂PO₄, 1.2; MgSO₄·7H₂O, 1.2; and glucose, 11.1. Suramin, 174UTP, ATP, αβ-meATP, ADP, U46619, xanthine amine con-175gener (XAC) and SCH58261 (7-(2-phenylethyl)-5-amino-1762-(2-furyl)-pyrazolo-[4,3-e]-1,2,4-triazolo[1,5-c]pyrimidine) 177were purchased from Sigma (Poole, Dorset, UK), while DUP 178697 (5-bromo-2-(4-fluorophenyl)-1793-[4-(me thylsulfonyl)phenyl]-thiophene), pyridoxal 180 phosphate-6-azo(benzene-2,4-disulfonic acid) (PPADS), 181

Purinergic Signalling

MRS2578 $(N, N''-1, 4$ -butanediyl bis $(N'-[3-$
isothiocynatophenyl] thiourea)), MRS2179 (2'-deoxy-N ⁶ -
methyladenosine 3',5'-bisphosphate tetrasodium salt),
MRS2768 (uridine-5'-tetraphosphate δ -phenyl ester
tetrasodium salt) and substance P were purchased from Tocris
Biosciences Ltd. (Bristol, UK). NF449 (4,4',4",4'
"-[carbonylbis(imino-5,1,3-benzenetriyl-bis(carbonylimino))]
tetrakis-1,3-benzenedisulfonic acid) was purchased from
Calbiochem-Merck4Biosciences. U46619 was dissolved in
ethanol at 10 mM stock concentration. PPADS, suramin,
$\alpha\beta$ -meATP, ATP, ADP, UTP, NF449, MRS2179, MRS2768
and substance P were dissolved in distilled water. DUP 697,
XAC, MRS2578 and SCH58261 were dissolved in DMSO at
10 mM stock concentration.

The contractions to ATP, $\alpha\beta$ -meATP and UTP were measured 197198from the stabilised U46619-induced response and were expressed in grams, while the relaxations to ATP and ADP 199were expressed as a percentage of the U46619-induced con-200traction. Data were expressed as log concentration-response 201 202 plots. Values for all figures refer to mean \pm standard error of the mean (SEM) with 95 % confidence. Results were com-Q1 203 204pared by one- or two-way ANOVA with Bonferroni's post hoc 205test or unpaired Student's t test (Prism, GraphPad, San Diego, CA, USA). Differences were considered to be significant 206when the P value was <0.05. N expresses the number of 207208animals.

209 Results

Effect of purine and pyrimidine nucleotides on vascular tonein porcine-isolated pancreatic arteries

212To investigate the effect of purine and pyrimidine nucleotide 213agonists on porcine pancreatic arteries, $\alpha\beta$ -meATP (10 nM to 100 µM), ATP (1 µM to 10 mM), UTP (10 µM to 1 mM), 214ADP (1 µM to 1 mM) and MRS2768 (100 nM to 30 µM) 215216 were applied after preconstriction with U46619. The responses to ATP and $\alpha\beta$ -meATP were found to be desensitised 217rapidly. Therefore, they were applied at single concentrations 218219(one concentration per tissue segment). The responses to UTP, 220ADP and MRS2768 did not desensitise rapidly; thus, cumulative concentration-response curves were generated. ATP, 221222 $\alpha\beta$ -meATP, UTP and MRS2768 induced concentrationdependent contraction with a potency order of $\alpha\beta$ -meATP > 223MRS2768 > ATP \geq UTP (P<0.001, two-way ANOVA; 224Fig. 1a). The response to ATP was biphasic, since its contrac-225226tion was followed by a relaxation (Fig. 1b) which was equipotent to the concentration-dependent relaxation pro-227duced by ADP (Fig. 1a). The efficacies of ATP and $\alpha\beta$ -228



Fig. 1 a Concentration-dependent contraction of ATP, $\alpha\beta$ -meATP, UTP and MRS2768, a selective P2Y₂ agonist, and relaxation of ADP and ATP in U46619-preconstricted porcine pancreatic arteries (n=7–12). **b** Typical trace showing the biphasic response to ATP (contraction followed by relaxation). Data are presented as mean ± SEM

meATP in inducing contraction were similar and greater than 229 those of UTP or MRS2768. The relaxation to ADP and ATP at 230 the highest concentration of the agonists used (1 mM) was 231 similar at 4.5 ± 0.5 g (n=10) and 5.5 ± 0.2 g (n=7), respective-232 ly; there was no significant difference between these re-233 sponses (Fig. 1a). UTP, MRS2768 and $\alpha\beta$ -meATP did not 234 elicit vasorelaxation. 235

Characterisation of responses to ATP and $\alpha\beta$ -meATP	236
in U46619-preconstricted porcine-isolated pancreatic	237
arteries	238

