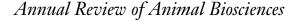
# ANNUAL REVIEWS



Reproductive and Metabolic Health Following Exposure to Environmental Chemicals: Mechanistic Insights from Mammalian Models

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## **Keywords**

environmental chemicals, reproduction, metabolic health, animal models, epigenetics, transgenerational

#### **Abstract**

The decline in human reproductive and metabolic health over the past 50 years is associated with exposure to complex mixtures of anthropogenic environmental chemicals (ECs). Real-life EC exposure has varied over time and differs across geographical locations. Health-related issues include declining sperm quality, advanced puberty onset, premature ovarian insufficiency, cancer, obesity, and metabolic syndrome. Prospective animal studies with individual and limited EC mixtures support these observations and provide a means to investigate underlying physiological and molecular mechanisms. The greatest impacts of EC exposure are through programming of the developing embryo and/or fetus, with additional placental effects reported in eutherian mammals. Single-chemical effects and mechanistic studies, including transgenerational epigenetic inheritance, have been undertaken in rodents. Important translational models of human exposure are provided by companion animals, due to a shared environment, and sheep

exposed to anthropogenic chemical mixtures present in pastures treated with sewage sludge (biosolids). Future animal research should prioritize EC mixtures that extend beyond a single developmental stage and/or generation. This would provide a more representative platform to investigate genetic and underlying mechanisms that explain sexually dimorphic and individual effects that could facilitate mitigation strategies.

#### 1. INTRODUCTION

Over the last 50 years, a decline in human reproductive and metabolic health has been seen across the world. Exposure to the complex low-level mixtures of anthropogenic chemicals that are ubiquitous within the modern environment appears to be a contributory factor. The structural similarity of some of these environmental chemicals (ECs) to endogenous hormones allows them to directly activate and/or block endogenous hormone receptors, resulting in the modification/disruption of normal endocrine regulation health (1, 2). Where the structure of an EC resembles a sex hormone, or where the actions of a hormone are sex specific, these direct effects of ECs can be sexually dimorphic. ECs can also impact health indirectly, through pathways that alter reproductive and metabolic processes such as oxidative stress and DNA damage, apoptosis, inflammation, and epigenetic regulation, and again, such effects can be sexually differentiated. Human epidemiological studies have provided extensive evidence on the effects of exposure to certain ECs and some EC mixtures (e.g., cigarette smoking) and the developmental programming of long-term health. However, the nature of everyday EC exposure is complex and ever changing, which poses great challenges in determining exposure risk. To address this, animal models have been used to study the effects of both single and mixed EC exposure on various health outcomes. This review critiques key studies that have used animals to gain insights into mechanisms (including epigenetics) through which ECs can alter metabolic and reproductive health, and how animal models can be used to address unanswered questions relating to effects of exposure to low-level EC mixtures.

#### 2. OVERVIEW OF EC EXPOSURE

The environment contains a plethora of ECs from various sources. These include the socalled persistent organic pollutants (POPs), such as the pesticide dichlorodiphenyltrichloroethane (DDT); polychlorinated biphenyls (PCBs, which were used as heat-transfer fluids and as dielectric and coolant fluids); polyfluoroalkyl substances (PFAS, found in firefighting foams and textiles); polybrominated compounds, e.g., polybrominated diphenyl ethers (PBDEs, often used as flame retardants); dioxins (a by-product of the manufacturing of pesticides and construction materials); and polychlorinated dibenzodioxins (e.g., polychlorinated dibenzo-p-dioxins, generated as a consequence of the incomplete combustion of chlorine-containing substances) (3,4). Other ECs include organophosphate pesticides (e.g., parathion and malathion); polycyclic aromatic hydrocarbons (PAHs) from combustion of fossil fuels; plasticizers such as bisphenol A (BPA) and phthalate esters, for instance, diethylhexyl phthalate (DEHP) (5); pharmaceuticals (e.g., antibiotics, chemotherapeutics, psychotropics); and natural and synthetic hormones (6-8). Once released, ECs can (a) become widely distributed throughout the environment as a result of natural processes, (b) persist for exceptionally long periods of time (even after their production and use has been discontinued), (c) accumulate in living organisms including humans, (d) bioaccumulate to greater concentrations at higher trophic levels, and (e) induce toxic effects on both humans and wildlife.

Since the 1950s, more than 140,000 new chemicals have been produced, and the number and quantity of anthropogenic chemicals released into the environment have increased simultaneously.

ECs are readily detectable in air, water, and soil (7), and although their distribution patterns show some geographical variability, typically due to local regional prohibition of their production and use, most countries show contamination from a wide but similar range of industrial/anthropogenic chemicals (8). The exposure of humans and domestic animal and wildlife species to a mixture of ECs is, therefore, inevitable. Despite a global EC monitoring plan initiated to monitor worldwide trends in the concentrations of POPs listed under the Stockholm Convention (2, 9), there is little or no environmental assessment of the singular or, potentially more importantly, additive/ cumulative effects of complex EC mixtures that mirror real-life EC exposure. Traditionally, chemical risk assessments are based on the no-observed-adverse-effect level (NOAEL), which is determined by testing the effects of individual chemicals in rodents or cell-based assays. Although NOAELs are invaluable for individual chemical toxicological assessment, they can be orders of magnitude higher than that of human EC exposure. Furthermore, the toxicological profile of a mixture of ECs cannot be determined from NOAELs of each component chemical in the mixture. Studies of the potential risk posed by exposure to mixtures of ECs have used mathematical models based on the individual chemicals within a mixture (10). However, they typically do not account for synergistic or antagonistic interactions between chemicals and the cumulative/additive/synergistic effects of ECs present in mixtures that are below the NOAEL.

#### 3. GLOBAL METABOLIC AND REPRODUCTIVE HEALTH ISSUES

Over the last 50 years, there has been a striking increase in global obesity (9, 11), and by 2025, 20% of the world population is predicted to be obese. In association with this obesity pandemic, there has been a dramatic increase in the incidence of metabolic syndrome (MetS), which is an associated cluster of clinical metabolic symptoms including increased blood pressure, high blood sugar, lower abdominal fat deposition, and abnormal cholesterol or triglyceride levels (12–15). MetS affects an estimated ~35% of adults and 50% of those aged 60 years or older in the United States (16). MetS is a societal health concern, as it predisposes individuals to insulin resistance, hyperglycemia, dyslipidemia, and cardiovascular disease (CVD) and dramatically increases the risk of type 2 diabetes, metabolic dysfunction—associated steatotic liver disease (MASLD), stroke, chronic kidney disease, and certain cancers, all of which have major health and economic implications (17, 18). Of the component conditions, diabetes is of note, as it alone affects ~9% of the world's population (>422 million adults), and in the United States 30 million adults (9.4% of the population) are estimated to be diabetic, with a further 84 million (34% of population) being prediabetic (16).

These dramatic changes in metabolic health are occurring alongside a parallel and alarming decline in human fertility/fecundity and increase in reproductive health problems (19–21). The human fertility rate declined from 5 to 2.5 births per woman between 1960 and 2015 (22), and worldwide an estimated 60–80 million couples seek medical help with reproduction (23). In the United States, the percentage of women of reproductive age that have difficulty becoming pregnant has risen from 8% to 12% since the early 1980s (24). The problems do not lie predominantly within one sex, as both male (25) and female (24) fertility are in decline. In women, the incidence of ovarian dysgenesis syndrome (26), which encompasses premature ovarian failure, ovarian cancer, polycystic ovarian syndrome (PCOS), and reduced fertility, has risen dramatically (27). In men, sperm counts declined globally by up to 50% between 1938 and 2011 (28, 29). A cross-sectional population-based study also reported that semen quality in 25% of Finnish and 35% of Danish men (30) has fallen below the World Health Organization reference for normal fertility (30–55 × 10<sup>6</sup> sperm/mL) (31). These changes in sperm quality/output are occurring alongside increased congenital male reproductive malformations (e.g., cryptorchidism and hypospadias) and

testicular cancer, which exist under the umbrella of testicular dysgenesis syndrome (TDS), the reported incidence of which has increased over the last 50 years, particularly among men of European descent (31).

The etiology of impaired metabolic and reproductive health is undoubtedly multifactorial. Socioeconomic factors, diet, and a sedentary lifestyle impact metabolic health (32), and societal changes, such as increased maternal age at first birth (33), are likely to contribute to the observed decline in fertility. The interrelationship between adverse metabolic and reproductive outcomes [for example, the association between MetS and both decreased male (34) and female (35) reproductive health and fertility] and the link between MetS and the most common form of female reproductive endocrine infertility, PCOS (36, 37), beg the question of whether common physiological mechanisms underlie or link changes in reproductive and metabolic health.

### 3.1. EC Exposure and Declining Human Metabolic Health

Baillie-Hamilton (38) reported associations between exposure to ECs found in pesticides, solvents, plastics, and flame retardants and increased weight gain. Additional ECs have been recognized subsequently to alter hormonal pathways that regulate lipid metabolism, stimulate adipocyte differentiation, and predispose individuals to obesity and/or related metabolic disorders. Such ECs can be classified as obesogens and/or metabolism-disrupting chemicals (39-41). In 2015, a review published by the Endocrine Society found strong evidence for ECs' role in the etiology of metabolic diseases (42), and several subsequent epidemiological studies have provided intriguing links between ECs and metabolic disease. For example, diabetes has been linked to DDT and its metabolite DDE, dioxins, PCBs, and BPA exposure (43-45). The latter chemical has also been associated with obesity and insulin resistance (46) and MASLD (47). Phthalates (e.g., DEHP) have also been linked to MetS (48-50). Intriguingly, exposure to some ECs, including phthalates (51), BPA (52), multiple PCB congeners (53), PFAS chemicals (54), and organochlorine pesticides (55), may also contribute toward increased CVD risk. In terms of everyday exposure to ECs, recent evidence has highlighted air pollution as a possible contributory factor to metabolic diseases. Air pollutants include mixtures of ECs such as benzo(a)pyrene (the main marker of PAH presence), PCBs, sulfur dioxide, nitrogen dioxide, carbon monoxide, organic compounds (organic solvents and dioxins), and heavy metals, often produced by transport and industrial processes. Air pollution is associated with diabetes, CVD, and MASLD (56, 57). Together, these studies suggest that a diverse array of ECs may individually, or collectively, play a significant role in the programming and pathophysiology of various metabolic diseases.

# 3.2. EC Exposure and Declining Human Reproductive Health

Several specific ECs have been linked with female reproductive health problems (58), TDS, and a decline in male reproductive function (42, 59). Estrogenic ECs (60) such as bisphenols (e.g., BPA) (61) and per- and polyfluorinated alkyl substances (PFAS) (62) are directly associated with breast cancer; however, causality has not been established (63). EC exposure has also been indirectly linked with breast cancer risk via effects on other aspects of female reproductive health, including precocious onset of puberty (60). Although female infertility is a major and growing concern, due to factors such as advanced reproductive age, 15–30% of cases remain unexplained. However, recognized contributory factors such as premature ovarian insufficiency, endometriosis, and PCOS have all been associated with EC exposure (64–67). Exposure to phthalate esters variably affects puberty onset; some studies have shown premature thelarche (68) and early/precocious puberty in girls (69–71), whereas others report delayed pubarche without thelarche (72) or no effect (73).

As with the effects in females, the relationship between adult EC exposure and male reproductive health and function is similarly complex. For example, BPA exposure has been negatively associated with sperm concentration, sperm motility, and total sperm count (74–76). Phthalate [DEHP, di(*n*-butyl) phthalate (DBP)] exposure has an inverse relationship with anogenital distance (AGD) (77); a negative association with sperm concentration and sperm motility (78); a positive association with semen volume, progressive motility (78), and anopenile distance (79); and no association with semen quality (80). Serum concentrations of PCB congeners PCB-118 and PCB-77 have both negative and positive associations with semen volume and progressive motility (81). DEHP and PCB-153 reduce human sperm motility and increase DNA fragmentation (82), and PBDEs are both without effect on semen parameters (80) and negatively associated with sperm concentration, total sperm count, and progressive motility and viability (83).

Fertility in both sexes has been reported to be negatively impacted by air pollution (84), potentially due to ECs such as benzo(a)pyrene (the main marker of PAH presence), PCBs, sulfur dioxide, nitrogen dioxide, carbon monoxide, organic compounds (organic solvents and dioxins), and heavy metals, which are produced by transport and industrial processes. In males, industrial air pollution is associated with decreased fertility (85, 86) and reduced sperm quality (number and motility) (84, 87–89). In females, air pollution can lead to reduced odds of positive in vitro fertilization pregnancy outcomes (90) and is associated with increased risk of stillbirth (91). Additionally, exposure to particulate matter, also found in air pollution, is associated with reduced fecundability (92), decreased antral follicle count, and anti-Müllerian hormone levels, suggesting a negative impact on ovarian reserve (93). However, as with many of the studies of the effects of individual ECs, the relevance of these studies relative to real-life EC exposure is limited by heterogeneity among studies, low numbers of participants, the small numbers of pollutants analyzed, and analytical methods.

