



## 12 **Summary**

13 There is some evidence that West Nile virus (WNV), which causes encephalomyelitis in equids,  
14 is an emerging disease in Europe. The aim of this study was to perform a systematic review  
15 and meta-analysis to analyse seroprevalence studies of West Nile virus in equids in European  
16 countries between 2001 and 2018. Two electronic databases, PubMed and Scopus, were  
17 searched for relevant publications published from 2001 to 2018 using predetermined  
18 keywords. A total of 1484 papers was initially found. After applying the eligibility criteria, 39  
19 papers were finally included in the systematic review. Analysis of 28,089 equids from 16  
20 European countries revealed a pooled seroprevalence of 8% (95% CI 5–12%,  $P < 0.001$ ,  $I^2 =$   
21 99.3%) in Europe. The pooled seroprevalence was slightly higher in Mediterranean basin  
22 countries than other countries and when calculated for samples collected between 2001 and  
23 2009 compared to 2010 to 2018. Differences in study design (e.g. sampling associated with  
24 recent outbreaks of WNV) contributed to a high degree of variability among studies. Further  
25 studies with harmonized study design and reporting of the results are recommended to better  
26 estimate and monitor European seroprevalence of West Nile virus in equids.

27

28 **Keywords:** horses, equids, Europe, Mediterranean basin, seroprevalence, WNV.

## 29 Introduction

30 West Nile virus (WNV) is a mosquito-borne zoonotic virus from the genus *Flavivirus* and family  
31 *Flaviviridae* (Anon, 2017). Its transmission cycle involves birds and mosquitoes, especially  
32 from the *Culex* species, which act as vectors of the virus. Several vertebrate species can be  
33 infected by the virus, but mammals, particularly humans and equids, are considered “dead-  
34 end” hosts as they do not usually develop sufficiently high levels of viraemia for transmission  
35 to blood-feeding mosquitoes (Komar, 2000). The virus was first isolated from a human being  
36 in 1937 in the West Nile region of Uganda (Smithburn et al., 1940). Since the first reported  
37 case of West Nile virus in horses in Egypt in 1963 (Schmidt & Mansoury, 1963), the disease  
38 has expanded in range causing significant human and animal health issues. An example of  
39 these are the outbreaks reported in EU member countries since the start of the 2020  
40 transmission season and as of 10 September 2020, 173 human cases of WNV infection and 15  
41 deaths (Greece, Spain, Italy and Romania) and 60 outbreaks among equids (Spain, Italy,  
42 France, Portugal and Germany) have been reported through the European Animal Disease  
43 Notification [Systemervisee](#) (ADNS).

44 Several genetic lineages of the virus have been found, but isolates from lineages 1 and 2 have  
45 mainly been responsible for the disease in humans and equids in European countries, with  
46 lineage 1 predominant until the mid-2000s (Ciccozzi et al., 2013; Long, 2014). As a neurotropic  
47 virus causing encephalomyelitis, clinical signs in horses include ataxia, paralysis of the limbs,  
48 prolonged recumbency, muscle fasciculations, and abnormal mentation (Long, 2014). The  
49 mortality rate in horses has been estimated at 35–45% (Long, 2014). However, studies  
50 suggest that only around 10% of infected horses present neurological signs (Gardner et al.,  
51 2007). For diagnosis, laboratory testing is necessary to confirm the infection as the  
52 neurological signs are not pathognomonic for the disease. Treatment is mostly supportive as  
53 there are no known effective antiviral medications (Long, 2014). An equine WNV vaccine was  
54 first licensed in the USA in 2003<sup>5</sup>, and further types of WNV vaccines<sup>s</sup> have since been  
55 approved for use in horses, but an equine WNV vaccine was not licensed for use in Europe  
56 until 2009. Due to the low viral titres in horses, *ante mortem* PCR-based detection of viral RNA  
57 is unreliable (Kleiboeker et al., 2004). Therefore, suspected cases of WNV infection are usually  
58 confirmed by IgM capture ELISA and/or measuring seroconversion using a plaque-reduction  
59 neutralization test (PRNT). Equine WNV-specific IgM antibodies are usually detectable from

60 around 8 days post-infection (so most horses with encephalitis test positive at the time that  
61 clinical signs are first observed) and remain detectable for up to 3 months (Beck et al., 2017).  
62 Neutralising (IgG) antibodies are detectable in equine serum by 2 weeks post-infection and  
63 can persist for more than 1 year. The OIE Terrestrial Manual (OIE, 2018) suggests that IgG  
64 indirect and competitive ELISAs, virus neutralisation test (VNT) or PRNT are suitable methods  
65 for determining prevalence of infection. However, as ELISA methods are less specific, where  
66 related flaviviruses co-circulate with WNV, positive results obtained should be confirmed by  
67 PRNT or VNT and testing against other flaviviruses in parallel. Other flaviviruses detected in  
68 Europe include Bagaza virus (BAGV), louping ill virus (LIV), tick-borne encephalitis virus (TBEV)  
69 and Usutu virus (USUV) (Llorente et al., 2015; Long, 2014).

70 The aim of this study was to conduct a systematic review and meta-analysis to analyse the  
71 prevalence reported on WNV studies in equids in Europe from the year 2001 to 2018  
72 inclusive, to compare the prevalence in countries of the Mediterranean basin with other  
73 European countries and to evaluate the prevalence of two periods: from 2001 to 2009 and  
74 from 2010 to 2018.

## 75 **Materials and methods**

### 76 **Search strategy**

77 A systematic search strategy was performed in the databases PubMed and Scopus to identify  
78 all published studies reporting the prevalence of WNV in equids in Europe from 1 January  
79 2001 to 20 March 2019 (the date the search was performed). The following key words and  
80 Boolean operators (“AND” and “OR”) were used: (prevalence OR incidence OR frequency OR  
81 occurrence OR detection OR identification OR isolation OR characterization OR investigation)  
82 AND (WNV OR West Nile virus OR Flavivirus) AND (horse OR equine OR equid OR donkey OR  
83 mule OR foal). In Scopus, the search terms were applied to the title, abstract and the  
84 keywords. In PubMed, the search terms were applied in all fields. No language restrictions  
85 were applied. Retrieved searches were entered into a Microsoft Excel (2018) file.

86 The reference lists of the selected publications were reviewed manually to identify all  
87 potential studies that could have been missed in the two databases.

### 88 **Eligibility criteria**

89 Inclusion criteria were divided into two categories: inclusion criteria related to the literature  
90 search and inclusion criteria inherent to the studies. First, the studies had to be published

91 between 1 January 2001 and 20 March 2019 and the full text had to be available in English,  
92 Spanish or French. In addition, studies had to be prospective or retrospective serosurveys  
93 with animal level prevalence and animals of the genus *Equus* (excluding zebra) reported,  
94 carried out in a European country and have performed a VNT and/or PRNT to confirm the  
95 specificity of antibodies detected by ELISA.

96 Studies were excluded if the titles and abstracts were not relevant to the subject of interest,  
97 did not fulfil the above eligibility criteria, had data missing or duplicated data published in  
98 another included study.

### 99 **Study selection and data extraction**

100 In the first screening of all searched studies, duplicates were eliminated. The titles and  
101 abstracts of all retrieved studies were then independently screened by two authors (MBCM  
102 and MB) to identify potentially relevant studies. When the study could not be assessed from  
103 the title and abstract, the full text was screened. The full text of the studies retained after the  
104 first screening were further scanned independently and in a standardized manner by two  
105 authors (MBCM and MB) applying the eligibility criteria.

106 After the eligibility assessment process, data were extracted independently by two authors  
107 (MBCM and MB) and classified in three categories: general data related to the study, data  
108 related to the diagnostic techniques and data related to the animals. Any disagreements that  
109 arose between the authors was resolved through discussion with a third author (JMD). All the  
110 extracted data were summarized in a Microsoft Excel (2018) file.

111 The general data related to the study were: title, first author's name, name of the journal,  
112 year of publication, database where the study was identified (PubMed or Scopus), type of  
113 study (i.e. prospective or retrospective serosurvey), language, country and region, sampling  
114 protocol (e.g. convenience or random sample), year and season of testing. The data related  
115 to the diagnostic techniques were: initial serological test to detect the presence of antibody  
116 (immunoglobulin G), type of confirmatory test, strain of the confirmatory test and additional  
117 serological tests performed. The data related to the animals included: number of equids,  
118 mean age, sex, breed, vaccination, clinical signs.

119 The total number of equids tested and the number testing positive specifically for WNV  
120 antibodies (and without a reported history of vaccination) were also extracted independently  
121 by two authors (JMD and OTO) and any disagreement confirmed by a third author (MB).

## 122 **Data analysis and presentation**

123 Statistical meta-analysis of the proportion of WNV antibody-positive animals was conducted  
124 and a forest plot generated using *metaprop* in STATA 16 (Nyaga et al., 2014). Subgroup meta-  
125 analysis was done for the Mediterranean and non-Mediterranean countries of Europe to  
126 investigate if greater prevalence was reported in Mediterranean countries where the climate  
127 is more favourable for mosquitoes. As well, analysis of two balanced 9-year-period subgroups  
128 (2001-2009 and 2010-2018) were performed. This allowed the impact of the introduction of  
129 a WNV vaccine on seroprevalence studies to be determined. Estimates from individual studies  
130 were transformed using the Freeman-Tukey double arcsine transformation to stabilize the  
131 variance. Heterogeneity was assessed using the I squared statistic ( $I^2$ ). A funnel plot to assess  
132 publication bias was generated and outliers identified using R (R Core Team, 2014).

## 133 **Maintenance of study standard**

134 This study has been performed in accordance with guidelines for meta-analysis of  
135 observational studies (MOOSE statement) and preferred reporting items for systematic  
136 reviews and meta-analyses (PRISMA statement) (Moher et al., 2015; Stroup et al., 2000).

## 137 **Results**

### 138 **Search results and study selection**

139 From the initial database search, 1484 potentially relevant publications were identified of  
140 which 663 were found in Scopus and 821 in PubMed. After removing the duplicates, the title  
141 and abstract of 950 studies were screened. Of the 104 studies that remained, 65 studies were  
142 excluded for reasons listed in Figure 1. The lack of a confirmatory test to measure neutralising  
143 antibodies was one of the main reasons for exclusion (n=13). The other exclusion factors  
144 were: review articles, type of study, language other than English, French or Spanish,  
145 insufficient data, full text not available, duplicated data, type of study and year of study.

146 Finally, a total of 39 publications satisfied the inclusion criteria and were included in the  
147 systematic review, of which 38 were in English and one in French. Of the 39 publications, 3  
148 were found in Scopus, 8 in PubMed and 28 in both databases. No additional studies that  
149 satisfied the inclusion criteria were found in the reference lists of selected studies. Table 1  
150 presents the studies included in the systematic review.

### 151 **Study characteristics**

152 Of the 39 studies included in the systematic review, the majority (n=36) were prospective  
153 serosurveys; 3 studies were retrospective. In 14 studies (35.9%), it was stated that the equine  
154 serum samples were taken randomly. It was assumed that in the other studies, convenience  
155 samples were obtained. In total, 28,089 equids were tested, of which 375 were donkeys or  
156 mules. The prevalence was described only in horses in 34 studies, only in donkeys and mules  
157 in one study (García-Bocanegra et al., 2012c) and in both horses and donkeys in 3 studies  
158 (Bosiljka et al., 2013; Ozkul et al., 2013; Raleigh et al., 2012) The mean number of equids  
159 sampled in each study was 720 with a wide range (68 to 5178).

160 Of the 16 European countries in which studies were conducted, 7 (Albania, Croatia, Spain,  
161 France, Italy, Portugal and Turkey) are part of the Mediterranean basin (Figure 2). The highest  
162 number of studies was found for Spain (n=9), followed by France and Serbia (n=4). Prevalence  
163 data were available for both date ranges in the following ten countries: Croatia, Czech  
164 Republic, France, Germany, Ireland, Poland, Portugal, Serbia, Spain and Turkey (34 studies).  
165 Twelve of the studies were carried out in the first date range (2001–2009) and 19 in the  
166 second period (2010–2018). In one study (Raleigh et al., 2012), prevalence data were  
167 separated in the two periods of time. There were five studies that started in the first period  
168 and finished in the second period and two for which the year(s) of sampling was not specified.  
169 In the majority of studies (n=24), samples were first screened by ELISA and some or all of the  
170 positive-testing samples were confirmed by testing for WNV-specific neutralising antibodies.  
171 In 10 studies, a neutralisation test was performed without prior screening by ELISA and in one  
172 study both ELISA and VNT were used to screen the samples. In three studies, the initial  
173 screening test was either agar gel immunodiffusion (AGID), immunofluorescence antibody  
174 test (IFAT), or multiplex immuno-assay (MIA). Additional tests performed included western  
175 blot and haemagglutination inhibition (HI) test.

176 The virus strain used for the VNT or PRNT was described in 24 studies. The strain 'Eg101' was  
177 used in 12 studies, 'New York (NY99)' in 7 studies and 'Israel 1998 (IS98-ST1)' in one study.  
178 These three strains belong to genetic lineage 1. Only five studies used genetic lineage 2  
179 strains: 'Austrian' (n=3), 'Hungary 578/2010' (n=1) or unspecified (n=1). Of the five studies  
180 that used genetic lineage 2 strains, one study also used a strain from genetic lineage 1. The  
181 remaining studies (n=15) did not specify the strain used.

182 In 18 of the 39 studies, samples were additionally screened for neutralising antibodies to  
183 other flaviviruses; TBEV only in 2, USUV only in 6, USUV and TBEV in 9 and Bagaza virus in 1.  
184 Positive titres were detected against TBEV or USUV in five and seven studies, respectively. In  
185 four studies, some samples had similar titres against both WNV and USUV.

186 Of the 39 selected studies, only 3 described minimal demographic data (age, sex, breed) and  
187 8 reported whether or not any of the tested equids were vaccinated or showed any clinical  
188 signs.

189 More than half of the studies (n=23) reported the season of year when the animals were  
190 sampled; the majority of the studies were carried out in autumn (September to November).

### 191 **Meta-analysis of West Nile virus seroprevalence**

192 The pooled seroprevalence was 8% (95% CI 5–12%) with substantial heterogeneity ( $I^2 =$   
193 99.3%) (Figure 3). Pooled seroprevalence was slightly higher in Mediterranean (9%, 95% CI 5–  
194 14%) than non-Mediterranean countries (7%, 95% CI 3–14) and in the first sampling period  
195 (8%, 95% CI 2–17%) than in the second sampling period (7%, 95% CI 4–10%) (Table 2). A funnel  
196 plot (Figure 4) did not identify significant publication bias. However, two studies (Calistri et  
197 al., 2010; Petrović et al., 2014) were identified as outliers in the Studentized residual test.

### 198 **Discussion**

199 This systematic review sought to highlight important trends in WNV seroprevalence in equids  
200 in Europe; prevalence data of WNV reported in 39 studies of equids in Europe were analysed  
201 from the year 2001 to the year 2018. The pooled seroprevalence obtained was 8% (95% CI 5–  
202 12%). However, few studies reported using random sampling methods, therefore caution  
203 must be applied when generalising the seroprevalence estimates to the target population.  
204 The substantial heterogeneity ( $I^2 = 99.3%$ ) meant that meaningful conclusions could not be  
205 drawn about differences in seroprevalence between Mediterranean and non-Mediterranean  
206 countries of the two periods evaluated.

207 There was no evidence that small studies with small effect sizes were missing. However, the  
208 two studies that were identified as outliers (Calistri et al., 2010; Petrović et al., 2014) were  
209 also the two studies with the highest seroprevalence: 39% and 49%, respectively. The study  
210 by Calistri et al. (2010) was associated with investigation of an outbreak of WNV. Similarly, in  
211 the study by Petrović et al. (2014), samples were collected from horses in November and  
212 December 2012 after the first human outbreak of WNV reported in Serbia, which started in



213 August 2012. Other studies with high seroprevalence were also associated with recent  
214 outbreaks.

215 The quality of data reporting varied between studies, for example, only three studies provided  
216 information on animal characteristics such as age, sex and breed, each of which could  
217 influence risk of exposure to and/or susceptibility to WNV infection for example, older  
218 animals are more likely to have been exposed to virus. Reporting these demographic data is  
219 important for comparisons are to be made between the outcomes of different studies.  
220 Recruitment criteria are important in understanding disease transmission in mobile animal  
221 populations such as horses where animals may have been exposed to the virus somewhere  
222 other than the study location. Furthermore, vaccination status became important after an  
223 equine WNV vaccine was first licensed in Europe in 2009. For example, Ziegler et al. (2012)  
224 found four samples positive for WNV antibodies in a study conducted in Germany, but three  
225 of these were from vaccinated horses (and were therefore removed from the seroprevalence  
226 estimation in this study) and one was from a horse from Hungary. However, vaccination status  
227 was only reported in 8 studies although 19 were conducted on samples collected after 2009.  
228 ELISA is often the assay of choice for conducting seroepidemiological studies because it is  
229 simple, sensitive, rapid and often commercially available. However, due to extensive cross-  
230 reactivity between antibodies raised against different flaviviruses, the ELISA can yield false  
231 positive results where different flaviviruses co-circulate (Beck et al., 2013). Therefore, this  
232 systematic review only included studies that used virus / plaque reduction neutralisation tests  
233 to confirm positive samples, however 21 out of 39 studies did not test in parallel other  
234 flaviviruses, which could have introduced errors in their prevalence estimation. The issue of  
235 cross-reactivity in ELISA was illustrated in some of the studies, for example Berxholi et al.  
236 (2013) found that two of seven samples that were positive in ELISA but negative in WNV VNT  
237 were positive for TBEV antibodies. Similarly, Ziegler et al. (2013a) found that four samples  
238 that were positive by ELISA but negative by WNV VNT were positive for TBEV (but not for  
239 USUV). One of the included studies (Lupulovic et al., 2011) was the first to report neutralising  
240 antibodies to USUV in horses, however, as the PRNT titres were 120 and 90 for WNV and  
241 USUV, respectively, they were not able to conclude whether this represented cross-reactive  
242 antibodies or prior exposure to both viruses. Calistri et al. (2010) mention that USUV was  
243 circulating in Italy in the year before samples were obtained in their study, but they did not

244 test for USUV antibodies. In most cases, neutralisation tests were positive for one virus only  
245 or titres were markedly higher (e.g. at least 2-fold) for one virus. However, neutralisation tests  
246 were not always discriminatory, particularly where VNT titres were low (Jiménez-Clavero et  
247 al., 2007). Furthermore, Vanhomwegen et al. (2017) concluded that of 21 samples that were  
248 positive for flavivirus antibodies, 11 were specifically positive for WNV, 2 for USUV and 1 for  
249 TBEV while 8 were positive for an unidentified flavivirus (1 of which they reported as positive  
250 for both WNV and an unidentified flavivirus). Ziegler et al. (2013a) reported four samples that  
251 were positive by WNV ELISA but VNT negative with WNV and USUV, which although this could  
252 also be related with due to the lower sensitivity of VNT compared to ELISA (Beck et al., 2017).  
253 Furthermore, a high prevalence of tick-borne encephalitis virus (TBEV) has been reported in  
254 other studies in some European countries (Rushton et al., 2013). Therefore, there is evidence  
255 of non WNV flaviviruses circulating in Europe, which should be taken into account when  
256 performing a serosurvey study.

257 Although most studies did not specify if seropositivity was caused by lineage 1 or 2 WNV  
258 strains, this is probably not critical, because it is not possible to tell by which virus lineage the  
259 immune response was elicited as the two strains differ only in three amino acids within  
260 domain III of the E protein, against which most neutralizing antibodies are directed (Berxholi  
261 et al., 2013).

262 Most studies were performed in autumn, probably after the beginning of the transmission  
263 season and occurrence of outbreaks in the summer. This should not affect the results of the  
264 meta-analysis other than the already mentioned bias caused by association of a serosurvey  
265 study after regional outbreaks, which could falsely increase the prevalence estimation.

266 Despite the fact that WNV transmission and outbreaks in equids occurs in EU countries, the  
267 limitations detected in this study precluded the evaluation of an increase in seroprevalence  
268 over time. An analysis of the number of reported outbreaks could be useful to determine the  
269 re-emerging status of this virus, however this was not the scope of this study.

270 Horses have been suggested as useful sentinels for WNV surveillance. However, the true  
271 seroprevalence of WNV in European equids remains uncertain due to variation in study design  
272 and reporting, and difficulty discriminating between cross-reactive antibodies. Standardised  
273 seroprevalence studies are critical to better understand the current epidemiological status of  
274 WNV in Europe and to monitor future changes.

275

## 276 **Conflict of interest statement**

277 No conflict of interest to declare by the authors.

278

## 279 **Ethics statement**

280 The authors confirm that the ethical policies of the journal, as noted on the journal's author  
281 guidelines page, have been adhered to. No ethical approval was required as this is a review  
282 article with no original research data.

283

## 284 **Data availability statement**

285 Data sharing is not applicable to this article as no new data were created or analyzed in this  
286 study.

287

## 288 **References**

- 289 Abad-Cobo, A., Llorente, F., Barbero, M. D. C., Cruz-López, F., Forés, P., & Jiménez-Clavero, M.  
290 A. (2017). Serosurvey Reveals Exposure to West Nile Virus in Asymptomatic Horse  
291 Populations in Central Spain Prior to Recent Disease Foci. *Transbound Emerg Dis*,  
292 64(5), 1387-1392. doi:10.1111/tbed.12510
- 293 Alba, A., Allepuz, A., Napp, S., Soler, M., Selga, I., Aranda, C., . . . Busquets, N. (2014). Ecological  
294 surveillance for West Nile in Catalonia (Spain), learning from a five-year period of  
295 follow-up. *Zoonoses Public Health*, 61(3), 181-191. doi:10.1111/zph.12048
- 296 Anon. (2017). Chapter 29 - Flaviviridae. In N. J. MacLachlan & E. J. Dubovi (Eds.), *Fenner's*  
297 *Veterinary Virology (Fifth Edition)* (pp. 525-545). Boston: Academic Press.
- 298 Bakonyi, T., Ferenczi, E., Erdélyi, K., Kutasi, O., Csörgő, T., Seidel, B., . . . Nowotny, N. (2013).  
299 Explosive spread of a neuroinvasive lineage 2 West Nile virus in Central Europe,  
300 2008/2009. *Vet Microbiol*, 165(1-2), 61-70. doi:10.1016/j.vetmic.2013.03.005
- 301 Barbić, L., Listeš, E., Katić, S., Stevanović, V., Madić, J., Starešina, V., . . . Savini, G. (2012).  
302 Spreading of West Nile virus infection in Croatia. *Vet Microbiol*, 159(3-4), 504-508.  
303 doi:10.1016/j.vetmic.2012.04.038
- 304 Barbić, L., Vilibić-Čavlek, T., Listeš, E., Stevanović, V., Gjenero-Margan, I., Ljubin-Sternak, S., .  
305 . . Savini, G. (2013). Demonstration of Usutu virus antibodies in horses, Croatia. *Vector*  
306 *Borne Zoonotic Dis*, 13(10), 772-774. doi:10.1089/vbz.2012.1236
- 307 Barros, S. C., Ramos, F., Fagulha, T., Duarte, M., Henriques, A. M., Waap, H., . . . Fevereiro, M.  
308 (2017). West Nile virus in horses during the summer and autumn seasons of 2015 and  
309 2016, Portugal. *Vet Microbiol*, 212, 75-79. doi:10.1016/j.vetmic.2017.11.008
- 310 Barros, S. C., Ramos, F., Fagulha, T., Duarte, M., Henriques, M., Luís, T., & Fevereiro, M. (2011).  
311 Serological evidence of West Nile virus circulation in Portugal. *Vet Microbiol*, 152(3-4),  
312 407-410. doi:10.1016/j.vetmic.2011.05.013

- 313 Bażanów, B., Jansen van Vuren, P., Szymański, P., Stygar, D., Fracka, A., Twardón, J., . . .  
314 Pawęska, J. T. (2018). A Survey on West Nile and Usutu Viruses in Horses and Birds in  
315 Poland. *Viruses*, *10*(2). doi:10.3390/v10020087
- 316 Beck, C., Jiménez-Clavero, M. A., Leblond, A., Durand, B., Nowotny, N., Leparç-Goffart, I., . . .  
317 Lecollinet, S. (2013). Flaviviruses in Europe: complex circulation patterns and their  
318 consequences for the diagnosis and control of West Nile disease. *Int J Environ Res*  
319 *Public Health*, *10*(11), 6049-6083. doi:10.3390/ijerph10116049
- 320 Beck, C., Lowenski, S., Durand, B., Bahuon, C., Zientara, S., & Lecollinet, S. (2017). Improved  
321 reliability of serological tools for the diagnosis of West Nile fever in horses within  
322 Europe. *PLoS neglected tropical diseases*, *11*(9), e0005936-e0005936.  
323 doi:10.1371/journal.pntd.0005936
- 324 Berxholi, K., Ziegler, U., Rexhepi, A., Schmidt, K., Mertens, M., Korro, K., . . . Groschup, M. H.  
325 (2013). Indigenous West Nile virus infections in horses in Albania. *Transbound Emerg*  
326 *Dis*, *60 Suppl 2*, 45-50. doi:10.1111/tbed.12141
- 327 Bosiljka, D., Vasić, A., Rogožarski, D., Vojinović, D., Elezović-Radovanović, M., Manić, M., . . .  
328 Gligić, A. (2013). Seroepizootiological-epidemiological investigation and mapping of  
329 west nile infection in the republic of Serbia. *Acta Veterinaria*, *63*(5-6), 569-579.  
330 doi:10.2298/AVB1306569D
- 331 Busani, L., Capelli, G., Cecchinato, M., Lorenzetto, M., Savini, G., Terregino, C., . . . Marangon,  
332 S. (2011). West Nile virus circulation in Veneto region in 2008-2009. *Epidemiology and*  
333 *Infection*, *139*(6), 818-825. doi:10.1017/S0950268810001871
- 334 Busquets, N., Laranjo-González, M., Soler, M., Nicolás, O., Rivas, R., Talavera, S., . . . Napp, S.  
335 (2019). Detection of West Nile virus lineage 2 in North-Eastern Spain (Catalonia).  
336 *Transboundary and Emerging Diseases*, *66*(2), 617-621. doi:10.1111/tbed.13086
- 337 Cabre, O., Durand, J. P., Prangé, A., Gomez, J., Maurizi, L., Tolou, H., & Davoust, B. (2005).  
338 West Nile virus infection: serological investigation among horses in France and in  
339 Africa. *Médecine tropicale : revue du Corps de santé colonial*, *65*(5), 439-443.
- 340 Calistri, P., Giovannini, A., Savini, G., Monaco, F., Bonfanti, L., Ceolin, C., . . . Lelli, R. (2010).  
341 West nile virus transmission in 2008 in north-eastern Italy. *Zoonoses and Public*  
342 *Health*, *57*(3), 211-219. doi:10.1111/j.1863-2378.2009.01303.x
- 343 Ciccozzi, M., Peletto, S., Cella, E., Giovanetti, M., Lai, A., Gabanelli, E., . . . Zehender, G. (2013).  
344 Epidemiological history and phylogeography of West Nile virus lineage 2. *Infect Genet*  
345 *Evol*, *17*, 46-50. doi:10.1016/j.meegid.2013.03.034
- 346 Csank, T., Drzewnioková, P., Korytár, L., Major, P., Gyuranecz, M., Pistl, J., & Bakonyi, T. (2018).  
347 A Serosurvey of Flavivirus Infection in Horses and Birds in Slovakia. *Vector Borne*  
348 *Zoonotic Dis*, *18*(4), 206-213. doi:10.1089/vbz.2017.2216
- 349 Durand, B., Dauphin, G., Zeller, H., Labie, J., Schuffenecker, I., Murri, S., . . . Zientara, S. (2005).  
350 Serosurvey for West Nile virus in horses in southern France. *Vet Rec*, *157*(22), 711-  
351 713. doi:10.1136/vr.157.22.711
- 352 Ergunay, K., Gunay, F., Erisoz Kasap, O., Oter, K., Gargari, S., Karaoglu, T., . . . Ozkul, A. (2014).  
353 Serological, molecular and entomological surveillance demonstrates widespread  
354 circulation of West Nile virus in Turkey. *PLoS Negl Trop Dis*, *8*(7), e3028.  
355 doi:10.1371/journal.pntd.0003028
- 356 García-Bocanegra, I., Arenas-Montes, A., Jaén-Téllez, J. A., Napp, S., Fernández-Morente, M.,  
357 & Arenas, A. (2012c). Use of sentinel serosurveillance of mules and donkeys in the  
358 monitoring of West Nile virus infection. *Vet J*, *194*(2), 262-264.  
359 doi:10.1016/j.tvjl.2012.04.017

360 García-Bocanegra, I., Arenas-Montes, A., Napp, S., Jaén-Téllez, J. A., Fernández-Morente, M.,  
361 Fernández-Molera, V., & Arenas, A. (2012a). Seroprevalence and risk factors  
362 associated to West Nile virus in horses from Andalusia, Southern Spain. *Vet Microbiol*,  
363 *160*(3-4), 341-346. doi:10.1016/j.vetmic.2012.06.027

364 García-Bocanegra, I., Jaén-Téllez, J. A., Napp, S., Arenas-Montes, A., Fernández-Morente, M.,  
365 Fernández-Molera, V., & Arenas, A. (2012b). Monitoring of the West Nile virus  
366 epidemic in Spain between 2010 and 2011. *Transbound Emerg Dis*, *59*(5), 448-455.  
367 doi:10.1111/j.1865-1682.2011.01298.x

368 Gardner, I. A., Wong, S. J., Ferraro, G. L., Balasuriya, U. B., Hullinger, P. J., Wilson, W. D., . . .  
369 MacLachlan, N. J. (2007). Incidence and effects of West Nile virus infection in  
370 vaccinated and unvaccinated horses in California. *Vet Res*, *38*(1), 109-116.  
371 doi:10.1051/vetres:2006045

372 Hubálek, Z., Ludvíková, E., Jahn, P., Trembl, F., Rudolf, I., Svobodová, P., . . . Staššíková, Z.  
373 (2013). West Nile Virus equine serosurvey in the Czech and Slovak republics. *Vector*  
374 *Borne Zoonotic Dis*, *13*(10), 733-738. doi:10.1089/vbz.2012.1159

375 Hubálek, Z., Wegner, E., Halouzka, J., Tryjanowski, P., Jerzak, L., Šikutová, S., . . . Wlodarczyk,  
376 R. (2008). Serologic survey of potential vertebrate hosts for West Nile virus in Poland.  
377 *Viral Immunol*, *21*(2), 247-253. doi:10.1089/vim.2007.0111

378 Jiménez-Clavero, M. A., Llorente, F., Sotelo, E., Soriguer, R., Gómez-Tejedor, C., & Figuerola,  
379 J. (2010). West Nile virus serosurveillance in horses in Donana, Spain, 2005 to 2008.  
380 *Vet Rec*, *167*(10), 379-380. doi:10.1136/vr.c3155

381 Jiménez-Clavero, M. A., Gómez-Tejedor, C., Rojo, G., Soriguer, R., & Figuerola, J. (2007).  
382 Serosurvey of West Nile virus in equids and bovids in Spain. *Vet Rec*, *161*(6), 212.  
383 doi:10.1136/vr.161.6.212

384 Kleiboeker, S. B., Loiacono, C. M., Rottinghaus, A., Pue, H. L., & Johnson, G. C. (2004).  
385 Diagnosis of West Nile virus infection in horses. *J Vet Diagn Invest*, *16*(1), 2-10.  
386 doi:10.1177/104063870401600102

387 Komar, N. (2000). West Nile viral encephalitis. *Rev. sci. tech. Off. int. Epiz.*, *19*, 166-176.

388 Llorente, F., Pérez-Ramírez, E., Fernández-Pinero, J., Elizalde, M., Figuerola, J., Soriguer, R. C.,  
389 & Jiménez-Clavero, M. Á. (2015). Bagaza virus is pathogenic and transmitted by direct  
390 contact in experimentally infected partridges, but is not infectious in house sparrows  
391 and adult mice. *Veterinary research*, *46*(1), 93-93. doi:10.1186/s13567-015-0233-9

392 Long, M. T. (2014). West Nile virus and equine encephalitis viruses: new perspectives. *Vet Clin*  
393 *North Am Equine Pract*, *30*(3), 523-542. doi:10.1016/j.cveq.2014.08.009

394 Lupulovic, D., Martín-Acebes, M. A., Lazic, S., Alonso-Padilla, J., Blázquez, A. B., Escribano-  
395 Romero, E., . . . Saiz, J. C. (2011). First serological evidence of West Nile virus activity  
396 in horses in Serbia. *Vector Borne Zoonotic Dis*, *11*(9), 1303-1305.  
397 doi:10.1089/vbz.2010.0249

398 Madić, J., Savini, G., Di Gennaro, A., Monaco, F., Jukić, B., Kovač, S., . . . Listeš, E. (2007).  
399 Serological evidence for West Nile virus infection in horses in Croatia. *Vet Rec*, *160*(22),  
400 772-773. doi:10.1136/vr.160.22.772

401 Maquart, M., Dahmani, M., Marié, J. L., Gravier, P., Leparç-Goffart, I., & Davoust, B. (2017).  
402 First Serological Evidence of West Nile Virus in Horses and Dogs from Corsica Island,  
403 France. *Vector Borne Zoonotic Dis*, *17*(4), 275-277. doi:10.1089/vbz.2016.2024

404 Medić, S., van den Hoven, R., Petrović, T., Lupulović, D., & Nowotny, N. (2014). Serological  
405 evidence of West Nile virus infection in the horse population of northern Serbia. *J*  
406 *Infect Dev Ctries*, *8*(7), 914-918. doi:10.3855/jidc.3885

407 Moher, D., Shamseer, L., Clarke, M., Ghersi, D., Liberati, A., Petticrew, M., . . . Group, P.-P.  
408 (2015). Preferred reporting items for systematic review and meta-analysis protocols  
409 (PRISMA-P) 2015 statement. *Systematic Reviews*, 4(1), 1. doi:10.1186/2046-4053-4-1  
410 Monaco, F., Lelli, R., Teodori, L., Pinoni, C., Di Gennaro, A., Polci, A., . . . Savini, G. (2010). Re-  
411 emergence of West Nile virus in Italy. *Zoonoses Public Health*, 57(7-8), 476-486.  
412 doi:10.1111/j.1863-2378.2009.01245.x

413 Nyaga, V. N., Arbyn, M., & Aerts, M. (2014). Metaprop: a Stata command to perform meta-  
414 analysis of binomial data. *Archives of public health = Archives belges de sante publique*,  
415 72(1), 39-39. doi:10.1186/2049-3258-72-39

416 OIE. (2018). West Nile fever. In *OIE Terrestrial Manual* (8 ed., pp. 697-710). Paris, France: OIE.

417 Ozkul, A., Ergunay, K., Koysuren, A., Alkan, F., Arsava, E. M., Tezcan, S., . . . Us, D. (2013).  
418 Concurrent occurrence of human and equine West Nile virus infections in Central  
419 Anatolia, Turkey: the first evidence for circulation of lineage 1 viruses. *Int J Infect Dis*,  
420 17(7), e546-551. doi:10.1016/j.ijid.2013.02.005

421 Ozkul, A., Yildirim, Y., Pinar, D., Akcali, A., Yilmaz, V., & Colak, D. (2006). Serological evidence  
422 of West Nile Virus (WNV) in mammalian species in Turkey. *Epidemiol Infect*, 134(4),  
423 826-829. doi:10.1017/S0950268805005492

424 Petrović, T., Lazić, S., Lupulović, D., Lazić, G., Bugarski, D., Vidanović, D., . . . Petrić, D. (2014).  
425 Serological study on WNV presence in horses in Vojvodina after the human outbreak  
426 in Serbia in 2012. *Archives of Biological Sciences* 66, 473-481.

427 Pradier, S., Sandoz, A., Paul, M. C., Lefebvre, G., Tran, A., Maingault, J., . . . Leblond, A. (2014).  
428 Importance of wetlands management for West Nile Virus circulation risk, Camargue,  
429 Southern France. *Int J Environ Res Public Health*, 11(8), 7740-7754.  
430 doi:10.3390/ijerph110807740

431 R Core Team. (2014). R: A language and environment for statistical computing. Retrieved from  
432 <http://www.R-project.org/>

433 Raleigh, P. J., Sammin, D. J., Connell, J., Markey, B. K., & O'Connor, M. (2012). Surveillance for  
434 antibodies to West Nile virus in Ireland. *Vet Rec*, 170(7), 180. doi:10.1136/vr.100333

435 [Rushton, J.O., Lecollinet, S., Hubálek, Z., Svobodová, P., Lussy, H., & Nowotny, N. \(2013\). Tick-](#)  
436 [borne Encephalitis virus in horses, Austria, 2011. \*Emerg Infect Dis\*, 19\(4\), 635-637.](#)  
437 [doi:10.3201/eid1904.121450](#)

438 Schmidt, J. R., & Mansoury, H. K. E. (1963). Natural and Experimental Infection of Egyptian  
439 Equines with West Nile Virus. *Annals of Tropical Medicine & Parasitology*, 57(4), 415-  
440 427. doi:10.1080/00034983.1963.11686194

441 Smithburn, K. C., Hughes, T. P., Burke, A. V., & Paul, J. H. (1940). A neurotropic virus isolated  
442 from the blood of a native of Uganda. *Am. J. Trop. Med. Hyg*, 20, 471-492.

443 Stroup, D. F., Berlin, J. A., Morton, S. C., Olkin, I., Williamson, G. D., Rennie, D., . . . Thacker, S.  
444 B. (2000). Meta-analysis of observational studies in epidemiology: a proposal for  
445 reporting. Meta-analysis Of Observational Studies in Epidemiology (MOOSE) group.  
446 *Jama*, 283(15), 2008-2012. doi:10.1001/jama.283.15.2008

447 Vanhomwegen, J., Beck, C., Desprès, P., Figuerola, A., García, R., Lecollinet, S., . . . Serra-Cobo,  
448 J. (2017). Circulation of Zoonotic Arboviruses in Equine Populations of Mallorca Island  
449 (Spain). *Vector Borne Zoonotic Dis*, 17(5), 340-346. doi:10.1089/vbz.2016.2042

450 Weissenböck, H., Hubálek, Z., Halouzka, J., Pichlmair, A., Maderner, A., Fagner, K., . . .  
451 Nowotny, N. (2003). Screening for West Nile virus infections of susceptible animal  
452 species in Austria. *Epidemiol Infect*, 131(2), 1023-1027.  
453 doi:10.1017/s0950268803001031

454 Ziegler, U., Angenvoort, J., Klaus, C., Nagel-Kohl, U., Sauerwald, C., Thalheim, S., . . . Groschup,  
455 M. H. (2013a). Use of competition ELISA for monitoring of West Nile virus infections  
456 in horses in Germany. *Int J Environ Res Public Health*, *10*(8), 3112-3120.  
457 doi:10.3390/ijerph10083112  
458 Ziegler, U., Seidowski, D., Angenvoort, J., Eiden, M., Müller, K., Nowotny, N., & Groschup, M.  
459 H. (2012). Monitoring of West Nile virus infections in Germany. *Zoonoses Public*  
460 *Health*, *59 Suppl 2*, 95-101. doi:10.1111/zph.12015  
461 Ziegler, U., Skrypnyk, A., Keller, M., Staubach, C., Bezymennyi, M., Damiani, A. M., . . .  
462 Groschup, M. H. (2013b). West nile virus antibody prevalence in horses of Ukraine.  
463 *Viruses*, *5*(10), 2469-2482. doi:10.3390/v5102469  
464  
465

Table 1. Characteristics of studies included in the systematic review

Publication	Country <sup>1</sup>	No. positive	No. tested	Seroprevalence (%)	Year	Screening test	Tests for other flaviviruses
1. Abad-Cobo et al. 2017	Spain (ES)	5	369	1.36	2011–2013	ELISA	USUV
2. Alba et al. 2014	Spain (ES)	0	178	0	2011	ELISA	
3. Bakonyi et al. 2013	Hungary (HU)	79	276	28.62	2009	IFAT	
4. Barbić et al. 2012	Croatia (HR)	72	2098	3.43	2010–2011	ELISA	TBEV + USUV
5. Barbić et al. 2013	Croatia (HR)	48	1380	3.48	2011	ELISA	TBEV + USUV
6. Barros et al. 2011	Portugal (PT)	40	1313	3.05	2004–2010	ELISA	
7. Barros et al. 2017	Portugal (PT)	18	989	1.82	2011–2016	ELISA	
8. Bażanów et al. 2018	Poland (PL)	62	411	15.09	2012–2013	VNT	USUV
9. Berxholi et al. 2013	Albania (AL)	37	167	22.16	N.S.	ELISA & VNT	TBEV
10. Bosiljka et al. 2013	Serbia (RS)	45	1199 <sup>2</sup>	3.75	2008–2012	AGID	
11. Busani et al. 2011	Italy (IT)	348	2528	13.77	2008 & 2009	ELISA	TBEV + USUV
12. Busquets et al. 2019	Spain (ES)	9	138	6.52	2017 & 2018	ELISA	BAGV
13. Cabre et al. 2005	France (FR)	0	94	0	2003	ELISA	
14. Calistri et al. 2010	Italy (IT)	794	2030	39.11	2008	PRNT	
15. Csank et al. 2018	Slovakia (SK)	10	145	6.90	2013	ELISA	TBEV + USUV
16. Durand et al. 2005	France (FR)	304	906	33.55	2003	ELISA	
17. Ergunay et al. 2014	Turkey (TR)	48	389	12.34	2011–2013	PRNT	
18. García-Bocanegra et al. 2012a	Spain (ES)	36	510	7.06	2010	ELISA	
19. García-Bocanegra et al. 2012b	Spain (ES)	12	109	11.01	2010–2011	ELISA	
20. García-Bocanegra et al. 2012c	Spain (ES)	12	165 <sup>3</sup>	7.27	2011	ELISA	
21. Hubálek et al. 2008	Poland (PL)	0	78	0	2006	PRNT	USUV



22. Hubálek et al. 2013	Czechia (CZ) & Slovakia (SK)	19	395	4.81	2008–2011	PRNT	TBEV + USUV
23. Jiménez-Clavero et al. 2007	Spain (ES)	13	157	8.28	2005	VNT	USUV
24. Jiménez-Clavero et al. 2010	Spain (ES)	0	68*	0	2008	VNT	USUV
25. Lupulovic et al. 2011	Serbia (RS)	42	349	12.03	2009–2010	ELISA	USUV
26. Madić et al. 2007	Croatia (HR)	4	980	0.41	2010–2011	ELISA	
27. Maquart et al. 2017	France (FR)	9	96	9.38	2014	ELISA	USUV
28. Medić et al. 2014	Serbia (RS)	72	252	28.57	2007–2011	ELISA	
29. Monaco et al. 2010	Italy (IT)	271	770	35.19	2008	VNT	TBEV + USUV
30. Ozkul et al. 2006	Turkey (TR)	36	299 <sup>4</sup>	12.04	N.S.	PRNT	
31. Ozkul et al. 2013	Turkey (TR)	57	180	31.67	2011	PRNT	
32. Petrović et al. 2014	Serbia (RS)	64	130	49.23	2012	ELISA	
33. Pradier et al. 2014	France (FR)	143	1159	12.34	2007–2008	ELISA	
34. Raleigh et al. 2012	Ireland (IE)	0	490 <sup>5</sup>	0	2005–2006 (n=90) & 2010 (n=400)	ELISA	
35. Vanhomwegen et al. 2017	Spain (ES)	11	172	6.40	2011–2012	MIA	TBEV + USUV
36. Weissenböck et al. 2003	Austria (AT)	0	350	0	2001	PRNT	
37. Ziegler et al. 2012	Germany (DE)	1	1282	0.08	2007–2009	ELISA	TBEV + USUV
38. Ziegler et al. 2013a	Germany (DE)	2	5178	0.04	2010–2012	ELISA	TBEV + USUV
39. Ziegler et al. 2013b	Ukraine (UA)	42	310	13.55	2010–2011	ELISA	TBEV

<sup>1</sup>Two letter ISO country code

N.S., not specified

AGID, agar gel immunodiffusion; IFAT, immunofluorescence antibody test; ELISA, enzyme-linked immunosorbent assay; MIA, multiplex immunoassay; PRNT, plaque reduction test; VNT, virus neutralization test.

BAGV, Bagaza virus; TBEV, tick-borne encephalitis virus; USUV, Usutu virus

<sup>2</sup>1133 horses and 66 donkeys; <sup>3</sup>82 donkeys and 83 mules; <sup>4</sup>259 horses and 40 mules; <sup>5</sup>386 horses and 104 donkeys

\*Only results from samples collected in 2008 were included in the meta-analysis

**Table 2.** Pooled seroprevalence of WNV in Europe

		No. positive	No. tested	% (95% CI)	D.F.	I <sup>2</sup> (%)
Region	Mediterranean	2327	17,244	9 (5–14)	24	99.1
	Non-Mediterranean	438	10,845	7 (3–14)	13	99.0
Sampling period	2000–2009	1953	9788	8 (2–17)	12	99.4
	2010–2018	521	14,327	7 (4–10)	19	98.3
Overall		2765	28,089	8 (5–12)	38	99.3

## Figure legends

**Figure 1.** Flow diagram of article selection for West Nile prevalence in equids in Europe

**Figure 2.** Map showing European countries for which data were included in the systematic review. Created using <https://mapchart.net/europe.html> with different colour shading used for Mediterranean and non-Mediterranean countries and depth of shading indicating number of studies performed in each country.

**Figure 3.** Forest plot showing the pooled estimated seroprevalence (ES) of West Nile virus among equids in Europe. Horizontal lines represent 95% confidence intervals (CIs). Each square box denotes the seroprevalence rate point estimate and the area is proportional to the weight of the study.

**Figure 4.** Funnel plot of standard error by Freeman-Tukey double arcsine transformed proportion for all studies (n=39)