Effect of suramin, PPADS and $\alpha\beta$ -meATP 239Responses to ATP and $\alpha\beta$ -meATP were characterised 240using the non-selective P2 receptor antagonists, suramin 241(100 µM) and PPADS (10 µM). Both suramin and PPADS 242significantly attenuated the contractions evoked by ATP 243(1 mM) and $\alpha\beta$ -meATP (1 μ M) (Fig. 2a, b). These con-244centrations of ATP and $\alpha\beta$ -meATP were chosen since 245they produced robust and submaximal responses, and for 246 $\alpha\beta$ -meATP, the concentration was close to the half max-247imal effective concentration (EC₅₀) value [mean EC_{50} 248Q2 value was 1.6 µM (95 % confidence interval (CI), 1.05 249to 2.53 μ M; n = 8; Fig. 1a)]. The relaxation to ATP was not 250affected by suramin or PPADS (Fig. 2c). Since $\alpha\beta$ -251meATP induces desensitisation of P2X receptors more 252

261

253readily than ATP because it is broken down more slowly254than ATP [5], the responses to ATP and $\alpha\beta$ -meATP were255studied in the presence of $\alpha\beta$ -meATP, in which $\alpha\beta$ -256meATP (1 µM) was added 10 min prior the addition of257U46619. As seen in Fig. 2a, b, the contractions to ATP and258 $\alpha\beta$ -meATP were reduced in the presence of the



Fig. 2 Effect of suramin (100 μ M), PPADS (10 μ M) and desensitisation by $\alpha\beta$ -meATP (1 μ M) on contractions to **a** ATP (1 mM) and **b** $\alpha\beta$ meATP (1 μ M) and **c** on the relaxation to ATP in U46619-preconstricted porcine pancreatic arteries. PPADS, suramin and $\alpha\beta$ -meATP reduced the contractions of **a** ATP and **b** $\alpha\beta$ -meATP (**P<0.01; ***P<0.001, oneway ANOVA with Bonferroni's post hoc test, responses of ATP or $\alpha\beta$ meATP vs their responses in the presence of PPADS, suramin or $\alpha\beta$ meATP, n =6–9). **c** The relaxation to ATP was not significantly different in the absence or presence of PPADS, suramin or $\alpha\beta$ -meATP (n=7). Data are presented as mean ± SEM

desensitising agent, while the relaxation to ATP was not 259 affected (Fig. 2c). 260

Effect of NF449, a selective P2X1 receptor antagonist

Contractile responses to $\alpha\beta$ -meATP suggest an expres-262sion of P2X1 receptors in porcine pancreatic arteries 263(Fig. 2b). In turn, the involvement of P2X1 receptors in 264contraction to ATP seems likely because contraction was 265significantly blocked by $\alpha\beta$ -meATP (Fig. 2a). The re-266sponses to ATP and $\alpha\beta$ -meATP were studied further in 267the presence of NF449 (10 µM), a P2X1 receptor-268selective antagonist. The contractions to ATP and $\alpha\beta$ -269meATP were inhibited in the presence of NF449 (Fig. 3). 270Effect of endothelium removal 271

The response to ATP was tested after the endothelium 272had been removed. The contraction and the relaxation 273induced by ATP (Fig. 4) were statistically not significantly 274different in the absence or presence of the endothelium. 275Similarly, removal of the endothelium had no effects on 276the contractions to KCl, U46619 or $\alpha\beta$ -meATP; for ex-277ample, the contraction to 75 mM KCl was 9.5±0.5 g in 278endothelium-intact arteries, while it was 9 ± 0.5 g in 279endothelium-denuded arteries (n=7-9). The contraction 280



Fig. 3 Effect of NF449 (10 μ M), a selective P2X1 receptor antagonist, on contractions to **a** ATP (1 mM) and **b** $\alpha\beta$ -meATP (1 μ M), in U46619-preconstricted porcine pancreatic arteries. NF449 reduced the effects of **a** ATP and **b** $\alpha\beta$ -meATP (****P*<0.001, unpaired Student's *t* test, *n*=10–13). Data are presented as mean \pm SEM

Purinergic Signalling



Fig. 4 Effect of removal of the endothelium on **a** contraction and **b** relaxation to ATP (1 mM) in U46619-preconstricted porcine pancreatic arteries. The effect of the removal of endothelium on the contraction or relaxation of ATP was not significantly different (n=9-11). Data are presented as mean \pm SEM

281to 10–100 nM U46619 was 5.5 ± 0.5 g in endothelium-282intact arteries, while it was 5.8 ± 0.6 g in endothelium-283denuded arteries (n=12-14). The contraction to 1 μM284 $\alpha\beta$ -meATP was 3.2 ± 0.6 g in endothelium-intact arteries,285while it was 3 ± 0.6 g in endothelium-denuded arteries (n=2866); there was no significant difference between these287responses.

