

1 **Short Communication**

2 **Mycobacteriosis outbreak caused by *Mycobacterium avium* subsp. *avium* involving**
3 **five porcine fattening farms detected through slaughterhouse surveillance**

4 Bernat Pérez de Val¹, Llorenç Grau-Roma², Joaquim Segalés^{1,2}, Mariano Domingo^{1,2},
5 Enric Vidal^{1,*}

6 ¹Centre de Recerca en Sanitat Animal (CRESA), UAB-IRTA 08193 Bellaterra, Barcelona,
7 Catalonia, Spain

8 ²Servei de Diagnòsti de Patologia Veterinària (SDPV), Departament de Sanitat i d'Anatomia
9 Animals, Universitat Autònoma de Barcelona, 08193 Bellaterra, Barcelona, Catalonia, Spain

10

11 **Bernat Pérez de Val**, BSc, MSc,

12

13 **Llorenç Grau-Roma**, DVM, PhD, DipECVP

14

15 **Joaquim Segales**, DVM, PhD, DipECVP

16

17 **Mariano Domingo**, DVM, PhD, DipECVP

18

19 **Enric Vidal**, DVM, PhD

20

21

22

23 E-mail for correspondence: enric.vidal@cresa.uab.cat

24

25

26

27

28 **Abstract**

29 Between December 2010 and January 2011 a number (n=20) of cases were submitted to the
30 Slaughterhouse Support Service (Servei de Suport a Escorxadors, SESC-CReSA), consisting of
31 grossly nodular granulomatous and caseous lesions in pig carcasses from five different farms.
32 Lesions involved lymph nodes, lungs, liver and spleen.

33 Histopathological examination showed multifocal to coalescent, granulomatous and
34 necrotizing splenitis, hepatitis, pneumonia and lymphadenitis. The presence of acid-fast bacilli
35 in some cases revealed that it was a mycobacteriosis.

36 Bacteriological analysis was performed to confirm the diagnosis and identify the aetiological
37 agent (to rule out it was from the *M. tuberculosis* complex mycobacteria, which includes
38 species causing human and animal tuberculosis). The identification of culture isolates by PCR
39 confirmed the growth of *M. avium* complex. Further sequencing analysis determined it was *M.*
40 *avium*. subsp. *avium*.

41 The most likely source of the outbreak was considered to be the feed which shared the five
42 farms, which might have been contaminated with *M. avium* subsp. *avium* (common pathogen
43 in poultry and other birds). The fact that most of the animals presented a clear involvement of
44 abdominal viscera is consistent with an oral route of infection.

45

46

47 *Keywords: mycobacteriosis, tuberculosis, pigs, Mycobacterium avium, slaughterhouse*

48

49

50 Animal tuberculosis (TB), caused by *Mycobacterium tuberculosis* complex (MTBC) species, is a
51 chronic zoonotic disease mainly affecting cattle, but also can cause disease in a wide range of
52 animal hosts and humans (OIE 2009). The wild boar (*Sus scrofa*) is the third animal species
53 after cattle and goats in the number of MTBC isolates in Spain (Rodriguez-Campos and others
54 2012), where is considered to be the main wild reservoir of TB (Naranjo and others 2008;
55 Garcia-Bocanegra and others 2012). In addition, domestic pigs (*Sus scrofa domestica*)
56 represented the 1% of Spanish MTBC isolates from animals in the period 1996-2011
57 (Rodriguez-Campos and others 2012). Moreover, recent TB outbreaks in domestic pigs due to
58 MTBC have been also reported in Italy (Di Marco and others 2012). There are, however, other
59 non-tuberculous mycobacteria that are non-zoonotic pathogens but can be opportunistic
60 causing similar pathologies in swine.