# 3.3. Programming of Metabolic and Reproductive Health Outcomes by Developmental EC Exposure

The concept that the early developmental environment can program adult disease is well established (94) and supported by both human epidemiological and animal studies (95–98). Research into the developmental origins of health and disease (DOHaD) has focused largely on maternal nutrition and/or stress experienced during pregnancy and consequences for offspring cardiovascular and metabolic health (99–103). Studies also indicate that reproductive development and function can be affected (104). DOHaD-related health effects may be detected in the fetus and persist until adulthood. Other effects may not become apparent until adulthood. Irrespective of timing, these effects usually occur due to epigenetic programming (103, 105) (discussed below). Emerging evidence also suggests that poor metabolic and reproductive health arise as a consequence of events experienced by our parents, grandparents, or even earlier generations (106, 107).

A well-publicized example of EC-induced DOHaD relates to the gestational exposure of women to the synthetic estrogen diethylstilbesterol (DES), as their offspring have a high lifetime risk of a range of adverse health outcomes including breast cancer (108). Basic clinical and epidemiological evidence also suggests that TDS may often originate during fetal life (109) as a consequence of in utero exposure to ECs (110). Although establishing the causation of EC exposure effects on health can be challenging in human studies, some of the most compelling evidence comes from cases where developmental EC exposure has occurred following an occupational or environmental accident. For example, men who were perinatally exposed to dioxin in Seveso, Italy (111), or prenatally exposed to PCBs and polychlorinated dibenzofurans in Taiwan (YuCheng accident) (112) exhibited reduced semen quality as adults. Other studies have reported increased

risks of genital malformations in children of workers occupationally exposed to pesticides (113), and high incidences of cryptorchidism are observed in geographical regions with intensive agriculture (114) or high levels of industrialization (115). Furthermore, maternal phthalate exposure is associated with impaired Leydig cell function (116). Developmental exposure to obesogenic ECs has also been linked to latent effects on metabolic health (117, 118). For example, maternal exposure to BPA and phthalates resulted in altered offspring growth and body mass index in childhood (119), and developmental perfluorooctanoic acid exposure may reduce fetal growth (120).

Finally, women who smoke during pregnancy provide a natural human paradigm to determine the latent effects of exposure to EC mixtures. Cigarette smoking increases the risk of various adverse pregnancy outcomes such as miscarriage, placenta previa, preeclampsia, and premature delivery (121). Importantly, chemicals contained in cigarettes can cross the placenta and have a negative impact on the fetus at the genetic and cellular level. Consequently, maternal smoking during pregnancy has increased our understanding of the long-term effects of early-life exposure to EC mixtures on offspring health. Longitudinal studies that have followed the offspring of mothers exposed to cigarette smoke have demonstrated that it affects reproductive and metabolic health of offspring both as children and as adults. For example, males whose mothers smoked during pregnancy have reduced semen quality and testis size in adulthood (122, 123), and female offspring exhibit altered ovarian development (124) and reduced fecundability (125). In parallel, maternal smoking is linked to intrauterine growth restriction and low birth weight for gestation age (126), as well as childhood and adult obesity (127, 128) and CVD risk in later life (129, 130). Given that exposure to individual chemicals (e.g., perfluorooctanoic acid) (131) and chemical mixtures (cigarette smoke) alters fetal growth, and that intrauterine growth restriction is a known risk factor for CVD in later life according to DoHaD, it is possible that the mechanisms that underlie fetal programming of health are sensitive to EC exposure as well as other maternal factors and that these share common features.

In summary, there is some compelling evidence linking human exposure to ECs, either as adults or during development, with alterations in metabolic and/or reproductive health. Many studies have focused on either accidental or occupational exposure to high levels of ECs or have reported associations between individual or limited mixtures of chemicals and metabolic and reproductive outcomes. However, by their very nature, such studies are associative and do not establish causation. Outcomes have also been variable and sometimes confounded by small study populations and other lifestyle variables. To complement human studies, therefore, use of prospective, controlled animal models of individual and mixed EC exposure has been necessary.

#### 4. MAMMALIAN MODELS FOR THE STUDY OF ECs

Animal models provide opportunities to assess the effects of dose and timing of EC exposure and to interrogate mechanisms of action of individual and combined ECs across the life course. Even with such paradigms, however, modeling real-life mixed EC exposure remains challenging. Importantly, EC exposure and effects of EC exposure may be influenced by species-specific differences in absorption, metabolism, and elimination of ECs, and in the duration of developmental stages (e.g., differences in gestation length), which affects the relative duration of periods of EC exposure and could affect the nature of their effects. The use of multiple models, and comparison across animal models, allows the individual strengths of each model to be utilized and accounted for and significantly advances our understanding of the possible health effects of EC exposure.

# 4.1. Companion Animals as Sentinel Species

Because companion animals share our home environment, cats and dogs could be used as sentinel species for the effects of human EC exposure (132, 133). As with humans, companion animal EC

exposure occurs through food, air, dust, and water and has been linked to both adverse reproductive and metabolic outcomes (133). In cats, high levels of ECs such as PBDEs (flame retardants) are thought to result from contact and grooming and have been linked with thyroid dysfunction (134), metabolic disorders, and cancer (135), whereas other ECs, such as PFAS, have been linked to obesity (136). In dogs, a 30% decline in progressive sperm motility has been reported over a 26-year period alongside an increased incidence of cryptorchidism (137, 138). These canine data are of note because they were obtained from a single laboratory and thus lack some of the inherent confounders in human meta-analyses of the effects of ECs on TDS. In addition, ECs have been detected in dog testes at concentrations able to inhibit sperm motility (dog and human), and as with humans, testicular EC profiles vary geographically (82, 139). Some comparative studies done with companion animal species have reported that metabolic disease can be linked with high PFAS concentrations in both cats and dogs (133).

# 4.2. The Sheep Biosolids Sentinel and Animal Model

Another animal model that reflects human EC exposure is the biosolids-exposed sheep model. Biosolids is the solid waste generated in wastewater treatment plants. It is a nutrient-rich organic matter and is used globally as an agricultural fertilizer. Indeed, biosolids use is recommended by the US Environmental Protection Agency and through the European Sewage Sludge (biosolids) Directive. Approximately 55% of the 18 million dry metric tonnes of treated sewage sludge produced annually in the United States is applied to agricultural land (140). Because biosolids originates from human activity, it contains a broad mixture of ECs, including those derived from personal-care products, pharmaceuticals, pesticides, flame retardants, detergents, BPA, PCBs, PBDEs, PAHs, organochloride pesticides, and pharmaceuticals and ECs in waste from industrialized manufacturing processes (141-145). Application of biosolids to land results in increased EC concentrations in soil (143-145) and in tissues and blood samples collected from sheep grazed on biosolids-treated pastures (146-149). It therefore constitutes an exposure paradigm that reflects the human exposome and has inadvertently provided an excellent model of real-life human EC exposure. Over 25 years, this model has been used in a series of studies investigating the impact of gestational exposure to EC mixtures across different life stages (fetal, neonatal, prepubertal, adult) of the offspring. Initial investigations focused on the reproductive axis (150-158), but studies have also documented effects on behavior (159), bone density (160), thyroid structure (160), liver function (161), and more recently metabolic parameters in both exposed mothers and their offspring (162-164).

Relative to reproductive health, exposure of male fetuses to ECs by maternal grazing on biosolids-treated pasture either throughout or for an 80-day period during early, mid-, or late gestation results in an abnormal testicular phenotype, a characteristic of which is reduced numbers of Sertoli cells (158). The developmental and transcriptomic changes observed in gestational day (GD) 140 male fetuses indicate an overall antiandrogenic effect (165, 166). In female fetuses, GD80 and GD110, there is a reduction in transitional ovarian follicles (158, 166), and EC exposure for 80 days during mid- and late gestation appears to result in greater effects, with animals exhibiting a reduced AGD indicative of an overall androgenization effect (167–169). Male and female fetal lambs exposed to ECs via biosolids, either throughout or during the final third of gestation, have altered expression of drug-metabolizing genes. This could account for greater effects of transient versus continuous exposure across gestation observed in various outcomes in this model (166, 170). EC exposure via biosolids in both male and female fetuses (GD140) also alters the gonadotropin-releasing hormone neurosecretory system along with other key neuroendocrine regulators (e.g., kisspeptin, galanin receptors) in the hypothalamus and pituitary, thus illustrating hypothalamic-pituitary–gonadal axis sensitivity at all three levels (152, 153, 168). When considering fetal effects

of ECs, these could be mediated indirectly through the mother, as EC exposure induces changes in the maternal metabolome, in her steroid profile, and in the expression of mediators of oxidative stress (155, 171).

In later life, pubertal timing is affected in lambs born to mothers exposed to biosolids immediately prior to and through gestation; it is advanced in male but delayed in female offspring (164). Furthermore, when the EC exposure period was extended by the grazing of lambs exposed during gestation through to puberty, a subset of the adult males exhibited testicular abnormalities, including Sertoli cell–only tubules and reduced numbers of germ cells (162, 163). Beyond puberty, 11-month-old (adult) male offspring exposed in utero exhibited two metabolic profiles indicative of different degrees of susceptibility to EC exposure (150, 157). In summary, these extensive studies illustrate that exposure to biosolids during gestation adversely affects fetal, neonatal, pubertal, and adult reproductive and/or metabolic function (**Figure 1**).

#### 4.3. Rodents

Historically, most EC exposure studies have used rodent models (172). This reflects their ease of management, short gestation length, controlled genetics, the capacity to explore chemical effects at the population level, transgenerational studies, and the use of transgenic and knockout mice

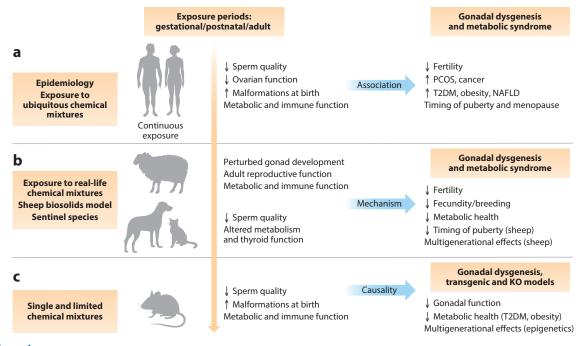


Figure 1

Mammalian models for the assessment of EC effects on reproductive and metabolic function. (a) Human studies are largely epidemiology based; temporal trends in gonadal dysgenesis and metabolic syndromes have been reported. (b) Real-life chemical effects can be explored in a highly characterized sheep model of exposure to EC mixtures present in biosolids derived from human sewage/ effluent. Companion animal sentinels (i.e., dog, cat) reflect reproductive and metabolic changes in the human. Both models have shown a decline in fertility, fecundity, and puberty timing (sheep), and multigenerational studies in sheep are ongoing. (c) Rodent models of gonadal dysgenesis, transgenics, and KO strains are excellent tools to investigate causality, including multigenerational effects. Abbreviations: EC, environmental chemical; KO, knockout; NAFLD, nonalcoholic fatty liver disease; PCOS, polycystic ovarian syndrome; T2DM, type 2 diabetes mellitus.

to explore the role of EC-sensitive genes in biological pathways. Despite some obvious caveats around the use of polytocous rodents when extrapolating EC impacts to humans, the contribution of rodent studies to the understanding of EC action is substantial.

Given that EC exposure is associated with TDS in humans (see Section 3), some earlier studies focused on mechanisms underlying a comparable TDS phenotype in a murine model. Landmark studies identified an early programming window (gestational days 15.5–19.5) in male rats, which corresponds to the period between 8 and 14 weeks in humans, as during this period exposure of rats to the antiandrogen flutamide induces both cryptorchidism and hypospadias (173), characteristics of the TDS phenotype widely reported in the human. Intriguingly, exposure of the female rat fetus to androgens during this window increased AGD length comparable to that in males, illustrating a similar masculinization programming window to androgens in utero. In humans, TDS is associated with reduced AGD. Notably, in the rat, exposure of the male fetus to the plasticizer DBP is associated with reduced testosterone and AGD only when exposure occurs during the equivalent androgenization window outlined above (165, 174, 175). Rodent models have, therefore, allowed us to interrogate the effects of ECs and EC mixtures during periods of known developmental sensitivity (165, 176).

Rodent models have also been used to identify developmental periods of sensitivity and potential underlying mechanisms involved in premature ovarian insufficiency, reduced fertility, PCOS, and endometriosis. Because the timing of fetal ovarian development in the rodent differs to that in the human, with early follicle development occurring postpartum, this presents some experimental opportunities, but with caveats. Such models provide access to follicle development stages that occur prenatally in the human. However, the route of EC exposure is via placental transfer in humans but is predominantly oral in postnatal rodents. Complementary in vitro approaches using excised and cultured ovaries to study the effects of mixed EC exposure on follicle development have been underused.

