

Accepted Manuscript

Functional Categorisation of Dietary Fibre in Foods: Beyond 'Soluble' vs 'Insoluble'

Michael J. Gidley, Gleb E. Yakubov

PII: S0924-2244(18)30049-9

DOI: <https://doi.org/10.1016/j.tifs.2018.12.006>

Reference: TIFS 2381

To appear in: *Trends in Food Science & Technology*

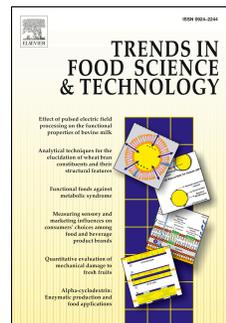
Received Date: 19 January 2018

Revised Date: 28 July 2018

Accepted Date: 20 December 2018

Please cite this article as: Gidley, M.J, Yakubov, G.E, Functional Categorisation of Dietary Fibre in Foods: Beyond 'Soluble' vs 'Insoluble', *Trends in Food Science & Technology*, <https://doi.org/10.1016/j.tifs.2018.12.006>.

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.



1 **Abstract**

2 *Background*

3 Diets rich in dietary fibre are associated with multiple health benefits, but there is often only
4 a restricted understanding of the mechanisms underlying these associations. This limits the
5 ability to select or design foods for specific nutritional purposes. Traditionally, the diverse
6 physical and chemical forms of dietary fibre have only been categorised as either soluble or
7 insoluble.

8 *Scope and Approach*

9 In this review, the physicochemical properties that have been proposed to be responsible for
10 the biological functionality of dietary fibres in the digestive tract are summarised and
11 classified. The extent to which these properties follow naturally from categorisation into
12 soluble vs insoluble forms are then assessed. Based on this analysis, a new approach to
13 functional categorisation of dietary fibres is proposed.

14 *Key Findings and Conclusions*

15 The physicochemical properties of dietary fibre components that are relevant to digestive
16 tract functionality can be classified under the headings of binding, structuring, and transport
17 barriers. Major nutritional outcomes such as control of macronutrient digestion or the nature
18 of residual digesta that are available for fermentation by the large intestinal microbiota
19 depend on combinations of these physicochemical properties in ways which are not readily
20 reflected by a soluble vs insoluble fibre definition. An alternative approach is proposed based
21 on 2D mapping of dietary fibre materials as a function of molecule/particle size and local
22 density. This effectively separates diverse fibre materials and can be linked semi-
23 quantitatively with functionally-important properties.

24

1 **Functional Categorisation of Dietary Fibre in Foods: Beyond ‘Soluble’ vs ‘Insoluble’**

2

3 *Michael J Gidley* & Gleb E Yakubov*

4

5 ARC Centre of Excellence in Plant Cell Walls, Centre for Nutrition and Food Sciences,

6 Queensland Alliance for Agriculture and Food Innovation (MJG) and School of Chemical

7 Engineering (GEY), The University of Queensland, St. Lucia, Brisbane, QLD 4072, Australia

8 * Corresponding author.

9 Phone: +61 7 3365 2145. Email address: m.gidley@uq.edu.au (M. J. Gidley)

10

11

12

13

14

15

16

17

18

19

20

21

22

23

24

25 **Keywords:** plant cell wall; hydrocolloid; gastrointestinal digesta; gut microbiota; particle

26 size; molecular density

27 1. Introduction

28 *1.1 Dietary fibre intake is associated with good health outcomes*

29 Results from a number of large prospective cohort studies have shown clear associations
30 between dietary fibre intake and reduced risks of all-cause mortality, cardiovascular diseases,
31 diabetes and cancers of the digestive tract (Anderson et al, 2009; Chuang et al, 2012,
32 Threapleton et al, 2013). As most dietary fibre is in the form of foods derived from cereals,
33 fruits, vegetables, legumes and nuts, this is reflected in consensus health advice around the
34 world that a diet rich in plant-based foods provides the best dietary protection against non-
35 communicable diseases. Some studies have attempted to identify specific protective effects of
36 fibre from each of cereals, vegetables and fruits. This is more challenging because most
37 people eat all three food types, but the analysis to date suggests that there may be some
38 differences between these broad classes, with cereal fibre being particularly protective (Park,
39 Subar, Hollenbeck & Schatzkin, 2011; Huang, Xu, Lee, Cho & Qi, 2015; Aune et al, 2016).
40 Whilst these epidemiological studies can be statistically powerful and have a place in
41 deriving population-level dietary guidelines, they show correlations not causations. It is
42 therefore frequently identified that greater mechanistic understanding of the protective
43 actions of fibre is needed in order to provide more tailored dietary advice and guide the
44 design of formulated food with optimised nutritional benefit (Chuang et al, 2012; Gidley
45 2013; Jones 2013; Grundy et al, 2016; Capuano, 2017).

46 Hypotheses for the protective action of at least some fibres against diabetes, cardiovascular
47 disease and colon cancer have been proposed (Gidley 2013; Jones 2013), with a focus on
48 carbohydrate (diabetes) and lipid/sterol (cardiovascular disease) metabolism, food intake
49 limitation (satiety), and/or large intestinal microbiota (colon cancer). However, there is a
50 large gap between whole-of-diet data, analysed at the population level to derive correlations,

51 and mechanistic studies that typically focus on single ingredients. One of the challenges in
52 bridging this gap is the lack of a coherent framework for connecting relevant measurable
53 properties of specific fibre components with the mechanisms by which they may influence
54 health outcomes as diverse as microbiome modulation, nutrient uptake rates, gastrointestinal
55 passage rates, and satiety.