Effect of XAC, an adenosine receptor antagonist
 The relaxation to ATP was investigated in the presence
 of a non-selective adenosine receptor antagonist; XAC
 (10 μM) had no effect on the contraction evoked by
 ATP (Fig. 5a), while it reduced significantly the relaxation
 to ATP (Fig. 5b).

294 Characterisation of response to UTP in U46619 295 preconstricted porcine-isolated pancreatic arteries

296	•	Effect of suramin, PPADS, $\alpha\beta$ -meATP and MRS2578, a
297		selective P2Y ₆ receptor antagonist
298		The contraction to UTP was examined in the presence
299		of suramin (100 μM), PPADS (10 μM), αβ-meATP



Fig. 5 Effect of XAC (10 μ M) on **a** contraction and **b** relaxation to ATP (1 mM) in U46619-preconstricted porcine pancreatic arteries. **a** XAC had no effect on the contraction to ATP (n=8-10), and **b** XAC reduced the relaxation to ATP (***P<0.001, unpaired Student's *t* test, n=8-10). Data are presented as mean \pm SEM

(1 µM) and MRS2578 (10 µM). Suramin and PPADS 300 significantly reduced the contraction to UTP (Fig. 6). 301 while the UTP responses were not affected after P2X 302 receptor desensitisation in the presence of $\alpha\beta$ -meATP 303 $(1 \mu M)$ or in the presence of a selective P2Y₆ receptor 304 antagonist (MRS2578); for example, the contraction to 3051 mM UTP was 1.8±0.2 g in the absence of MRS2578 306 (n=7), while it was 2.1 ± 0.2 g in the presence of 307

Fig. 6 Effect of suramin (100 μ M) and PPADS (10 μ M) on contraction to UTP in U46619-preconstricted porcine pancreatic arteries. With suramin and PPADS, effect of UTP concentration (*F*=16.77 and *F*=12.38, respectively, ****P*<0.001), suramin and PPADS reduced the contraction evoked by UTP (*F*=14.47 and *F*=12.48, respectively, ****P*<0.001, two-way ANOVA; *n*=9–12). Data are presented as mean ± SEM

308		MRS2578 ($n=6$); there was no significant difference be-
309		tween these responses.
310	•	Effect of endothelium removal
311		The effects of UTP were studied after the endothelium
312		had been removed. The contraction induced by UTP was
313		significantly attenuated in the endothelium-denuded arter-
314		ies (Fig. 7).
315	•	Effect of DUP 697, a cyclooxygenase-2 inhibitor
316		Because the contraction to UTP was largely
317		endothelium-dependent, the contraction was studied in
318		the presence of DUP 697, a cyclooxygenase-2 (COX-2)
319		inhibitor, since COX-2 facilitates the release of agents
320		which are responsible for endothelium-dependent contrac-
321		tion. DUP 697 (3 μ M) diminished the response to UTP
322		(Fig. 8) to a similar extent as removal of the endothelium
323		(Fig. 7), while DUP 697 did not alter the contraction to
324		U46619 (the preconstriction agent) or the contraction to
325		ATP (data not shown).

Characterisation of response to ADP in U46619 preconstricted porcine-isolated pancreatic arteries

Effect of MRS2179, a P2Y₁ receptor selective antagonist,
 and of endothelium removal

The relaxation to ADP in pancreatic arteries was stud-330 331ied in the presence of MRS2179 (10 µM) and after the endothelium had been removed. The relaxation to ADP 332 was reduced slightly but significantly in the presence of 333 334 MRS2179 (Fig. 9a) and in the endothelium-denuded arteries (Fig. 9b), which indicates the involvement of $P2Y_1$ 335 336 receptors and the endothelium in ADP-mediated relaxa-337 tion of porcine pancreatic arteries.

Effect of XAC, an adenosine receptor antagonist, and
 SCH58261, a selective adenosine A_{2A} receptor antagonist
 The relaxation to ADP was investigated in the presence

of XAC (10 µM). The relaxation to ADP was largely



Fig. 7 Effect of removal of the endothelium on contraction to UTP in U46619-preconstricted porcine pancreatic arteries. Effect of UTP concentration (F=11.91, ***P<0.001) and removal of endothelium reduced the contraction evoked by UTP (F=43, ***P<0.001, two-way ANOVA; n=10–12). Data are presented as mean ± SEM

341



4

Fig. 8 Effect of DUP 679 (3 μ M), a cyclooxygenase-2 inhibitor, on contraction to UTP in U46619-preconstricted porcine pancreatic arteries. Effect of UTP concentration (*F*=8.48, ****P*<0.001) and DUP 679 reduced the contraction evoked by UTP (*F*=50.8, ****P*<0.001, two-way ANOVA; *n*=8–12). Data are presented as mean ± SEM

reduced in the presence of this inhibitor which indicates 342 the involvement of adenosine receptors (Fig. 10). To find 343 out about the adenosine subtype involved in the relaxation 344 to ADP, the response to ADP was investigated in the 345 presence of SCH58261, a selective adenosine A_{2A} receptor antagonist. This antagonist significantly inhibited the 347 relaxation to ADP, to a similar extent as seen with XAC 348



Fig. 9 Effect of a MRS2179 (10 μ M) and b the removal of the endothelium on relaxation to ADP in U46619-preconstricted porcine pancreatic arteries. With MRS2179 and in endothelium-denuded arteries, effect of ADP concentration (*F*=21.42 and 16.77, respectively, ****P*<0.001) and MRS2179 and removal of endothelium reduced the contraction evoked by ADP (*F*=21.42 and *F*=32.04, respectively, ****P*<0.001, two-way ANOVA; *n*=10–12). Data are presented as mean ± SEM

Purinergic Signalling



Fig. 10 Effect of XAC (10 μ M), a non-selective adenosine receptor antagonist, and SCH58261 (1 μ M), a selective adenosine A_{2A} receptor antagonist, on relaxation to ADP in U46619-preconstricted porcine pancreatic arteries. With XAC and SCH58261, effect of ADP concentrations (*F*=7.14 and *F*=6.08, respectively, ****P*<0.001), XAC and SCH58261 reduced the relaxation evoked by ADP (*F*=71.19 and *F*=58.16, respectively, ****P*<0.001, two-way ANOVA; *n*=9–14). Data are presented as mean ± SEM

349	(Fig. 10). This showed that the relaxation to ADP involved
350	A_{2A} adenosine receptors.

351 Discussion

352The current report has provided evidence for the functional expression of contractile P2X1, P2Y2 and P2Y4 receptors and 353vasorelaxant P2Y1 and A2A adenosine receptors in porcine 354pancreatic arteries. These receptors are sensitive to the extra-355cellular nucleotides ATP (P2X1), UTP (P2Y2 and P2Y4) and 356 ADP (P2Y₁ and A_{2A}). The contraction to ATP was endothe-357 lium independent, while UTP induced an endothelium-358dependent contraction which may involve P2Y₂ and/or 359 360 P2Y₄ receptors. The relaxation to ADP involved the endothelium and P2Y₁ receptors and A_{2A} adenosine receptors. 361

362 A vasoconstrictor response elicited by ATP has been reported in a number of different arteries [21-23]. ATP may also 363 364 induce vasorelaxation depending on the experimental conditions (level of pre-tone) and relative expression of relevant 365366 vasocontractile and vasorelaxant receptors [24, 25]. In porcine 367 pancreatic arteries, ATP induced a biphasic response consisting of a contraction followed by a relaxation 368(Fig. 1b). Since the contraction to ATP was rapidly 369370 desensitising, non-cumulative concentration-response curves 371were investigated. The contractions to ATP and $\alpha\beta$ -meATP were reduced in the presence of suramin, PPADS, $\alpha\beta$ -meATP 372373 (a desensitiser of P2X1 receptors) and NF449 (a P2X1 selective antagonist) (Figs. 2a, b and 3a, b), which indicates that a 374large part of the contraction to ATP could be attributed to the 375activation of P2X1 receptors. Moreover, the contractile effect 376 377 of $\alpha\beta$ -meATP is consistent with the expression of P2X1 378receptors in porcine pancreatic arteries. $\alpha\beta$ -meATP was more 379 potent than ATP in eliciting vasoconstriction most likely due to its greater stability [5]. Since the contraction to ATP was not changed after the endothelium had been removed (Fig. 4a), the expression of P2X1 receptors was shown to be on the vascular smooth muscle cells (VSMCs). This is consistent with the abundant expression of P2X1 receptors on VSMCs of most tissues [7].

ATP-induced vasorelaxation was not affected after the 386 endothelium had been removed or in the presence of suramin 387 or PPADS, which suggests that the relaxation to ATP was not 388 due to its action at P2Y receptors. However, the relaxation to 389ATP was significantly inhibited in the presence of XAC, 390 which suggested an involvement of adenosine receptors 391 expressed on VSMCs of the pancreatic arteries; it is likely 392that this is due to the activity of adenosine derived from ATP 393 metabolism by ecto-nucleotidase 5'-triphosphate 394 diphosphohydrolase (ENTPDase) enzymes followed by the 395 activity of CD37 and ecto-5'-nucleotidase enzymes [26]. Sim-396 ilarly, in rat coronary arteries, the relaxation to ATP involved 397 P1 receptors, although there was an additional involvement of 398 P2Y receptors [24]. In the current study, further investigation 399 of the adenosine receptor subtypes involved in the relaxation 400 to ATP is required. We and others have shown previously a 401 slow relaxation in response to $\alpha\beta$ -meATP in rat mesenteric 402 arteries, subsequent to contraction [27-29], but we did not 403 observe this in the present study in the porcine pancreatic 404 arteries. 405

The vasoconstriction to UTP did not desensitise quickly; 406 therefore, cumulative concentration-response curves were 407 used to study the effect of UTP on pancreatic arteries. This 408 contraction was significantly inhibited by suramin and 409PPADS (Fig. 6), and there was a reduction of the response 410 after the removal of the endothelium (Fig. 7). That would 411 indicate for the first time an endothelium-dependent vasocon-412striction evoked by UTP. UTP is known to be active at P2Y₂, 413 $P2Y_4$ and $P2Y_6$ receptors [30]. The expression of these recep-414 tors in the endothelium and smooth muscle of vessels has been 415reported [31]. Since MRS2578 was not able to alter the 416 contraction to UTP, this indicates that UTP had no action at 417P2Y₆ receptors. There are currently no commercially available 418 selective antagonists for either P2Y₂ or P2Y₄ receptors. How-419 ever, we believe that UTP acted at $P2Y_4$ receptors since the 420contraction to UTP was significantly inhibited by both endo-421thelium removal and in the presence of DUP 697, but re-422sponses to ATP were unaffected. UTP-induced contraction 423 may also be mediated by P2Y2 receptors, since MRS2768 424 which is a selective agonist at P2Y₂ receptors and displays no 425affinity for P2Y₄ or P2Y₆ receptors was able to evoke a 426 contraction in pancreatic arteries [32] (Fig. 1a). 427

UTP-induced vasoconstriction has been documented in a 428 number of arteries including rat pulmonary arteries in which 429 the contraction was attributed to $P2Y_2$ receptors, and in rabbit 430 basilar arteries in which the contraction to UTP was due to 431 action of $P2Y_4$ receptors [33, 34]. UTP produced an 432

433endothelium-dependent relaxation in rabbit pulmonary arteries and in rat mesenteric arterial bed, but the receptor subtypes 434were undefined [22, 35]. In bovine middle cerebral arterial 435strips, UTP had a dual response, and it induced a contraction 436437 in endothelium-denuded arteries, but a relaxation in 438 endothelium-intact arteries [36]. The absence of 439endothelium-dependent or endothelium-independent relaxation to UTP and some other nucleotides in rat renal arteries 440was reported [37], which is consistent with the current study 441 since there was no evidence of a UTP-mediated relaxation in 442443 porcine pancreatic arteries. Hence, porcine pancreatic arteries appear not to express relaxant P2Y2 and/or P2Y4 receptors. 444

To investigate the mechanism underlying the contraction 445mediated by UTP in pancreatic arteries, the response to UTP 446 447 was examined in the presence of DUP 697. As seen in Fig. 8, 448 the endothelium-dependent contraction was attenuated in the presence of the selective COX-2 inhibitor. Endothelial cells 449 450can release endothelium-derived contractile factors (EDCFs), 451which may include thromboxane A₂, prostaglandin $F_{2\alpha}$, leukotrienes and endothelin-1. Thromboxane A2 and prostaglan-452din $F_{2\alpha}$ are released from the endothelium due to the activity 453454 of COX-2 [38, 39]. The reduction of the contraction to UTP in 455the presence of DUP 697 indicated the involvement of thromboxane A_2 and prostaglandins in the contraction to UTP. 456These agents, after being released from the endothelium, 457458may act on their receptors on VSMCs to cause contraction [39]. The different effects of DUP 697 on responses to UTP 459460 and ATP further suggest that they are acting on different 461receptors.

The relaxation to ADP did not desensitise rapidly; there-462fore, cumulative concentration-response curves were used to 463study the effect of ADP on pancreatic arteries. The relaxation 464was significantly attenuated by MRS2179, a selective P2Y₁ 465receptor antagonist (Fig. 9a). In addition, the relaxation to 466 ADP was reduced after the endothelium had been removed, 467 by a similar extent as observed in the presence of the 468MRS2179 (Fig. 9b). This may suggest that P2Y₁ receptors 469470 are expressed on the endothelium. Indeed, a number of reports 471 show that $P2Y_1$ receptors are expressed on the endothelium 472and are responsible for the relaxation of arteries, including rat 473 thoracic aortic and porcine mesenteric arteries [40, 41]. The 474relaxation to ADP in our study was largely reduced in the 475presence of XAC and SCH58261 (adenosine receptor antag-476 onists). Adenosine receptors may be expressed on the endo-477 thelium or the vascular smooth muscle [42]. Since XAC and SCH58261 produced a greater reduction in the relaxation to 478479ADP than the inhibition induced by removal of the endothelium (Fig. 10), this suggests that relaxation to ADP involves 480 A_{2A} adenosine receptors expressed, at least in part, on 481 VSMCs. The mechanism by which ADP would produce 482483 adenosine to act at the adenosine receptors is still to be 484 elucidated. The simplest explanation is that it is broken down by ENTPDases and by CD37 enzymes to adenosine [26]. 485

Alternatively, as suggested in porcine coronary arteries, 486ADP mediates a relaxation via a mechanism that involves 487 ADP-evoked adenosine release and the subsequent activation 488 of A_{2A} receptors [20]. In contrast to the porcine pancreatic 489vessels, ADP in rat pancreatic arteries induced a contraction at 490 a high concentration (1 mM); this contraction was similar to 491that produced by ATP and was much lower than the contrac-492 tion induced by $\alpha\beta$ -meATP [43]. Further investigation is 493required to determine the involvement of endothelium-494derived relaxing factors or endothelium-derived 495hyperpolarising factors released from the endothelium in the 496 ADP-induced relaxation. 497

Reduction in pancreatic blood flow has been observed in 498acute and chronic pancreatitis and some other pancreatic 499diseases [44, 45], implicating pancreatic tissue perfusion as 500an important factor in the pathogenesis of pancreatic diseases 501and symptoms. There is increasing evidence for the role of 502purinergic signalling in the pathophysiology of the pancreas 503[2]. Hence, drugs designed to target specific components of 504purinergic system may be of relevance to the management of 505pancreatitis, cystic fibrosis, pancreatic cancer and diabetes. 506

In summary, the functional expression of P2X1 and A_{2A} 507 adenosine receptors on VSMCs and P2Y2 and/or P2Y4 recep-508 tors on the endothelium of porcine pancreatic arteries was 509indicated in the current study. Activation of P2X1 receptors 510by ATP or $\alpha\beta$ -meATP induced a vasoconstriction, and UTP 511acts at P2Y₂ and/or P2Y₄ receptors to induce a contraction. 512ADP and ATP activate A2A adenosine receptors to induce 513relaxation, together with an action of ADP on P2Y1 receptors. 514Pancreatic arteries appear to lack vasorelaxant P2Y₂ and/or 515P2Y₄ receptors. 516

AcknowledgmentsWe would like to thank the Damascus University in517Syria for funding the project.518

References

- 520Q3
- 1. Novak I (2008) Purinergic receptors in the endocrine and exocrine pancreas. Purinergic Signal 4(3):237–253
 521
- 2. Burnstock G, Novak I (2012) Purinergic signalling in the pancreas in health and disease. J Endocrinol 213(2):123–141 524
- Khakh BS, Burnstock G, Kennedy C, King BF, North RA, Séguéla P
 tal (2001) International union of pharmacology. XXIV. Current status of the nomenclature and properties of P2X receptors and their subunits. Pharmacol Rev 53(1):107–118
- 4. Abbracchio MP, Burnstock G, Boeynaems J-M, Barnard EA, Boyer
 JL, Kennedy C et al (2006) International union of pharmacology
 LVIII: update on the P2Y G protein-coupled nucleotide receptors:
 from molecular mechanisms and pathophysiology to therapy.
 Pharmacol Rev 58(3):281–341
 533
- 5. Kasakov L, Burnstock G (1982) The use of the slowly degradable 534 analog, α,β -methylene ATP, to produce desensitisation of the P2purinoceptor: effect on non-adrenergic, non-cholinergic responses of the guinea-pig urinary bladder. Eur J Pharmacol 86(2):291–294 537

668

- Burnstock G (2006) Pathophysiology and therapeutic potential of purinergic signaling. Pharmacol Rev 58(1):58–86
- Kügelgen I (2008) Pharmacology of mammalian P2X and P2Y receptors. Biotrend Reviews, no. 3
- Communi D, Govaerts C, Parmentier M, Boeynaems J-M (1997) Cloning of a human purinergic P2Y receptor coupled to phospholipase C and adenylyl cyclase. J Biol Chem 272(51):31969–31973
- 9. Qi A-D, Zambon AC, Insel PA, Nicholas RA (2001) An arginine/ glutamine difference at the juxtaposition of transmembrane domain 6
 and the third extracellular loop contributes to the markedly different nucleotide selectivities of human and canine P2Y₁₁ receptors. Mol Pharmacol 60(6):1375–1382
- Léon C, Hechler B, Vial C, Leray C, Cazenave J-P, Gachet C (1997) The
 P2Y₁ receptor is an ADP receptor antagonized by ATP and expressed in
 platelets and megakaryoblastic cells. FEBS Lett 403(1):26–30
- 11. Bodor ET, Waldo GL, Hooks SB, Corbitt J, Boyer JL, Harden TK
 (2003) Purification and functional reconstitution of the human P2Y₁₂
 receptor. Mol Pharmacol 64(5):1210–1216
 - Marteau F, Le Poul E, Communi D, Communi D, Labouret C, Savi P
 et al (2003) Pharmacological characterization of the human P2Y₁₃
 receptor. Mol Pharmacol 64(1):104–112
- 13. Waldo GL, Harden TK (2004) Agonist binding and Gq-stimulating
 activities of the purified human P2Y₁ receptor. Mol Pharmacol 65(2):
 426–436
- 14. Nicholas RA, Lazarowski ER, Watt WC, Li Q, Boyer J, Harden TK
 (1996) Pharmacological and second messenger signalling selectivities of cloned P2Y receptors. J Auton Pharmacol 16(6):319–323
 - 15. Carter RL, Fricks IP, Barrett MO, Burianek LE, Zhou Y, Ko H et al (2009) Quantification of G_i-mediated inhibition of adenylyl cyclase activity reveals that UDP is a potent agonist of the human P2Y₁₄ receptor. Mol Pharmacol 76(6):1341–1348
 - 9 16. Coutinho-Silva R, Parsons M, Robson T, Lincoln J, Burnstock G
 0 (2003) P2X and P2Y purinoceptor expression in pancreas from streptozotocin-diabetic rats. Mol Cell Endocrinol 204(1–2):141–154
 - 17. Hillaire-Buys D, Chapal J, Petit P, Loubatières-Mariani M-M (1991)
 Dual regulation of pancreatic vascular tone by P2X and P2Y
 purinoceptor subtypes. Eur J Pharmacol 199(3):309–314
- 575 18. Hillaire-Buys D, Dietz S, Chapal J, Petit P, Loubatieres-Mariani MM
 576 (1999) Involvement of P2X and P2U receptors in the constrictor
 577 effect of ATP on the pancreatic vascular bed. Journal de la Societe
 578 de Biologie 193(1):57–61
- 579 19. Alsaqati M, Chan SLF, Ralevic R (2011) Characterisation of the response to ADP in porcine isolated pancreatic arteries. http://www. 581 pa2online.org/abstracts/vol8issue1abst028p.pdf
- 20. Rayment SJ, Ralevic V, Barrett DA, Cordell R, Alexander SPH
 (2007) A novel mechanism of vasoregulation: ADP-induced relaxation of the porcine isolated coronary artery is mediated via adenosine
 release. FASEB J 21(2):577–585
- 586 21. Kügelgen I, Häussinger D, Starker K (1987) Evidence for a vasoconstriction-mediating receptor for UTP, distinct from the P2 purinoceptor, in rabbit ear artery. Naunyn Schmiedebergs Arch Pharmacol 336(5):556–560
- 22. Ralevic V, Burnstock G (1991) Effects of purines and pyrimidines on
 the rat mesenteric arterial bed. Circ Res 69(6):1583–1590
- 23. Rubino A, Burnstock G (1996) Evidence for a P2-purinoceptor
 mediating vasoconstriction by UTP, ATP and related nucleotides in
 the isolated pulmonary vascular bed of the rat. Br J Pharmacol
 118(6):1415–1420
- 596 24. Korchazhkina O, Wright G, Exley C (1999) Intravascular ATP and coronary vasodilation in the isolated working rat heart. Br J 598 Pharmacol 127(3):701–708
- 599 25. Ralevic V, Burnstock G (1996) Relative contribution of P2U- and
 600 P2Y-purinoceptors to endothelium-dependent vasodilatation in the
 601 golden hamster isolated mesenteric arterial bed. Br J Pharmacol
 602 117(8):1797–1802

- 26. Zimmermann H (2000) Extracellular metabolism of ATP and other
nucleotides. Naunyn Schmiedebergs Arch Pharmacol 362(4–5):299–
309603
604
- 27. Stanford SJ, Mitchell JA (1998) ATP-induced vasodilatation in the rat isolated mesenteric bed exhibits two apparent phases. Br J 607 Pharmacol 125:94P 608
- 28. Ralevic V (2001) Mechanism of prolonged vasorelaxation to ATP in the rat isolated mesenteric arterial bed. Br J Pharmacol 132(3):685– 692
 610 611
- 29. Ralevic V (2002) The involvement of smooth muscle P2X612receptors in the prolonged vasorelaxation response to purine613nucleotides in the rat mesenteric arterial bed. Br J Pharmacol614135(8):1988–1994615
- 30. Burnstock G, Williams M (2000) P2 purinergic receptors: modula-
tion of cell function and therapeutic potential. J Pharmacol Exp Ther
295(3):862–869616
617
618
- 31. Burnstock G (2007) Purine and pyrimidine receptors. CMLS 64(12): 619 1471–1483 620
- 32. Ko H, Carter RL, Cosyn L, Petrelli R, de Castro S, Besada P et al (2008) Synthesis and potency of novel uracil nucleotides and derivatives as P2Y₂ and P2Y₆ receptor agonists. Bioorg Med Chem 16(12):6319–6332 624
- 33. Hartley SA, Kato K, Salter KJ, Kozlowski RZ (1998) Functional
 evidence for a novel suramin-insensitive pyrimidine receptor in rat
 small pulmonary arteries. Circ Res 83(9):940–946
 627
- 34. Miyagi Y, Zhang JH (2004) α,β-Methylene ATP enhances P2Y4 628 contraction of rabbit basilar artery. Am J Physiol Heart Circ Physiol 286(4):H1546–H1551 630
 35. Qasabian RA, Schvvens C, Owe-Young R, Killen JP, Macdonald PS, 631
- 35. Qasabian RA, Schyvens C, Owe-Young R, Killen JP, Macdonald PS,
Conigrave AD et al (1997) Characterization of the P2 receptors in
rabbit pulmonary artery. Br J Pharmacol 120(4):553–558633
- 36. Miyagi Y, Kobayashi S, Nishimura J, Fukui M, Kanaide H (1996)
 Dual regulation of cerebrovascular tone by UTP: P2U receptormediated contraction and endothelium-dependent relaxation. Br J
 Pharmacol 118(4):847–856
 37. Knight GE, Oliver-Redgate R, Burnstock G (2003) Unusual absence
- 37. Knight GE, Oliver-Redgate R, Burnstock G (2003) Unusual absence
 of endothelium-dependent or -independent vasodilatation to purines
 or pyrimidines in the rat renal artery. Kidney Int 64(4):1389–1397
 640
- 38. Mombouli JV, Vanhoutte PM (1993) Purinergic endothelium dependent and -independent contractions in rat aorta. Hypertension
 22(4):577–583
 643
- 39. Wong SL, Leung FP, Lau CW, Au CL, Yung LM, Yao X et al (2009)
 Cyclooxygenase-2-derived prostaglandin F2alpha mediates
 endothelium-dependent contractions in the aortae of hamsters with
 increased impact during aging. Circ Res 104(2):228–235
 647
- 40. Dol-Gleizes F, Mares AM, Savi P, Herbert JM (1999) Relaxant effect 648 of 2-methyl-thio-adenosine diphosphate on rat thoracic aorta: effect 649 of clopidogrel. Eur J Pharmacol 367(2–3):247–253 650
 41. Alefishat E, Alexander S, Ralevic V (2010) Effect of palmitoyl CoA 651
- 41. Alefishat E, Alexander S, Ralevic V (2010) Effect of palmitoyl CoA 651
 on ADP-evoked vasorelaxations in porcine isolated coronary and mesenteric arteries. http://www.fasebj.org/cgi/content/meeting_abstract/24/1_MeetingAbstracts/lb426 653
 42. Schulte G, Fredholm BB (2003) Signalling from adenosine receptors 655
- Schulte G, Fredholm BB (2003) Signalling from adenosine receptors to mitogen-activated protein kinases. Cell Signal 15(9):813–827
- 43. Chapal J, Loubatieres-Mariani MM (1983) Evidence for purinergic
 receptors on vascular smooth muscle in rat pancreas. Eur J Pharmacol
 87(4):423–430
 659
- 44. Satoh A, Shimosegawa T, Satoh K, Ito H, Kohno Y, Masamune A
 et al (2000) Activation of adenosine A₁-receptor pathway induces
 edema formation in the pancreas of rats. Gastroenterology 119(3):
 829–836
 663
- 45. Nguyen NC, Taalab K, Osman MM (2010) Decreased blood flow with increased metabolic activity: a novel sign of pancreatic tumor aggressiveness. Clin Cancer Res Off J Am Assoc Cancer Res 16(1): 367, author reply 567

656

AUTHOR QUERIES

AUTHOR PLEASE ANSWER ALL QUERIES.

- Q1. Kindly check the inserted spelled out form 'standard error of the mean' of the abbreviation 'SEM' if correct.
- Q2. Kindly check the inserted spelled out form 'half maximal effective concentration' of the abbreviation 'EC₅₀' if correct.
- Q3. Kindly provide access date for references 'Alsaqati et al. [19]' and 'Alefishat et al. [41]' in the list.

.d'Ale