Rodents have also been instrumental in determining EC effects linked to metabolic dysfunction (177), obesity and type 2 diabetes mellitus, and MetS (178, 179). One specific example is that of perinatal BPA exposure, which has been shown to induce changes in DNA methylation (candidate gene and global). Specifically, pathway analyses indicate changes in DNA methylation of metabolic and neural signaling genes (179). Knockout mice provide ideal tools for elucidating reproductive or metabolic mechanisms. For example, comparisons of Ahr<sup>+/+</sup> to Ahr<sup>-/-</sup> mice clearly demonstrate the adverse consequences of dioxin exposure on liver metabolism (177). That is, hepatotoxicity was dependent on the presence of the Ahr receptor. Endocrine-/metabolism-disrupting chemicals can interfere with several additional signaling pathways for which knockout mice are available. These include the estrogen, retinoid X, androgen, and several other receptor-mediated pathways (180). Thus, rodents are ideal experimental tools to delineate molecular mechanisms associated with effects of ECs on reproduction and metabolism (**Figure 1**).

#### 5. MECHANISMS OF ACTION

The deleterious effects of ECs on both metabolic and reproductive function can be mediated centrally (i.e., within the hypothalamic–pituitary axis) and/or locally (i.e., within specific tissues and organs). For example, the homeorhetic regulation of basal metabolic rate, appetite, and energy metabolism by thyroid hormones is mediated both centrally and peripherally (including the gravid uterus) (181) (**Figure 2**). The consequences of in utero exposure and their contributions to the DOHaD effects of ECs are discussed earlier in this article. EC-disrupted thyroid hormone–directed signaling in the liver largely affects lipid and glucose metabolism (182), whereas perturbed thyroid hormone–directed signaling in the pancreas mostly influences

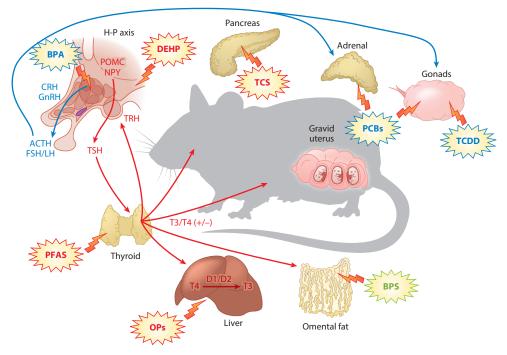


Figure 2

Targeted effects of ECs impact multiple organ and tissue systems, including the gravid uterus in pregnant females. Depending on exposure timing and duration, these effects can be limited to founder (F0) individuals or extend to F1 and subsequent generations following in utero exposure. Illustrative examples are provided firstly for effects of specific ECs on the regulation of pleiotropic thyroid hormones (181) involved in the homeorhetic and homeostatic regulation of cellular metabolism and appetite. These effects can occur centrally (e.g., DEHP) (264, 265), acting on neural networks (e.g., POMC, NPY) controlling food intake or on TRH, which regulates the pituitary release of TSH. ECs can also act directly on the thyroid [e.g., PFAS (266)] to affect T4 and T3 production or locally at target organs and tissues [e.g., OPs in the liver (182) and TCS in the pancreas (183)]. Importantly, many of these local effects include the dysregulation of iodothyronine deiodinases (e.g., D1 and D2) that coordinate T4-to-T3 conversions in a tissue-specific manner. Also depicted are EC effects (e.g., BPA) on the hypothalamic-pituitary production and release of gonadotrophins (FSH/LH) (267) and ACTH (268), which act on gonads and adrenal glands, respectively, to disrupt function and dysregulate steroid production. ECs can also have direct effects on tissues, which, although apparent from in vivo studies [e.g., effects of dioxins (TCDD) on ovarian function (185)], are best demonstrated by either cell or organ culture experiments [e.g., PCBs on gonadal and adrenal cells (269) and BPS on adipocytes (270)]. Blue arrows: H-P hormone actions on reproductive tissues. Red arrows: target organs/tissues affected by pleiotropic thyroid hormones. Bursts depict action of environmental chemicals, which can be either direct or indirect (acting via the H-P axis or the thyroid gland). Abbreviations: ACTH, adrenocorticotrophic hormone; BPA, bisphenol A; BPS, bisphenol S; CRH, corticotropin-releasing hormone; DEHP, diethylhexyl phthalate; EC, environmental chemical; FSH, follicle-stimulating hormone; GnRH, gonadotropin-releasing hormone; H-P, hypothalamic-pituitary; LH, luteinizing hormone; NPY, neuropeptide Y; OPs, organophosphates; PCBs, polychlorinated biphenyls; PFAS, polyfluoroalkyl substances; POMC, proopiomelanocortin; T3, triiodothyronine; T4, thyroxine; TCDD, 2,3,7,8-tetrachlorodibenzo-p-dioxin; TCS, triclosan; TRH, thyrotropin-releasing hormone; TSH, thyroid-stimulating hormone.

glucose-stimulated insulin secretion (183). Likewise, ECs can affect both positive and negative feedback mechanisms operating within the hypothalamic–pituitary–gonadal axis to affect reproductive development/function (152, 153) as well as autocrine and paracrine mechanisms within gonads that regulate steroidogenesis and germ-cell maturation (184, 185) (**Figure 2**).

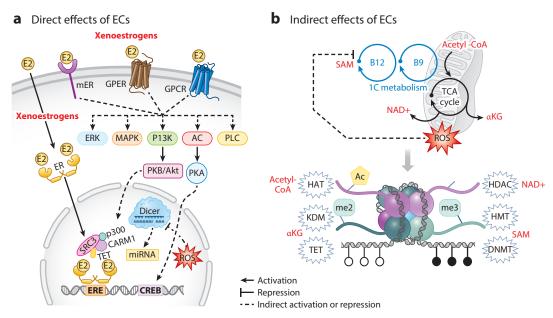


Figure 3

Examples of (a) direct and (b) indirect modes of action for ECs in the epigenetic regulation of gene expression. (a) Direct actions of ECs are best characterized for nuclear receptor superfamily members; the example presented is for the ER depicting genomic and nongenomic signaling related to transcriptional activation. Xenoestrogens (e.g., BPA, DES) can substitute for estrogen binding to cytoplasmic/nuclear receptors at gene-promotor ERE and could thus epigenetically alter the coordinated recruitment of chromatin modifiers and coactivators (e.g., SRC3, p300, CARM1, and TET) required for transcriptional activation, further modified by the actions of miRNAs (e.g., miR-494). Nongenomic epigenetic actions of xenoestrogens can be mediated by any combination of three membrane-bound receptors (i.e., mER, GPER, or GPCR) to permanently alter intracellular protein kinase cascades involving ERK, MAPK, P13K, AC, and/or PLC signaling. The relative contribution of these pathways and details regarding EC-induced heritable changes in chromatin configuration related to estrogen signaling are the subject of ongoing investigations by several groups. (b) Indirect actions of ECs can arise due to modifications in intermediary metabolism that alter the availability of metabolites that serve as substrates or cofactors for enzymes involved in chromatin modification. These include acetyl-CoA-dependent HATs, NAD+-dependent HDAC (e.g., Sirtuins), SAM-dependent HMT and DNMT; and αKG-dependent histone (KDM) and TET demethylases (271, 272). Many genes encoding enzymes involved in 1C, as well as associated epigenetic regulators, are modulated either directly or indirectly by androgen or estrogen receptors (as depicted in panel a) (202). Exposure to ECs such as heavy metals and EDCs can also lead to the excessive generation of ROS, which, in turn, can (i) induce DNA breaks and dysregulate associated base-repair mechanisms; (ii) directly impede/alter chromatin modification (e.g., carbonylation and glutathionylation) as well as perturbing 1C metabolism; and (iii) inhibit Dicer activity, (a) thus impairing miRNA maturation (217). Open and solid lollipops represent unmethylated and methylated CpGs, respectively, on DNA. Abbreviations: αKG, α-ketoglutarate; 1C, one carbon; B9, folate; B12, cobalamin; BPA, bisphenol A; CARM1, coactivator-associated arginine methyltransferase 1; DES, diethylstilbesterol; DNMT, DNA methyltransferase; EC, environmental chemical; EDC, endocrine-disrupting compound; ER, estrogen receptor; ERE, estrogen-response element; ERK, extracellular signal regulated kinase; GPCR, G protein-coupled receptor; GPER, G protein-coupled estrogen receptor; HAT, histone acetyltransferase; HDAC, histone deacetylase; HMT, histone methyltransferase; KDM, lysine demethylase; MAPK, mitogen-activated protein kinase; miRNA, microRNA; NAD, nicotinamide adenine dinucleotide; P13K, phosphoinositide 3-kinase; PKA, protein kinase A; PKB, protein kinase B; PLC, phospholipase C; ROS, reactive oxygen species; SAM, S-adenosylmethionine; SRC3, steroid receptor coactivator-3; TCA, tricarboxylic acid; TET, ten-eleven translocation.

One mode of action of ECs is to mimic the behavior of natural hormones. Xenoestrogens such as BPA, for example, mimic the effects of estradiol by binding to both nuclear (i.e., estrogen, androgen, and glucocorticoid) and membrane (i.e., GPER) receptors (**Figure 3**). In so doing, they interact with several transcription factors (e.g., PPAR $\gamma$ , C/EBP) to alter adipose and liver homeostasis (186). The antiandrogen effects of BPA also operate in this way, leading to impaired testicular

#### **EPIGENETIC PROGRAMMING**

The term epigenetics refers to heritable, yet reversible, changes in gene function that arise in the absence of an alteration to DNA sequence. These changes relate principally to covalent modifications in DNA and associated proteins that are directed to an extent by noncoding RNAs (197, 198). Heritable in this context relates to the mitotic transmission of such modifications from parent to daughter cells during cell division. It follows that epigenetic processes are central to directing cell fate and lineage determination during postfertilization development. However, as discussed later, there is some debate concerning the extent to which modifications can be propagated through meiosis (i.e., intergenerationally) in mammals due to the double wave of epigenetic reprogramming that occurs during early embryogenesis and later in primordial germ cells (199). Notwithstanding, sweeping epigenetic modifications occur around the time of conception when, during syngamy, two terminally differentiated cells (i.e., the sperm and egg) give rise to a totipotent zygote. Epigenetic memory is important for guiding cell fate during later developmental stages, and this is achieved partially through preference for concordant DNA methylation (200). However, early embryonic cells operate with reduced fidelity of methyl-group transfer, rendering them more susceptible to environmentally induced perturbations in DNA methylation. Consequently, it has been argued that the periconceptional period is the development stage most sensitive to potential modifications due to extraneous factors such as environmental chemicals (201, 202), although recognized later stages of development [e.g., the masculinization programming window (165)] may also be epigenetically sensitive to specific agents.

physiology and reduced spermatogenesis. ECs also act indirectly by increasing oxidative stress and DNA damage, leading to apoptosis. Many serve as oxidants and impact on mitochondrial function, generating reactive oxygen species and other reactive compounds (187). Oxidative stress during pregnancy can lead to birth defects (188), impairing reproductive function, cardiometabolic health, and vascular dysfunction in CVD (189–191). Apoptosis and inflammation are hypothesized to play a role in these processes. For example, the apoptotic/inflammation regulatory molecule NFkB has been implicated in both testicular germ cell apoptosis (192, 193) and inflammation involved in endothelial damage and atherosclerosis (194) in response to chemical exposure. However, more fundamental mechanisms of action underpinning the harmful effects of ECs pertain to the epigenetic regulation of gene expression, in particular, long-term (including transgenerational) adverse developmental effects observed following EC exposure during pregnancy and/or infancy.

# 5.1. EC-Induced Epigenetic Effects

A series of systematic reviews published in the last decade have summarized comprehensively the epigenetic consequences of EC exposure at different stages of development (i.e., in utero, during infancy, and in adulthood) (172, 173, 175, 195, 196; see the sidebar titled Epigenetic Programming). These reviews indicated that most studies related to humans and rodents and the reported effects were associative in nature (particularly in humans). Changes in DNA methylation were the most frequently investigated epigenetic modifications, although recent studies have begun to explore associations with noncoding (nc)RNAs (particularly small ncRNAs) and post-translational modifications to histones. Despite heterogeneity between cited studies, EC exposure, including air pollution and cigarette smoking during pregnancy, along with an extensive list of recognized ECs including heavy metals, POPs (e.g., dioxins, PCBs), and known carcinogens, consistently induced epigenetic modifications to chromatin in a range of cell and tissue types.

More recently, there has been an attempt to identify homologous DNA methylation reprogramming following lead and phthalate (DEHP) exposure in mice and humans, with the aim of enhancing the translational value of mice in toxicology studies (203). Mouse databases originating

from the National Institute of Environmental Health Sciences-sponsored TaRGET II program (204) were compared to four human cohorts with previously published DNA methylation data. In mice, lead and DEHP exposure during pregnancy and up to weaning led to a significant number of differentially methylated genes (~750) in both male and female offspring, which corresponded to ~35,000 differentially methylated cytosines on the human EPIC array. Although there was little overlap with reported differentially methylated regions in mice, the regions that did overlap included three imprinted loci (i.e., KCNQ1, CDKNIC, and CMTM1) (discussed later). The main conclusion to be drawn from this study is that, although animals serve as great models to establish and understand the epigenetic basis of adverse phenotypic effects following exposure to ECs, they do not represent direct comparators for humans. Instead, the power of animal studies (rodents in particular) lies in their ability to go beyond association analyses to undertake prospective interventional studies (e.g., gene knockout/in), to establish causality and mode of action, and to assess long-term (including transgenerational) epigenetic and phenotypic effects of EC exposure. To this end, transgenic mice have been used to explore aspects of germline epigenetic inheritance in a paternal obesity paradigm (205-207), and the adverse metabolic effects to offspring of feeding high-fat diets to sires were recapitulated when purified small ncRNAs (identified following sperm analyses) were injected into zygotes prior to embryo transfer (208). However, these approaches have yet to be exploited in the context of EC exposure.

# 5.2. Mechanisms Underlying EC-Induced Epigenetic Modifications

ECs could induce epigenetic modifications to chromatin via several means, although they have yet to be investigated fully. From what is known, mechanisms can be categorized as either direct or indirect and, related to this, global or gene specific in nature (209) (Figure 3). However, it is important to distinguish chromatin-based mechanisms that transiently facilitate transcription from those that are genuinely epigenetic; the latter term infers that any modifications to chromatin are heritable (210; see the sidebar titled Epigenetic Programming). Steroid receptors, for example, can transiently activate transcription through recruitment of several coregulators to facilitate histone modification, chromatin remodeling, and general transcription machinery stabilization (211). ECs can induce heritable (epigenetic) modifications to these processes, but the precise mechanisms involved have not yet been established.

A case in point is the estrogen receptor (ER), a subclass of the nuclear receptor superfamily of proteins (212), which in most cases binds directly (or indirectly) via AP-1 or SP-1 transcription factors to estrogen-responsive elements in DNA of target genes (213) (**Figure 3***a*). Upon ER binding, the process of rendering chromatin more accessible to transcription factors generally involves the sequential recruitment of a cohort of chromatin modifiers and coactivators commencing with SRC-3, which recruits p300 (a histone acetyltransferase) and CARM1 (coactivator-associated arginine methyltransferase 1). This is followed by either passive or active demethylation of DNA, the latter step involving TET (ten-eleven translocation) enzymes. A further layer of complexity involves actions of small ncRNAs (e.g., miR-494) that can modulate these processes. However, endogenous estrogens and xenoestrogens (e.g., BPA, DES, PCBs) can also bind to membrane-bound ERs, and so epigenetic modifications to chromatin can be induced via both genomic and nongenomic pathways, although the relative importance of each is uncertain (214). Although the recent development of transgenic mice lacking either membrane or nuclear ERs will provide valuable insights in this regard (215), our understanding of how these events lead to heritable changes to chromatin remains to be determined.

Indirect effects of ECs can be mediated in part through actions on intermediary metabolism operating, for example, in and around the mitochondrion (**Figure 3***b*). This can arise due to

EC-induced oxidative stress and/or modifications in levels of metabolites that serve as substrates or cofactors for enzymes involved in the epigenetic modification of chromatin and/or coding and ncRNAs (216). For example, the excessive generation of reactive oxygen and nitrogen species following EC exposure can interfere with the activity of enzymes involved in chromatin methylation and acetylation, thus inducing DNA damage and activating base-repair mechanisms (217). It can also lead to the oxidation of enzymes involved in one carbon (1C) metabolism or induce excessive glutathione production, thus depleting S-adenosyl methionine and chromatin methylation. Recent examples of indirect effects include exposure to the carcinogenic and neurotoxic element cobalt, which was found to downregulate the expression of the m<sup>6</sup>A demethylase ALKBH5, leading to an enrichment of both hypermethylated and hypomethylated transcripts linked to neurodegenerative diseases (218). Confirming a nonspecific role for 1C metabolism, folic acid supplementation of pregnant rats exposed to a mixture of POPs mitigated observed adverse effects on sperm microRNA profiles in offspring over four successive generations (219).

## 5.3. Genomic Imprinting

Although most mammalian genes are expressed by both parental alleles, a small number of genes [recent estimates for the mouse list  $\geq$ 388 (220)] are expressed by only one allele in a parent-of-origin-specific manner (see the sidebar titled Mechanisms Directing Mono-Allelic Gene Expression). These are referred to as imprinted genes. The importance of genomic imprinting in the context of mammalian development resides in the fact that uniparental embryos generally fail to develop much beyond implantation. Errors in genomic imprinting result in aberrant placental and fetal development in mammals (221, 222) but can also lead to lifelong disorders, including Angelman, Prader–Willi, and Beckwith–Wiedemann syndromes in humans (222) and related large offspring syndrome in cattle and sheep (223–225).

#### MECHANISMS DIRECTING MONO-ALLELIC GENE EXPRESSION

Parent-of-origin-specific gene expression is regulated by the inheritance of differentially methylated imprinting control regions (ICRs), together with other germline differentially methylated regions (DMRs). The establishment of these ICRs/DMRs during gametogenesis is carefully choreographed, with the transient presence of histone variants [e.g., histone 3 lysine 4 di/tri-methylation (H3K4me2/3), H3K27me3, and H3K36me3 (220, 226-228)] operating in cohort with local transcriptional events specific to male and female germlines. These serve to direct allele-specific methylation actioned by the methyltransferase complex DNMT3A-DNMT3L (229). For specific imprinted genes, PIWI-interacting RNAs, along with other short- and long-noncoding RNAs, serve as part of the transcriptional machinery that regulates de novo methylation at DMRs (230). Normally, ICRs/DMRs escape the wave of genome-wide demethylation that occurs during preimplantation stages of embryo development (231). This arises through the combined actions of the maintenance DNA methyltransferase and its partner, ubiquitinlike with PHD and ring finger domains 1, together with two Krüppel-associated box domain zinc-finger proteins. These factors bind specifically to the methylated allele, thus protecting it from demethylation while directing methylation to the nascent allele arising following DNA replication during cell division (232-234). In addition, evidence is emerging for a key role of long-terminal-repeat retrotransposons (LTR) in both canonical and noncanonical (i.e., DNA methylation-independent) imprint establishment during gametogenesis and maintenance in embryonicand somatic-cell lineages, although noncanonical imprinting appears to be transient in nature (228). Nevertheless, LTR-guided imprinting imposes an additional level of complexity, particularly in outbred species such as humans, as these sequences are highly polymorphic and therefore likely to contribute to molecular and phenotypic differences observed between individuals.

Several putative molecular targets for ECs could lead to the erroneous expression of this subset of developmentally important genes. Indeed, rodent studies have identified effects of several ECs, including BPA; phthalates (e.g., DEHP); 2,3,7,8-tetrachlorodibenzo-p-dioxin; and the pesticide vinclozolin on DNA methylation and expression of imprinted genes. However, reported outcomes (including phenotypic) are variable and mechanistic details sparse, focusing mostly on canonical modes of action (reviewed in 235). In these studies, the imprinted genes concerned were preselected and include those for which previous evidence of more general environmentally induced dysregulation in methylation and expression exists (e.g., Igfr2, Peg3, Snrpn, Igf2/H19) (232, 233). Several factors likely contributed to the variability in effects observed, including rodent strain (236), specific ECs concerned, exposure timing (i.e., gametogenesis versus early embryogenesis versus later stages of gestation and/or infancy), dose (pharmacological versus physiological), and route of administration (e.g., oral versus hypodermal). Importantly, to date, most rodent studies investigating EC effects on genomic imprinting have focused on single chemicals or small mixtures of chemicals. Also, it will be necessary to establish the full extent of sex specificity in responses (at both a molecular and physiological level) for these genes and to determine both canonical and noncanonical mechanisms of dysregulated genomic imprinting.

Investigations into the effects of EC-induced perturbations in imprinted genes in large, outbred mammalian species are limited. However, several studies in humans, spanning a wide range of geographical locations (and ethnicities), have reported errors in genomic imprinting following EC exposure during pregnancy (235). Although observational and associative in nature [being mostly linked to cord blood or urinary levels of phthalates or BPA (237–240)], these studies demonstrate that real-life EC-induced errors in genomic imprinting are detectable in human populations. Extending this further, evidence exists for sex-specific effects on DNA methylation at the H19 ICR and IGF2 DMR2 in early second-trimester human fetal livers following in utero exposure to cigarette chemicals (241). Importantly, these sex-specific effects are associated with depleted levels of hepatic cobalt and vitamin B12, together with altered expression of DNMT1 and transcripts for several enzymes involved in 1C metabolism.

# 5.4. Transgenerational Epigenetic Inheritance

Overall, a compelling body of evidence from rodent studies supports the notion of transgenerational epigenetic and phenotypic inheritance following parental exposure to ECs (196, 249) (see the sidebar titled Mechanisms of Germline Epigenetic Transmission). A recent systematic review identified 43 EC-related articles reporting transgenerational epigenetic inheritance in rodents (196). No equivalent human or large animal studies were found in this search. Cited rodent studies involved a range of (mostly) single chemicals with known endocrine-disrupting effects including atrazine, BPA, phthalates, DDT, dioxins, and vinclozolin. F0 exposure to ECs occurred mainly during pregnancy, although a few studies considered F0 male exposure prior to mating, e.g., to methoxychlor (250). Most studies provided evidence of epigenetic transmission via the male germ line, reporting a range of chromatin modifications and altered populations of small ncRNAs in sperm and/or somatic cells. Some studies could confirm only intergenerational inheritance—i.e., F1 paternal [e.g., benzo(a)pyrene (251)] and F2 maternal [e.g., BPA (252)] transmission—whereas others found that differences in DNA methylation and the incidence of cryptorchidism had diminished by F4 [e.g., F0 pregnancy exposure to DEHP (253)].

To the best of our knowledge, there is currently no equivalent evidence of EC-induced transgenerational epigenetic inheritance in large outbred species such as humans and farm animals (196, 197, 254). Such effects were shown to attenuate DEHP-induced transgenerational epigenetic inheritance of TDS in FVB/N relative to C57BL/6J strains of mice (255). Genetic

#### MECHANISMS OF GERMLINE EPIGENETIC TRANSMISSION

A long-held belief was that due to the significant extent of germline-chromatin remodeling that takes place in mammals, epigenetic modifications acquired during the lifetime of an individual are erased and thus not passed on to the next generation. Such a contention adheres to the concepts of evolutionary biologist August Weismann, who distinguished the "immortal" germline from the "disposable" soma. Breaching the Weismann barrier requires a mechanism(s) by which acquired epimutations can escape germline erasure and be passed on to successive generations. Furthermore, to satisfy the definition of transgenerational epigenetic inheritance in mammals, this mechanism(s) must persist for at least two generations to confirm paternal transmission and three or more generations to confirm maternal transmission (242).

Research findings in the past 10–15 years are beginning to elucidate such mechanisms. Several have now been proposed for germline epigenetic inheritance incorporating direct replicative and indirect reconstructive modes of transmission (243, 244). Examples of the former include DNA methylation, particularly in the context of genomic imprinting, and histone modifications that escape germline erasure. A significant proportion of sperm-derived histones are retained following fertilization, and these could serve to replicate posttranslational modifications acquired during spermatogenesis (245). Among the putative mechanisms underpinning reconstructive transmission in mammals, the role of both maternally and paternally inherited noncoding RNAs has gained the greatest traction (246, 247). However, separating replicative from reconstructive modes of transmission is problematic, and emerging evidence indicates that they operate synergistically to direct epigenetic transgenerational inheritance, as witnessed in gestating female rats exposed to either the agricultural pesticide vinclozolin or the pesticide dichlorodiphenyltrichloroethane (248).

contributions to epigenetic and phenotypic outcomes have contributed to the controversy surrounding the significance of the few studies that have considered transgenerational epigenetic inheritance in outbred animal species and humans (199).

# 6. ASSESSING RISK OF ECs ON METABOLIC AND REPRODUCTIVE HEALTH

There are several major challenges to understanding the effects of real-life EC mixture exposure on public and environmental health. Core to this is the fact that the effects of ECs, when present within a mixture, do not necessarily reflect the sum of the effects of the individual chemicals/chemical types, which can be closely defined by laboratory studies. Indeed, when in a mixture, ECs may exhibit additive, synergistic, or even antagonistic effects, making prediction of effects highly complex (256). Many published laboratory studies document the effects of acute, high-dose EC exposure, whereas real-life EC exposure is typically chronic and not to overtly toxic levels of ECs. Extrapolation from such laboratory studies to predict what might be the effects of an EC mixture is also difficult, because EC dose-response profiles are often nonmonotonic (257). Finally, although some physiological effects may be mediated by the EC itself, they can also arise in response to in vivo metabolites of the EC, which may exhibit effects and work via different modes of action relative to their parent EC (1). For example, the plasticizer DEHP and its primary metabolite MEHP [mono-(2-ethylhexyl) phthalate] exhibit pro- and antiandrogenic activities, respectively (258).

When considering ECs as a mediator of DOHaD, an additional complication relates to the developmental time at which EC exposure occurs. The timing and coordination of normal fetal growth and development rely on complex signaling (endocrine and nonendocrine) that controls processes such as organogenesis, steroidogenesis, neuronal development, and metabolism. Therefore, the impacts of EC exposure will depend on the developmental stage at which it occurs.

For example, in the developing rat, exposure to an androgenic EC between embryonic day 15.5 and 19.5, which corresponds to weeks 8-14 of human gestation, would result in masculinization of reproductive tissues/structures (259), and blockade of normal androgen signaling is likely to result in male offspring with a TDS-like phenotype. In contrast, exposing rat pups to an androgenic EC, such as DBP, between embryonic day 13.5 and 15.5, i.e., before the masculinization window, does not affect the external appearance of the male genitalia but affects other aspects of reproductive function, including germ cell development (260). The exact mechanisms through which these developmental time-sensitive effects of ECs occur have not been characterized extensively but are likely to be the result of interactive effects of ECs, aberrant endocrine signaling, and the induction of molecular and/or epigenetic changes. Critically, however, real-life EC exposure is continual, although the balance of ECs that may have androgenic/antiandrogenic/estrogenic/antiestrogenic effects may vary across gestation and across other developmentally important periods. Finally, the effects of EC exposure may be affected by genetic factors, i.e., the genetic background and associated pharmacogenomic differences between study subjects/populations. This is a particular issue when considering laboratory studies that have used specific inbred strains or rats and mice in which inconsistent results may be strain dependent (257, 261-263) but may not represent the effects seen in a wild or outbred population.

#### 7. CONCLUSION

Obesity, metabolic disease, alterations in pubertal timing, infertility, and CVD in humans are all linked to EC exposure. However, given the epidemiological basis of human studies, none have definitively established causation, and the mechanism(s) (including epigenetic) that underlie these pathologies remain to be elucidated fully. Our understanding of how ECs affect health is based largely on epidemiological studies in humans and wildlife (not considered in this article) combined with traditional toxicological (risk) assessments in animal models. Choice of appropriate animal models requires an understanding of species differences in lifespan, development, and physiology. The lack of genetic diversity in many laboratory species, although acceptable for controlled toxicological studies, renders them less translationally relevant with respect to effects of more general EC exposure. Future research should extend beyond single-chemical effects to prioritize real-life EC mixtures across multiple developmental stages and/or generations in outbred animal species. This approach would provide more representative platforms to investigate sexually dimorphic and individual effects of EC exposure. However, establishing the mechanistic basis for effects of EC mixtures will require a combined approach using in vivo and targeted in vitro (e.g., cell culture, organ-on-a-chip) models. Demonstrating causality and mode of action will facilitate the development of One Health strategies to mitigate the harmful developmental effects of EC exposure.

#### DISCLOSURE STATEMENT

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#### LITERATURE CITED

Sifakis S, Androutsopoulos VP, Tsatsakis AM, Spandidos DA. 2017. Human exposure to endocrine disrupting chemicals: effects on the male and female reproductive systems. *Environ. Toxicol. Pharmacol.* 51:56–70

- Nadal A, Quesada I, Tuduri E, Nogueiras R, Alonso-Magdalena P. 2017. Endocrine-disrupting chemicals and the regulation of energy balance. Nat. Rev. Endocrinol. 13(9):536–46
- Hung H, Katsoyiannis AA, Guardans R. 2016. Ten years of global monitoring under the Stockholm Convention on Persistent Organic Pollutants (POPs): trends, sources and transport modelling. *Environ. Pollut.* 217:1–3
- Secr. Stockholm Conv. 2019. What are POPs? https://chm.pops.int/Convention/ThePOPs/tabid/ 673/language/en-US/Default.aspx
- McFarland VA, Clarke JU. 1989. Environmental occurrence, abundance, and potential toxicity of polychlorinated biphenyl congeners: considerations for a congener-specific analysis. *Environ. Health Perspect.* 81:225–39
- Wilkinson JL, Boxall ABA, Kolpin DW, Leung KMY, Lai RWS, et al. 2022. Pharmaceutical pollution of the world's rivers. PNAS 119(8):e2113947119
- Aris AZ, Shamsuddin AS, Praveena SM. 2014. Occurrence of 17α-ethynylestradiol (EE2) in the environment and effect on exposed biota: a review. Environ. Int. 69:104–19
- 8. Arcand-Hoy L, Nimrod A, Benson W. 1998. Endocrine-modulating substances in the environment: estrogenic effects of pharmaceutical products. *Int. 7. Toxicol.* 17(2):139–58
- Ng M, Fleming T, Robinson M, Thomson B, Graetz N, et al. 2014. Global, regional, and national prevalence of overweight and obesity in children and adults during 1980–2013: a systematic analysis for the Global Burden of Disease Study 2013. *Lancet* 384(9945):766–81
- Elcombe CS, Evans NP, Bellingham M. 2022. Critical review and analysis of literature on low dose exposure to chemical mixtures in mammalian in vivo systems. Crit. Rev. Toxicol. 52(3):221–38
- Arroyo-Johnson C, Mincey KD. 2016. Obesity epidemiology worldwide. Gastroenterol. Clin. N. Am. 45(4):571–79
- 12. Saklayen MG. 2018. The global epidemic of the metabolic syndrome. Curr. Hypertens. Rep. 20(2):12
- Aguilar M, Bhuket T, Torres S, Liu B, Wong RJ. 2015. Prevalence of the metabolic syndrome in the United States, 2003–2012. JAMA 313(19):1973–74
- NCD Risk Factor Collab. 2016. Worldwide trends in diabetes since 1980: a pooled analysis of 751 population-based studies with 4.4 million participants. *Lancet* 387(10027):1513–30
- NCD Risk Factor Collab. 2016. Trends in adult body-mass index in 200 countries from 1975 to 2014:
   a pooled analysis of 1698 population-based measurement studies with 19.2 million participants. *Lancet* 387(10026):1377–96
- Cent. Dis. Control. 2020. National diabetes statistics report. Rep., Cent. Dis. Control, Atlanta. https://www.cdc.gov/diabetes/pdfs/data/statistics/national-diabetes-statistics-report.pdf
- Grundy SM, Brewer HB Jr., Cleeman JI, Smith SC Jr., Lenfant C. 2004. Definition of metabolic syndrome: report of the National Heart, Lung, and Blood Institute/American Heart Association conference on scientific issues related to definition. *Arterioscler: Thromb. Vasc. Biol.* 24(2):e13–18
- 18. Alberti KG, Eckel RH, Grundy SM, Zimmet PZ, Cleeman JI, et al. 2009. Harmonizing the metabolic syndrome: a joint interim statement of the International Diabetes Federation Task Force on Epidemiology and Prevention; National Heart, Lung, and Blood Institute; American Heart Association; World Heart Federation; International Atherosclerosis Society; and International Association for the Study of Obesity. Circulation 120(16):1640–45
- Woodruff TJ. 2011. Bridging epidemiology and model organisms to increase understanding of endocrine disrupting chemicals and human health effects. J. Steroid Biochem. Mol. Biol. 127(1–2):108–17
- Blomberg Jensen M, Priskorn L, Jensen TK, Juul A, Skakkebaek NE. 2015. Temporal trends in fertility rates: a nationwide registry based study from 1901 to 2014. PLOS ONE 10(12):e0143722
- Levine H, Jorgensen N, Martino-Andrade A, Mendiola J, Weksler-Derri D, et al. 2017. Temporal trends in sperm count: a systematic review and meta-regression analysis. Hum. Reprod. Update 23(6):646–59
- UN Dep. Econ. Soc. Aff. 2022. World population prospects 2022: summary of results. Rep., UN Dep. Econ. Soc. Aff., New York. https://www.un.org/development/desa/pd/sites/www.un.org.development.desa.pd/files/wpp2022\_summary\_of\_results.pdf
- Rutstein M, Shah I. 2004. Infecundity, infertility, and childlessness in developing countries. DHS Compar. Rep., ORC Macro, Calverton, MD

- Chandra A, Copen CE, Stephen EH. 2014. Infertility service use in the United States: data from the National Survey of Family Growth, 1982–2010. Natl. Health Stat. Rep. 2014(73):1–21
- Kumar N, Singh AK. 2015. Trends of male factor infertility, an important cause of infertility: a review of literature. J. Hum. Reprod. Sci. 8(4):191–96
- Johansson HKL, Svingen T, Fowler PA, Vinggaard AM, Boberg J. 2017. Environmental influences on ovarian dysgenesis—developmental windows sensitive to chemical exposures. *Nat. Rev. Endocrinol.* 13(7):400–14
- 27. Buck Louis GM, Cooney MA, Peterson CM. 2011. The ovarian dysgenesis syndrome. J. Dev. Orig. Health Dis. 2(1):25–35
- Carlsen E, Giwercman A, Keiding N, Skakkebaek NE. 1992. Evidence for decreasing quality of semen during past 50 years. BM7 305(6854):609–13
- Swan SH, Elkin EP, Fenster L. 2000. The question of declining sperm density revisited: an analysis of 101 studies published 1934–1996. Environ. Health Perspect. 108(10):961–66
- Virtanen HE, Jorgensen N, Toppari J. 2017. Semen quality in the 21<sup>st</sup> century. Nat. Rev. Urol. 14(2):120– 30
- Rodprasert W, Virtanen HE, Sadov S, Perheentupa A, Skakkebaek NE, et al. 2019. An update on semen
  quality among young Finnish men and comparison with Danish data. Andrology 7(1):15–23
- World Health Organ. 2024. Obesity and overweight. Fact Sheet, World Health Organ., Geneva. http://www.who.int/en/news-room/fact-sheets/detail/obesity-and-overweight
- OECD (Organ. Econ. Co-op. Dev.). 2017. SF2.3: Age of mothers at childbirth and age-specific fertility.
   Fam. Database, OECD, Paris, updated March 31, 2017. https://www.oecd.org/content/dam/oecd/en/data/datasets/family-database/sf\_2\_3\_age\_mothers\_childbirth.pdf
- Goulis DG, Tarlatzis BC. 2008. Metabolic syndrome and reproduction: I. Testicular function. Gynecol. Endocrinol. 24(1):33–39
- 35. Al Awlaqi A, Alkhayat K, Hammadeh ME. 2016. Metabolic syndrome and infertility in women. *Int. J. Womens Health Reprod. Sci.* 4(3):89–95
- Madani T, Hosseini R, Ramezanali F, Khalili G, Jahangiri N, et al. 2016. Metabolic syndrome in infertile women with polycystic ovarian syndrome. Arch. Endocrinol. Metab. 60(3):199–204
- Vryonidou A, Paschou SA, Muscogiuri G, Orio F, Goulis DG. 2015. Mechanisms in endocrinology: metabolic syndrome through the female life cycle. Eur. J. Endocrinol. 173(5):R153–63
- 38. Baillie-Hamilton PF. 2002. Chemical toxins: a hypothesis to explain the global obesity epidemic. *J. Altern. Complement. Med.* 8(2):185–92
- Heindel JJ, Blumberg B, Cave M, Machtinger R, Mantovani A, et al. 2017. Metabolism disrupting chemicals and metabolic disorders. Reprod. Toxicol. 68:3–33
- 40. Heindel JJ, vom Saal FS, Blumberg B, Bovolin P, Calamandrei G, et al. 2015. Parma consensus statement on metabolic disruptors. *Environ. Health* 14:54
- Grun F, Blumberg B. 2006. Environmental obesogens: organotins and endocrine disruption via nuclear receptor signaling. Endocrinology 147(6 Suppl.):S50–55
- 42. Gore AC, Chappell VA, Fenton SE, Flaws JA, Nadal A, et al. 2015. EDC-2: the Endocrine Society's second scientific statement on endocrine-disrupting chemicals. *Endocr. Rev.* 36(6):E1–E150
- Lee D-H, Lee I-K, Porta M, Steffes M, Jacobs D. 2007. Relationship between serum concentrations
  of persistent organic pollutants and the prevalence of metabolic syndrome among non-diabetic adults:
  results from the National Health and Nutrition Examination Survey 1999–2002. *Diabetologia* 50:1841–
  51
- Gasull M, Pumarega J, Téllez-Plaza M, Castell C, Tresserras R, et al. 2012. Blood concentrations of persistent organic pollutants and prediabetes and diabetes in the general population of Catalonia. *Environ. Sci. Technol.* 46(14):7799–810
- Roos V, Ronn M, Salihovic S, Lind L, van Bavel B, et al. 2013. Circulating levels of persistent organic pollutants in relation to visceral and subcutaneous adipose tissue by abdominal MRI. Obesity 21(2):413– 18
- 46. Wang T, Li M, Chen B, Xu M, Xu Y, et al. 2012. Urinary bisphenol A (BPA) concentration associates with obesity and insulin resistance. *J. Clin. Endocrinol. Metab.* 97(2):E223–E27

- 47. Sangwan S, Bhattacharyya R, Banerjee D. 2024. Plastic compounds and liver diseases: whether bisphenol A is the only culprit. *Liver Int.* 44(5):1093–105
- Stahlhut RW, Van Wijngaarden E, Dye TD, Cook S, Swan SH. 2007. Concentrations of urinary phthalate metabolites are associated with increased waist circumference and insulin resistance in adult US males. Environ. Health Perspect. 115(6):876–82
- 49. Kim JH, Park HY, Bae S, Lim Y-H, Hong Y-C. 2013. Diethylhexyl phthalates is associated with insulin resistance via oxidative stress in the elderly: a panel study. *PLOS ONE* 8(8):e71392
- Perez-Diaz C, Uriz-Martínez M, Ortega-Rico C, Leno-Duran E, Barrios-Rodríguez R, et al. 2024.
   Phthalate exposure and risk of metabolic syndrome components: a systematic review. *Environ. Pollut.* 340(1):122714
- 51. Lind PM, Lind L. 2011. Circulating levels of bisphenol A and phthalates are related to carotid atherosclerosis in the elderly. *Atherosclerosis* 218(1):207–13
- Lang IA, Galloway TS, Scarlett A, Henley WE, Depledge M, et al. 2008. Association of urinary bisphenol A concentration with medical disorders and laboratory abnormalities in adults. JAMA 300(11):1303–10
- Lind PM, van Bavel B, Salihovic S, Lind L. 2012. Circulating levels of persistent organic pollutants (POPs) and carotid atherosclerosis in the elderly. *Environ. Health Perspect.* 120(1):38–43
- Schillemans T, Donat-Vargas C, Åkesson A. 2024. Per- and polyfluoroalkyl substances and cardiometabolic diseases: a review. Basic Clin. Pharmacol. Toxicol. 134(1):141–52
- Mohammadkhani MA, Shahrzad S, Haghighi M, Ghanbari R, Mohamadkhani A. 2023. Insights into organochlorine pesticides exposure in the development of cardiovascular diseases: a systematic review. *Arch. Iran. Med.* 26(10):592–99
- Bonanni LJ, Wittkopp S, Long C, Aleman JO, Newman JD. 2024. A review of air pollution as a driver of cardiovascular disease risk across the diabetes spectrum. Front. Endocrinol. 15:1321323
- 57. Kutlar Joss M, Boogaard H, Samoli E, Patton AP, Atkinson R, et al. 2023. Long-term exposure to trafficrelated air pollution and diabetes: a systematic review and meta-analysis. *Int. 7. Public Health* 68:1605718
- Hassan S, Thacharodi A, Priya A, Meenatchi R, Hegde TA, et al. 2024. Endocrine disruptors: unravelling the link between chemical exposure and women's reproductive health. *Environ. Res.* 241:117385
- Vabre P, Gatimel N, Moreau J, Gayrard V, Picard-Hagen N, et al. 2017. Environmental pollutants, a possible etiology for premature ovarian insufficiency: a narrative review of animal and human data. Environ. Health 16:37
- Vandenberg LN. 2021. Endocrine disrupting chemicals and the mammary gland. Adv. Pharmacol. 92:237–77
- Liu G, Cai W, Liu H, Jiang H, Bi Y, Wang H. 2021. The association of bisphenol A and phthalates with risk of breast cancer: a meta-analysis. Int. J. Environ. Res. Public Health 18(5):2375
- Cong X, Liu Q, Li W, Wang L, Feng Y, et al. 2023. Systematic review and meta-analysis of breast cancer risks in relation to 2,3,7,8-tetrachlorodibenzo-p-dioxin and per- and polyfluoroalkyl substances. Environ. Sci. Pollut. Res. Int. 30(37):86540–55
- 63. Rochester JR. 2013. Bisphenol A and human health: a review of the literature. Reprod. Toxicol. 42:132-55
- Coiplet E, Courbiere B, Agostini A, Boubli L, Bretelle F, Netter A. 2022. Endometriosis and environmental factors: a critical review. J. Gynecol. Obstet. Hum. Reprod. 51(7):102418
- Campbell S, Raza M, Pollack AZ. 2016. Perfluoroalkyl substances and endometriosis in US women in NHANES 2003–2006. Reprod. Toxicol. 65:230–35
- Wang B, Zhang R, Jin F, Lou H, Mao Y, et al. 2017. Perfluoroalkyl substances and endometriosis-related infertility in Chinese women. *Environ. Int.* 102:207–12
- Ao J, Zhang R, Huo X, Zhu W, Zhang J. 2024. Environmental exposure to legacy and emerging perand polyfluoroalkyl substances and endometriosis in women of childbearing age. Sci. Total Environ. 907:167838
- Colon I, Caro D, Bourdony CJ, Rosario O. 2000. Identification of phthalate esters in the serum of young Puerto Rican girls with premature breast development. *Environ. Health Perspect.* 108(9):895–900
- Hashemipour M, Kelishadi R, Amin MM, Ebrahim K. 2018. Is there any association between phthalate exposure and precocious puberty in girls? *Environ. Sci. Pollut. Res.* 25(14):13589–96

- Srilanchakon K, Thadsri T, Jantarat C, Thengyai S, Nosoognoen W, Supornsilchai V. 2017. Higher
  phthalate concentrations are associated with precocious puberty in normal weight Thai girls. J. Pediatr.
  Endocrinol. Metab. 30(12):1293–98
- Golestanzadeh M, Riahi R, Kelishadi R. 2020. Association of phthalate exposure with precocious and delayed pubertal timing in girls and boys: a systematic review and meta-analysis. *Environ. Sci. Process Impacts* 22(4):873–94
- Frederiksen H, Sørensen K, Mouritsen A, Aksglaede L, Hagen C, et al. 2012. High urinary phthalate concentration associated with delayed pubarche in girls. Int. 7. Androl. 35:216–26
- 73. Lomenick JP, Calafat AM, Melguizo Castro MS, Mier R, Stenger P, et al. 2010. Phthalate exposure and precocious puberty in females. *7. Pediatr*: 156(2):221–25
- 74. Adoamnei E, Mendiola J, Vela-Soria F, Fernández MF, Olea N, et al. 2018. Urinary bisphenol A concentrations are associated with reproductive parameters in young men. *Environ. Res.* 161:122–28
- Hu W, Dong T, Wang L, Guan Q, Song L, et al. 2017. Obesity aggravates toxic effect of BPA on spermatogenesis. Environ. Int. 105:56–65
- Radwan M, Wielgomas B, Dziewirska E, Radwan P, Kałużny P, et al. 2018. Urinary bisphenol A levels and male fertility. Am. 7. Men's Health 12(6):2144–51
- 77. Radke EG, Braun JM, Meeker JD, Cooper GS. 2018. Phthalate exposure and male reproductive outcomes: a systematic review of the human epidemiological evidence. *Environ. Int.* 121:764–93
- Chen Q, Yang H, Zhou N, Sun L, Bao H, et al. 2017. Phthalate exposure, even below US EPA reference doses, was associated with semen quality and reproductive hormones: prospective MARHCS study in general population. *Environ. Int.* 104:58–68
- Arbuckle TE, Agarwal A, MacPherson SH, Fraser WD, Sathyanarayana S, et al. 2018. Prenatal exposure to phthalates and phenols and infant endocrine-sensitive outcomes: the MIREC study. *Environ. Int.* 120:572–83
- Albert O, Huang JY, Aleksa K, Hales BF, Goodyer CG, et al. 2018. Exposure to polybrominated diphenyl
  ethers and phthalates in healthy men living in the greater Montreal area: a study of hormonal balance
  and semen quality. *Environ. Int.* 116:165–75
- 81. Paul R, Moltó J, Ortuño N, Romero A, Bezos C, et al. 2017. Relationship between serum dioxin-like polychlorinated biphenyls and post-testicular maturation in human sperm. *Reprod. Toxicol.* 73:312–21
- 82. Sumner RN, Tomlinson M, Craigon J, England GCW, Lea RG. 2019. Independent and combined effects of diethylhexyl phthalate and polychlorinated biphenyl 153 on sperm quality in the human and dog. *Sci. Rep.* 9(1):3409
- Yu YJ, Lin BG, Liang WB, Li LZ, Hong YD, et al. 2018. Associations between PBDEs exposure from house dust and human semen quality at an e-waste areas in South China—a pilot study. *Chemosphere* 198:266–73
- 84. Conforti A, Mascia M, Cioffi G, De Angelis C, Coppola G, et al. 2018. Air pollution and female fertility: a systematic review of literature. *Reprod. Biol. Endocrinol.* 16(1):117
- 85. Ramsay JM, Fendereski K, Horns JJ, VanDerslice JA, Hanson HA, et al. 2023. Environmental exposure to industrial air pollution is associated with decreased male fertility. *Fertility Steril.* 120(3 Pt. 2):637–47
- Zhang J, Cai Z, Ma C, Xiong J, Li H. 2020. Impacts of outdoor air pollution on human semen quality: a meta-analysis and systematic review. *Biomed. Res. Int.* 2020:7528901
- 87. Deng Z, Chen F, Zhang M, Lan L, Qiao Z, et al. 2016. Association between air pollution and sperm quality: a systematic review and meta-analysis. *Environ. Pollut.* 208:663–69
- Perin PM, Maluf M, Czeresnia CE, Januário DANF, Saldiva PHN. 2010. Impact of short-term preconceptional exposure to particulate air pollution on treatment outcome in couples undergoing in vitro fertilization and embryo transfer (IVF/ET). 7. Assist. Reprod. Genet. 27:371–82
- Green RS, Malig B, Windham GC, Fenster L, Ostro B, Swan S. 2009. Residential exposure to traffic and spontaneous abortion. *Environ. Health Perspect*. 117(12):1939–44
- Zeng X, Jin S, Chen X, Qiu Y. 2020. Association between ambient air pollution and pregnancy outcomes in patients undergoing in vitro fertilization in Chengdu, China: a retrospective study. *Environ. Res.* 184:109304
- 91. Faiz AS, Rhoads GG, Demissie K, Kruse L, Lin Y, Rich DQ. 2012. Ambient air pollution and the risk of stillbirth. *Am. J. Epidemiol.* 176(4):308–16

- Slama R, Bottagisi S, Solansky I, Lepeule J, Giorgis-Allemand L, Sram R. 2013. Short-term impact of atmospheric pollution on fecundability. *Epidemiology* 24(6):871–79
- Wieczorek K, Szczęsna D, Radwan M, Radwan P, Polańska K, et al. 2024. Exposure to air pollution and ovarian reserve parameters. Sci. Rep. 14:461
- Hanson MA, Gluckman PD. 2014. Early developmental conditioning of later health and disease: Physiology or pathophysiology? *Physiol. Rev.* 94(4):1027–76
- Colborn T, Saal FSV, Soto AM. 1993. Developmental effects of endocrine-disrupting chemicals in wildlife and humans. Environ. Health Perspect. 101(5):378–84
- Ideraabdullah FY, Belenchia AM, Rosenfeld CS, Kullman SW, Knuth M, et al. 2019. Maternal vitamin D deficiency and developmental origins of health and disease (DOHaD). 7. Endocrinol. 241(2):R65–80
- Gentner MB, O'Connor Leppert ML. 2019. Environmental influences on health and development: nutrition, substance exposure, and adverse childhood experiences. Dev. Med. Child Neurol. 61(9):1008–14
- Hoffman DJ, Reynolds RM, Hardy DB. 2017. Developmental origins of health and disease: current knowledge and potential mechanisms. Nutr. Rev. 75(12):951–70
- Barker DJ, Gluckman PD, Godfrey KM, Harding JE, Owens JA, Robinson JS. 1993. Fetal nutrition and cardiovascular disease in adult life. *Lancet* 341(8850):938–41
- Barker DJ, Osmond C. 1986. Infant mortality, childhood nutrition, and ischaemic heart disease in England and Wales. Lancet 1(8489):1077–81
- Barker DJ, Winter PD, Osmond C, Margetts B, Simmonds SJ. 1989. Weight in infancy and death from ischaemic heart disease. *Lancet* 2(8663):577–80
- 102. Barker DJP. 1990. The fetal and infant origins of adult disease. Br. Med. 7. 301(6761):1111
- 103. Almond D, Currie J. 2011. Killing me softly: the fetal origins hypothesis. J. Econ. Perspect. 25(3):153-72
- Sinclair KD. 2018. When maternal periconceptional diet affects neurological development, it's time to think. PNAS 115(31):7852–54
- Brehm E, Flaws JA. 2019. Transgenerational effects of endocrine-disrupting chemicals on male and female reproduction. *Endocrinology* 160(6):1421–35
- Li L-X, Chen L, Meng X-Z, Chen B-H, Chen S-Q, et al. 2013. Exposure levels of environmental endocrine disruptors in mother-newborn pairs in China and their placental transfer characteristics. PLOS ONE 8(5):e62526
- Chen M-L, Chang C-C, Shen Y-J, Hung J-H, Guo B-R, et al. 2008. Quantification of prenatal exposure and maternal-fetal transfer of nonylphenol. *Chemosphere* 73(1):S239–S45
- Hoover RN, Hyer M, Pfeiffer RM, Adam E, Bond B, et al. 2011. Adverse health outcomes in women exposed in utero to diethylstilbestrol. N. Engl. 7. Med. 365(14):1304–14
- Dieckmann KP, Skakkebaek NE. 1999. Carcinoma in situ of the testis: review of biological and clinical features. Int. J. Cancer 83(6):815–22
- Skakkebaek N, Rajpert-De Meyts E, Main K. 2001. Testicular dysgenesis syndrome: an increasingly common developmental disorder with environmental aspects: opinion. *Hum. Reprod.* 16:972–78
- 111. Mocarelli P, Gerthoux PM, Needham LL, Patterson DG Jr., Limonta G, et al. 2011. Perinatal exposure to low doses of dioxin can permanently impair human semen quality. *Environ. Health Perspect.* 119(5):713–18
- Guo YL, Hsu PC, Hsu CC, Lambert GH. 2000. Semen quality after prenatal exposure to polychlorinated biphenyls and dibenzofurans. *Lancet* 356(9237):1240–41
- Weidner IS, Møller H, Jensen TK, Skakkebaek NE. 1998. Cryptorchidism and hypospadias in sons of gardeners and farmers. Environ. Health Perspect. 106(12):793–96
- 114. García-Rodríguez J, García-Martín M, Nogueras-Ocaña M, de Dios Luna-del-Castillo J, Espigares García M, et al. 1996. Exposure to pesticides and cryptorchidism: geographical evidence of a possible association. *Environ. Health Perspect.* 104(10):1090–95
- 115. Kim SC, Kwon SK, Hong YP. 2011. Trends in the incidence of cryptorchidism and hypospadias of registry-based data in Korea: a comparison between industrialized areas of petrochemical estates and a non-industrialized area. *Asian J. Androl.* 13(5):715–18
- Henriksen LS, Frederiksen H, Jørgensen N, Juul A, Skakkebæk NE, et al. 2023. Maternal phthalate exposure during pregnancy and testis function of young adult sons. Sci. Total Environ. 871:161914

- Janesick A, Blumberg B. 2011. Minireview: PPARγ as the target of obesogens. J. Steroid Biochem. Mol. Biol. 127(1–2):4–8
- 118. Legler J. 2013. An integrated approach to assess the role of chemical exposure in obesity. *Obesity* 21(6):1084–85
- Philips EM, Jaddoe VWV, Trasande L. 2017. Effects of early exposure to phthalates and bisphenols on cardiometabolic outcomes in pregnancy and childhood. Reprod. Toxicol. 68:105–18
- Johnson PI, Sutton P, Atchley DS, Koustas E, Lam J, et al. 2014. The Navigation Guide—evidencebased medicine meets environmental health: systematic review of human evidence for PFOA effects on fetal growth. *Environ. Health Perspect.* 122(10):1028–39
- Hayashi K, Matsuda Y, Kawamichi Y, Shiozaki A, Saito S. 2011. Smoking during pregnancy increases risks of various obstetric complications: a case-cohort study of the Japan Perinatal Registry Network database. J. Epidemiol. 21(1):61–66
- 122. Jensen TK, Jørgensen N, Punab M, Haugen TB, Suominen J, et al. 2004. Association of in utero exposure to maternal smoking with reduced semen quality and testis size in adulthood: a cross-sectional study of 1,770 young men from the general population in five European countries. Am. J. Epidemiol. 159(1):49–58
- 123. Jensen MS, Mabeck L, Toft G, Thulstrup AM, Bonde JP. 2005. Lower sperm counts following prenatal tobacco exposure. *Hum. Reprod.* 20(9):2559–66
- 124. Fowler PA, Childs AJ, Courant F, MacKenzie A, Rhind SM, et al. 2014. In utero exposure to cigarette smoke dysregulates human fetal ovarian developmental signalling. Hum. Reprod. 29(7):1471–89
- Weinberg CR, Wilcox AJ, Baird DD. 1989. Reduced fecundability in women with prenatal exposure to cigarette smoking. Am. J. Epidemiol. 129(5):1072–78
- Horta BL, Victora CG, Menezes AM, Halpern R, Barros FC. 1997. Low birthweight, preterm births and intrauterine growth retardation in relation to maternal smoking. *Paediatr. Perinat. Epidemiol.* 11(2):140– 51
- 127. Ino T. 2010. Maternal smoking during pregnancy and offspring obesity: meta-analysis. *Pediatr. Int.* 52(1):94–99
- Oken E, Levitan E, Gillman M. 2008. Maternal smoking during pregnancy and child overweight: systematic review and meta-analysis. Int. J. Obes. 32(2):201–10
- 129. Mamun AA, O'Callaghan MJ, Williams GM, Najman JM. 2012. Maternal smoking during pregnancy predicts adult offspring cardiovascular risk factors—evidence from a community-based large birth cohort study. PLOS ONE 7(7):e41106
- Huang R-C, Burke V, Newnham J, Stanley F, Kendall G, et al. 2007. Perinatal and childhood origins of cardiovascular disease. Int. J. Obes. 31(2):236–44
- 131. Blake BE, Fenton SE. 2020. Early life exposure to per- and polyfluoroalkyl substances (PFAS) and latent health outcomes: a review including the placenta as a target tissue and possible driver of peri- and postnatal effects. *Toxicology* 443:152565
- 132. Sumner RN, Harris IT, Van der Mescht M, Byers A, England GCW, Lea RG. 2020. The dog as a sentinel species for environmental effects on human fertility. *Reproduction* 159(6):R265–R76
- Pocar P, Grieco V, Aidos L, Borromeo V. 2023. Endocrine-disrupting chemicals and their effects in pet dogs and cats: an overview. *Animals* 13(3):378
- 134. Mensching DA, Slater M, Scott JW, Ferguson DC, Beasley VR. 2012. The feline thyroid gland: A model for endocrine disruption by polybrominated diphenyl ethers (PBDEs)? J. Toxicol. Environ. Health A. 75(4):201–12
- 135. Ali N, Malik RN, Mehdi T, Eqani SA, Javeed A, et al. 2013. Organohalogenated contaminants (OHCs) in the serum and hair of pet cats and dogs: biosentinels of indoor pollution. *Sci. Total Environ.* 449:29–36
- Bost PC, Strynar MJ, Reiner JL, Zweigenbaum JA, Secoura PL, et al. 2016. U.S. domestic cats as sentinels for perfluoroalkyl substances: Possible linkages with housing, obesity, and disease. *Environ. Res.* 151:145– 53
- 137. Lea RG, Byers AS, Sumner RN, Rhind SM, Zhang Z, et al. 2016. Environmental chemicals impact dog semen quality in vitro and may be associated with a temporal decline in sperm motility and increased cryptorchidism. *Sci. Rep.* 6:31281

- 138. Levine H, Jorgensen N, Martino-Andrade A, Mendiola J, Weksler-Derri D, et al. 2023. Temporal trends in sperm count: a systematic review and meta-regression analysis of samples collected globally in the 20th and 21st centuries. *Hum. Reprod. Update* 29(2):157–76
- 139. Sumner RN, Byers A, Zhang Z, Agerholm JS, Lindh L, et al. 2021. Environmental chemicals in dog testes reflect their geographical source and may be associated with altered pathology. *Sci. Rep.* 11(1):7361
- 140. Sharma B, Sarkar A, Singh P, Singh RP. 2017. Agricultural utilization of biosolids: a review on potential effects on soil and plant grown. *Waste Manag.* 64:117–32
- 141. Venkatesan AK, Halden RU. 2014. Contribution of polybrominated dibenzo-p-dioxins and dibenzofurans (PBDD/Fs) to the toxic equivalency of dioxin-like compounds in archived biosolids from the U.S. EPA's 2001 national sewage sludge survey. *Environ. Sci. Technol.* 48(18):10843–49
- 142. Evans NP, Bellingham M, Sharpe RM, Cotinot C, Rhind SM, et al. 2014. Reproduction Symposium: Does grazing on biosolids-treated pasture pose a pathophysiological risk associated with increased exposure to endocrine disrupting compounds? *J. Anim. Sci.* 92(8):3185–98
- 143. Zhang Z, Le Velly M, Rhind SM, Kyle CE, Hough RL, et al. 2015. A study on temporal trends and estimates of fate of bisphenol A in agricultural soils after sewage sludge amendment. Sci. Total Environ. 515–16:1–11
- 144. Rhind SM, Kyle CE, Ruffie H, Calmettes E, Osprey M, et al. 2013. Short- and long-term temporal changes in soil concentrations of selected endocrine disrupting compounds (EDCs) following single or multiple applications of sewage sludge to pastures. *Environ. Pollut.* 181:262–70
- 145. Rhind SM, Smith A, Kyle CE, Telfer G, Martin G, et al. 2002. Phthalate and alkyl phenol concentrations in soil following applications of inorganic fertiliser or sewage sludge to pasture and potential rates of ingestion by grazing ruminants. J. Environ. Monit. 4(1):142–48
- 146. Rhind SM, Kyle CE, Kerr C, Osprey M, Zhang ZL. 2011. Effect of duration of exposure to sewage sludge-treated pastures on liver tissue accumulation of persistent endocrine disrupting compounds (EDCs) in sheep. Sci. Total Environ. 409(19):3850–56
- 147. Rhind SM, Kyle CE, Mackie C, McDonald L. 2009. Accumulation of endocrine disrupting compounds in sheep fetal and maternal liver tissue following exposure to pastures treated with sewage sludge. 7. Environ. Monit. 11(8):1469–76
- 148. Rhind SM, Kyle CE, Mackie C, Telfer G. 2007. Effects of exposure of ewes to sewage sludge-treated pasture on phthalate and alkyl phenol concentrations in their milk. *Sci. Total Environ.* 383(1–3):70–80
- 149. Rhind SM, Kyle CE, Telfer G, Duff EI, Smith A. 2005. Alkyl phenols and diethylhexyl phthalate in tissues of sheep grazing pastures fertilized with sewage sludge or inorganic fertilizer. *Environ. Health Perspect.* 113(4):447–53
- Bellingham M, Amezaga MR, Mandon-Pepin B, Speers CJ, Kyle CE, et al. 2013. Exposure to chemical cocktails before or after conception—the effect of timing on ovarian development. *Mol. Cell. Endocrinol.* 376(1–2):156–72
- 151. Bellingham M, Fiandanese N, Byers A, Cotinot C, Evans NP, et al. 2012. Effects of exposure to environmental chemicals during pregnancy on the development of the male and female reproductive axes. Reprod. Domest. Anim. 47(Suppl. 4):15–22
- 152. Bellingham M, Fowler PA, Amezaga MR, Rhind SM, Cotinot C, et al. 2009. Exposure to a complex cocktail of environmental endocrine-disrupting compounds disturbs the kisspeptin/GPR54 system in ovine hypothalamus and pituitary gland. *Environ. Health Perspect.* 117(10):1556–62
- 153. Bellingham M, Fowler PA, Amezaga MR, Whitelaw CM, Rhind SM, et al. 2010. Foetal hypothalamic and pituitary expression of gonadotrophin-releasing hormone and galanin systems is disturbed by exposure to sewage sludge chemicals via maternal ingestion. J. Neuroendocrinol. 22(6):527–33
- 154. Bellingham M, Fowler PA, MacDonald ES, Mandon-Pepin B, Cotinot C, et al. 2016. Timing of maternal exposure and foetal sex determine the effects of low-level chemical mixture exposure on the foetal neuroendocrine system in sheep. *J. Neuroendocrinol.* 28(12). https://doi.org/10.1111/jne.12444
- 155. Bellingham M, McKinnell C, Fowler PA, Amezaga MR, Zhang Z, et al. 2012. Foetal and post-natal exposure of sheep to sewage sludge chemicals disrupts sperm production in adulthood in a subset of animals. Int. 7. Androl. 35(3):317–29
- Fowler PA, Bellingham M, Sinclair KD, Evans NP, Pocar P, et al. 2012. Impact of endocrine-disrupting compounds (EDCs) on female reproductive health. Mol. Cell. Endocrinol. 355(2):231–39

- Fowler PA, Dora NJ, McFerran H, Amezaga MR, Miller DW, et al. 2008. In utero exposure to low doses
  of environmental pollutants disrupts fetal ovarian development in sheep. Mol. Hum. Reprod. 14(5):269–80
- 158. Paul C, Rhind SM, Kyle CE, Scott H, McKinnell C, Sharpe RM. 2005. Cellular and hormonal disruption of fetal testis development in sheep reared on pasture treated with sewage sludge. *Environ. Health Perspect*. 113(11):1580–87
- Erhard HW, Rhind SM. 2004. Prenatal and postnatal exposure to environmental pollutants in sewage sludge alters emotional reactivity and exploratory behaviour in sheep. Sci. Total Environ. 332(1–3):101–8
- Hombach-Klonisch S, Danescu A, Begum F, Amezaga MR, Rhind SM, et al. 2013. Peri-conceptional changes in maternal exposure to sewage sludge chemicals disturbs fetal thyroid gland development in sheep. Mol. Cell. Endocrinol. 367(1–2):98–108
- 161. Filis P, Walker N, Robertson L, Eaton-Turner E, Ramona L, et al. 2019. Long-term exposure to chemicals in sewage sludge fertilizer alters liver lipid content in females and cancer marker expression in males. Environ. Int. 124:98–108
- 162. Thangaraj SV, Zeng L, Pennathur S, Lea R, Sinclair KD, et al. 2023. Developmental programming: impact of preconceptional and gestational exposure to a real-life environmental chemical mixture on maternal steroid, cytokine and oxidative stress milieus in sheep. Sci. Total Environ. 900:165674
- 163. Thangaraj SV, Kachman M, Halloran KM, Sinclair KD, Lea R, et al. 2023. Developmental programming: Preconceptional and gestational exposure of sheep to a real-life environmental chemical mixture alters maternal metabolome in a fetal sex-specific manner. Sci. Total Environ. 864:161054
- 164. Ghasemzadeh-Hasankolaei M, Elcombe CS, Powls S, Lea RG, Sinclair KD, et al. 2024. Preconceptional and in utero exposure of sheep to a real-life environmental chemical mixture disrupts key markers of energy metabolism in male offspring. J. Neuroendocrinol. 36(1):e13358
- Sharpe RM. 2020. Androgens and the masculinization programming window: human-rodent differences. Biochem. Soc. Trans. 48(4):1725–35
- Lea RG, Mandon-Pépin B, Loup B, Poumerol E, Jouneau L, et al. 2022. Ovine fetal testis stage-specific sensitivity to environmental chemical mixtures. *Reproduction* 163(2):119–31
- Elcombe CS, Monteiro A, Ghasemzadeh-Hasankolaei M, Evans NP, Bellingham M. 2021. Morphological and transcriptomic alterations in neonatal lamb testes following developmental exposure to low-level environmental chemical mixture. *Environ. Toxicol. Pharmacol.* 86:103670
- 168. Elcombe CS, Monteiro A, Elcombe MR, Ghasemzadeh-Hasankolaei M, Sinclair KD, et al. 2022. Developmental exposure to real-life environmental chemical mixture programs a testicular dysgenesis syndrome-like phenotype in prepubertal lambs. *Environ. Toxicol. Pharmacol.* 94:103913
- 169. Elcombe CS, Monteiro A, Ghasemzadeh-Hasankolaei M, Padmanabhan V, Lea R, et al. 2023. Developmental exposure to a real-life environmental chemical mixture alters testicular transcription factor expression in neonatal and pre-pubertal rams, with morphological changes persisting into adulthood. Environ. Toxicol. Pharmacol. 100:104152
- 170. Lea RG, Amezaga MR, Loup B, Mandon-Pépin B, Stefansdottir A, et al. 2016. The fetal ovary exhibits temporal sensitivity to a 'real-life' mixture of environmental chemicals. Sci. Rep. 6:22279
- 171. Evans NP, Bellingham M, Elcombe CS, Ghasemzadeh-Hasankolaei M, Lea RG, et al. 2023. Sexually dimorphic impact of preconceptional and gestational exposure to a real-life environmental chemical mixture (biosolids) on offspring growth dynamics and puberty in sheep. *Environ. Toxicol. Pharmacol.* 102:104257
- 172. Palma-Gudiel H, Cirera F, Crispi F, Eixarch E, Fananas L. 2018. The impact of prenatal insults on the human placental epigenome: a systematic review. *Neurotoxicol. Teratol.* 66:80–93
- 173. Suhaimi NF, Jalaludin J, Abu Bakar S. 2021. Deoxyribonucleic acid (DNA) methylation in children exposed to air pollution: a possible mechanism underlying respiratory health effects development. Rev. Environ. Health 36(1):77–93
- 174. van den Driesche S, Kilcoyne KR, Wagner I, Rebourcet D, Boyle A, et al. 2017. Experimentally induced testicular dysgenesis syndrome originates in the masculinization programming window. JCI Insight 2(6):e91204
- 175. Goodman S, Chappell G, Guyton KZ, Pogribny IP, Rusyn I. 2022. Epigenetic alterations induced by genotoxic occupational and environmental human chemical carcinogens: an update of a systematic literature review. *Mutat. Res. Rev. Mutat. Res.* 789:108408

- 176. Conley JM, Lambright CS, Evans N, Cardon M, Medlock-Kakaley E, et al. 2021. A mixture of 15 phthalates and pesticides below individual chemical no observed adverse effect levels (NOAELs) produces reproductive tract malformations in the male rat. *Environ. Int.* 156:106615
- Zhang L, Hatzakis E, Nichols RG, Hao R, Correll J, et al. 2015. Metabolomics reveals that aryl hydrocarbon receptor activation by environmental chemicals induces systemic metabolic dysfunction in mice. *Environ. Sci. Technol.* 49(13):8067–77
- Marraudino M, Bonaldo B, Farinetti A, Panzica G, Ponti G, Gotti S. 2019. Metabolism disrupting chemicals and alteration of neuroendocrine circuits controlling food intake and energy metabolism. Front. Endocrinol. 9:766
- 179. Anderson OS, Kim JH, Peterson KE, Sanchez BN, Sant KE, et al. 2017. Novel epigenetic biomarkers mediating bisphenol A exposure and metabolic phenotypes in female mice. *Endocrinology* 158(1):31–40
- Heindel JJ, Howard S, Agay-Shay K, Arrebola JP, Audouze K, et al. 2022. Obesity II: establishing causal links between chemical exposures and obesity. *Biochem. Pharmacol.* 199:115015
- Cicatiello AG, Di Girolamo D, Dentice M. 2018. Metabolic effects of the intracellular regulation of thyroid hormone: old players, new concepts. Front. Endocrinol. 9:474
- 182. Peluso T, Nittoli V, Reale C, Porreca I, Russo F, et al. 2023. Chronic exposure to chlorpyrifos damages thyroid activity and imbalances hepatic thyroid hormones signaling and glucose metabolism: dependency of T3-FOXO1 axis by hyperglycemia. *Int. 7. Mol. Sci.* 24(11):9582
- 183. Hua X, Cao XY, Wang XL, Sun P, Chen L. 2017. Exposure of pregnant mice to triclosan causes insulin resistance via thyroxine reduction. *Toxicol. Sci.* 160(1):150–60
- 184. Saradha B, Vaithinathan S, Mathur P. 2008. Single exposure to low dose of lindane causes transient decrease in testicular steroidogenesis in adult male Wistar rats. *Toxicology* 244(2–3):190–97
- 185. Hall A, Mattison D, Singh N, Chatzistamou I, Zhang J, et al. 2023. Effect of TCDD exposure in adult female and male mice on the expression of miRNA in the ovaries and testes and associated reproductive functions. Front Toxicol. 5:1268293
- 186. Cimmino I, Fiory F, Perruolo G, Miele C, Beguinot F, et al. 2020. Potential mechanisms of bisphenol A (BPA) contributing to human disease. *Int. J. Mol. Sci.* 21(16):5761
- 187. Duarte-Hospital C, Tête A, Brial F, Benoit L, Koual M, et al. 2021. Mitochondrial dysfunction as a hallmark of environmental injury. *Cells* 11(1):110
- 188. Pizzino G, Irrera N, Cucinotta M, Pallio G, Mannino F, et al. 2017. Oxidative stress: harms and benefits for human health. Oxidative medicine and cellular longevity. 2017:8416763
- Hussain T, Metwally E, Murtaza G, Kalhoro DH, Chughtai MI, et al. 2024. Redox mechanisms of environmental toxicants on male reproductive function. Front. Cell Dev. Biol. 12:1333845
- 190. Dong R, Chen J, Zheng J, Zhang M, Zhang H, et al. 2018. The role of oxidative stress in cardiometabolic risk related to phthalate exposure in elderly diabetic patients from Shanghai. *Environ. Int.* 121:340–48
- Lemini C, Silveyra P, Segovia-Mendoza M. 2024. Cardiovascular disrupting effects of bisphenols, phthalates, and parabens related to endothelial dysfunction: review of toxicological and pharmacological mechanisms. *Environ. Toxicol. Pharmacol.* 107:104407
- 192. Rasoulpour RJ, Boekelheide K. 2005. NF-κB is activated in the rat testis following exposure to mono-(2-ethylhexyl) phthalate. *Biol. Reprod.* 72(2):479–86
- 193. Zhao X-F, Wang Q, Ji Y-L, Wang H, Liu P, et al. 2011. Fenvalerate induces germ cell apoptosis in mouse testes through the Fas/FasL signaling pathway. *Arch. Toxicol.* 85:1101–8
- 194. Song H, Park J, Bui PT, Choi K, Gye MC, et al. 2017. Bisphenol A induces COX-2 through the mitogenactivated protein kinase pathway and is associated with levels of inflammation-related markers in elderly populations. *Environ. Res.* 158:490–98
- Ruiz-Hernandez A, Kuo CC, Rentero-Garrido P, Tang WY, Redon J, et al. 2015. Environmental chemicals and DNA methylation in adults: a systematic review of the epidemiologic evidence. Clin. Epigenetics 7(1):55
- 196. Van Cauwenbergh O, Di Serafino A, Tytgat J, Soubry A. 2020. Transgenerational epigenetic effects from male exposure to endocrine-disrupting compounds: a systematic review on research in mammals. Clin. Epigenetics 12:65

- Sinclair KD, Rutherford KM, Wallace JM, Brameld JM, Stoger R, et al. 2016. Epigenetics and developmental programming of welfare and production traits in farm animals. Reprod. Fertil. Dev. 28:1443– 78
- 198. Cedar H, Bergman Y. 2009. Linking DNA methylation and histone modification: patterns and paradigms. *Nat. Rev. Genet.* 10(5):295–304
- Horsthemke B. 2018. A critical view on transgenerational epigenetic inheritance in humans. Nat. Commun. 9:2973
- Choi M, Genereux DP, Goodson J, Al-Azzawi H, Allain SQ, et al. 2017. Epigenetic memory via concordant DNA methylation is inversely correlated to developmental potential of mammalian cells. PLOS Genet. 13(11):e1007060
- Steegers-Theunissen RP, Twigt J, Pestinger V, Sinclair KD. 2013. The periconceptional period, reproduction and long-term health of offspring: the importance of one-carbon metabolism. *Hum. Reprod. Update* 19(6):640–55
- Clare CE, Brassington AH, Kwong WY, Sinclair KD. 2019. One-carbon metabolism: linking nutritional biochemistry to epigenetic programming of long-term development. *Annu. Rev. Anim. Biosci.* 7:263–87
- 203. Petroff RL, Dolinoy DC, Wang K, Montrose L, Padmanabhan V, et al. 2024. Translational toxi-coepigenetic meta-analyses identify homologous gene DNA methylation reprogramming following developmental phthalate and lead exposure in mouse and human offspring. *Environ. Int.* 186:108575
- Wang T, Pehrsson EC, Purushotham D, Li D, Zhuo X, et al. 2018. The NIEHS TaRGET II Consortium and environmental epigenomics. Nat. Biotechnol. 36(3):225–27
- Pepin AS, Lafleur C, Lambrot R, Dumeaux V, Kimmins S. 2022. Sperm histone H3 lysine 4 trimethylation serves as a metabolic sensor of paternal obesity and is associated with the inheritance of metabolic dysfunction. *Mol. Metab.* 59:101463
- Siklenka K, Erkek S, Godmann M, Lambrot R, McGraw S, et al. 2015. Disruption of histone methylation in developing sperm impairs offspring health transgenerationally. Science 350(6261):aab2006
- Lismer A, Siklenka K, Lafleur C, Dumeaux V, Kimmins S. 2020. Sperm histone H3 lysine 4 trimethylation is altered in a genetic mouse model of transgenerational epigenetic inheritance. *Nucleic Acids. Res.* 48(20):11380–93
- Chen Q, Yan M, Cao Z, Li X, Zhang Y, et al. 2016. Sperm tsRNAs contribute to intergenerational inheritance of an acquired metabolic disorder. Science 351(6271):397–400
- Alavian-Ghavanini A, Rüegg J. 2018. Understanding epigenetic effects of endocrine disrupting chemicals: from mechanisms to novel test methods. Basic Clin. Pharmacol. Toxicol. 122(1):38–45
- John RM, Rougeulle C. 2018. Developmental epigenetics: phenotype and the flexible epigenome. Front. Cell Dev. Biol. 6:130
- Yi P, Yu X, Wang Z, O'Malley BW. 2021. Steroid receptor-coregulator transcriptional complexes: new insights from CryoEM. Essays Biochem. 65(6):857–66
- Porter BA, Ortiz MA, Bratslavsky G, Kotula L. 2019. Structure and function of the nuclear receptor superfamily and current targeted therapies of prostate cancer. Cancers 11(12):1852
- Rawłuszko-Wieczorek AA, Romanowska K, Nowicki M. 2022. Chromatin modifiers—coordinators of estrogen action. Biomed. Pharmacother. 153:113548
- Walker CL. 2016. Minireview: epigenomic plasticity and vulnerability to EDC exposures. Mol. Endocrinol. 30(8):848–55
- Rosenfeld CS, Cooke PS. 2019. Endocrine disruption through membrane estrogen receptors and novel pathways leading to rapid toxicological and epigenetic effects. J. Steroid Biochem. Mol. Biol. 187:106–17
- Menezo YJ, Silvestris E, Dale B, Elder K. 2016. Oxidative stress and alterations in DNA methylation: two sides of the same coin in reproduction. *Reprod. Biomed. Online* 33(6):668–83
- Rubio K, Hernandez-Cruz EY, Rogel-Ayala DG, Sarvari P, Isidoro C, et al. 2023. