Apparent diffusion coefficient for genetic

characterisation of untreated adult gliomas:

a meta-analysis stratified by methods

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Abstract

Background:

Isocitrate dehydrogenase (IDH) mutation and chromosome 1p19q genotyping have become fundamental to the prognostic grouping of adult diffuse gliomas. Apparent diffusion coefficient (ADC) values may enable non-invasive prediction of glioma molecular status. The purpose of this systematic review and meta-analysis was to investigate the diagnostic accuracy of ADC for IDH and 1p19q genotyping, considering measurement techniques and tumour grade.

Methods:

A systematic search of PubMed and Cochrane Library databases was performed in December 2024. Studies were grouped according to the ADC parameter measured and the measurement techniques used. A meta-analysis was performed, supplemented by Egger's regression testing. The quality of studies was assessed with the QUADAS-2 tool.

Results:

Thirty-three studies including a total of 4297 patients fulfilled the inclusion criteria. IDH mutation and 1p19q deletion status were assessed by 30 and 14 studies respectively. Pooled area under the curve (AUC) values for the prediction of an IDH mutation and 1p19q codeletion ranged from 0.743 (0.680-0.805) to 0.804 (0.689-0.919), and 0.678 (0.614-0.741) to 0.692 (0.600-0.783). No significant differences were identified between regional and volumetric measurements, between ADCmean and ADCmin values, or comparing normalised and raw ADC data.

Conclusion:

This meta-analysis supports ADC as an imaging biomarker in untreated gliomas, specifically to predict IDH status. ROI measurement, particularly by a single ADC_{mean}, is rapid, reproducible and appears statistically equivalent

to volumetric read outs. We found no evidence for superior diagnostic accuracy by ADC normalisation. Published ADC thresholds have been summarised for consideration of prospective testing across institutions.

Keywords

Glioma, glioblastoma, isocitrate dehydrogenase (IDH), apparent diffusion coefficient, diffusion-weighted imaging

Key points

- Apparent diffusion coefficient (ADC) values support the prediction of glioma IDH status.
- Regional and volumetric ADC performance was equivalent in meta-analysis.
- ADC thresholds are proposed for sensitive identification of glioblastoma genetics.

Importance of the study

Predicting brain tumour genotypes has become an important objective in radiological diagnosis. This is particularly the case for identifying molecular glioblastoma, which may otherwise be at risk of inequitable low grade triage. Multiple studies have proposed apparent diffusion coefficient values as a biomarker of diffuse glioma IDH and 1p19 status. This systematic review and meta-analysis examined the entire available literature on the subject, including a variety of different measurement methods. Thresholds for prospective research and clinical trial application are proposed.

Introduction

Gliomas represent the most common primary malignancy of the central nervous system (CNS) in adults and are frequently incurable (1–3). Molecular markers of prognostic relevance have become fundamental in the diagnosis of gliomas as defined in the World Health Organization (WHO) 2021 Classification of CNS Tumors (4). Diffuse gliomas are divided into three genetic groups based on the presence of an isocitrate dehydrogenate gene mutation (IDH-mutant, IDH^{mut}), with or without chromosome 1p19q codeletion (1p19q^{codel}) (4). Glioblastoma (GBM) is the most lethal type of glioma, characterised by absence of an IDH mutation (IDH-wildtype, IDH^{wt}) and malignant histology (WHO grade 4) (4). In contrast, most IDH-mutant tumours are low-grade gliomas (WHO grades 2-3), divided into IDH^{mut}/1p19q^{retained} astrocytomas and IDH^{mut}/1p19q^{codel} oligodendrogliomas (2,4,5).

Glioma genotyping is essential for risk stratification and to guide clinical management. GBM is treated by resection followed by radiotherapy and temozolomide chemotherapy (6–9). Maximising tumor resection prolongs the survival of GBM, which creates an argument for prompt identification (5,8,10). A proportion of IDH^{wt} tumours display histological low-grade features but belong to the molecular class of GBM, requiring radical treatment with a risk of comparably poor outcomes (1,2,4). On the contrary, the survival in IDH^{mut} WHO grade 4 tumours tends to be longer than in GBM (1). In IDH^{mut} astrocytomas, postoperative tumor volume is independently associated with survival, whereas 1p19q^{codel} oligodendrogliomas preferentially respond to chemotherapy (11). A preoperative prediction of IDH status could help better utilise sequencing resources in situations where IDH sequencing is not routine for all gliomas and/or where geographical inequities contribute to diagnostic delays (12,13).

Diffusion-weighted imaging (DWI) performed with b-values of 0 s/mm² and 1000 s/mm² (sometimes with an additional b-value of 500 s/mm²) is widely integrated into clinical glioma MRI protocols (14,15). From this, apparent diffusion coefficient (ADC) maps are calculated to estimate the magnitude of diffusion in each image voxel (14). ADC values have been negatively correlated with glioma cellularity in most studies but other factors, including matrix composition, influence ADC (16,17). Several studies reported higher ADC values in IDH^{mut} gliomas compared to IDH^{wt} tumours, which may enable non-invasive prediction of glioma genotype (18–20). However, it is unknown whether measurement methods influence the accuracy of ADC results for glioma molecular diagnosis. Much of the published literature on the diagnostic accuracy of ADC values for characterising gliomas predates the WHO 2021 Classification of CNS Tumors (4). Specifically, older studies highlighted differences in ADC parameters between highgrade and low-grade gliomas, without reporting on genetic status (21). This raises the possibility of a WHO grade influence on ADC diagnostic accuracy.

The purpose of this systematic review and meta-analysis was to investigate the diagnostic accuracy of ADC for glioma IDH and 1p19 status prediction by measurement techniques and considering the possible influence of glioma WHO grade.

Methods

A literature review and meta-analysis were conducted according to the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) guidelines (22). The meta-analysis component of the research was prospectively registered on 23rd July 2024 with the University of Nottingham Repository (http://doi.org/10.17639/nott.7439)(23).

Information sources and search strategy

A systematic search of Pubmed and Cochrane databases was commenced on 1st September 2023 and last updated on 21st December 2024 to identify studies reporting the diagnostic accuracy of ADC for glioma IDH and 1p19q status prediction. To capture the genotyping era, a filter was applied to only include studies published since 2013 (10 years prior to the commencement of the analysis). Full details of the search terms are provided in the Supplementary Material.

Eligibility criteria

Inclusion criteria were: original research in diffuse glioma (WHO grades 2-4), DWI/ADC or mean diffusivity/ADC calculated from diffusion tensor imaging (DTI) performed on glioma patients pre-treatment, assessment of the diagnostic or prognostic value of one or more diffusion parameters for the purpose of glioma grouping (e.g. WHO grade, genotype), quantitative measurements described without or alongside histogram

parameters or advanced computation, and studies no more than 10 years retrospect to capture the genotyping era (24).

Exclusion criteria were: no diffusion-weighted (DWI/ADC or diffusion-tensor imaging [DTI]) sequence interpretation, animal/laboratory measurements, studies confined to paediatric gliomas (defined as <5 adult cases), review articles, case reports of <5 cases, conference abstracts, no English full text, any previous treatment (surgery, radiotherapy, chemotherapy) and tumour types other than diffuse glioma.

Study selection and data collection process

The titles and abstracts of all studies identified by the search were uploaded into the Rayyan online systematic review platform (25). Each abstract was independently screened by two reviewers (F.B. and J.S.). Following unblinding of each reviewer's screening results, conflicts were resolved through consensus. Candidate full texts were independently reviewed by the same reviewers against the inclusion and exclusion criteria with conflicts resolved after unblinding. Each reviewer (F.B. and J.S.) extracted data from all included studies into Microsoft Excel (Microsoft Excel for Mac, Version 16.88, Microsoft Corporation). The complete data extraction was compared, and discrepancies were resolved in consensus discussion with two senior authors (S.T. and N.S.).

Data items

Items extracted consisted of author details and publication year, study design, research purpose, patient number, age, sex, microscopic WHO grade(s) and histopathological diagnoses, immunohistochemistry methods, IDH and 1p19q status, DWI acquisition details, diffusion parameter(s) measured, measurement methods (e.g. region of interest [ROI] defined as a single slice measurement(s) or volume of interest [VOI] defined as a measurement(s) obtained from multiple image slices) and interobserver testing, where published. The WHO numerical grade is Arabic throughout the manuscript in keeping with the latest WHO 2021 convention. The original grading nomenclature has been retained where older Roman grades were used in research publications. The key outcome measure was the receiver operating characteristic [ROC] area under the curve [AUC] value of the diffusion parameter(s) used for IDH or 1p19q genotyping of gliomas.

Study risk of bias assessment

Risk of bias was assessed using the Quality Assessment of Diagnostic Accuracy Studies-2 (QUADAS-2) tool (26). The QUADAS-2 questions were defined through a planning consultation between the authors and are listed in the Supplementary Material. Studies which did not report consecutive or random enrolments, were assigned high risk of bias in the patient selection domain. If more than one reference standard was used, a high risk of bias was assigned in the reference standard domain. For the flow and timing domain, any interval between the index and reference tests greater than two weeks was labelled as high risk of bias. If the interval between the index and reference tests was not specified, this was labelled as unclear risk of bias. Each study was independently assessed by two reviewers (F.B and J.S.) with disagreements resolved through consensus with one senior author present (N.S.).

Statistical analysis and synthesis methods

A meta-analysis was performed to estimate the diagnostic performance of ADC measurements. For studies with 95% confidence intervals (CIs) but not standard error (SE) values, the 95% CIs were used to calculate the SE values by dividing the CI range by 3.92 as specified in the Cochrane Handbook for Systematic Reviews of Interventions (27). For studies in which AUC values were reported without an SE value or CI, the corresponding authors were emailed three times to supply the missing data. If SE and/or CI values remained unconfirmed, an estimate of the SE value was calculated using Equation II described by Cortes and colleagues (28).

Studies were grouped according to the ADC parameter measured (e.g. minimum ADC (ADCmin) or mean ADC (ADCmean)), and the method of measurement (ROI or VOI). Studies which marked ROIs in the visually perceived lowest regions of the ADC map were coded as ADCmin. Studies which described placing a ROI across the largest axial tumour cross section were classified as ADCmean. Studies taking an average ADC measurement from multiple ROIs placed within a tumour, but not targeting the lowest regions of the ADC map, were also classified as ADCmean. Studies, which targeted the visually perceived lowest ADC regions were classed as ADCmin, even where these lowest ADC values were averaged. This grouping served the purpose of meta-analysis to align methods as closely as possible, irrespective of individual publication nomenclature. The data were further grouped based on whether absolute or normalised (e.g. to contralateral normal-appearing white matter [CNAWM]) ADC measurements were recorded.

The meta-analysis was performed using the 'Meta-analysis' function on JASP software (JASP Team 2024, version 0.18.3 [Apple Silicon]). The fixed effects model was initially used with Cochran's Q statistic to test for residual heterogeneity, whereby a threshold of p<0.05 indicated significant residual heterogeneity. In the case of significant residual heterogeneity, the maximum likelihood model was applied instead. Results are displayed in forest plots with summary estimates presented. Funnel plots were produced and 'Egger's regression test' was performed to assess for plot asymmetry if at least 5 studies were available for analysis. To analyse the influence of WHO grade on AUC values, a linear regression was performed using the quoted AUC values and proportion of WHO grade 4 tumours in each cohort where at least 5 studies reported a specific ADC parameter. Linear regression was performed using GraphPad Prizm Cloud (GraphPad Software, Boston, Massachusetts USA, www.graphpad.com).

Results

Study selection and overview

The database searches yielded 808 unique studies, of which 33 were eligible for inclusion in this review. A PRISMA flow diagram summarising the study selection process is provided in Figure 1.

Thirty-three studies including a total of 4297 patients were included in the final analysis with a mean of 130 patients per study and range of 11 to 475. All 33 studies were retrospective. Across the 33 studies, 30 studies assessed prediction of IDH mutation status (29–58), and 14 studies assessed prediction of 1p19q codeletion status (47–57,59–61). Tables 1-3 summarise the data extracted from each of the 33 included studies grouped according to whether they assessed IDH mutation, both IDH mutation and 1p19q codeletion, or 1p19q codeletion alone.

Cohort composition

The analysed WHO grades varied across the included studies. The most common cohort mixes were WHO grade 2, 3 gliomas (n=11) (30–33,47–51,59,61), and WHO grade 2, 3, 4 gliomas (n=10) (38–41,53–58). A breakdown of studied WHO grades is provided in the Supplementary Material.

MRI field strength

Three studies reported using 1.5T MRI (37,44,61), 20 studies used 3.0T MRI (29,32,35,36,38,39,41–43,46,47,50,51,53–57,59,60), and 9 studies used a combination of 1.5T and 3.0T MRI from multiple vendors (30,31,33,34,40,45,48,52,58). The MRI field strength was not reported in one study (49).

ADC measurement

Eighteen studies used ROI-based measurements (32–34,36,38–42,44–47,50,55,58–60), 13 studies used VOI-based measurements (29,35,37,48–54,56,57,61), and two studies used both ROI and VOI methods (30,31). Of the studies using VOI methods, four studies used automated tumour segmentation techniques (48,49,53,56), whilst the remaining 11 studies described manual whole tumour segmentation (29–31,35,37,50–52,54,57,61). For tumour segmentation, four studies used T2-weighted (T2w) sequences (30,31,50,51), one study used T2-FLAIR (FLAIR) (54),

one study used T2w and FLAIR (52), one study used T1-weighted (T1w) or T2w (37), and one used T1w alone (35). Three studies did not state what sequences were used for tumour segmentation (29,57,61). Studies were broadly consistent in excluding calcified, cystic, haemorrhagic, or necrotic regions of tumours from measurements. Of the 20 studies using ROI methods of ADC measurement, 14 studies assessed minimum ADC values (ADCmin) (30,32–34,36,41,42,44–47,55,59,60) and 12 studies assessed mean ADC values (ADCmean) (31,33,36,38–40,44,45,49,50,58,60). Nine studies did not describe exact ROI definitions for ADC measurements (31,34,37,43,48–50,52,53).

Fifteen studies described measuring normalised ADC values using a comparative ROI (29– 33,36,38,39,44,46,47,55,57,59,60). CNAWM with location not further specified (NFS) was used as a comparison in nine studies (29,33,36,38,39,46,55,59,60), whilst contralateral normal-appearing centrum semiovale and contralateral normal-appearing posterior limb of internal capsule were listed in four (30,31,44,57) and two (32,47) studies respectively.

Eleven studies described methods of two observers working in consensus to mark ROIs or VOIs (35,36,38,39,41,44,56,57,59–61). Thirteen studies provided data on intraclass correlation coefficient (ICC) values of ADC measurements (29–31,33,42,45,46,50–53,55,58). The ICC values of ADC measurements were greater than 0.80 in 12 of the 13 studies (29–31,33,42,46,50–53,55,58). One study reported ICC values of 0.532 and 0.598 for ROI-based ADCmean and ADCmin measurements respectively in a cohort of WHO 4 gliomas (45). In two studies ADC measurements were performed by one observer (34,50).

Meta-analysis

Studies were grouped for meta-analysis based on the ADC parameter which they assessed. The primary groups were ADCmean (n=19) (30,31,33,36,38–40,44,45,49–54,56,58,60,61), ADCmin (n=14) (29,30,32–34,36,38,41,42,44,45,47,51,55), rADCmean (n=6) (30,31,33,36,39,60), and rADCmin (n=9)

(30,32,33,36,44,46,47,55,59). Studies which provided AUC values for other ADC metrics, but not one of the 4 previously listed ADC parameters, are described separately (n=4) (35,37,48,50). The groupings of studies by ADC parameters is provided in the Supplementary Material.

IDH

ADCmean

Eight studies used ROI methods, and seven studies used VOI methods to measure ADCmean values to classify glioma IDH mutation status. The cohorts of the eight studies using ROI methods comprised of WHO 1-4 (n=1)(36), WHO 2-3 (n=2)(30,33), WHO 2-4 (n=3)(39,40,58), and WHO 4 gliomas (n=2)(44,45). The seven studies using VOI methods involved cohorts of WHO 2-3 (n=2)(30,39), WHO 2-4 (n=4)(31,53,54,56), and WHO 3 gliomas (n=1)(52).

Two studies by Maynard et al. (33) and Thust et al. (31) were excluded from meta-analysis to avoid pseudoreplication as these reported data on the same cohort of gliomas as another study in the meta-analysis (30). A study by Cheng and colleagues was also excluded from the meta-analysis despite specifying ADCmean values (56), because of retrospective fusion ROI measurements to biopsy sites, which differs from the ROI and VOI methods used in all other studies (56). Cheng et al. reported an AUC value of 0.500 in their cohort of 10 WHO 2-4 gliomas (56).

The pooled AUC values for ROI and VOI methods were 0.760 (0.701-0.818) (I^2 =72.623) and 0.799 (0.719-0.879) (I^2 =76.737) as shown in Figure 2 (Panel A and B respectively).

rADCmean

Five studies used ROI methods, and two studies used VOI methods to measure rADCmean values to classify glioma IDH mutation status. The cohorts of the five studies using ROI methods comprised of WHO 1-4 (n=1)(36), WHO 2-3 (n=3) (30,31,33), and WHO 2-4 gliomas (n=1) (39). Both studies using VOI methods analysed cohorts of WHO 2-3 gliomas (30,31).

Two studies were again excluded to avoid pseudoreplication (31,33). Due to this, three studies contributed to the meta-analysis of ROI methods, and no meta-analysis was possible for rADCmean studies using VOI methods. The pooled AUC value for studies using ROI methods was 0.778 (0.687-0.870) (I²=79.330) as shown in Figure 2 (Panel C). Using VOI methods, Thust et al. 2021 reported an rADCmean AUC value of 0.82 (0.76-0.88) in a cohort of 283 WHO 2 and 3 gliomas (30).

ADCmin

Eleven studies used ROI methods, and three studies used VOI methods to measure ADCmin values to classify IDH mutation status. In three of the eleven ROI studies, the generation of ADCmin measurements involved 3 or more individual ROI placements with ADC averaging (36,44,55). The cohorts of the studies using ROI methods comprised of WHO 1-4 (n=1) (36), WHO 2-3 (n=4) (30,32,33,47), WHO 3 (n=1) (34), WHO 2-4 (n=2) (41,55), WHO 3-4 (n=1) (42), and WHO 4 gliomas (n=2) (44,45). Of the two studies using VOI methods, Thust et al. assessed a cohort of WHO 2-3 gliomas, whilst Villanueva-Meyer at al. assessed a cohort of WHO 2 gliomas (29,30).

The studies by Maynard et al. (33), and Xiong et al. (47) were excluded from the ROI meta-analysis to avoid pseudoreplication, because their cohorts overlapped with those of other studies in the analysis (30,32). Studies using ROI methods gave a pooled AUC of 0.743 (0.680-0.805) (I^2 =76.055) (Figure 2, Panel D), whilst studies using VOI methods gave a pooled AUC of 0.804 (0.689-0.919) (I^2 =89.969) (Figure 2, Panel E).

Eight studies used ROI methods (30,32,33,36,44,46,47,55), and one study used VOI methods (30) to measure rADCmin values to classify glioma IDH mutation status. In 3 studies, ADCmin measurements involved 3 or more individual ROI placements for averaging (36,44,47,55). The eight studies using ROI methods, analysed cohorts of WHO 1-4 (n=1) (36), WHO 2-3 (n=5) (30,32,33,46,47), WHO 2-4 (n=1) (55), and WHO 4 gliomas (n=1) (44). Thust et al. was the only study to use VOI rADCmin methods with a reported AUC of 0.72 (0.66-0.79) (30).

One study by Maynard et al. was again excluded to avoid pseudoreplication (33) with another study in the meta-analysis (30). The study by Xiong and colleagues (47) was also excluded to avoid pseudoreplication, as the study cohort (n=84) appeared to overlap with the cohort of another study (n=90) included in the meta-analysis (51). The pooled AUC values for the remaining six studies which used ROI methods was 0.802 (0.72-0.877) (I²=83.523) as shown in Figure 2 (Panel F).

1p19q

ADCmean

Six studies assessed ADCmean values as a classifier of glioma 1p19q codeletion status, of which five used VOI methods (51,53,54,56,61) and one used ROI methods (60).

Of the 5 studies using VOI methods, Nuessle et al.(54), Liu et al. (51), and Su et al. (53) only assessed the prediction of 1p19q codeletion amongst IDH^{mut} gliomas whilst Latysheva et al., used a cohort comprising 71 WHO 2-3 gliomas of which 33 were oligodendrogliomas and 38 were astrocytomas (note molecular status was not reported for all) (61). Cheng et al. used a cohort of WHO 2-4 gliomas which included 6 IDH^{wt}, 5 IDH^{mut}/1p19q^{retained}, and 2 IDH^{mut}/1p19q^{codel} gliomas, however due to their methods of retrospectively identifying ROIs from surgical biopsy

sites by fusing intraoperative MRI with pre-operative imaging, this study was not included in the meta-analysis (56). Cheng et al. reported an AUC of 0.916 using this method (56). The pooled AUC of the remaining 4 studies using VOI methods was 0.692 (0.600-0.783) (I²=44.948) (forest plot provided in Supplementary Material).

Cui et al. were the only to use ROI methods in a cohort of 35 WHO 2 gliomas, of which 33 were IDH-mut, reporting an AUC of 0.820 for the prediction of 1p19q codeletion status (60).

rADCmean

One study provided an AUC value for rADCmean as a predictor of 1p19q codeletion status. Cui et al. used ROI methods to measure rADCmean values in a cohort of 35 WHO 2 gliomas, of which 3 were IDH^{wt} and 32 were IDH^{mut}, reporting an AUC value of 0.81 (0.67-0.95)(60).

ADCmin

One study assessed ADCmin values as a classifier of 1p19q codeletion status. Ma et al. reported an AUC value of 0.68 (0.57-0.80) when using the average ADC value of three ROIs placed on the visually perceived lowest regions of the ADC map to identify 1p19q codeletion amongst IDH^{mut} gliomas (55).

rADCmin

Three studies measured rADCmin values using ROI methods to predict 1p19q codeletion status giving a pooled AUC value of 0.678 (0.614-0.741) ($I^2=0$) (forest plot provided in Supplementary Material) (47,55,59). Ma et al. and Yang et al. assessed this amongst IDH-mut gliomas only whilst Xiong et al. used a cohort of 84 oligodendroglial tumours (47,55,59).

Summary of studies not included in the prior groupings

Two studies reported ADC entropy as the best performing ADC parameter for the classification of IDH mutation status. Su et al. used VOI methods to measure ADC entropy values in a cohort of 52 WHO 3 gliomas, obtaining an AUC of 0.724 (0.572-0.845) (35). Gihr et al. similarly used VOI methods in a cohort of 87 WHO 1-4 gliomas and reported an AUC value of 0.804 (0.6849-0.9231) (37).

Aliotta et al. reported AUC values for 75th percentile and 50th percentile ADC values in the classification of IDH mutation and 1p19q codeletion status, respectively (48). Using automated tumour volume segmentation, in a cohort of 41 WHO 2-3 gliomas, 75th percentile ADC achieved an AUC of 0.81 (0.78-0.84) for the classification of IDH mutation status and 50th percentile ADC achieved and AUC of 0.83 (0.80-0.86) for the classification of 1p19q codeletion (48).

Cho et al. measured median tumour rADC values using VOI methods in a cohort of non-enhancing WHO 2-3 gliomas (57). They reported AUC values of 0.848 for distinguishing IDH^{mut} astrocytomas from all other gliomas, 0.805 for distinguishing between IDH^{mut} astrocytomas and IDH^{mut} oligodendrogliomas, and 0.883 for distinguishing IDH^{mut} astrocytomas from IDH^{wt} gliomas (57).

Lee. et al assessed whether ADC values could be used to distinguish IDH^{mut},1p19q^{retained} gliomas from IDH^{wt} gliomas in a cohort of 110 WHO 2 and 3 gliomas. They reported 10th percentile ADC values, measured using ROI methods, provided an AUC of 0.751 (0.617-0.886) (50).

Aliotta et al. assessed whether IDH^{mut}/1p19^{retained} gliomas could be distinguished from all other gliomas using ADC histogram parameters derived from automatically segmented tumour volumes (49). In a validation set of

93 WHO 2 and 3 gliomas from The Cancer Imaging Archive (TCIA) database, both ADCmean and volume of ADC >1.5 provided an AUCs of 0.81 (CIs not provided) (49).

Influence of WHO grade on ADC performance for IDH genotyping

The linear regression analysis of reported AUC values for each ADC parameter (with at least 5 observations) and the proportion of WHO 4 gliomas in the study cohorts showed no significant association between AUC and WHO grade (p>0.05 for all ADC parameters). Full results are provided in the Supplementary Material.

ADC thresholds

ADC threshold values for IDH genotyping were proposed by multiple studies included in the meta-analyses, with values below the threshold denoting IDH^{wt} status in each cohort. There were 34 instances of threshold recommendations reported across the included publications, for which descriptive statistics are listed in Table 4. The maximum (Max) value specifies the threshold at which sensitivity for IDH^{wt} status would be maximised across the included studies.

Risk of bias in studies

Egger's regression analysis for funnel plot asymmetry

Funnel plots were created, and Egger's regression analysis was performed for the ADCmean (ROI), ADCmean (VOI), ADCmin (ROI), and rADCmin (ROI) predictions of IDH mutation status as these groups contained at least 5 studies. Funnel plots can be found in the Supplementary Material. Egger's regression analysis revealed no significant funnel plot asymmetry for each of the ADC parameters (ADCmean (ROI) z = -1.897 (p=0.058), ADCmean (VOI) z = -1.502 (p=0.133), ADCmin (ROI) z = -1.449 (p=0.147), rADCmin (ROI) z = -1.850 (p=0.064)).

The results of the QUADAS-2 tool risk of bias and applicability assessments, as well as information on individual studies are provided in the Supplementary Material. All studies were retrospective. Five studies reported enrolling a consecutive or random sample of patients, whilst the remaining 28 studies were unclear regarding patient selection methods. Two studies reported using one observer to obtain ADC measurements with no consensus or comparison with a second observer. In one study, one observer obtained all ADC values with a smaller subset of cases being reviewed by a second observer. Seven studies did not specify methods of testing for IDH mutation or 1p19q deletion. Twenty-seven studies did not specify the time between the index and reference tests. Nine studies were deemed to be at high risk of bias in the flow and timing domain due to either not all patients receiving the same reference standard (e.g. in some studies 1p19q co-deletion was assessed with either fluorescent immunocytochemistry or chromosomal analysis), or not all patients being included in the final analysis (e.g. due to missing data on IDH mutation status). In two studies, the interval between the reference and index test was reported as less than 1 year, which was also allocated high risk of bias.

Discussion

Defining molecular status has become central to the prognostic grouping of diffuse gliomas (4). MRI genotyping has the potential to impact the timing and extent of tumour resection (62), including to accelerate radical therapy for non-contrast-enhancing glioblastoma stages (8), which may otherwise receive a low-grade working diagnosis (63). Based on numerous studies, which proposed ADC as an imaging biomarker in glioma, we investigated the performance of ADC parameters for IDH and 1p19q genotyping.

This systematic review highlights that the published literature is spread across several methods of ADC measurement. Firstly, methods varied between ROI and VOI approaches. The meta-analysis indicates only minor differences between the diagnostic performance of regional and volumetric values measurements (AUC ROI 0.760

vs VOI 0.799 for ADC_{mean} and ROI 0.743 vs VOI 0.804 for ADC_{min}). The similarity between the diagnostic accuracy of ROI and VOI ADC for the classification of IDH mutation status has been documented previously (64,65). Volumetric measurements appear optimal through capture of all representative tissue, but we found no statistical evidence for their superiority. Regional measurements are much easier to obtain and require no software beyond standard MRI viewing equipment. Thus, ROI ADC values may serve as the fastest approach with the advantage of being widely available.

For the meta-analysis, studies were grouped by measurement extent (ROI vs. VOI), by ADC parameter (mean vs. min) and according to whether ADC values were absolute or normalised, whereby each item could influence quantitative results. Surprisingly, the diagnostic performance of ADC for IDH genotyping is similar across all of these method differences. Comparing the 95% CIs of the pooled AUC values revealed no statistically significant differences between ROI and VOI measurements of ADC_{mean}, rADC_{mean}, aDC_{min}, or rADC_{min}. Drawing one ADCmean ROI across the largest glioma cross-section may be perceived as easier than deciding on the visually lowest ADC parts of a tumour. Furthermore, in several (3 of 11) studies using ROI ADC_{min} measurements, these involved outlining 3 or more regions for averaging.

Applying the maximum threshold proposed by studies for a chosen ADC method would represent the most sensitive strategy for identifying IDH^{wt} status (at the expense of specificity over using the median). For ROI ADCmean, this would correspond to $1.3 \times 10^{-3} \text{ mm}^2/\text{s}$ (n=7 studies), or alternatively an rADCmean threshold of 1.75 could be memorable for its similarity to perfusion thresholding, although this is based on fewer (n=3) studies (66). ROI ADCmin values < 1.1 $\times 10^{-3} \text{ mm}^2/\text{s}$ or ROI rADCmin values <1.4 should prompt suspicion for IDH^{wt} genetics.

Encouragingly, the ICC calculations exceeded 0.80 in 12 of 13 studies which reported these, confirming high interobserver agreement for ADC (29–31,33,42,45,46,50–53,55,58). The only study reporting lower ICC values (0.53 for ROI ADCmean and 0.60 for ROI ADCmin) was conducted in WHO grade 4 gliomas, potentially reflecting a

complexity in marking ROIs within necrotic tumours (45). Twenty-four of 33 studies excluded necrotic, cystic, haemorrhagic, or calcified glioma components, where present. This seems justified based on their potential to confound tumour measurements; however little data exists on the performance of ADC according to such exclusions. In one study by Lewis et al. using ADC texture analysis, the performance for IDH typing was slightly improved by excluding necrotic gliomas (67).

The pooled AUC estimates were lower for 1p19q genotyping than IDH genotyping, with estimates of 0.692 (0.600-0.783) for volumetric ADC mean measurements, and 0.678 (0.614-0.741 for regional rADCmin measurements. Moreover, in 8 of 9 studies, either only IDH^{mut} gliomas were examined, or study cohorts including mostly IDH^{mut} gliomas were examined. This markedly limits the generalisability of 1p19q results for clinical practice. Glioma IDH genotyping is essential due to the strong association with survival, whereas prognosis differences are often less pronounced for IDH^{mut} subgroups, although 1p19q status may influence systemic therapy (6).

Normalisation of ADC values is aimed at reducing MRI scanner and acquisition-related variations in ADC measurements. If normalisation is performed, the centrum semiovale represents a preferable target (68). No significant differences were observed between the pooled AUCs of normalised ADC measurements and raw ADC measurements. Several studies provided AUCs for both absolute and normalised ADC values. Du et al., Thust et al. (2018 and 2021), and Liu S et al. all reported similar AUCs when using regional measurements of ADCmean and rADCmean (30,31,36,39). Likewise, no significant AUC differences were identified in the 6 studies which assessed absolute and normalised regional ADCmin measurements (30,32,36,44,47,55).

The possible effect of WHO grade distribution in the different study cohorts was explored based on the hypothesis that developing necrosis may confound ADC measurements in WHO grade 4 glioblastoma, possibly more so than in solid IDH^{wt} early disease stages (69). However, many studies that analysed cohorts of different WHO grades did not report diagnostic accuracy separately for each WHO grade (38–41,53–56,58). The small number of studies

within each meta-analysis together with limited cohort details, precluded a further subgroup analysis. It is however noteworthy that one research group stratified WHO grade 2-3 gliomas by enhancement pattern (non-enhancing, solid-patchy enhancing and rim-enhancing with necrosis) with an observation that ROI ADC values appeared strongly associated with IDH genotype in non-enhancing and solid-patchy enhancing tumours, but not in necrotic tumours (30).

Limitations

Our research was limited by several factors. Heterogeneity exists within the meta-analyses with l² values ranging from 0% to 89.969%. All evidence included in this review originates from retrospective research. ADC maps from 1.5T and 3T ADC MRI scanners of different vendors contributed to the analysis with potential to impact quantification of absolute ADC values. Several studies omitted reporting on the blinding of observers to histopathological data, and some studies did not state the CIs of the AUC values. Studies in which the ADC values were combined with other imaging parameters (e.g. visual parameters on anatomical MRI) had to be excluded, if the AUC values for ADC were not provided separately.

To facilitate meta-analysis, studies were grouped according to the final consensus of 4 reviewers (F.B., J.S., NS., S.T), however this may not reflect the entirety of methodological differences. Funnel plots could only be produced for analyses including five or more studies, and in a few instances SE values were deducted by a mathematical formula (28). We did not explore the potential impact of manual (30) versus automated (48,49,53,56) glioma segmentation on ADC results, and this may represent a topic for further study where the availability of automated volumetric ADC extraction will likely grow. The future use of ADC values for glioma molecular diagnosis will most likely benefit from integration with other parameters such as visual features, physiological MRI metrics and age (18,33). Furthermore, the presence of necrosis has been associated with IDHwt status and may predict this

irrespective of ADC (70). In the future, automated segmentation may facilitate the integration of volumetric ADC measurements into clinical workflows (71). Furthermore, prospective research is required to validate proposed ADC thresholds.

Practical guidance

Whilst considering limitations of the presented data, and meta-analysis generally, it appears possible to arrive at preliminary guidance on how to perform and use ADC measurements in clinical practice. Firstly, obtaining a measurement is preferable to qualitative inspection, which is an unreliable predictor of IDH status with poor interobserver agreement (72). The placement of a circular ROI to measure ADCmean in the largest solid tumour cross-section (or in the largest solid tumour focus, where this is not an entire cross-section due to necrosis) is deemed suitable with a view to workflow integration. If preferred, normalised values, or alternatively ADCmin measurements, can be justified where their use is already established in local practice. We refer to Figure 1 in Maynard et al. 2020 for an example of glioma and normal white matter ROI placements, with the note that these should be interpreted as draft guidance (33). In support of the identification of high-risk disease, it is suggested to align measurements of ADC towards sensitive IDHwt identification, particularly for lesions of perceived 'low grade' morphology. Radiological assessments should consider further factors, including age and differentials of diffuse glioma.

Conclusion

This meta-analysis supports ADC as an imaging biomarker in untreated gliomas, specifically to predict IDH status. Published ADC thresholds have been summarised and should be considered for prospective testing. ROI measurement, particularly a single ADCmean, is rapid, reproducible and appears statistically equivalent to volumetric read outs. We found no evidence for altered diagnostic accuracy through ADC normalisation. Future research should aim to formulate numerical thresholds across multiple institutions.

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Conflict of interest

The authors declare no conflict of interest for the work presented.

Contributions

ST and NS conceptualised the review topic and created the initial draft protocol. ST, NS, FB and JS were involved in reviewing and refining the research protocol. FB and JS performed the screening, data extraction and QADAS-2 assessments. ST and NS led the consensus reviews. CT defined the statistical methods. All authors contributed to the results analysis, interpretation and manuscript.

S.T. and F.B. are the guarantors of the research.

Data availability

A summary of the data extracted from all included studies is provided in the Supplementary Material together with the QUADAS-2 assessment, funnel plots and linear regression. Further information can be made available upon reasonable request. No original study data were generated in this research.

References

- Pinson H, Silversmit G, Vanhauwaert D, Vanschoenbeek K, Okito JPK, Vleeschouwer S De, et al.
 Epidemiology and survival of adult-type diffuse glioma in Belgium during the molecular era.
 Neuro Oncol. 2024 Jan;26:191–202.
- Ostrom QT, Price M, Neff C, Cioffi G, Waite KA, Kruchko C, et al. CBTRUS Statistical Report: Primary Brain and Other Central Nervous System Tumors Diagnosed in the United States in 2015-2019. Neuro Oncol [Internet]. 2022;24(Suppl 5):V1–95. Available from: https://pubmed.ncbi.nlm.nih.gov/36196752/
- Sharifian MJ, Igland J, Klungsøyr K, Engeland A, Zhou A, Bjørge T. Incidence trends of adult glioma in Norway and its association with occupation and education: A registry-based cohort study. Cancer Epidemiol. 2024 Apr;89.
- Louis DN, Perry A, Wesseling P, Brat DJ, Cree IA, Figarella-Branger D, et al. The 2021 WHO Classification of Tumors of the Central Nervous System: a summary. Neuro Oncol [Internet]. 2021;23(8):1231–51. Available from: https://dx.doi.org/10.1093/neuonc/noab106
- Jusue-Torres I, Lee J, Germanwala A V., Burns TC, Parney IF. Effect of Extent of Resection on Survival of Patients with Glioblastoma, IDHeWild-Type, WHO Grade 4 (WHO 2021): Systematic Review and Meta-Analysis. World Neurosurg [Internet]. 2023 Mar 1 [cited 2024 Sep 21];171:e524. Available from: /pmc/articles/PMC10030177/
- 6. Van Den Bent MJ, Brandes AA, Taphoorn MJB, Kros JM, Kouwenhoven MCM, Delattre JY, et al. Adjuvant procarbazine, lomustine, and vincristine chemotherapy in newly diagnosed anaplastic oligodendroglioma: Long-term follow-up of EORTC brain tumor group study 26951. Journal of

Clinical Oncology [Internet]. 2013 Jan 20 [cited 2024 Sep 21];31(3):344–50. Available from: https://ascopubs.org/doi/10.1200/JCO.2012.43.2229

- Karschnia P, Young JS, Dono A, Häni L, Sciortino T, Bruno F, et al. Prognostic validation of a new classification system for extent of resection in glioblastoma: A report of the RANO resect group. Neuro Oncol [Internet]. 2023;25(5):940–54. Available from: https://dx.doi.org/10.1093/neuonc/noac193
- Karschnia P, Dietrich J, Bruno F, Dono A, Juenger ST, Teske N, et al. Surgical management and outcome of newly diagnosed glioblastoma without contrast enhancement (low-grade appearance): a report of the RANO resect group. Neuro Oncol [Internet]. 2024;26(1):166–77. Available from: https://dx.doi.org/10.1093/neuonc/noad160
- Molinaro AM, Hervey-Jumper S, Morshed RA, Young J, Han SJ, Chunduru P, et al. Association of Maximal Extent of Resection of Contrast-Enhanced and Non-Contrast-Enhanced Tumor With Survival Within Molecular Subgroups of Patients With Newly Diagnosed Glioblastoma. JAMA Oncol [Internet]. 2020;6(4):495–503. Available from:

https://pubmed.ncbi.nlm.nih.gov/32027343/

 Kreth FW, Thon N, Simon M, Westphal M, Schackert G, Nikkhah G, et al. Gross total but not incomplete resection of glioblastoma prolongs survival in the era of radiochemotherapy. Ann Oncol [Internet]. 2013;24(12):3117–23. Available from:

https://pubmed.ncbi.nlm.nih.gov/24130262/

van der Vaart T, Wijnenga MMJ, van Garderen K, Dubbink HJ, French PJ, Smits M, et al.
 Differences in the Prognostic Role of Age, Extent of Resection, and Tumor Grade between
 Astrocytoma IDHmt and Oligodendroglioma: A Single-Center Cohort Study. Clin Cancer Res. 2024
 Sep;30:3837–44.

- Lasocki A, Roberts-Thomson SJ, Gaillard F. Radiogenomics of adult intracranial gliomas after the 2021 World Health Organisation classification: a review of changes, challenges and opportunities. Quant Imaging Med Surg. 2023 Nov;13:7572–81.
- 13. BNOS Position Statement Guideline for tissue sampling of brain tumours. 2023 Aug.
- Patterson DM, Padhani AR, Collins DJ. Technology insight: water diffusion MRI--a potential new biomarker of response to cancer therapy. Nat Clin Pract Oncol [Internet]. 2008;5(4):220–33.
 Available from: https://pubmed.ncbi.nlm.nih.gov/18301415/
- Thust SC, Heiland S, Falini A, Jäger HR, Waldman AD, Sundgren PC, et al. Glioma imaging in Europe: A survey of 220 centres and recommendations for best clinical practice. Eur Radiol [Internet]. 2018;28(8):3306–17. Available from: https://pubmed.ncbi.nlm.nih.gov/29536240/
- Chen L, Liu M, Bao J, Xia Y, Zhang J, Zhang L, et al. The Correlation between Apparent Diffusion Coefficient and Tumor Cellularity in Patients: A Meta-Analysis. PLoS One [Internet].
 2013;8(11):e79008–e79008. Available from:

https://journals.plos.org/plosone/article?id=10.1371/journal.pone.0079008

- 17. Sadeghi N, Camby I, Goldman S, Gabius HJ, Balériaux D, Salmon I, et al. Effect of hydrophilic components of the extracellular matrix on quantifiable diffusion-weighted imaging of human gliomas: preliminary results of correlating apparent diffusion coefficient values and hyaluronan expression level. AJR Am J Roentgenol [Internet]. 2003 Jul 1 [cited 2024 Sep 21];181(1):235–41. Available from: https://pubmed.ncbi.nlm.nih.gov/12818866/
- Suh CH, Kim HS, Jung SC, Choi CG, Kim SJ. Imaging prediction of isocitrate dehydrogenase (IDH) mutation in patients with glioma: a systemic review and meta-analysis. Eur Radiol [Internet].
 2019;29(2):745–58. Available from: https://pubmed.ncbi.nlm.nih.gov/30003316/

- 19. Xing Z, Yang X, She D, Lin Y, Zhang Y, Cao D. Noninvasive Assessment of IDH Mutational Status in World Health Organization Grade II and III Astrocytomas Using DWI and DSC-PWI Combined with Conventional MR Imaging. AJNR Am J Neuroradiol [Internet]. 2017;38(6):1134–44. Available from: https://pubmed.ncbi.nlm.nih.gov/28450436/
- Lasocki A, Anjari M, Örs Kokurcan S, Thust SC. Conventional MRI features of adult diffuse glioma molecular subtypes: a systematic review. Neuroradiology [Internet]. 2021;63(3):353–62.
 Available from: https://pubmed.ncbi.nlm.nih.gov/32840682/
- 21. Tan WL, Huang WY, Yin B, Xiong J, Wu JS, Geng D. Can Diffusion Tensor Imaging Noninvasively Detect IDH1 Gene Mutations in Astrogliomas? A Retrospective Study of 112 Cases. AJNR Am J Neuroradiol [Internet]. 2014;35(5):920. Available from: /pmc/articles/PMC7964529/
- Page MJ, McKenzie JE, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD, et al. The PRISMA 2020 statement: an updated guideline for reporting systematic reviews. BMJ [Internet]. 2021;372.
 Available from: https://pubmed.ncbi.nlm.nih.gov/33782057/
- Bhatti F, Strobel J, Tench C, Grech-Sollars M, Sollman N, Thust S. Systematic Review Protocol.
 Diffusion-weighted image parameters for the characterisation of untreated gliomas: a systematic review of cohorts. Nottingham Research Data Management Repository. 2024.
- Eckel-Passow JE, Lachance DH, Molinaro AM, Walsh KM, Decker PA, Sicotte H, et al. Glioma
 Groups Based on 1p/19q, IDH , and TERT Promoter Mutations in Tumors. New England Journal of
 Medicine [Internet]. 2015;372(26):2499–508. Available from: https://www.nejm.org/doi/full/10.1056/NEJMoa1407279
- 25. Ouzzani M, Hammady H, Fedorowicz Z, Elmagarmid A. Rayyan-a web and mobile app for systematic reviews. Syst Rev [Internet]. 2016;5(1). Available from: https://www.rayyan.ai/cite/

- Whiting PF, Rutjes AWS, Westwood ME, Mallett S, Deeks JJ, Reitsma JB, et al. QUADAS-2: a revised tool for the quality assessment of diagnostic accuracy studies. Ann Intern Med [Internet].
 2011;155(8):529–36. Available from: https://pubmed.ncbi.nlm.nih.gov/22007046/
- 27. Higgins J, Thomas J, Chandler J, Cumpston M, Li T, Page M, et al. Cochrane Handbook for
 Systematic Reviews of Interventions version 6.5. In: https://training.cochrane.org/handbook.
 2024.
- 28. Cortes C, information MMA in neural, 2004 undefined. Confidence intervals for the area under the ROC curve. proceedings.neurips.ccC Cortes, M MohriAdvances in neural information processing systems, 2004•proceedings.neurips.cc [Internet]. Available from: https://proceedings.neurips.cc/paper/2004/hash/a7789ef88d599b8df86bbee632b2994d-Abstract.html
- 29. Villanueva-Meyer JE, Wood MD, Choi BS, Mabray MC, Butowski NA, Tihan T, et al. MRI Features and IDH Mutational Status in Grade II Diffuse Gliomas: Impact on Diagnosis and Prognosis. AJR Am J Roentgenol [Internet]. 2018;210(3):621. Available from: /pmc/articles/PMC5823758/
- Thust SC, Maynard JA, Benenati M, Wastling SJ, Mancini L, Jaunmuktane Z, et al. Regional and Volumetric Parameters for Diffusion-Weighted WHO Grade II and III Glioma Genotyping: A Method Comparison. AJNR Am J Neuroradiol [Internet]. 2021;42(3):441–7. Available from: https://pubmed.ncbi.nlm.nih.gov/33414227/
- 31. Thust SC, Hassanein S, Bisdas S, Rees JH, Hyare H, Maynard JA, et al. Apparent diffusion coefficient for molecular subtyping of non-gadolinium-enhancing WHO grade II/III glioma: volumetric segmentation versus two-dimensional region of interest analysis. Eur Radiol [Internet]. 2018;28(9):3779–88. Available from: https://pubmed.ncbi.nlm.nih.gov/29572636/

- 32. Xiong J, Tan WL, Pan JW, Wang Y, Yin B, Zhang J, et al. Detecting isocitrate dehydrogenase gene mutations in oligodendroglial tumors using diffusion tensor imaging metrics and their correlations with proliferation and microvascular density. J Magn Reson Imaging [Internet]. 2016;43(1):45–54. Available from: https://pubmed.ncbi.nlm.nih.gov/26016619/
- 33. Maynard J, Okuchi S, Wastling S, Al Busaidi A, Almossawi O, Mbatha W, et al. World Health Organization Grade II/III Glioma Molecular Status: Prediction by MRI Morphologic Features and Apparent Diffusion Coefficient. Radiology [Internet]. 2020;296(1):111–21. Available from: https://pubmed.ncbi.nlm.nih.gov/32315266/
- 34. Wasserman JK, Nicholas G, Yaworski R, Wasserman AM, Woulfe JM, Jansen GH, et al. Radiological and pathological features associated with IDH1-R132H mutation status and early mortality in newly diagnosed anaplastic astrocytic tumours. PLoS One [Internet]. 2015;10(4). Available from: https://pubmed.ncbi.nlm.nih.gov/25849605/
- 35. Su CQ, Lu SS, Zhou MD, Shen H, Shi HB, Hong XN. Combined texture analysis of diffusionweighted imaging with conventional MRI for non-invasive assessment of IDH1 mutation in anaplastic gliomas. Clin Radiol [Internet]. 2019;74(2):154–60. Available from: https://pubmed.ncbi.nlm.nih.gov/30391048/
- 36. Du N, Zhou X, Mao R, Shu W, Xiao L, Ye Y, et al. Preoperative and Noninvasive Prediction of
 Gliomas Histopathological Grades and IDH Molecular Types Using Multiple MRI Characteristics.
 Front Oncol [Internet]. 2022;12:1. Available from: /pmc/articles/PMC9196247/
- Gihr G, Horvath-Rizea D, Kohlhof-Meinecke P, Ganslandt O, Henkes H, Härtig W, et al. Diffusion
 Weighted Imaging in Gliomas: A Histogram-Based Approach for Tumor Characterization. Cancers
 (Basel) [Internet]. 2022;14(14):3393. Available from: /pmc/articles/PMC9321540/

- Springer E, Cardoso PL, Strasser B, Bogner W, Preusser M, Widhalm G, et al. MR Fingerprinting-A Radiogenomic Marker for Diffuse Gliomas. Cancers (Basel) [Internet]. 2022;14(3). Available from: https://pubmed.ncbi.nlm.nih.gov/35158990/
- Liu S, Zhang Y, Kong Z, Jiang C, Wang Y, Zhao D, et al. Feasibility of evaluating the histologic and genetic subtypes of WHO grade II-IV gliomas by diffusion-weighted imaging. BMC Neurosci [Internet]. 2022;23(1). Available from: /pmc/articles/PMC9720933/
- 40. Kamble AN, Agrawal NK, Koundal S, Bhargava S, Kamble AN, Joyner DA, et al. Imaging-based stratification of adult gliomas prognosticates survival and correlates with the 2021 WHO classification. Neuroradiology [Internet]. 2023;65(1):41–54. Available from: https://pubmed.ncbi.nlm.nih.gov/35876874/
- Xie Y, Li S, Shen N, Gan T, Zhang S, Liu WV, et al. Assessment of Isocitrate Dehydrogenase 1
 Genotype and Cell Proliferation in Gliomas Using Multiple Diffusion Magnetic Resonance
 Imaging. Front Neurosci [Internet]. 2021;15:783361. Available from: /pmc/articles/PMC8645648/
- 42. Cindil E, Sendur HN, Cerit MN, Erdogan N, Celebi F, Dag N, et al. Prediction of IDH Mutation
 Status in High-grade Gliomas Using DWI and High T1-weight DSC-MRI. Acad Radiol [Internet].
 2022;29 Suppl 3:S52–62. Available from: https://pubmed.ncbi.nlm.nih.gov/33685792/
- 43. Lee S, Choi SH, Ryoo I, Yoon TJ, Kim TM, Lee SH, et al. Evaluation of the microenvironmental heterogeneity in high-grade gliomas with IDH1/2 gene mutation using histogram analysis of diffusion-weighted imaging and dynamic-susceptibility contrast perfusion imaging. J Neurooncol [Internet]. 2015;121(1):141–50. Available from: https://pubmed.ncbi.nlm.nih.gov/25205290/
- 44. Halefoglu AM, Camurcuoglu E, Tanik C, Kizilkaya O, Yilmaz A. Predictive role of magnetic resonance imaging in the distinction of isocitrate dehydrogenase (IDH) mutant grade 4

astrocytomas versus glioblastomas. Acta radiol [Internet]. 2023;64(6):2074–86. Available from: https://journals.sagepub.com/doi/10.1177/02841851231165282?url_ver=Z39.88-2003&rfr_id=ori%3Arid%3Acrossref.org&rfr_dat=cr_pub++0pubmed

- 45. Uetani H, Azuma M, Khant ZA, Watanabe Y, Kudo K, Kadota Y, et al. Importance of Age and Noncontrast-Enhancing Tumor as Biomarkers for Isocitrate Dehydrogenase–Mutant Glioblastoma: A Multicenter Study. J Comput Assist Tomogr [Internet]. 2023;47(4):659. Available from: /pmc/articles/PMC10348614/
- Xing Z, Zhang H, She D, Lin Y, Zhou X, Zeng Z, et al. IDH genotypes differentiation in glioblastomas using DWI and DSC-PWI in the enhancing and peri-enhancing region. https://doi.org/101177/0284185119842288 [Internet]. 2019;60(12):1663–72. Available from: https://journals.sagepub.com/doi/10.1177/0284185119842288
- 47. Xiong J, Tan W, Wen J, Pan J, Wang Y, Zhang J, et al. Combination of diffusion tensor imaging and conventional MRI correlates with isocitrate dehydrogenase 1/2 mutations but not 1p/19q genotyping in oligodendroglial tumours. Eur Radiol [Internet]. 2016;26(6):1705–15. Available from: https://pubmed.ncbi.nlm.nih.gov/26396108/
- Aliotta E, Nourzadeh H, Batchala PP, Schiff D, Lopes MB, Druzgal JT, et al. Molecular Subtype
 Classification in Lower-Grade Glioma with Accelerated DTI. AJNR Am J Neuroradiol [Internet].
 2019;40(9):1458–63. Available from: https://pubmed.ncbi.nlm.nih.gov/31413006/
- Aliotta E, Dutta SW, Feng X, Tustison NJ, Batchala PP, Schiff D, et al. Automated apparent diffusion coefficient analysis for genotype prediction in lower grade glioma: association with the T2-FLAIR mismatch sign. J Neurooncol [Internet]. 2020;149(2):325–35. Available from: https://pubmed.ncbi.nlm.nih.gov/32909115/

- 50. Lee MK, Park JE, Jo Y, Park SY, Kim SJ, Kim HS. Advanced imaging parameters improve the prediction of diffuse lower-grade gliomas subtype, IDH mutant with no 1p19q codeletion: added value to the T2/FLAIR mismatch sign. Eur Radiol [Internet]. 2020;30(2):844–54. Available from: https://pubmed.ncbi.nlm.nih.gov/31446467/
- Liu D, Gao SX, Liao HF, Xu JM, Wen M. A Comparative Study of 2 Different Segmentation
 Methods of ADC Histogram for Differentiation Genetic Subtypes in Lower-Grade Diffuse Gliomas.
 Biomed Res Int [Internet]. 2020;2020. Available from: https://pubmed.ncbi.nlm.nih.gov/33062706/
- 52. Hong EK, Choi SH, Shin DJ, Jo SW, Yoo RE, Kang KM, et al. Comparison of Genetic Profiles and Prognosis of High-Grade Gliomas Using Quantitative and Qualitative MRI Features: A Focus on G3 Gliomas. Korean J Radiol [Internet]. 2021;22(2):233–42. Available from: https://pubmed.ncbi.nlm.nih.gov/32932560/
- 53. Su X, Yang X, Sun H, Liu Y, Chen N, Li S, et al. Evaluation of Key Molecular Markers in Adult Diffuse Gliomas Based on a Novel Combination of Diffusion and Perfusion MRI and MR Spectroscopy. J Magn Reson Imaging [Internet]. 2024;59(2):628–38. Available from: https://pubmed.ncbi.nlm.nih.gov/37246748/
- 54. Nuessle NC, Behling F, Tabatabai G, Vega SC, Schittenhelm J, Ernemann U, et al. ADC-Based Stratification of Molecular Glioma Subtypes Using High b-Value Diffusion-Weighted Imaging. J Clin Med [Internet]. 2021;10(16). Available from: https://pubmed.ncbi.nlm.nih.gov/34441747/
- 55. Ma X, Cheng K, Cheng G, Li C, Lyu J, Lan Y, et al. Apparent Diffusion Coefficient as Imaging Biomarker for Identifying IDH Mutation, 1p19q Codeletion, and MGMT Promoter Methylation Status in Patients With Glioma. J Magn Reson Imaging [Internet]. 2023;58(3):732–8. Available from: https://pubmed.ncbi.nlm.nih.gov/36594577/

- 56. Cheng Y, Song S, Wei Y, Xu G, An Y, Ma J, et al. Glioma Imaging by O-(2-18F-Fluoroethyl)-L-Tyrosine PET and Diffusion-Weighted MRI and Correlation With Molecular Phenotypes, Validated by PET/MR-Guided Biopsies. Front Oncol [Internet]. 2021;11. Available from: https://pubmed.ncbi.nlm.nih.gov/34912706/
- 57. Cho NS, Sanvito F, Le VL, Oshima S, Teraishi A, Yao J, et al. Diffusion MRI is superior to quantitative T2-FLAIR mismatch in predicting molecular subtypes of human non-enhancing gliomas. Neuroradiology. 2024 Dec;
- 58. Zhang HW, Zhang HB, Liu XL, Deng HZ, Zhang YZ, Tang XM, et al. Clinical Assessment of Magnetic Resonance Spectroscopy and Diffusion-Weighted Imaging in Diffuse Glioma: Insights Into Histological Grading and IDH Classification. Can Assoc Radiol J. 2024 Nov;75:868–77.
- 59. Yang X, Lin Y, Xing Z, She D, Su Y, Cao D. Predicting 1p/19q codeletion status using diffusion-, susceptibility-, perfusion-weighted, and conventional MRI in IDH-mutant lower-grade gliomas. Acta radiol [Internet]. 2021;62(12):1657–65. Available from: https://journals.sagepub.com/doi/10.1177/0284185120973624?url_ver=Z39.88-2003&rfr_id=ori%3Arid%3Acrossref.org&rfr_dat=cr_pub++0pubmed
- 60. Cui Y, Ma L, Chen X, Zhang Z, Jiang H, Lin S. Lower apparent diffusion coefficients indicate distinct prognosis in low-grade and high-grade glioma. J Neurooncol [Internet]. 2014;119(2):377–85.
 Available from: https://link.springer.com/article/10.1007/s11060-014-1490-6
- Latysheva A, Emblem KE, Brandal P, Vik-Mo EO, Pahnke J, Røysland K, et al. Dynamic susceptibility contrast and diffusion MR imaging identify oligodendroglioma as defined by the 2016 WHO classification for brain tumors: histogram analysis approach. Neuroradiology [Internet]. 2019;61(5):545–55. Available from: https://pubmed.ncbi.nlm.nih.gov/30712139/

- Wijnenga MMJ, French PJ, Dubbink HJ, DInjens WNM, Atmodimedjo PN, Kros JM, et al. The impact of surgery in molecularly defined low-grade glioma: an integrated clinical, radiological, and molecular analysis. Neuro Oncol [Internet]. 2018 Jan 10 [cited 2024 Sep 21];20(1):103–12. Available from: https://dx.doi.org/10.1093/neuonc/nox176
- 63. Metellus P, Coulibaly B, Colin C, De Paula AM, Vasiljevic A, Taieb D, et al. Absence of IDH mutation identifies a novel radiologic and molecular subtype of WHO grade II gliomas with dismal prognosis. Acta Neuropathol [Internet]. 2010 Nov 16 [cited 2024 Sep 21];120(6):719–29. Available from: https://link.springer.com/article/10.1007/s00401-010-0777-8
- 64. Thust SC, Maynard JA, Benenati M, Wastling SJ, Mancini L, Jaunmuktane Z, et al. Regional and Volumetric Parameters for Diffusion-Weighted WHO Grade II and III Glioma Genotyping: A Method Comparison. AJNR Am J Neuroradiol [Internet]. 2021 Mar 1 [cited 2024 Apr 5];42(3):441–7. Available from: https://pubmed.ncbi.nlm.nih.gov/33414227/
- 65. Thust SC, Hassanein S, Bisdas S, Rees JH, Hyare H, Maynard JA, et al. Apparent diffusion coefficient for molecular subtyping of non-gadolinium-enhancing WHO grade II/III glioma: volumetric segmentation versus two-dimensional region of interest analysis. Eur Radiol [Internet]. 2018 Sep 1 [cited 2024 Apr 9];28(9):3779–88. Available from: https://pubmed.ncbi.nlm.nih.gov/29572636/
- 66. Law M, Oh S, Johnson G, Babb JS, Zagzag D, Golfinos J, et al. Perfusion magnetic resonance imaging predicts patient outcome as an adjunct to histopathology: A second reference standard in the surgical and nonsurgical treatment of low-grade gliomas. Neurosurgery. 2006 Jun;58:1099–107.
- 67. Lewis MA, Ganeshan B, Barnes A, Bisdas S, Jaunmuktane Z, Brandner S, et al. Filtration-histogram based magnetic resonance texture analysis (MRTA) for glioma IDH and 1p19q genotyping. Eur J

Radiol [Internet]. 2019 Apr 1 [cited 2024 Sep 21];113:116–23. Available from:

https://pubmed.ncbi.nlm.nih.gov/30927935/

- 68. Cho NS, Hagiwara A, Sanvito F, Ellingson BM. A multi-reader comparison of normal-appearing white matter normalization techniques for perfusion and diffusion MRI in brain tumors. Neuroradiology [Internet]. 2023 Mar 1 [cited 2024 Sep 21];65(3):559. Available from: /pmc/articles/PMC9905164/
- 69. Yee PP, Wang J, Chih SY, Aregawi DG, Glantz MJ, Zacharia BE, et al. Temporal radiographic and histological study of necrosis development in a mouse glioblastoma model. Front Oncol. 2022 Oct 14;12:993649.
- Lasocki A, Buckland ME, Drummond KJ, Wei H, Xie J, Christie M, et al. Conventional MRI features can predict the molecular subtype of adult grade 2–3 intracranial diffuse gliomas.
 Neuroradiology. 2022 Dec;64:2295–305.
- 71. Wu J, Thust S. Article in press. American Journal of Neuroradiology. 2025;
- 72. Hyare H, Rice L, Thust S, Nachev P, Jha A, Milic M, et al. Modelling MR and clinical features in grade II/III astrocytomas to predict IDH mutation status. Eur J Radiol. 2019 May;114:120–7.

Figure legends:

Figure 1: PRISMA flow diagram

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Figure 2: Forest plots showing pooled estimate of AUC values from studies using each ADC parameter as a predictor of IDH mutation status

A = ADCmean (ROI methods), B= ADCmean (VOI methods), C = rADCmean (ROI methods), D = ADCmin (ROI methods), E = ADCmin (VOI methods), F = rADCmin (ROI methods). Asterix (*) denotes studies where standard error was not provided and therefore estimated using a formula. Circumflex (^) denotes studies where 3 or more ROI measurements were averaged to determine ADC_{min} values.

Tables

Table 1: Summary of studies assessing IDH mutation only.

Summary of study author, year of publication, main research purpose, composition of study cohort (WHO grade, IDH status, 1p19q status, sex), methods of ADC measurements, and key results for studies assessing IDH mutation only. Studies are listed from low to higher WHO grade(s).

Abbreviations: ADC: Apparent Diffusion Coefficient, AUC: Area under the curve, cMRI: Conventional MRI, CNAWM: Contralateral normal appearing white matter, CNS: Central nervous system, Codel: Codeletion, DG: Diffuse gliomas, DTI: Diffusion tensor imaging, DWI: Diffusion weighted imaging, F: Female, GBM: Glioblastoma multiforme, HGGs: High grade gliomas, IDH: Isocitrate dehydrogenase, IQR: Interquartile range, LGGs: Low grade gliomas, M: Male MRI: Magnetic Resonance Imaging, Mut: Mutant, NAWM: Normal appearing white matter, NS: Not stated, ROI: Region of interest, Sens: Sensitivity, Spec: Specificity, T1w: T1-weighted imaging. T2w: T2-weighted imaging, VOI: Volume of interest, Wt: Wild-type.

Table 2: Summary of studies assessing IDH mutation and 1p19q codeletion

Summary of study author, year of publication, main research purpose, composition of study cohort (WHO grade, IDH status, 1p19q status, sex), methods of ADC measurements, and key results for studies assessing both IDH mutation and 1p19 codeletion.

Abbreviations: ADC: Apparent Diffusion Coefficient, AUC: Area under the curve, cMRI: Conventional MRI, CNAWM: Contralateral normal appearing white matter, CNS: Central nervous system, Codel: Codeletion, DG: Diffuse gliomas, DTI: Diffusion tensor imaging, DWI: Diffusion weighted imaging, F: Female, GBM: Glioblastoma multiforme, HGGs: High grade gliomas, IDH: Isocitrate dehydrogenase, IQR: Interquartile range, LGGs: Low grade gliomas, M: Male, MRI: Magnetic Resonance Imaging, Mut: Mutant, NAWM: Normal appearing white matter, NS: Not stated, ROI: Region of interest, Sens: Sensitivity, Spec: Specificity

T1w: T1 weighted imaging. T2w: T2-weighted imaging, VOI: Volume of interest, Wt: Wild-type.

Table 3: Summary of studies assessing 1p19q codeletion alone

Summary of study author, year of publication, main research purpose, composition of study cohort (WHO grade, IDH status, 1p19q status, sex), methods of ADC measurements, and key results for studies assessing 1p19 codeletion only.

Abbreviations: ADC: Apparent Diffusion Coefficient, AUC: Area under the curve, Codel: Codeletion, DG: Diffuse gliomas, DTI: Diffusion tensor imaging, DWI: Diffusion weighted imaging, F: Female, IDH: Isocitrate dehydrogenase, M: Male, MRI: Magnetic Resonance Imaging, Mut: Mutant, ROI: Region of interest, Sens: Sensitivity, Spec: Specificity, VOI: Volume of interest, Wt: Wild-type.

Table 4: Proposed ADC thresholds for IDH genotyping according to studies included in the meta-analyses.

N: number, SD=standard deviation, Min=minimum, Max=maximum. ADC values in units of *10⁻³ mm²/s. rADC values have no unit.

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Table 1: Summary of studies assessing IDH mutation only.

Summary of study author, year of publication, main research purpose, composition of study cohort (WHO grade, IDH status, 1p19q status, sex), methods of ADC measurements, and key results for studies assessing IDH mutation only. Studies are listed from low to higher WHO grade(s).

Abbreviations: ADC: Apparent Diffusion Coefficient, AUC: Area under the curve, cMRI: Conventional MRI, CNAWM: Contralateral normal appearing white matter, CNS: Central nervous system, Codel: Codeletion, DG: Diffuse gliomas, DTI: Diffusion tensor imaging, DWI: Diffusion weighted imaging, F: Female, GBM: Glioblastoma multiforme, HGGs: High grade gliomas, IDH: Isocitrate dehydrogenase, IQR: Interquartile range, LGGs: Low grade gliomas, M: Male MRI: Magnetic Resonance Imaging, Mut: Mutant, NAWM: Normal appearing white matter, NS: Not stated, ROI: Region of interest, Sens: Sensitivity, Spec: Specificity

T1w: T1-weighted imaging. T2w: T2-weighted imaging, VOI: Volume of interest, Wt: Wild-type.

Study	Research	WHO	Cohort size and	Δσe	ROL or VOL	Results
- Stady						incounts and incou
	purpose	Grades	composition	(years),	methods	
				Sex		
Villanueva-	To identify MRI	2	100 WHO 2.	IDH-wt	VOI	- ADCmin: AUC 0.905 (0.830–
Meyer JE et	markers		(IDH-wt = 22, IDH-	Median 58.	Tumour	0.954), Cut off <0.9x10 ⁻³ , Sens
al 2018	predictive of IDH		mut = 65)	IDH-mut	delineated on	91% Spec 76% p<0.001
	predictive of ibit		mat 037	1011 mat	defineated off	51%, 5pee 7 %, p (0.001.
	mutational			Median 41.	all axial slices	
	status in grade II			Sex NS	to calculate	
	status in grude in			Sex 115		
	diffuse gliomas				min, mean and	
	(DGs) and				max ADC of	
	(_ 30) and					
	evaluate the				tumour.	
	complementary				Necrosis cysts	
	complementaly				14CC1 0313, Cy313,	

	roles of MRI				haemorrhage,	
	features and IDH				vessels avoided	
	mutational					
	status to better					
	predict					
	outcomes for					X
	these patients.					
Thust SC et	To compare	23	283	Median 30	BOI and VOI	- VOLADCmin: Cut off 0.81 x10
-1 2021	volumetrie and	2,5	()			3000 ² /a Same C0 40(Same C0 20)
al. 2021	volumetric and		(WHO 2 and 3)	IQK 33-53.	Regional ADC	3mm²/s, Sens 68.4%, Spec 60.3%,
	regional ADC		(IDH-wt = 79, IDH-	164M, 119F	measurements:	AUC 0.68 (0.61-0.75).
	measurement		mut 1p19q-retained		3, small, 30-	 VOI rADCmin (5th percentile): Cut
	techniques for		= 104, IDH-mut		40mm ² ROIs	off 1.08, Sens 68.4%, Spec 61.3%,
	glioma		1p19q-codel = 100)		placed in	AUC 0.72 (0.66-0.79).
	genotyping with				visually	- VOI ADCmean: Cut off 1.19 x10 ⁻
	a focus on IDH				perceived	³ mm ² /s, Sens 77.2%, Spec 64.2%,
	status				lowest ADC	AUC 0.78 (0.72-0.84).
	prediction.				portions of	- VOI rADCmean: Cut off 1.60,
					glioma,	Sens 86.8%, Spec 60.8%, AUC
			25		remaining in	0.82 (0.76-0.88).
					solid	- ROI ADCmin: Cut off 1.07 x10 ⁻
					component.	³ mm ² /s, Sens 82.3%, Spec 61.3%,
					ADCmean: 1	AUC 0.79 (0.73-0.85).
					large ROI	- ROI rADCmin: Cut off 1.40, Sens
					placed on	85.5%, Spec 62.3%, AUC 0.81
					largest axial	(0.76-0.86).
					tumour cross	- ROI ADCmean: Cut off 1.34 x10 ⁻
					section.	³ mm ² /s, Sens 84.8%, Spec 60.3%,
					Calcium, cysts,	AUC 0.81 (0.75-0.86).
					haemorrhage,	- ROI rADCmean: Cut off 1.75,
					vessels avoided	Sens 86.8%, Spec 62.3%. AUC
					from ROIs	0 83 (0 77-0 88)
						0.03 (0.77 0.00).
						P-0.001 101 all.

					tumour	
					segmentation	
					incorporating	
					entire T2-	
					weighted signal	
					abnormality.	
						, iP
Thust SC et	To investigate if	2,3	44	IDH-wt	ROI and VOI	- ADCmean (VOI): Cut off
al. 2018	quantitative ADC		(WHO 2 = 26, WHO	Mean 53+/-	ROI placed on	1201(x10 ⁻⁶ mm²/s), sens 0.83,
	measurements		3 = 18)	14.	largest tumour	spec 0.86, AUC 0.85.
	can predict		(IDH-wt = 14, IDH-	IDH-mut	cross section,	- rADCmean (VOI): Cut off 1.65,
	genetic subtypes		mut 1p19q-retained	1p19-	sparing the	sens 0.80, spec 0.92, AUC 0.86.
	of non-		= 16, IDH-mut	retained	tumour	- ADCmean (ROI) 1st observer: Cut
	gadolinium-		1p19q-codel = 14)	Mean	margin.	off 1.83, sens 0.86, spec 1.00,
	enhancing			33.9+/-8.6.	VOI: whole	AUC 0.93.
	gliomas,			IDH-mut	tumour	- ADCmean (ROI) 2nd observer:
	comparing whole			1p19q-	segmentation	Cut off 1.76, sens 0.86, spec 0.91,
	tumour against	X	0	codel Mean	incorporating	AUC 0.88.
	single slice			38.9+/-8.3.	entire T2-	
	analysis.	V		22M, 22F	weighted signal	
	C				abnormality.	
Xiong J et	To assess	2,3	90	WHO II	ROI	- ADCmin: Cut off 0.81, Sens
al. 2016	whether DTI		(WHO 2 = 54, WHO	Mean 40+/-	4-6 ROIs placed	78.7%, Spec 79.2%, AUC 0.77.
	metrics could aid		3 = 36).	10.	in solid	- rADCmin: Cut off 1.19, Sens
	the noninvasive		(IDH-mut = 67, IDH-	WHO III	tumour. The	80.9%, Spec 76.9%, AUC 0.80.
	detection of IDH		wt = 23)	Mean 46+/-	lowest ADC	
	mutations and		(Oligodendroglioma	11	from the ROIs	
	their correlations		= 29, Anaplastic	42M, 38F	drawn by two	
	with tumor		oligodendroglioma		observers was	
	proliferation and		= 24,		averaged and	

	microvascular		Oligoastrocytoma =		used as the as	
	density (MVD) in		25)		the minimum	
	oligodendroglial				ADC value.	
	tumours.				ROIs also	
					placed in	
					peritumoural	X
					region to	
					calculate	
					peritumoural	
					ADC.	
					Calcification,	
					cysts,	
					haemorrhage,	
				0	necrosis	
					avoided.	
Maynard J	To evaluate	2,3	339 (Study sample =	Study	ROI	- rADCmean: AUC 0.83.
et al. 2020	clinically		290, test sample =	sample:	1) 3 ROIs (30-	- ADCmin: AUC 0.78.
	available MRI		49)	Median 10,	40mm ²) placed	- rADCmin: AUC 0.8.
	parameters for	X	(WHO 2 and 3)	IQR 33-52,	in visually	- ADCmean: AUC 0.81.
	predicting IDH		(Study sample: IDH-	Range 17-	perceived	
	status in patients	X	wt = 82, IDH-mut	77.	lowest ADC	
	with glioma.		1p19q-retained =	169M,121F.	portions of	
			107, IDH-mut	Test sample	each tumour;	
	\mathbf{O}		1p19q-codel = 101)	age and sex	lowest ROI	
			(Test sample: IDH-	NS.	mean ADC	
			wt = 9, IDH-mut		measurement	
			1p19q-retained =		designated as	
			21, IDH-mut 1p19q-		ADCmin.	
			codel = 19)		2) One large	
					ROI placed to	
					cover the	
					largest axial	

					tumour cross	
					section; used	
					as ADCmean.	
					Tumour	
					margins.	
					necrosis	
					haomorrhago	
					colsification	
					avoided.	
Wasserman	To determine	3	37 WHO 3.	Mean 68,	ROI	- ADCmin: Cut off 0.950 x10 ⁻
JK et al.	whether		(Anaplastic	Range 20-	Small ROI	³ mm ² /s, Sens 76.9%, Spec 65.2%,
2015	pathological		astrocytoma = 28:	81.	(25mm²)	AUC 0.711 (0.534-0.887),
	and/or		IDH R132H-mut =	16M, 21F	placed in	p=0.033.
	radiological		12, IDH-wt = 16.		region of	
	variables exist		Anaplastic		lowest	
	that can reliably		oligoastrocytoma =		apparent ADC,	
	distinguish IDH1-		9: IDH R132H-mut		by visual	
	R132H-positive		=6, IDH-wt =3)		inspection, to	
	from IDH1-		75		determine	
	R132H-negative				ADCmin values.	
	tumours and to					
	identify variables					
	associated with					
	early mortality.					
Su CQ et al.	To examine	3	52 WHO 3.	Mean =	VOI	- ADCentropy, AUC 0.724 (0.572-
2019	whether texture		(IDH-mut n=21: 11	47.8+/-12,	Tumour	0.845), Cut off >5.763, Sens
	analysis of DWI		anaplastic	Range 18-	manually	71.4%, Spec 76%.
	combined with		astrocytoma, 10	72	outlined on	
	conventional		anaplastic	25M, 21F	contrast-	
	MRI could non-		oligodendroglioma.		enhanced	
	invasively predict		IDH-wt n=25 (13		T1WI as areas	
	IDH1 mutational		anaplastic		of abnormal	
	1					

	status in		astrocytoma, 12		enhancement	
	anaplastic		anaplastic		and non-	
	gliomas.		oligodendroglioma)		enhancing	
					tumour.	
					Vessels,	
					necrosis,	×
					oedema	
					avoided.	
Du N et al.	To explore the	1,2,3,4	166	Mean	ROI	- ADCmin: AUC 0.653 (0.561-
2022	correlation		(WHO 1= 12, WHO	51.1+/-	1) ADCmin: 3	0.745), Cut off 0.98, Sens 45.83,
	between MRI		2 = 31, WHO 3 = 18,	15.9, Range	different 20-	Spec 83.04.
	morphological		WHO 4 = 105)	14-85.	30mm ² ROIs	- ADCmean: AUC 0.643 (0.555-
	characteristics,		(IDH-wt = 112, IDH-	92M, 74F	placed on	0.731), Cut off 1.05, Sens 75.00,
	ADC parameters		mut = 48)	\mathbf{O}	visually	Spec 58.04.
	and pathological		*No IDH status for 6	NO.	determined	- rADCmin: AUC 0.656 (0.566-
	grade and IDH		patients.		lowest ADC;	0.746), Cut off 1.14, Sens 62.50,
	gene phenotypes				mean taken as	Spec 66.96.
	of gliomas.				ADCmin.	- rADCmean: AUC 0.652 (0.562-
			25		2) ADCmean:	0.742), Cut off 1.40, Sens 70.83,
		\sim			ROI plotted as	Spec 59.82.
					large as	
					possible on	
					largest	
					transverse	
					cross-section	
					of tumour.	
					Cysts,	
					calcification,	
					necrosis,	
					vessels	
					avoided.	

Gihr G et	To investigate (I)	1,2,3,4	82	WHO I+II	VOI	- ADC Entropy, AUC 0.8040
al. 2022	the potential of		(WHO 1 = 7, WHO 2	Mean 34.	Tumour	(0.6849–0.9231), p<0.0001. Cut
	ADC histogram		= 19, WHO 3 = 11,	WHO III+IV	volumes	off <5.488, Sens 0.73. Spec 0.97.
	analysis for		WHO 4 = 45)	Mean 62.	manually	- ADCmax, AUC 0.7314 (0.6054–
	distinguishing		(IDH-wt = 58, IDH-	34M, 48F	drawn in T1W	0.8573), p=0.0026.
	LGGs and HGGs		mut = 19)		or T2W images	- Skewness, AUC 0.7486 (0.6235-
	and (II) whether		*No IDH status for 5		along the	0.8737), p=0.0012.
	those		patients.		border of	
	parameters are				visible signal	
	associated with				alteration	
	Ki-67				(contrast	
	immunolabelling,				enhancing	
	the IDH1				region or T2W	
	mutation profile				hyperintense	
	and the MGMT			0	region) in	
	promoter				every slice of	
	methylation				detectable	
	profile.		$\mathbf{\cap}$		tumour.	
			2		Volume used	
		X	0		for histogram	
					analysis.	
Springer E		224	24	Moon 58 6	POL	- ADCmoon: AUC 0.875, p<0.001
springer E		2,3,4		Dange 22	ROIs marked	- ADCINEAII. AUC 0.875, p<0.001.
et al. 2022	Fingerprinting-			Range 23-	ROIS marked	
	derived 11 and		3 = 5, WHO 4 = 9)	//.	on 1) solid part	
	12 relaxation		(WHO2Diffuse	15M, 9F	of tumour with	
	maps to		Astrocytoma = 7:		and without	
	differentiate		IDH-mut = 6, IDH-wt		contrast	
	diffuse gliomas		= 1. WHO		enhancement	
	according to IDH		2Oligodendroglioma		(mean number	
	mutation.		IDH-mut 1p19q-		of ROIs per	
			codel = 3. WHO III		case = 2.7), 2)	
			Anaplastic		perilesional	

			astrocytoma = 4:		NAWM (less	
			IDH-mut = 3, IDH-wt		than or equal	
			= 1. WHO III		to 1cm from	
			anaplastic		tumour or	
			oligodendroglioma		peritumoral	
			IDH-mut 1p19q-		oedema), 3)	*
			codel = 1. WHO IV		perilesional	
			GBM n = 9: IDH-mut		NAWM less	
			= 1, IDH-wt = 8.)		than or equal	
					to 1 cm distant	
					from the tumor	
					or from	
					peritumoral	
					oedema, and	
				\mathbf{C}	4) contralateral	
					frontal lobe	
					NAWM.	
					Necrosis and	
			2		haemorrhage	
			0		avoided.	
	Touristic			Maria		
Liu S et al.		2,3,4		Mean	KUI	ADCmean: AUC 0.777
2022	feasibility of DWI		(WHO 2 = 36, WHO	44.3+/-	4 ROIs	(0.688,0.865), cut off 0.0012,
	metrics to		3 = 32, WHO 4 = 43)	12.1.	manually	sens 88.4, spec 67.7.
	predict the		(IDH-wt = 65, IDH-	58M, 53F	placed within	- rADCmean: AUC 0.836
	histologic		mut = 45)		solid	(0.757,0.914), cut off 1.60, sens
	subtypes and		*No IDH status for 1		components of	82.2, spec 80.0.
	genetic status of		patient.		tumours co-	
	gliomas				registered on	
	noninvasively.				T2WI.	
					Cysts, necrosis,	
					haemorrhage,	

					calcification	
					avoided.	
Kamble AN	Hypothesise that	2.3.4	475 (Training set =	Training	ROI	- ADCmean Training dataset: Cut
et al 2023	glioma can be		275 Validation set =	set.	ROI placed as	off 1 12 sens 82 1 spec 74 2
2023	stratified into 2		200)	Turne l	homogonously	
	stratified into 3				nomogenously	AUC 0.841, p <.0.0001.
	types using a		(WHO 2, 3, and 4)	Mean 47,	as possible to	- ADCmean Validation dataset: Cut
	flow chart of 4		(Training set: IDH-	Type II	calculate	off 1.20, sens 72.9, spec 64.9,
	yes/no		wt = 124, IDH-mut	Mean 45,	average	AUC 0.748, p <.0.0001.
	questions, which		1p19q-retained =	Type III	tumour ADC	J
	correlate with		54, IDH-mut 1p19q-	Mean 55.	after excluding	
	the 3 glioma		codel = 21)	152M,	tumour	
	types in the 2021		(Validation set: IDH-	122F.	necrosis.	
	WHO		wt = 106, IDH-mut	Validation	Necrosis	
	classification.		1p19q-retained =	set:	excluded.	
	Propose that		48, IDH-mut 1p19q-	Туре І		
	radiological		codel = 46)	Mean 45,		
	stratification		\mathbf{O}	Type II		
	would have		2	Mean 38,		
	prognostic			Type III		
	implications if			Mean 56.		
	correlated with	K		107M, 93F.		
	the WHO					
	classification.					
Xie Y et al.	To compare the	2,3,4	91	IDH-mut	ROI	WHO II and III tumours:
2021	efficacy of		(WHO 2 = 27, WHO	Median 53,	3-6 ROIs	- ADCmin: AUC 0.751, Sens 59.38,
	parameters from		3 = 20, WHO 4 = 43)	IQR 46.5-	manually	Spec 93.33, Cut off 1.084.
	multiple		(IDH-wt = 49, IDH-	58.	placed in solid	WHO IV tumours:
	diffusion		mut = 41)	IDH-wt	part of tumour	
	magnetic			Median 41,	parenchyma	- No significant difference found
	resonance			IQR 34.75-	(defined as	between diffusion imaging
	imaging for				contrast	parameters.



	histological				vessels	
	grading and				avoided.	
	isocitrate					
	dehydrogenase					
	(IDH)					
	classification in					*
	adult diffuse					
	gliomas.					² ²
Cindil E et	Evaluate the	3,4	56	IDH-mut	ROI	- ADCmin: AUC 0.686 (0.795-
al. 2022	diagnostic		(WHO 3 and 4)	Mean =	1-3 ROIs	0.950), Cut off 0.954, Sens 0.74,
	performance of		(GBM IDH-wt = 25,	49+/-17.	manually	Spec 0.66, PPV 0.77, NPV 0.58,
	DWI MRI		GBM IDH-mut = 10,	IDH-wt	placed on	Accuracy 0.68.
	parameters in		Anaplastic	Mean =	darkest areas	
	the non-invasive		astrocytoma IDH-wt	58+/-14.	on the tumour	
	prediction of IDH		= 10, Anaplastic	31M, 27F	core that	
	mutation status		astrocytoma IDH-		corresponded	
	in HGGs.		mut = 13))		to enhancing	
					tumour.	
			2		Lowest ROI	
					ADC value	
					used.	
	0				Calcium, cysts,	
					haemorrhage,	
					vessels	
					avoided.	
Lee S et al.	To explore the	3,4	52	Mean	VOI	- ADCmean, AUC 0.707 (0.564–
2015	difference		(WHO 3 = 15, WHO	49.81+/-	Tumour	0.825), Sens 50, Spec 91.7, cut
	between		4 = 37).	14.5, Range	borders	off>1333.42 (x10 ⁻⁶ mm²/s),
	isocitrate		(WHO III Anaplastic	22-72.	manually	p=0.0178.
	dehydrogenase		astrocytoma = 15:	32M, 20F	drawn in each	- ADC 10%, AUC 0.707 (0.564–
	(IDH)-1/2 gene		IDH-mut = 9, IDH-wt		section of co-	0.825), Sens 50, Spec 97.2, cut
	mutation-		= 6. WHO IV GBM =		registered	off >797 (x10 ⁻⁶ mm²/s), p=0.0250.

	positive and -		37: IDH-mut = 7,		T2WI.	- ADC 50%, AUC 0.690 (0.547-
	negative high-		IDH-wt = 30)		ADC histogram	0.825), Sens 43.7, Spec 91.7, cut
	grade gliomas				parameters	off >1,299 (x10 ⁻⁶ mm²/s),
	(HGGs) using				generated.	p=0.0256.
	histogram					
	analysis of ADC					×
	maps.					· · · · ·
Halefoglu	To investigate	4	170 WHO 4.	Mean	ROI	- ADCmean: Cut off $\leq 0.879 \times 10^{-10}$
AM et al.	whether MRI		(IDH-wt GBM = 146,	57.81+/-	3 ROIs of	³ mm ² /s, Sens 83.65%, Spec
2023	features can		IDH-mut	12.01.	similar size	76.19%, PPV 94.60%, NPV
	determine IDH		astrocytoma = 24).	103M, 67F	placed on	48.50%, AUC 0.866 (0.770-
	mutation in				visually	0.963), p< 0.01.
	HGG.				perceived	- ADCmin: Cutoff ≤0.765.67 x10 ⁻
					darkest regions	³ mm ² /s, Sens 77.88%, Spec
					of ADC map.	80.95%, PPV 95.30%, NPV
					Mean of 3 ROIs	42.50%, AUC 0.860 (0.760-
					used as	0.960), p< 0.01.
					ADCmin.	- rADCmin: Cut off $\leq 1.002 \times 10^{-10}$
			5		Method of	³ mm ² /s, Sens 91.35%, Spec
					ADCmean is	85.71%, PPV 96.90%, NPV
					unclear.	66.70%, AUC 0.939 (0.886-
					Cysts,	0.992), p< 0.01.
					calcification,	
					haemorrhage,	
					necrosis	
					avoided.	
Uetani H et	To investigate	4	327 WHO 4.	Mean 65,	ROI	- ADCmean Reader 1: Cutoff
al. 2023	the most useful		(IDH wt = 306, IDH	Range 24-	4 or more	≥1.014, Sens 55.0%, Spec 70.3%,
	clinical and MRI		mut = 21)	89	circular ROIs	Acc 69.3%, AUC 0.548 (0.383–
	parameters for			194M, 133F	placed within	0.712).
	differentiating				solid tumour,	- ADCmean Reader 2: Cutoff
					targeting	≥0.976, Sens 85.0%, Spec 40.7%,

	IDH mut and wt				regions with	Acc 43.5, AUC 0.61 (0.486–
	glioblastomas.				relatively low	0.734).
					ADC.	- ADCmin Reader 1: Cutoff ≥1.014,
					Necrosis,	Sens 45.0%, Spec 74.7%, Acc
					haemorrhage,	72.8%, AUC 0.533 (0.364–0.701).
					vessels	- ADCmin Reader 2: Cutoff ≥0.866,
					avoided.	Sens 75.0%, Spec 45.9%, Acc
						47. 7%, AUC 0.539 (0.412–0.665).
Xing Z et al.	To evaluate the	4	75 WHO 4.	IDH-mut	ROI	- rADCmin: AUC 0.703, cut off
2019	contribution of		(IDH-wt = 65, IDH-	Mean =	At least 5 non	0.98, Sens 90%, Spec 55.93%,
	DWI in the		mut = 10)	40.70+/-	overlapping	PPV 25.7%, NPV 97.10%.
	enhancing and			10.77.	ROIs placed in	
	peri-enhancing			IDH-wt	the solid	
	region for			Mean =	enhancing	
	discriminating			52.23+/-	portion of	
	IDH genotypes,			12.71.	tumour; mean	
	and the			41M, 34F	value of the	
	diagnostic values		\mathbf{O}		ROI of the	
	of combining		2		lowest ADC	
	two techniques	X	0		value was used	
	in the peri-				as ADCmin-	
	enhancing region	X			tumour	
	compared with				(ADCmin-t).	
	those in the				5 ROIs placed	
	enhancing				in the peri-	
	region.				tumoural, non-	
					enhancing	
					region; mean	
					value of the	
					ROI of the	
					lowest ADC	
					value was used	
					as ADCmin-	

			peritumoural	
			region	
			(ADCmin-p).	
			Necrosis, cysts,	
			haemorrhage,	
			vessels	
			avoided.	
P	Ś	20		

Table 2: Summary of studies assessing IDH mutation and 1p19q codeletion

Summary of study author, year of publication, main research purpose, composition of study cohort (WHO grade, IDH status, 1p19q status, sex), methods of ADC measurements, and key results for studies assessing both IDH mutation and 1p19 codeletion.

Abbreviations: ADC: Apparent Diffusion Coefficient, AUC: Area under the curve, cMRI: Conventional MRI, CNAWM: Contralateral normal appearing white matter, CNS: Central nervous system, Codel: Codeletion, DG: Diffuse gliomas, DTI: Diffusion tensor imaging, DWI: Diffusion weighted imaging, F: Female, GBM: Glioblastoma multiforme, HGGs: High grade gliomas, IDH: Isocitrate dehydrogenase, IQR: Interquartile range, LGGs: Low grade gliomas, M: Male, MRI: Magnetic Resonance Imaging, Mut: Mutant, NAWM: Normal appearing white matter, NS: Not stated, ROI: Region of interest, Sens: Sensitivity, Spec: Specificity

T1w: T1 weighted imaging. T2w: T2-weighted imaging, VOI: Volume of interest, Wt: Wild-type.

Study	Research	wнo	Cohort size and	Age,	ROI or VOI	Results
	purpose	Grades	composition	Sex	methods	
Xiong J et al. 2016	To explore the	2,3	84	Mean	ROI.	IDH
	correlations of		(WHO 2 = 50, WHO	41.5,	4-6 ROIs placed	- ADCmin: cut off 0.85, sens 77.8%,
	cMRI and DTI		3 = 34)	Range	in solid	spec 81.2%, PPV 94.2%, NPV
	values with the		(IDH-mut = 67, IDH-	24-60.	tumour. The	48.0%, AUC 0.82, p=0.001.
	1p/19		wt = 17, 1p19q-	40M,	lowest ADC	- rADCmin: cut off 1.19. sens
	codeletion and		codel = 60, 1p19q-	44F	from the ROIs	79.4% spec 81.2% PPV 94.3%
	IDH mutations		retained = 24)		drawn by the	NPV 50.0% AUC 0.83, p=0.002.
	in				two observers	10190
	oligodendroglial				was averaged	19134
	tumours.				and used as	
					the as the	
1		1	1	1	1	1

					minimum ADC	- ADCmin: cut off 1.13, AUC 0.63,
					value.	Sens 62.3%, Spec 70.0%, PPV
					Calcification,	83.7%, NPV 42.9%, p=0.315.
					cysts, necrosis,	
					haemorrhage	
					avoided.	×
			207			
Allotta E et al.	To develop an	2,3	227	Age	VOI.	IDHmut, 1p19q retained vs IDHwt
2020	ADC analysis-		(WHO 2 and 3,	NS.	Fully	and IDHmut, 1p19q codel.
	based approach		breakdown not	Sex	automated	- ADCmin (Internal dataset): Cut
	that can		provided.)	NS.	segmentation	off 0.8 x10 ⁻³ mm ² /s, Sens 0%, Spec
	automatically		(Internal set = 134:		using 3D-Unet	100%. AUC 0.42. p=0.04428.
	identify		IDH-wt = 31, IDH-		(Internal set)	- ADCmin (TCIA dataset): Cut off
	IDHmut-		mut 1p19q-codel =		and	$0.8 \times 10^{-3} \text{mm}^2/\text{s}$ Sons 0% Spoc
	noncodel LGG		54, IDH-mut 1p19q-		GLISTRboost	100% AUG 0.46 p=0.002
			retained = 49.)	U	(TCIA set)	100%, AUC 0.46, p=0.002.
			(TCIA set = 93: IDH-		algorithms.	- ADCmean (Internal dataset): Cut
			wt = 18, IDH-mut		Generated ADC	off 1.37 x10 ⁻³ mm ² /s, Sens 53%,
			1p19g-codel = 26.		histograms	Spec 91%, AUC 0.76, p<0.00001.
		0	IDH-mut 1p19g-			- ADCmean (TCIA dataset): Cut off
		KK	retained (0)			1.37 x10 ⁻³ mm ² /s, Sens 55%, Spec
			retaineu = 49.)			89%, AUC 0.81, p<0.00001.
Aliotta E et al.	To investigate	2,3	41	Mean	VOI.	IDH
2019	lower grade		(WHO 2 = 26, WHO	45.9,	Automated	- ADC75%: Sens 84+/-0.06 Snec
	glioma grading		3 = 15)	Range	segmentation,	
	using a machine		(IDH-wt = 15, IDH-	18-76.	including	n=0.008
	learning		mut 1p19q-	24M,	enhancing and	μ-0.008. 1510σ
	technique that		retained = 14, IDH-	17F.	non-enhancing	тртад
	estimates		mut 1p19q codel =		tumour, with	- ADC50%: Sens 81+/-0.06, Spec
	fractional		12)		DeepMedic.	0.73+/- 0.04, AUC 0.83+/-0.03,
	anisotropy from				Regions	p<0.001.
	accelerated				combined to	
	diffusion MR				generate	
	imaging scans				whole tumour	
	IIIIagiiig Scalis				whole tumour	

	containing only				volumes.	
	3 diffusion-				ADC	
	encoding				histograms	
	directions.				generated.	
Lee MK et al.	To assess the	2,3	110	Mean	ROI	IDH-mut 1p19q-retained vs IDH-wt
2020	diagnostic value		(WHO 2 = 45, WHO	47.4+/-	ROIs drawn to	- ADC10% AUC 0.751 (0.617-
	of adding the		3 = 65)	13.3.	encompass	0.886) Sens 84.2 Spec 63.6 Acc
	ADC and CBV to		(IDH-wt = 45, IDH-	56M,	entire	60.8 n=0.43
	the T2/FLAIR		mut 1p19q-	54F	hyperintense	No ADC parameters could
	mismatch sign		retained = 19, IDH-		lesion on FLAIR	distinguish IDH mut and IDH wt
	for		mut 1p19q codel =		images and	
	differentiation		46)		enhancing	tumours on multivariate analysis.
	of the IDH				solid tumour	
	mutation or			0	on cases with	
	1p/19q				contrast	
	codeletion.				enhancement.	
			À Ì		ADC	
					histograms	
)		generated.	
Liu D et al. 2020	To evaluate the	2,3	56	IDH	VOI	IDH
	diagnostic		(WHO 2 = 37, WHO	mut:	VOI1: Entire	- ADCmin VOI-1: Cut off 560, Sens
	performance of		3 = 19)	Mean	tumour	62.5%, Spec 87.5%, AUC 0.749.
	ADC histogram		(IDH-wt = 16, IDH-	41.5+/-	included.	- ADCmin VOI-2: Cut off 543, Sens
	parameters for		mut 1p19q-	10.5,	VOI2: Entire	62.5%, Spec 90.0%, AUC 0.831.
	differentiating		retained = 22, IDH-	Range	tumour,	IDH-mut 1p19q-codel vs IDH-mut
	the genetic		mut 1p19q codel =	23-66.	excluding	1p191-retained.
	subtypes in		18)	IDH	cystic and	
	lower-grade			wt:	necrotic	ADCmean VOI-1: Cut off 1546.32,
	diffuse gliomas			Mean	regions.	Sens 95.5%, Spec 55.6%, AUC
	and explore			=	ADC	0.715.
	which			51.9+/-		

	segmentation			16.0,	histograms	ADCmean VOI-2: Cut off 1387.97,
	method (ROI-1,			Range	generated.	Sens 81.8%, Spec 72.2%, AUC
	the entire			21-73.		0.758.
	tumor ROI;			27M,		
	ROI2, the tumor			29F.		
	ROI excluding					X
	cystic and					
	necrotic					
	portions)					
	performs					
	better.				.6	
	To such the	2	70	Maar	1101	
Hong EK et al.	To evaluate the	3	/6	Mean	VUI	IDH:
2021	association of		(WHO 3 = 76)	47.69,	lumour	- ADCmean: Cut off > 1.49, Acc
	MRI features		(IDH-mut = 47, IDH-	Range	 delineated on 	66.7%, Sens 66.7%, Spec 72.7%,
	with the major		wt = 29. 1p19q-	19-68.	axial slices to	AUC 0.67 (0.56–0.78), p=0.008.
	genomic		codel = 19, 1p19q-	47M,	contain high	1p19q:
	profiles and		retained = 57)	29F	signal intensity	
	prognosis of		(Anaplastic		lesions on	 No significant associations
	WHO grade III	. (astrocytoma = 57,		T2WI and	between ADC and 1p19q on
	gliomas		Anaplastic		FLAIR,	multivariable regression analysis.
	compared with		oligodendroglioma		including cystic	
	those of GBMs.		= 19)		and necrotic	
					regions.	
C					Multiplied by	
					slice thickness	
					and	
					intersection	
					gap to obtain	
					tumour	
					volume per	
					section then	
					summated to	

					obtain total	
					tumour	
					volume.	
					ADC	
					histograms	
					generated.	
Su X et al. 2023	To evaluate the	2,3,4	216 across test,	Mean	VOI	IDH:
	value of		training and	45.59.	Automated	- ADCmean (test cohort): Cut off >
	quantitative		validation set.	108M,	segmentation	1.630, Sens 93.8%, Spec 88.9%,
	MRI biomarkers		(WHO 2 ,3, and 4.	65F	with BraTumIA	AUC 0.913 (0.827–0.999).
	for the		Breakdown not		to include	 ADC15% (test cohort): Cut off >
	identification of		provided.)		enhancing and	1.186. Sens 93.8%. Spec 81.5%.
	IDH mutation		(IDH-wt = 127, IDH-		nonenhancing	AUC 0 888 (0 782-0 993)
	and 1p/19q		mut 1p19q-		tumour and	1n19g codeletion amongst IDH mut
	codeletion in		retained = 33, IDH-		necrosis then	gliomas
	adult patients		mut 1p19q codel =		core tumours	giomas.
	with diffuse		56)		obtained with	- ADCmean (training cohort): Cut
	glioma.		\mathbf{O}		registration	off > 1.397, Sens 100%, Spec
		.0			function in	18.8%, AUC 0.409 (0.139–0.624).
					FSL.	- ADC15% (training cohort): Cut off
					ADC	> 1.266, Sens 97.500%, Spec
	0.				histograms	18.8%, AUC 0.440 (0.230–0.651).
					generated.	
C						
Nuessle NC et al.	To investigate	2,3,4	97	Mean	VOI	IDH:
2021	the diagnostic		(WHO 2 = 37, WHO	51.6+/-	VOI manually	- ADCmean: AUC 0.883.
	performance of		3 = 28, WHO 4 =	15.3.	delineated	1p19g codeletion amongst IDH mut
	in vivo ADC-		32)	Sex	around entire	gliomas:
	based		(IDH-wt astrocytic =	NS.	tumour	5 normes.
	stratification of		44, IDH-mut		volume on	- ADCmean: AUC 0.699.
	integrated		astrocytic = 30,		FLAIR	
			1p19q-codel		sequences.	
					Necrosis,	

	molecular		oligodendrogliomas		oedema, and	
	glioma grades.		= 23)		vessels	
					avoided.	
Ma X et al. 2023	To investigate	2,3,4	159	Mean	ROI	IDH
	apparent		(WHO 2, 3, and 4.	47.6+/-	3 ROIs placed	- rADCmin: AUC 0.86 (0.80-0.92),
	diffusion		Breakdown not	14.4.	on visually	p<0.0001, Cut off 1.28, Sens
	coefficient		provided.)	93M,	perceived	69.2%. Spec 92.6%.
	(ADC) as		(IDH-wt GBM = 81,	66F	lowest regions	- ADCmin: AUC 0.84 (0.78–0.90).
	imaging		IDH-mut 1p19q-		of ADC map.	p<0.0001, Cut off 0.93 (x10 ⁻
	biomarker for		retained		Mean of ROI	³ mm ² /s). Sens 65.4%. Spec 91.4%.
	preoperatively		astrocytoma = 46,		ADC values	1p19q codeletion amongst IDH-mut
	identifying		IDH-mut 1p19q-		used as	gliomas
	glioma		codel		ADCmin.	
	genotypes		oligodendroglioma		Calcification,	- rADCmin: AUC 0.67 (0.56–0.79),
	based on the		= 32)		cysts,	p=0.009, Cut off 1.47, Sens
	2021 World				haemorrhage,	52.5%, Spec 81.2%.
	Health				necrosis	- ADCmin: AUC 0.68 (0.57–0.80) ,
	Organization		\bigcirc		avoided.	p=0.006, Cut off 1.17 (x10 ⁻
	(WHO)	. 0				³ mm ² /s), Sens 37.0%, Spec 100%.
	classification of					
	CNS tumors.					
Cheng Y et al.	To explore the	2,3,4	11	Age	VOI	IDH:
2021	correlation		(WHO 2 = 3, WHO	NS.	3D	- ADCmean: AUC 0.500, p>0.05.
	between the		3 = 4, WHO 4 = 4)	Sex	autocontouring	1p19q:
	molecular		(IDH-wt = 6, IDH-	NS.	segmentation.	
	phenotypes of		mut = 5. 1p19q-		Vessels	- ADCmean: AUC 0.916, p<0.05.
	glioma and ADC		codel = 2, 1p19q-		excluded.	
	values.		retained = 9.)			
Cho NS et al	To compare the	231	104 (note 105	Mean	VOL	IDH-mut astrocytoma vs IDH mut
2024		2,3,4		42		
2024	classification			42,		ongodendroglioma / IDHWt gliomas
	pertormance of		(WHO II = 61, WHO	Range	created by	

	normalized		III = 21, WHO IV =	22-79	voxel-wise	- Median rADC: AUC 0.848, cut off
	apparent		23)	59M,	dividision of	1.864, Sens 70.8%, Spec 85.0%,
	diffusion		(IDH-wt = 22, IDH-	45F	ADC by the	p<0.0001. IDH-mut astrocytoma
	coefficient with		mut = 83)"		mean ADC	vs IDH-mut oligodendroglioma
	percentage T2-		*Note: only		value of 3	IDH-mut astrocytoma vs IDH-mut
	FLAIR		included patients		spherical VOIs	oligodendroglioma
	mismatch-		with non-		in the	- Median rADC: AUC 0.805, cut off
	volume for		enhancing gliomas.		CNAWM.	1 864 Sens 70 8% Spec 94 4%
	differentiating				Tumour	n<0.0001
	between IDH-				segmentations	IDH-mut astrocytoma vs IDH-wt
	mutant				performed	glioma
	astrocytoma				manually by	Siona
	and other				one observer	- Median rADC: AUC 0.883, cut off
	glioma				and refined by	1.849, Sens 70.8%, Spec 95.5%,
	molecular			O	a semi-	p<0.0001.
	subtypes.				automated	
					thresholding	
			\frown		method using	
		0			Analysis of	
	X	C			Functional	
					Neurolmages	
	OX				software for	
					consistency	
C					prior to final	
					review by a	
					second	
					observer.	
					Cysts and CSF	
					excluded.	

Table 3: Summary of studies assessing 1p19q codeletion alone

Summary of study author, year of publication, main research purpose, composition of study cohort (WHO grade, IDH status, 1p19q status, sex), methods of ADC measurements, and key results for studies assessing 1p19 codeletion only.

Abbreviations: ADC: Apparent Diffusion Coefficient, AUC: Area under the curve, Codel: Codeletion, DG: Diffuse gliomas, DTI: Diffusion tensor imaging, DWI: Diffusion weighted imaging, F: Female, IDH: Isocitrate dehydrogenase, M: Male, MRI: Magnetic Resonance Imaging, Mut: Mutant, ROI: Region of interest, Sens: Sensitivity, Spec: Specificity, VOI: Volume of interest, Wt: Wild-type.

Study	Research purpose	WHO	Cohort size and	Age,	ROI or VOI methods	Results
		Grades	composition	Sex		
Yang X et	To explore whether	2,3	142	IDH-mut	ROI.	Identification of 1p19q
al. 2021	DWI can predict		(WHO 2 and 3.	1p19q	At least 5 ROIs placed	codeletion in IDH mut gliomas:
	1p19q codeletion	X	Breakdown not	retained:	in solid tumour. ROI	- nADC: Sens 76.71%, Spec
	status of IDH		provided.)	Mean	with lowest mean ADC	52.17%, PPV 88.30%, NPV
	mutant LGGs.	R	(IDH-mut 1p19q-	38.74+/-	used as ADCmin.	67.30%, AUC 0.71 (0.60-
			codel = 73, IDH-	10.09.	Avoided cysts,	0.79), p<0.001.
			mut 1p19q-	IDH-mut	haemorrhage,	
	\mathbf{O}		retained = 69)	1p19q-	necrosis.	
				codel:		
				Mean		
				44.94+/-		
				10.24.		
				80M,		
				62F		
			1			

Cui Y et	To investigate the	1,2,3,4	82	Mean	ROI.	Identification of 1p19q
al. 2014	correlation		(WHO 1 =1, WHO 2	44.16.	4 ROIs placed in solid	codeletion in 35 WHO II
	between tumour		= 35, WHO 3 = 22,	42M,	tumour. Mean of ROI	gliomas (32 IDHmut, 3 IDHwt):
	grade and		WHO 4 = 24)	40F	ADC values used.	- Mean ADC: Cut off 1,565
	prognostic		(Oligodendroglial =		Avoided CSF, cysts,	x10 ⁻⁶ mm²/s , Sens 72%, Spec
	biomarkers with		5, Oligoastrocytic =		necrosis, vessels.	88%, AUC 0.82 (0.68-0.97),
	ADC.		29, Astrocytic = 48)			p=0.003.
						- Mean nADC: Cut off 2.0,
						Sens 76.5%, Spec 88%, AUC
						0.81 (0.67-0.95), p=0.004.
					5	
Latysheva	To assess the value	2,3	71	Mean	VOI	Identification of 1p19q
A et al.	of DWI to		(WHO 2 = 42, WHO	48+/-	Tumour manually	codeletion in a cohort of
2019	characterize		3 = 29)	11.2.	delineated to include	astrocytomas and
	oligodendrogliomas		(Oligodendroglioma	35M,	enhancing and	oligodendrogliomas:
	and to distinguish		= 33, Astrocytoma	36F	nonenhancing regions	- Mean ADC: Cut off 1094
	them from		= 38)		on each axial slice.	x10 ⁻⁶ mm²/s , Sens 63% (54-
	astrocytomas.				Whole tumour	82), Spec 61% (51-83), PPV
			2		histogram distributions	65% (52; 81), NPV 73% (61;
		X			of ADC generated.	87), AUC 0.76, p=0.009.
					Cysts avoided.	
	0			I		I
	CY					
	\mathbf{C}					
0						
Y	~					

		ROI	ROI	ROI		VOI	VOI	VOI
	ROI ADCmin	rADCmin	ADCmean	rADCmean	VOI ADCmin	rADCmin	ADCmean	rADCmean
N studies	8	6	7	3	3	1	5	1
Median	0.95	1.17	1.05	1.6	0.81	1.08	1.38	1.6
SD	0.11	0.16	0.16	0.18	0.19	-	0.19	-
Min	0.77	0.98	0.88	1.4	0.54	-	1.19	-
Max	1.08	1.40	1.34	1.75	0.90	1.08	1.49	1.6
8			20					

F	ig	ur	e	1
	• •	••••	-	_



Figure 2

A: ADCmean (ROI metho	ds)	
Du N et al. 2022	⊢ ∎−−1	0.64 [0.56, 0.73]
Halefogiu AM et al. 2023		0.87 [0.77, 0.96]
Kamble AN et al. 2023		0.75 [0.65, 0.85]
Liu S et al. 2022		0.78 [0.69, 0.87]
Thust SC et al. 2021	-■-1	0.81 [0.76, 0.86]
Uetani H et al. 2023		0.61 [0.49, 0.73]
Zhang H et al. 2024*		0.81 [0.76, 0.86]
RE Model		0.76 [0.70, 0.82]
0	4 0.5 0.6 0.7 0.8 0.9 1.0	
	AUC Value	
B: ADCmean (VOI metho	ds)	
Hong EK et al. 2021		0.67 [0.56, 0.78]
Lee S et al. 2015		0.71 0.58, 0.84
Nuessle NC et al. 2021*	↓ ■	0.88 [0.81, 0.95]
Su X et al. 2023	⊢ ∎−−1	0.91 [0.83, 1.00]
Thust SC et al. 2021	H	0.78 [0.72, 0.84]
RE Model	_	0 80 10 72 0 88
KE MODEL		5.00 (0.72, 0.00)
(0.5 0.6 0.7 0.8 0.9 1.0	
0.100	AUC Value	
C: rADCmean (ROI meth)	005)	0.55 (0.55, 0.74)
Durivet al. 2022		0.65 [0.56, 0.74]
Thust SC at al. 2021		0.84 [0.76, 0.91]
Thust Sc et al. 2021		0.85 [0.78, 0.86]
RE Model	•	0.78 [0.69, 0.87]
0.5	05 07 08 09 10	
0.5	ALIC Value	
	riou runic	
D: ADCmin (ROI method:	<u>ت</u>	
Cindil E et al. 2022	⊢	0.69 [0.61, 0.76]
Du N et al. 2022^	—	0.65 [0.56, 0.74]
Halefoglu AM et al. 2023A	⊢ − −1	0.86 [0.76, 0.96]
Ma X et al. 2023^	⊢ ∎-1	0.86 [0.80, 0.92]
Thust SC et al. 2021	⊢∎⊣	0.79 [0.73, 0.85]
Uetani H et al. 2023	⊢	0.54 [0.41, 0.67]
Wasserman JK et al. 2015	H	0.71 [0.53, 0.89]
Xie Y et al. 2021*	⊢ •1	0.75 [0.65, 0.85]
Xiong J et al. 2016"	⊢ ∎−−1	0.77 [0.65, 0.89]
RE Model	_	0.74 (0.68, 0.81)
	· · · · · · · · · · · · · · · · · · ·	
	0.4 0.5 0.6 0.7 0.8 0.9 1.0	
	AUC Value	
E: ADCmin (VOI methods	U Contraction of the second se	
Thust SC et al. 2021		0.68 [0.61, 0.75]
Villanueva-Meyer JE et al. 2018		0.91 [0.84, 0.97]
Liu D et al. 2020-	· · · · ·	0.83 [0.70, 0.96]
RE Model		0.80 [0.69, 0.92]
	0.6 0.7 0.8 0.9 1.0	
	AUC Value	
F: rADCmin (ROI method	<u>n)</u>	
Du N et al. 2022^	—	0.66 [0.57, 0.75]
Halefoglu AM et al. 2023^	⊢∎-1	0.94 [0.89, 0.99]
Ma V as al. 20224	H B -1	0.84 [0.78, 0.90]
Ma X et al. 20230		
Thust SC et al. 2021	⊢ ∎-1	0.81 [0.76, 0.86]
Thust SC et al. 2021 Xing Z et al. 2019*		0.81 [0.76, 0.86] 0.70 [0.55, 0.86]
Ma X et al. 2023 Thust SC et al. 2021 Xing Z et al. 2019" Xiong J et al. 2016"		0.81 [0.76, 0.86] 0.70 [0.55, 0.86] 0.80 [0.68, 0.92]
Ma A et al. 2023 Thust SC et al. 2021 Xing J et al. 2019* Xiong J et al. 2016* RE Model		0.81 [0.76, 0.86] 0.70 [0.55, 0.86] 0.80 [0.68, 0.92] 0.80 [0.73, 0.88]
Ma A et al. 20239 Thust SC et al. 2019* Xiong J et al. 2016* RE Model		0.81 [0.76, 0.86] 0.70 [0.55, 0.86] 0.80 [0.68, 0.92] 0.80 [0.73, 0.88]
Ma K et al. 2023 Thust SC et al. 2021 Xing Z et al. 2019" Xiong J et al. 2016" RE Model		0.81 [0.76, 0.86] 0.70 [0.55, 0.86] 0.80 [0.68, 0.92] 0.80 [0.73, 0.88]