61 *M. avium* complex (MAC) comprises a number of bacterial species that are non-zoonotic
62 pathogens but with a different degree of pathogenicity and host preference (Álvarez and
63 others 2011). *M. avium* is subdivided in four subspecies: *M. avium* subsp. *avium* (MAA), *M.*
64 *avium* subsp. *silvaticum*, *M. avium* subsp. *paratuberculosis* (MAP), and *M. avium* subsp.
65 *hominisuis* (MAH). MAA is known to cause generalized granulomatous lesions in poultry and
66 wild birds, MAP is the causative agent of Johnes Disease in ruminants, while pigs are the
67 primary animal host for MAH (Thorel and others 2001; Mijs and others 2002; Agdestein and
68 others 2011; Álvarez and others 2011). However, pigs may also play a role as reservoirs of MAA
69 infection causing indistinguishable lesions from TB (Komijn and others 1999). Therefore,
70 mycobacterial species identification becomes crucial to determine the zoonotic nature of
71 outbreaks in pig farms with animals presenting TB-like lesions.

72 In December 2007, as an initiative of the Catalan Government's Health Protection Agency, the
73 Slaughterhouse Support Service (Servei de Suport a Escroxadors, SESC) was created within the
74 Animal Health Research Centre (Centre de Recerca en Sanitat Animal, CRESA). Its main

75 objective was to provide continuing education to meat inspectors and contribute in reaching
76 final diagnoses of slaughterhouse findings. Between December 2010 and January 2011, several
77 organs from a total of 20 pig cases coming from 5 different farms were submitted to SESC. The
78 lesions consisted of multifocal to coalescing whitish nodular lesions with caseous and partially
79 mineralized appearance, and affected mesenteric lymph nodes (LN), liver, spleen, mediastinal
80 LN and lung (see Figure 1). While lesions in organs of the abdominal cavity (mainly liver and
81 mesenteric LN) were observed in all pigs, lesions in the thoracic cavity (lungs and mediastinic
82 LN) were present in 12 pigs, coming from only 3 out of the 5 studied farms.

83 Histopathological examination of the lesions using haematoxylin and eosin (HE) routine
84 staining revealed multifocal, necrotizing and granulomatous splenitis, hepatitis, pneumonia
85 and lymphadenitis. Numerous multinucleated (Langhans) giant cells were observed. Ziehl-
86 Neelsen (ZN) staining revealed, in some of the cases, the presence of acid-fast bacilli indicating
87 that it was a mycobacteriosis (see Figure 2). Information on each of the outbreaks including
88 the organs examined and the different diagnostic techniques used are summarized in Table 1.

89 Consequently, a suspected TB was reported to local Animal and Human Health Authorities, and
90 biosafety measures (latex gloves and facial masks) were implemented for slaughterhouse
91 personnel.

92 Ruling out the infection caused by zoonotic mycobacteria was established as a priority.
93 Differential diagnosis was performed by means of bacteriological studies to identify the
94 ethological agent causing the lesions. Isolation was performed on Coletsos and Lowenstein-
95 Jensen selective media with pyruvate (bioMérieux España, Madrid, Spain). Thereafter DNA was
96 extracted from colonies by boiling them 10 min. at 100° C, and identification was performed
97 by means of a multiplex PCR specific for MTBC and MAC (Wilton 1992) followed by sequencing
98 of the DNA encoding 16S rRNA.

Commented [L1]: Correcte? N'hi ha un que, en base a la taula, no ho puc assegurar. Hi ha 19 fetges +, però potser el que fa 20 tenia lesions a LN mes i/o melsa?

Commented [L2]: correcte

Commented [L3]: Em sembla que en general, tot i que es tracti de short communications, les revistes solen demanar seguir l'estructura: Intro-M&M-Results-Discussion. Reconec però que desconec si el Vet Rec accepta "formats" diferent al habitual

99 The multiplex PCR of these colonies identified a non-tuberculous mycobacteria belonging to
100 the MAC. Sequencing and subsequent Basic Local Alignment Search Tool (BLAST®) analysis
101 (Altschul and others 1990) confirmed MAA in all cases.

102 Subsequent epidemiological investigation suggested that the most likely source of the
103 outbreak was the feed which was shared between the five different farms a few months
104 before the outbreak detection. Certain feed contents could have been contaminated with
105 MAA. Mycobacteriosis in pigs fed peat naturally contaminated with MAC has been previously
106 described (Matlova and others 2005; Agdestein and others 2011). In these infected pigs,
107 lesions were primarily found in the head and mesenteric LN. Accordingly, most of the animals
108 studied in the present outbreak showed a clear involvement of abdominal LN and viscera,
109 being strongly consistent with an oral route of infection.

110 Even though MAA is mainly isolated in birds and MAH is considered a human/porcine-type of
111 *M. avium* (Mijts and others 2002), a recent comparative study of MAA and MAH experimentally
112 infected pigs did not show significant differences in the ability of both pathogens to infect pigs
113 (Agdestein and others 2012). However, the authors demonstrated that only MAH was isolated
114 from pig faeces, causing a major animal-to-animal transmission by the faecal-oral route, which
115 could explain the higher incidence of infection caused by this subspecies in pigs as compared
116 to MAA (Agdestein and others 2012). Also, if MAA in pigs is not excreted by the faecal route,
117 feed contamination would be the most likely source of MAA-infection in the present outbreak.

118 Pigs are susceptible to both MTBC and MAC infections. The zoonotic risk of animals infected
119 with MTBC has been widely described (Rodwell and others 2008; Rodríguez and others 2009;
120 Torres-Gonzalez and others 2013). Nevertheless, severe MAC infections in humans have been
121 also reported, especially in immunosuppressed individuals, (Pavlik and others 2000; Biet and
122 others 2005; Mobius and others 2006). SESC proved to be an effective tool that allowed a

123 rapid diagnosis and molecular identification of the mycobacteriosis outbreak, leading to know
124 its associated risks for public health.

125

126 **Acknowledgements**

127 SESC (www.cresa.cat/blogs/sesc) is funded by the Agència de Protecció de la Salut (APSC)
128 (Catalan Public Health Protection Agency) from the Public Health Department of the Catalan
129 government (Generalitat de Catalunya). Lesions were indentified and documented by its
130 official veterinary meat inspectors. We are grateful to Ruben Cordon, Blanca Pérez, Aida Neira,
131 Zoraida Cervera, Maite Martin, Sierra Espinar, Marta Valle and Mariano Moreno of CReSA for
132 their technical support.

133

134

135

136

137

138

139

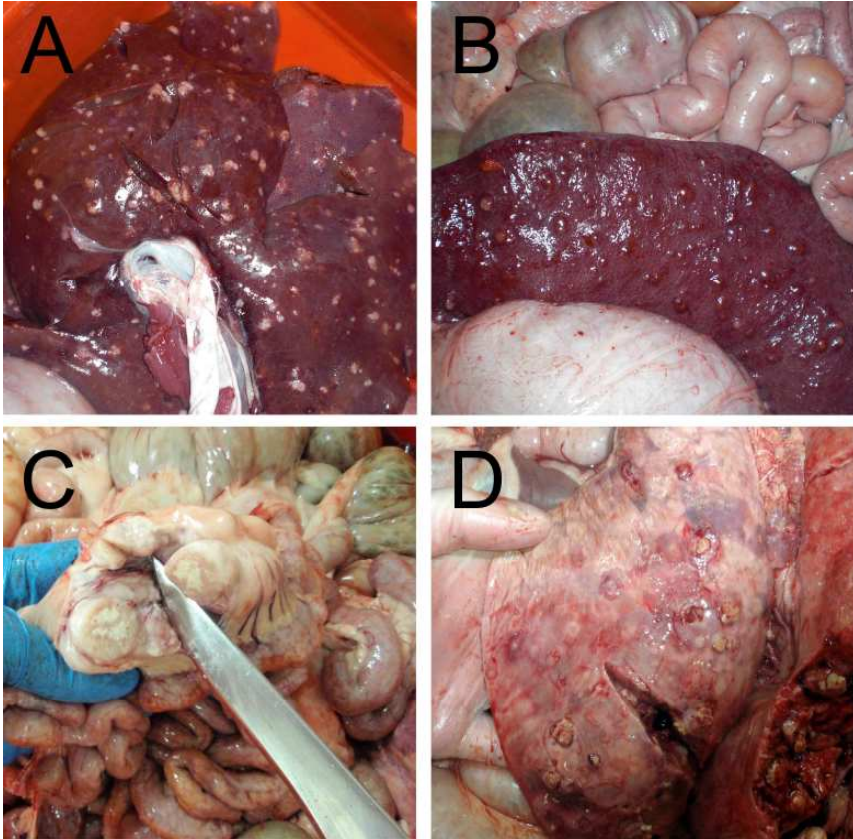
140

141

142

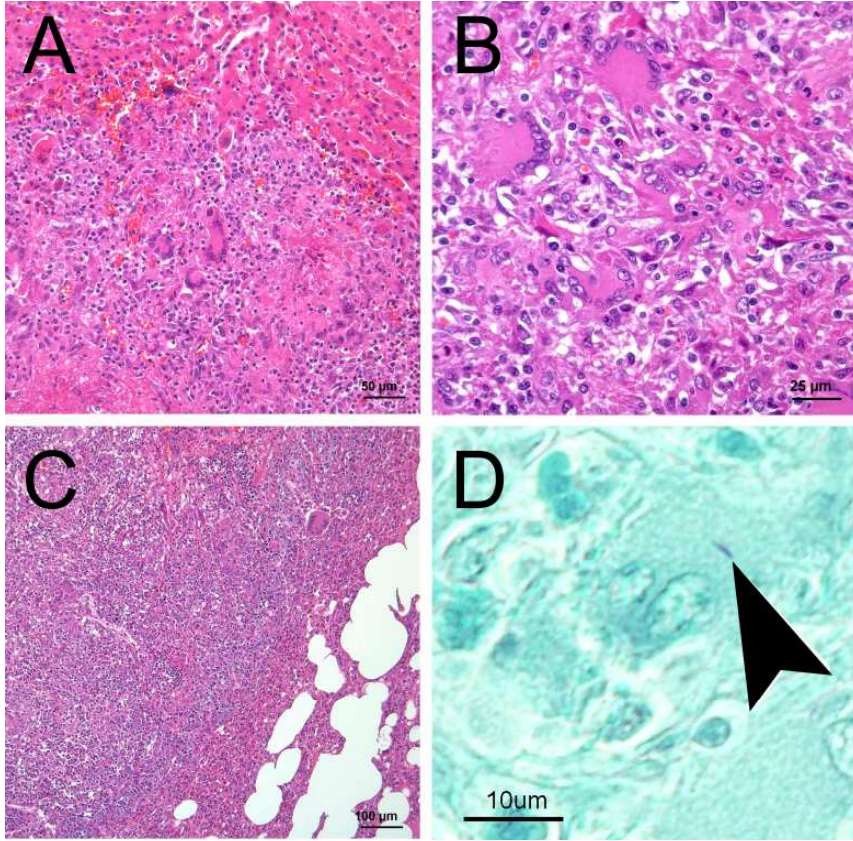
143

144 **Figures:**



145

146 **Figure 1.** Lesions consisting on granulomatous and caseous nodules, were observed in the
147 abdominal viscera affecting (A) liver, (B) spleen and (C) mesenteric lymph nodes. In some
148 cases, lesions were also observed in (D) thoracic cavity.



149

150 **Figure 2.** (A) Granulomatous hepatitis with necrosis foci, abundant macrophages and Langhans
151 cells. (B) Detail of Langhans cells in the splenic parenchyma. (C) In the lung, granulomatous foci
152 of inflammatory infiltrate were also appreciated. (D) Ziehl-Neelsen stain showed the presence
153 of a few acid-fast bacilli.

154

155

156

157

158

159

160 Tables:

161 Table 1: Information on the cases submitted and diagnostic techniques performed.

CASE	NO. OF AFFECTED ANIMALS	ORIGIN	REPORTED AFFECTED VISCERA	DIAGNOSTIC RESULTS
Case 1 December 2010	1	B	Liver (1/1) Mesenteric LN (1/1)	HP + (1/1) ZN + (1/1) Culture + (0/1)
Case 2 December 2010	1	A	Liver (1/1)	HP + (1/1) ZN + (1/1) Culture + (1/1)
Case 3 January 2011	4	C	Lungs (3/4) Spleen (2/4) Liver (4/4) Mesenteric LN (1/4)	HP + (4/4) ZN + (2/4) Culture + (4/4)
Case 4 January 2011	4	C and D	Liver (4/4) Spleen (4/4) Lungs (4/4) Mesenteric LN (4/4) Mediastinal LN (4/4)	HP + (4/4) ZN + (1/4) Culture (4/4)
Case 5 January 2011	10	E	Lungs (2/10) Mediastinal LN (5/10) Spleen (3/10) Liver (9/10) Mesenteric LN(3/10)	HP + (10/10) Culture + (10/10)

Commented [L4]: Casos 3, 4 i 5. Posaria els organs en el mateix ordre.

162

163 HP: Histopathology. ZN: Ziehl Neelsen's staining. LN: Lymph nodes. A to E: different farms
164 where the cases where originate.

165

166

167

168

169

170

171

172

173

174

- 176 AGDESTEIN A., JOHANSEN T. B., KOLBJORNSEN O., JORGENSEN A., DJONNE B. & OLSEN I.
177 (2012) A comparative study of mycobacterium avium subsp. avium and mycobacterium avium
178 subsp. hominissuis in experimentally infected pigs. BMC Veterinary Research 8, 11-6148-8-11
- 179 AGDESTEIN A., JOHANSEN T. B., POLACEK V., LIUM B., HOLSTAD G., VIDANOVIC D., ALEKSIC-
180 KOVACEVIC S., JORGENSEN A., ZULTAUSKAS J., NILSEN S. F. & DJONNE B. (2011) Investigation
181 of an outbreak of mycobacteriosis in pigs. BMC Veterinary Research 7, 63-6148-7-63
- 182 ALTSCHUL S. F., GISH W., MILLER W., MYERS E. W. & LIPMAN D. J. (1990) Basic local alignment
183 search tool. Journal of Molecular Biology 215, 403-410
- 184 ÁLVAREZ J., CASTELLANOS E., ROMERO B., ARANAZ A., BEZOS J., RODRÍGUEZ S., MATEOS A.,
185 DOMÍNGUEZ L. & DE JUAN L. (2011) Epidemiological investigation of a *mycobacterium avium*
186 subsp. *hominissuis* outbreak in swine. Epidemiology & Infection 139, 143
- 187 BIET F., BOSCHIROLI M. L., THOREL M. F. & GUILLOTEAU L. A. (2005) Zoonotic aspects of
188 mycobacterium bovis and mycobacterium avium-intracellulare complex (MAC). Veterinary
189 Research 36, 411-436
- 190 DI MARCO V., MAZZONE P., CAPUCCHIO M. T., BONIOTTI M. B., ARONICA V., RUSSO M.,
191 FIASCONARO M., CIFANI N., CORNELI S., BIASIBETTI E., BIAGETTI M., PACCIARINI M. L.,
192 CAGIOLA M., PASQUALI P. & MARIANELLI C. (2012) Epidemiological significance of the
193 domestic black pig (*sus scrofa*) in maintenance of bovine tuberculosis in sicily. Journal of
194 Clinical Microbiology 50, 1209-1218
- 195 GARCIA-BOCANEGRA I., PEREZ DE VAL B., ARENAS-MONTES A., PANIAGUA J., BOADELLA M.,
196 GORTAZAR C. & ARENAS A. (2012) Seroprevalence and risk factors associated to
197 mycobacterium bovis in wild artiodactyl species from southern spain, 2006-2010. PloS One 7,
198 e34908
- 199 KOMIJJN R. E., DE HAAS P. E., SCHNEIDER M. M., EGER T., NIEUWENHUIJS J. H., VAN DEN HOEK
200 R. J., BAKKER D., VAN ZIJLD ERVELD F. G. & VAN SOOLINGEN D. (1999) Prevalence of
201 mycobacterium avium in slaughter pigs in the netherlands and comparison of IS1245
202 restriction fragment length polymorphism patterns of porcine and human isolates. Journal of
203 Clinical Microbiology 37, 1254-1259
- 204 MATLOVA L., DVORSKA L., AYELE W. Y., BARTOS M., AMEMORI T. & PAVLIK I. (2005)
205 Distribution of mycobacterium avium complex isolates in tissue samples of pigs fed peat
206 naturally contaminated with mycobacteria as a supplement. Journal of Clinical Microbiology
207 43, 1261-1268
- 208 MIJS W., DE HAAS P., ROSSAU R., VAN DER LAAN T., RIGOUTS L., PORTAELS F. & VAN
209 SOOLINGEN D. (2002) Molecular evidence to support a proposal to reserve the designation
210 mycobacterium avium subsp. avium for bird-type isolates and 'M. avium subsp. hominissuis'
211 for the human/porcine type of M. avium. International Journal of Systematic and Evolutionary
212 Microbiology 52, 1505-1518
- 213 MOBIUS P., LENTZSCH P., MOSER I., NAUMANN L., MARTIN G. & KOHLER H. (2006)
214 Comparative macrorestriction and RFLP analysis of mycobacterium avium subsp. avium and

215 mycobacterium avium subsp. hominissuis isolates from man, pig, and cattle. Veterinary
216 Microbiology 117, 284-291

217 NARANJO V., GORTAZAR C., VICENTE J. & DE LA FUENTE J. (2008) Evidence of the role of
218 european wild boar as a reservoir of mycobacterium tuberculosis complex. Veterinary
219 Microbiology, 127, 1-9

220 OIE. (2009) Bovine tuberculosis. OIE Terrestrial Manual 2009 Chapter 2.4.7. Pages?

221 PAVLIK I., SVASTOVA P., BARTL J., DVORSKA L. & RYCHLIK I. (2000) Relationship between IS901
222 in the mycobacterium avium complex strains isolated from birds, animals, humans, and the
223 environment and virulence for poultry. Clinical and Diagnostic Laboratory Immunology 7, 212-
224 217

225 RODRÍGUEZ E., SÁNCHEZ L. P., PÉREZ S., HERRERA L., JIMÉNEZ M. S., SAMPER S. & IGLESIAS M.
226 J. (2009) Human tuberculosis due to mycobacterium bovis and M. caprae in Spain, 2004-2007.
227 The International Journal of Tuberculosis and Lung Disease 13, 1536-1541

228 RODRIGUEZ-CAMPOS S., GONZALEZ S., DE JUAN L., ROMERO B., BEZOS J., CASAL C., ALVAREZ
229 J., FERNANDEZ-DE-MERA I. G., CASTELLANOS E., MATEOS A., SAEZ-LLORENTE J. L., DOMINGUEZ
230 L., ARANAZ A. & SPANISH NETWORK ON SURVEILLANCE MONITORING OF ANIMAL
231 TUBERCULOSIS. (2012) A database for animal tuberculosis (mycoDB.es) within the context of
232 the Spanish national programme for eradication of bovine tuberculosis. Infection, Genetics and
233 Evolution : Journal of Molecular Epidemiology and Evolutionary Genetics in Infectious Diseases
234 12, 877-882

235 RODWELL T. C., MOORE M., MOSER K. S., BRODINE S. K. & STRATHDEE S. A. (2008)
236 Tuberculosis from mycobacterium bovis in binational communities, United States. Emerging
237 Infectious Diseases 14, 909-916

238 THOREL M. F., HUCHZERMEYER H. F. & MICHEL A. L. (2001) Mycobacterium avium and
239 mycobacterium intracellulare infection in mammals. Revue Scientifique Et Technique
240 (International Office of Epizootics) 20, 204-218

241 TORRES-GONZALEZ P., SOBERANIS-RAMOS O., MARTINEZ-GAMBOA A., CHAVEZ-MAZARI B.,
242 BARRIOS-HERRERA M. T., TORRES-ROJAS M., CRUZ-HERVERT L. P., GARCIA-GARCIA L., SINGH
243 M., GONZALEZ-AGUIRRE A., PONCE DE LEON-GARDUNO A., SIFUENTES-OSORNIO J. &
244 BOBADILLA-DEL-VALLE M. (2013) Prevalence of latent and active tuberculosis among dairy
245 farm workers exposed to cattle infected by mycobacterium bovis. PLoS Neglected Tropical
246 Diseases 7, e2177

247 WILTON S. & D. (1992) Detection and identification of multiple mycobacterial pathogens by
248 DNA amplification in a single tube. Genome Research 1, 269-273

249