56 *1.2 Dietary fibre is structurally and functionally diverse*

57 Dietary fibre in foods ranges from intact plant tissue pieces to small oligosaccharide
58 molecules. The boundaries of what is 'in' or 'out' of a definition of dietary fibre have been
59 debated for decades, but a consensus is now forming around a definition adopted by CODEX
60 in 2009. This definition is based on carbohydrate polymers that are not hydrolysed by the
61 endogenous enzymes in the small intestine of humans and are either (i) naturally occurring in
62 food, (ii) obtained from food raw materials by extraction, or (iii) synthetic carbohydrate
63 polymers. The key difference between type (i) and types (ii) and (iii) are that the latter are
64 qualified to only include those materials "which have been shown to have a physiological
65 effect of benefit to health as demonstrated by generally accepted scientific evidence to
66 competent authorities" (Jones, 2013). Thus, a clear distinction is drawn between endogenous
67 and extracted/synthetic carbohydrate polymers, which is consistent with health agency
68 dietary guidelines (based on prospective cohort studies) that focus on natural foods such as
69 whole grains, vegetables and fruit at the expense of those foods which are based on
70 recombination of refined ingredients. There are a number of questions of inclusion and
71 exclusion surrounding the CODEX definition. One is the minimum size (degree of
72 polymerisation; DP) which was initially set at DP10 with the option for individual countries
73 to reduce this to DP3. From a scientific and practical perspective (Jones, 2013), it seems
74 likely that DP3 will become the de facto standard. A second area is the lack of explicit
75 inclusion of lignin, which is not a carbohydrate polymer but is an intrinsic (but usually minor)

76 component of many edible plant tissues, and may contribute to health-related functional
77 properties.

78 Within each of the three broad classes of natural, extracted, and synthetic dietary fibres, there
79 is great structural diversity at the chemical as well as the physical structure level.
80 Categorisation of fibres in terms of chemical composition is often used. This has the benefit
81 of the analytical methods being robust, accurate and repeatable, but has major drawbacks in
82 that it does not usually include molecular size characterisation (so an oligosaccharide is
83 treated as equivalent to a polysaccharide), and does not usually distinguish between isolated
84 molecules and those which are part of a natural matrix, such as plant cell walls. The natural
85 heterogeneity of intrinsic fibre in plant-based foods at the polymer, cell wall, and tissue level
86 (Burton et al 2010) also provides many challenges in generating a sufficiently complete
87 molecular characterisation of dietary fibre components to address issues of nutritional
88 functionality.

89 In addition to diversity at the structural level, there is apparent diversity in the mechanisms
90 underlying nutritional functionality at all stages of digestive processing that make it
91 challenging to relate food composition to potential health outcomes (Capuano, 2017). For
92 example, the oral breakdown of solid plant-based foods through mastication can boost the
93 liberation of starch and/or sugar and thereby influence the rate of glucose absorption into the
94 blood (Ranawana, Monro, Mishra & Henry, 2010). Another example is the structuring
95 properties of dietary fibres that through modulation of rheological (flow) properties can
96 influence gastric residence time and therefore impact satiety as well as nutrient absorption
97 (Mackie, Bajka & Rigby, 2016). In addition, fibre components can bind or encapsulate
98 micronutrients controlling their bioaccessibility and hence modulate their bioavailability
99 (Padayachee et al, 2017). Finally, the rate of passage of digesta in the small intestine can be
100 increased by dietary fibres, potentially resulting in delayed nutrient uptake and the triggering

101 of the 'ileal brake' (van Avesaat, Troost, Ripken, Hendriks & Masclee, 2015), as well as
102 affecting the hydration of large intestinal contents, as exemplified by the faecal bulking
103 effect, which is greater for complex vegetable tissues than more refined fibres (Monro,
104 Mishra, Redman, Somerfield & Ng, 2016). By definition, dietary fibres are not digested by
105 human enzymes in the stomach or small intestine, and are therefore transported to the large
106 intestine where they can act as an energy source for the resident microbiota together with any
107 co-passenger micro- and macronutrients (Padayachee, Day, Howell & Gidley, 2017; Dhital,
108 Warren, Butterworth, Ellis & Gidley, 2017). The rate at which this fermentation occurs can
109 vary from very fast (with consequent potential for gastrointestinal discomfort) to very slow
110 (with consequent excretion of much of the fibre), largely dependent on the physical structure
111 of the digesta. The consequences for microbiome populations will also vary with fibre type,
112 but this is more likely to be due to chemical composition as specific microbial community
113 members can contribute the range of hydrolytic activities required to degrade specific
114 polysaccharide structures.

115 *1.3 Solubility is a limited indicator of dietary fibre functionality*

116 Apart from chemical structure, the other characteristic that has been traditionally used to
117 describe dietary fibre types is solubility. Typically, fibre solubility is evaluated after a food or
118 component has been digested under conditions related to those found in the gastrointestinal
119 tract (McCleary et al, 2012) and is separated from insoluble fibre by filtration or
120 centrifugation. Sometimes, solubility is assessed prior to in vitro digestion, and the
121 temperature regimes, centrifugation speeds or filtration cut-offs are often not standardised.
122 Nevertheless, there are clear examples of soluble fibres such as many low molecular weight
123 oligosaccharides and some polysaccharides, and similarly obvious insoluble fibres such as
124 cereal brans, fruit and vegetable skins. However, there is a large number of dietary fibre types
125 found in foods which either have elements of both soluble and insoluble fibre (e.g. cereal

126 flours; Comino, Collins, Lahnstein, Beahan & Gidley, 2014), or which have highly hydrated
127 but insoluble forms (e.g. fruit and vegetable purees; Padayachee, Day, Howell & Gidley,
128 2017). This range of solubilities creates a physical continuum stretching from easily soluble
129 fibres, to poorly soluble, swollen gel-like networks through to insoluble fibres.

130 Various nutritional functionalities are traditionally ascribed to either 'soluble' or 'insoluble'
131 fibre. Soluble fibre is often reported as increasing the 'viscosity' of digesta with consequent
132 effects on reducing gastric emptying and slowing nutrient absorption. The digesta flow
133 profile (rheology), however, depends on the applied stress, and can be shear thinning or
134 exhibit a yield stress behaviour (Lentle and Janssen, 2010). In the case of the former,
135 viscosity (resistance to shear deformation) reduces with applied stress, while in the case of
136 the yield stress fluid the onset of flow ('yield') occurs only above a critical value of the stress
137 ('yield stress'). In the context of foods, shear thinning is typically associated with high
138 molecular weight polymers in solution and yield stress behaviour with networks of food
139 particles. For dietary fibres, the 'viscosifying' effect thus can be due to increase in viscosity
140 or viscoelasticity for high molecular weight soluble polysaccharides, but also for hydrated but
141 insoluble materials such as oat bran or fruit and vegetable fibres, where yield stress behaviour
142 can emerge. Conversely, low molecular weight soluble fibres such as oligosaccharides would
143 not be expected to have any direct 'viscosifying' effect.

144 Further, digesta can demonstrate a significant degree of viscoelasticity (Shelat et al, 2015),
145 defined as the ratio of the loss modulus (viscous part) to the storage modulus (elastic part), as
146 well as exhibiting non-linear rheological effects which result in deviations between shear,
147 squeeze and extensional deformations, which are all present during gastrointestinal transit
148 (Lentle and Janssen, 2010).

149 The effect of comminution e.g. particle size reduction as a result of oral, gastric or intestinal
150 processing, is related to the ability of particles to adhere to each other. These interactions are
151 frequently driven by capillary forces, for example due to the presence of microscopic gas
152 bubbles, as well as due to interaction and bridging adhesion between surface polymer layers
153 of insoluble particles. The adhesive interaction promotes particle clustering resulting in the
154 formation of a cohesive semi-solid. Insoluble fibre, such as cereal brans and seeds, which
155 absorb water and form a polymer-rich interfacial layer can facilitate comminution, and are
156 often described as having the ability to promote the softening of digesta and support regular
157 bowel movements. By contrast, highly condensed or lignified tissues such as cereal hulls or
158 leaf stalks would not be expected to absorb water and thus would display weak adhesive
159 interactions that limits their ability to facilitate comminution.

160 An over-simplification that is sometimes made is that soluble fibres are readily fermented by
161 the resident microbiota but insoluble fibres are not. There are, however, many examples of
162 insoluble fibres e.g. from fruit, vegetable or cereal sources that are readily fermented, and
163 there are a few examples of soluble fibres whose chemistry is apparently so complex that
164 microbial enzymes are unable to hydrolyse them significantly (e.g. psyllium and other
165 mucilage gums). For the case of cereal flours and derived foods, it has recently been shown
166 that there is very similar fermentation behaviour for soluble and insoluble fibre fractions with
167 similar chemical compositions (Comino et al, 2018)

168 There is clearly a need for a more sophisticated way of categorising dietary fibres that is
169 linked to their nutritional functionality, as neither chemical composition nor fibre solubility
170 are sufficiently discriminatory.

171 *2. Dietary fibre functionality is linked to structuring, binding and/or barrier properties*

172 The main nutritional functionality of dietary fibres can be simplified to effects in the
173 digestive tract on:

- 174 - nutrient digestion and uptake rates
- 175 - residence times and passage rates
- 176 - fermentation products and microbiota populations,

177 but each of these is influenced by many different fibre physicochemical properties. For
178 example, nutrient digestion and uptake rates may be influenced by structuring effects that
179 limit the access of digestive enzymes to macronutrient substrates (protein, triglyceride,
180 starch) or the transport of hydrolysed products to the epithelial cell layer, where fibre effects
181 on the mucus may attenuate uptake (Mackie, Bajka & Rigby, 2016; Capuano, 2017).
182 Alternatively, macronutrient digestion may be limited by encapsulation within plant cellular
183 structures (Grundy et al, 2016) and food gels or by complexation with other food components
184 in condensed forms (Zhang, Dhital & Gidley, 2015) e.g. starch in wholemeal pasta. These
185 multiple approaches to achieving comparable outcomes suggest that there are underlying
186 properties that are more characteristic of individual fibre materials. We suggest that these
187 comprise:

- 188 - bulk structuring
- 189 - molecular binding
- 190 - transport barriers

191 Bulk structuring effects of a fibre relate to digesta rheology once interactions with other
192 components are taken into account, and are expected to influence e.g. digesta passage rate,
193 enzyme digestion rates, nutrient transport and fermentation kinetics. Molecular binding of
194 fibres with enzymes (Dhital, Gidley & Warren, 2015), micronutrients (Padayachee, Day,
195 Howell & Gidley, 2017), bile salts (Gunnness, Flanagan, Mata, Gilbert & Gidley, 2016),

196 mucins (Mackie, Bajka & Rigby, 2016; Sriamornsak & Wattanakorn, 2008; Meldrum,
197 Yakubov, Gartaula, McGuckin & Gidley, 2017) and bacteria (Gorham, Williams, Gidley &
198 Mikkelsen, 2016), as well as with other food components, are an under-appreciated feature of
199 many polymeric and particulate dietary fibres. These effects can contribute to all aspects of
200 digestion, passage and fermentation through e.g. reducing enzyme activities, preventing
201 micronutrient bioaccessibility, limiting absorption processes and affecting microbial
202 fermentation. Transport barriers act to separate micro- or macronutrients from other digesta
203 components and typically involve a locally dense structure that is sufficient to limit molecular
204 transport. Examples include encapsulating systems such as plant cells (Dhital, Bhattarai,
205 Gorham & Gidley, 2016) and food gels or condensed processed food forms such as
206 wholemeal pasta (Zou, Sissons, Warren, Gidley & Gilbert, 2016).

207 Whilst structuring, binding and barrier properties provide a reasonably comprehensive
208 framework for categorising the physicochemical properties important for nutritional
209 functionality of dietary fibres, this does not lead directly to classification of the properties of
210 individual types of fibre. For this, the characteristic structural features of fibres that
211 contribute to structuring, binding and barrier properties need to be identified.

212 Structuring (rheology) of dietary fibres in digesta can come from both soluble polymers and
213 swollen particles with both polymer/particle size and concentration being key determinants.
214 Binding phenomena will be expected to involve some specific chemical features. For
215 example the negative charge of pectins serves to enhance binding with positively charged
216 mucin (Sriamornsak & Wattanakorn, 2008; Meldrum, Yakubov, Gartaula, McGuckin &
217 Gidley, 2017) or anthocyanin (Phan, Flanagan, D'Arcy & Gidley, 2017), but reduces binding
218 with phenolic acids (Phan, Flanagan, D'Arcy & Gidley, 2017). More generally, local
219 molecular rigidity and/or density of fibre polymers should be expected to enhance binding
220 through presenting a structurally consistent surface. The key to maintaining an efficient

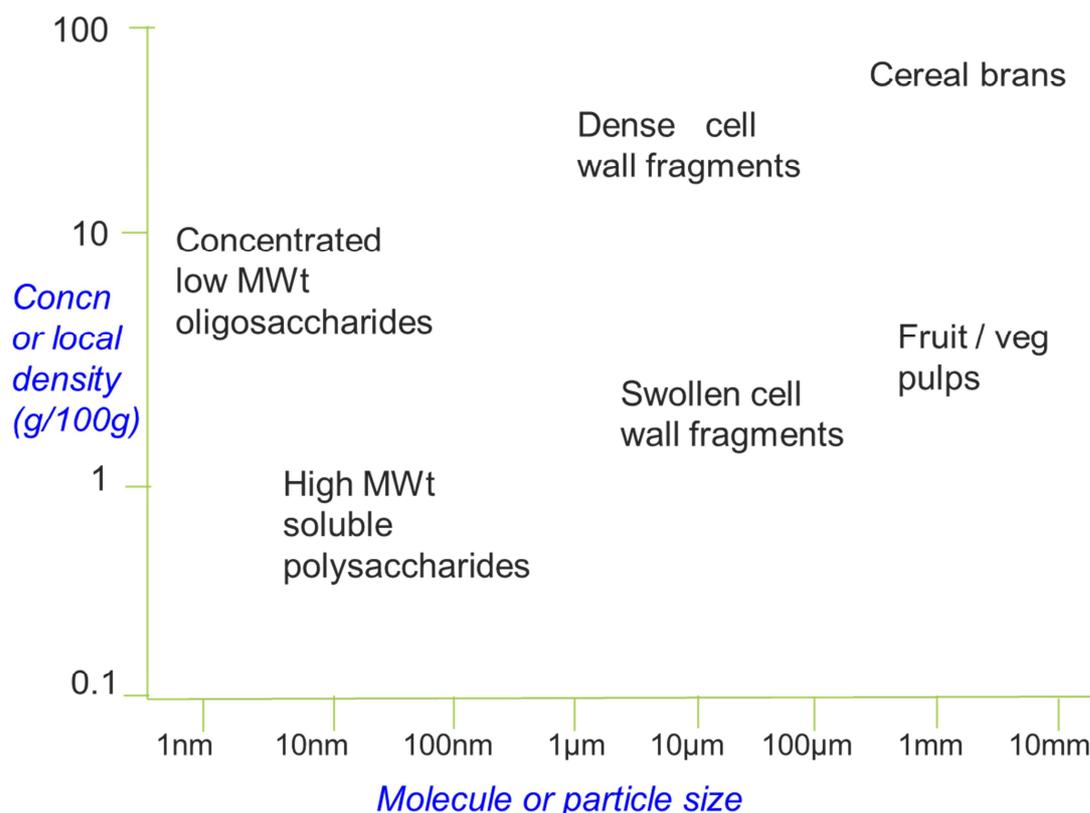
221 transport barrier is to reduce the effective pore size such that e.g. digestive enzymes are
222 retarded or prevented from crossing it. Both molecule/particle size and local density are
223 therefore important considerations. Based on this analysis, molecule/particle size and local
224 density/concentration are the key characteristics of fibre components that would be expected
225 to be related to structuring, binding and barrier properties and therefore to nutritional
226 functionality.

227

228 *3.1 Mapping dietary fibres as a function of size and local density*

229 We propose that a useful approach to categorising the physicochemical properties of diverse
230 types of dietary fibre is to map them by their size and their local density under application
231 conditions, typically fully hydrated. A size axis can cover both dissolved molecules
232 (hydrodynamic size) and particles, with dimensions ranging from about 1 nm for a
233 trisaccharide (the smallest molecule that can be classified as dietary fibre) up to the mm/cm
234 scale for large pieces of cereal bran or fruit/vegetable pulp. Although bulk concentration
235 could be used as another axis, it is argued above that local concentration or density is a more
236 appropriate measure for determining both binding and transport properties. Of course, for
237 dissolved molecules, concentration and local density are equivalent, it is only for particulate
238 materials that the two measures diverge. The range of concentration or density can range
239 from a practical lower level of about 0.1 g/100g up to highly condensed systems at close to
240 100g/100g. For both size and concentration/density, the wide range of possible values
241 suggests that a logarithmic rather than a linear scale would be appropriate for each axis. Such
242 a plot is shown in Figure 1, populated by selected examples of dietary fibre types. Apart from
243 the top left hand corner (which is bound by the physical solubility limit of oligo- or
244 polysaccharides), essentially the whole of the area in Figure 1 is sampled by different dietary

245 fibre types under realistic food and digestion conditions, giving the potential for a high level
 246 of differentiation between individual dietary fibres. We note also that during oral or digestive
 247 processing of food both size and local density may be altered by mechanical or (bio)chemical
 248 conditions, allowing the possibility of tracking changes across the plot illustrated in Figure 1.
 249



250
 251 Figure 1. Mapping of example types of dietary fibres against their molecular or particle size
 252 and concentration or local density. Positions of fibre types are illustrative and not intended to
 253 be quantitatively precise.

254
 255 *3.2 Size / density plots allow differentiation of solubility, viscosity, binding and fermentation*
 256 *properties*

257 To test the utility of size/density plots (Figure 1) to distinguish between fibre properties,
 258 approximate boundaries between soluble/insoluble, ‘flowing’/non-‘flowing’,

259 limited/extensive binding, and rapid/slow fermentation behaviours are illustrated
 260 schematically in Figure 2.

261 Solubility is limited by the size of polymers/colloids that can dissolve. This is in the 100 nm –
 262 1 μ m range for polysaccharides in water, above which entities would normally be expected to
 263 phase separate. As the conventional soluble fibre test is carried out under dilute conditions,
 264 the starting concentration would not be expected to influence the solubility markedly – hence
 265 the vertical boundary division (Figure 2A).

266

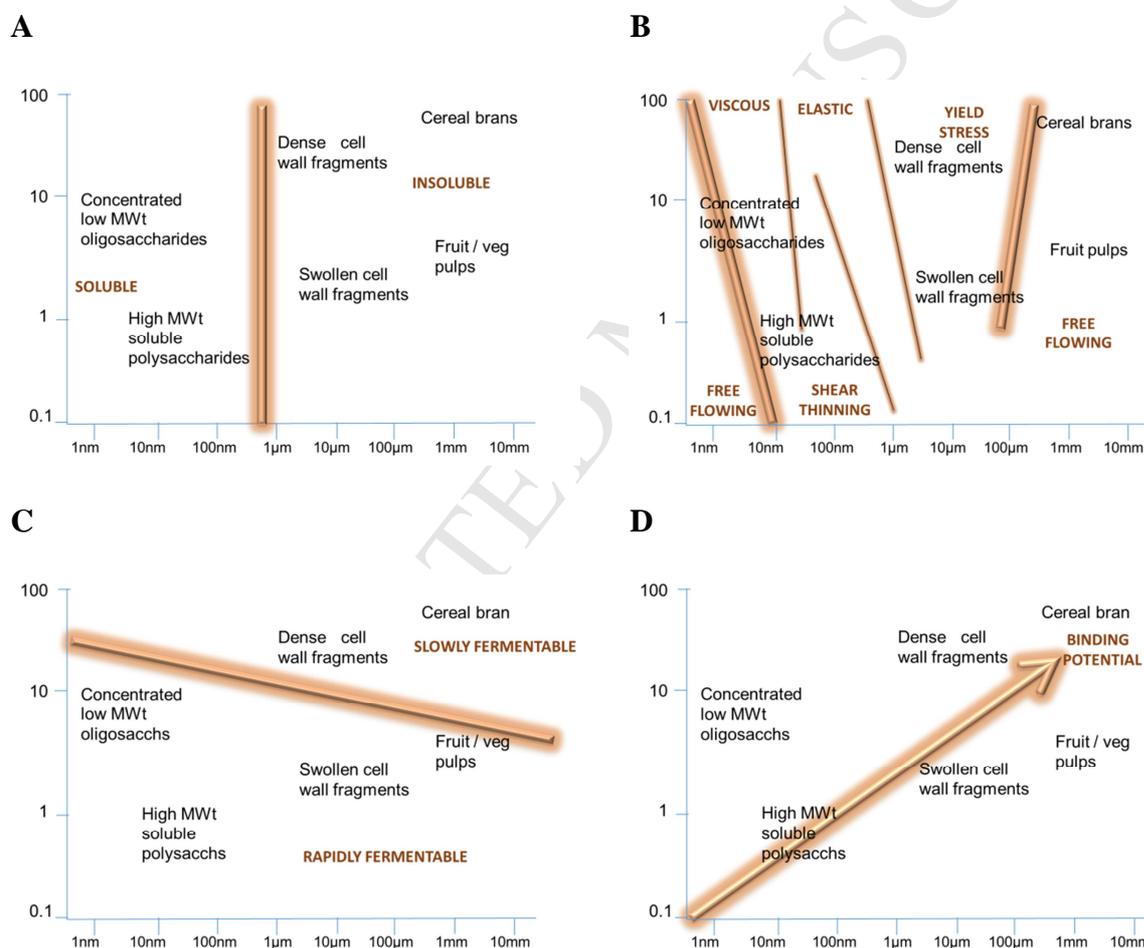


Figure 2. Illustrative expected variations in relevant properties as a function of molecular or particle size (horizontal axes) and concentration or local density (g/100g; vertical axes) in aqueous dietary fibre systems, A. Solubility, B. Flow Behaviour, C. Fermentation rate, D.

Binding potential. The boundary divisions are deliberately broad to emphasise the approximate nature of the size/density cut-offs, and linearity of boundaries is used for convenience.

267

268 Flow behaviour of fibre systems can arise from either polymers in solution or swollen
269 particles in suspension, with lower viscosity values (free flowing systems) for either low
270 molecular weight oligosaccharides in dilute solution or suspensions of relatively non-swollen
271 particles that sediment (Figure 2B). At intermediate values of fibre size, the key aspects are
272 elastic response typical of high molecular weight polymers, and yield stress behaviour
273 characteristic of concentrated suspensions. We note that there are of course many other
274 rheological parameters of relevance to the functionality of dietary fibres, with e.g. several
275 types of viscosity (shear, extensional, dynamic). However, each of these can be expected to
276 show systematic responses to fibre systems in different regions of the size/density map.

277 The fermentation rate of fibre systems is an important parameter because the rate of
278 fermentation is related to the site of fermentation within the large intestine, considering the
279 passage rate. Fermentation is limited under conditions of low water activity as would be
280 found for high concentrations of low molecular weight fibres, but these conditions are not
281 experienced in vivo. Alternatively, fermentation can be slow because the fibre substrate is
282 highly condensed, providing a barrier to efficient utilisation of carbohydrates inside particles
283 of e.g. lignocellulosic brans. As this effect is related to specific surface area, larger particles
284 of less local density should be expected to be fermented at similar rates as smaller particles
285 with higher density. Hence the position of the proposed boundary line in Figure 2C.

286 Binding of diverse molecules (micronutrients, enzymes, bile salts, mucins etc) to dietary
287 fibres can be driven by chemical specificity or surface interactions, so size and density are not

288 expected to be the only factors contributing to the extent and/or strength of binding.
289 Nevertheless, where chemical factors have been taken into account, it is expected that
290 molecules/particles of greater local density will provide more efficient binding than less
291 dense systems due to the larger surface energy of the former. Hence the broad directional
292 arrow in Figure 2D. For larger stiffer particles, we also expect surface roughness to have a
293 major influence that can dramatically increase the effective surface area leading to more
294 binding. In contrast, highly hydrated smaller fragments can show low binding due to lower
295 roughness, despite potentially higher nominal specific surface area.

296 Overall Figure 2 illustrates that a range of features important to dietary fibre functionality
297 have markedly different but systematic behaviours on the size/density plot. This highlights
298 the limited predictive value of categorising fibre as only soluble or insoluble, and suggests
299 that using size/density plots may be a more meaningful way of categorising dietary fibre
300 components such that nutritional functionality can be predicted.

301 *3.3 Challenges and future perspectives*

302 The proposed approach is a broad one, intending to capture all relevant types of dietary
303 fibres. Thus it is necessarily imprecise quantitatively, as generalising behaviour across
304 diverse biological sources and chemical structures would be expected to result in a range of
305 secondary effects on top of those due to size and local density. More quantitative and detailed
306 property maps for individual fibre types at different sizes and densities could in principle be
307 constructed, to compare behaviours between different fibre types. However, the nutritional
308 functionality and preventative health value of dietary fibre is also difficult to quantify
309 precisely, so we expect the maps to be more useful in a semi-quantitative form to compare
310 properties between chemically and biologically diverse dietary fibres.

311 One challenge in populating the maps will be in quantifying the two coordinates of size and,
312 particularly, local density for individual fibres. We note that the effective size of dissolved
313 oligosaccharides and polysaccharides can be obtained directly from measurements of intrinsic
314 viscosity or from size exclusion chromatography, both of which are related to the
315 hydrodynamic volume. The size distribution of particulate material can in general be readily
316 estimated by microscopy or fractional sieving. Local density is equivalent to concentration
317 for dissolved fibres, but is less easy to determine for particulate fibres. An average density
318 can be obtained from sedimentation volume (or hydration capacity) measurements as long as
319 interstitial volumes are taken into account. The bigger challenge is where there is e.g. an
320 intact cellular structure that has a relatively low average density but is bounded by a thin and
321 dense cell wall which provides an effective barrier (Dhital, Bhattarai, Gorham & Gidley,
322 2016). Further work is needed to provide realistic local density data for these types of
323 heterogeneous systems.

324 Once issues of quantification of individual fibre types have been addressed, it will be of
325 interest to consider how best to describe the various regions within the size/density map as a
326 way of communicating the diversity of dietary fibre functionality to consumers. There is a
327 large cohort of consumers who are eager to understand more about why a diet based on plant-
328 based foods, and therefore rich in dietary fibre, is the healthiest option.

329 A potential use for the proposed maps is to identify whether specific regions of the
330 size/density space are related to individual nutritional benefits of dietary fibre such as
331 cholesterol management, glucose absorption, blood lipid management, fermentation
332 throughout the large intestine, or whether these functionalities overlap in size/density
333 coordinates. If such relationships between size/density co-ordinates and nutritional properties
334 are suggested, then this can form the basis for clinical trials in which e.g. a single fibre source
335 is used with designed differences in size and density. It is possible that a diversity of map

336 locations for the range of positive nutritional functionalities ascribed to fibre will provide
337 evidence for why a diversity of plant-based foods and therefore fibre types is associated with
338 optimal health outcomes.

339 One challenge that will need to be addressed is the extent to which individual variation in
340 gastrointestinal physiology and microbiological fermentation over-rides the physical
341 properties of fibres discussed here. A second challenge will be to obtain sufficient data on the
342 physical state of fibres within the digestive tract in humans to understand the mechanisms
343 underlying relationships between ingested fibre size/density and nutritional outcomes. A third
344 challenge is how to simplify the concept for public health messaging, although this needs to
345 be first justified on the basis of property/nutrition correlations and then clinical intervention
346 trials.

347 *Acknowledgements*

348 We thank many members of the Australian Research Council Centre of Excellence in Plant
349 Cell Walls (CE110001008 which supported this work financially) for useful comments
350 during the development of the map concept.

351

352 *References*

353 Anderson, J. W., Baird, P., Davis, R. H. Jr., Ferreri, S., Knudtson, M., Koraym, A., Waters,
354 V., & Williams, C. L. (2009). Health benefits of dietary fiber. *Nutrition Reviews*, 67, 188-
355 205.

356 Burton, R. A., Gidley, M. J., & Fincher, G. B. (2010). Heterogeneity in the chemistry,
357 structure and function of plant cell walls. *Nature Chemical Biology*, 6, 724-732.

- 358 Aune, D., Keum, N., Giovannucci, E., Fadnes, L. T., Boffetta, P., Greenwood, D. C.,
359 Tonstad, S., Vatten, L. J., Riboli, E., & Norat, T. (2016). Whole grain consumption and risk
360 of cardiovascular disease, cancer, and all cause and cause specific mortality: systematic
361 review and dose-response meta-analysis of prospective studies. *British Medical Journal*, *353*,
362 i2716.
- 363 Capuano, E. (2017). The behavior of dietary fiber in the gastrointestinal tract determines its
364 physiological effect. *Critical Reviews in Food Science and Nutrition*, *57*, 3543-3564.
- 365 Chuang S-C et al. (2012) Fiber intake and total and cause-specific mortality in the European
366 Prospective Investigation into Cancer and Nutrition cohort. *American Journal of Clinical*
367 *Nutrition*, *96*, 164–74
- 368 Comino, P., Collins, H., Lahnstein, J., Beahan, C., & Gidley, M. J. (2014). Characterisation
369 of soluble and insoluble cell wall fractions from rye, wheat and hull-less barley endosperm
370 flours. *Food Hydrocolloids*, *41*, 219-226.
- 371 Comino, P., Williams, B. A., & Gidley, M. J. (2018). In vitro fermentation gas kinetics and
372 end-products of soluble and insoluble cereal flour dietary fibres are similar. *Food &*
373 *Function*, *9*, 898 - 905
- 374 Dhital, S., Bhattarai, R. R., Gorham, J., & Gidley, M. J. (2016). Intactness of cell wall
375 structure controls the in vitro digestion of starch in legumes. *Food & Function*, *7*, 1367-1379.
- 376 Dhital, S., Gidley, M. J., & Warren, F. J. (2015). Inhibition of α -amylase activity by
377 cellulose: Kinetic analysis and nutritional implications. *Carbohydrate Polymers*, *123*, 305-
378 312.

- 379 Dhital, S., Warren, F. J., Butterworth, P. J., Ellis, P. R., & Gidley, M. J. (2017). Mechanisms
380 of starch digestion by α -amylase—Structural basis for kinetic properties. *Critical Reviews in*
381 *Food Science and Nutrition*, 57, 875-892.
- 382 Gidley, M. J. (2013). Hydrocolloids in the digestive tract and related health implications.
383 *Current Opinion in Colloid & Interface Science*, 18, 371-378.
- 384 Gorham, J. B., Williams, B. A., Gidley, M. J., & Mikkelsen, D. (2016). Visualization of
385 microbe-dietary remnant interactions in digesta from pigs, by fluorescence in situ
386 hybridization and staining methods; effects of a dietary arabinoxylan-rich wheat fraction.
387 *Food Hydrocolloids*, 52, 952-962.
- 388 Grundy, M.-L., Edwards, C. H., Mackie, A. R., Gidley, M. J., Butterworth, P. R., & Ellis, P.
389 R. (2016). Re-evaluation of the mechanisms of dietary fibre and implications for
390 macronutrient bioaccessibility, digestion and postprandial metabolism. *British Journal of*
391 *Nutrition*, 116, 816-833.
- 392 Gunness, P., Flanagan, B. M., Mata, J. P., Gilbert, E. P., & Gidley, M. J. (2016). Molecular
393 interactions of a model bile salt and porcine bile with (1,3:1,4)- β -glucans and arabinoxylans
394 probed by ^{13}C NMR and SAXS. *Food Chemistry*, 197, 676-685.
- 395 Huang, T., Xu, M., Lee, A., Cho, S., & Qi L. (2015). Consumption of whole grains and cereal
396 fiber and total and cause-specific mortality: prospective analysis of 367,442 individuals.
397 *BMC Medicine*, 13, 59-67.
- 398 Jones, J. M. (2013). Dietary fiber future directions: Integrating new definitions and findings
399 to inform nutrition research and communication. *Advances in Nutrition*, 4, 8-15.

- 400 Lentle, R. G., & Janssen, P. W. M. (2010) Manipulating digestion with foods designed to
401 change the physical characteristics of digesta. *Critical Reviews in Food Science and*
402 *Nutrition*, 50, 130-145.
- 403 Mackie, A., Bajka, B., & Rigby, N. (2016). Roles for dietary fibre in the upper GI tract: The
404 importance of viscosity. *Food Research International*, 88, 234–238.
- 405 McCleary, B. V., DeVries, J. W., Rader, J. I., Cohen, G., Prosky, L., Mugford, D. C., &
406 Okuma, K. (2012). Determination of insoluble, soluble, and total dietary fiber (CODEX
407 definition) by enzymatic-gravimetric method and liquid chromatography: collaborative study.
408 *Journal of AOAC International*, 95, 824-844.
- 409 Meldrum, O. W., Yakubov, G. E., Gartaula, G., McGuckin, M. A., & Gidley, M. J. (2017).
410 Mucoadhesive functionality of cell wall structures from fruits and grains: Electrostatic and
411 polymer network interactions mediated by soluble dietary polysaccharides. *Scientific Reports*,
412 7, 15794.
- 413 Monro, J., Mishra, J., Redman, C., Somerfield, S., & Ng, J. (2016). Vegetable dietary fibres
414 made with minimal processing improve health-related faecal parameters in a valid rat model.
415 *Food & Function*, 7, 2645–2654.
- 416 Padayachee, A., Day, L., Howell, K., & Gidley, M. J. (2017). Complexity and health
417 functionality of plant cell wall fibers from fruits and vegetables. *Critical Reviews in Food*
418 *Science and Nutrition*, 57, 59-81.
- 419 Park, Y., Subar, A. F., Hollenbeck, A., & Schatzkin, A. (2011). Dietary fiber intake and
420 mortality in the NIH-AARP Diet and Health Study. *Archives of Internal Medicine*, 171,
421 1061-1068.

- 422 Phan, A. D. T., Flanagan, B. M., D'Arcy, B. R., & Gidley, M. J. (2017). Binding selectivity
423 of dietary polyphenols to different plant cell wall components: Quantification and
424 mechanism. *Food Chemistry*, *233*, 216-227.
- 425 Ranawana, V., Monro, J. A., Mishra, S., & Henry, C. J. K. (2010). Degree of particle size
426 breakdown during mastication may be a possible cause of interindividual glycemic
427 variability. *Nutrition Research*, *30*, 246–254.
- 428 Shelat, K. J, Nicholson, T., Flanagan, B. M., Zhang, D., Williams, B. A., & Gidley, M. J.
429 (2015). Rheology and microstructure characterisation of small intestinal digesta from pigs fed
430 a red meat-containing Western-style diet. *Food Hydrocolloids*, *44*, 300-308.
- 431 Sriamornsak, P., & Wattanakorn, N. (2008). Rheological synergy in aqueous mixtures of
432 pectin and mucin. *Carbohydrate Polymers*, *74*, 474–481.
- 433 Threapleton D. E., Greenwood, D. C., Evans, C. E. L., Cleghorn, C. L., Nykjaer, C.,
434 Woodhead, C., Cade, J. E., Gale, C. P., Burley, V. J. (2013). Dietary fibre intake and risk of
435 cardiovascular disease: systematic review and meta-analysis. *British Medical Journal*, *347*,
436 f6879.
- 437 van Avesaat, M., Troost, F. J., Ripken, D., Hendriks, H. F., & Masclee, A. A. M. (2015). Ileal
438 brake activation: macronutrient-specific effects on eating behavior? *International Journal of*
439 *Obesity*, *39*, 235-243.
- 440 Zhang, B., Dhital, S., & Gidley, M. J. (2015). Densely packed matrices as rate determining
441 features in starch hydrolysis. *Trends in Food Science & Technology*, *43*, 18-31.
- 442 Zou, W., Sissons, M., Warren, F. J., Gidley, M. J., & Gilbert, R. G. (2016). Compact
443 structure and proteins of pasta retard in vitro digestive evolution of branched starch molecular
444 structure. *Carbohydrate Polymers*, *152*, 441-449.

ACCEPTED MANUSCRIPT

Highlights

- Dietary fibre functionality can be related to physicochemical properties
- Bulk structuring, molecular binding and transport barriers all important
- Fibre categorisation on basis of solubility has limited links to functionality
- Molecule/particle size and local density each related to fibre properties
- New categorisation of dietary fibres proposed, based on size/density maps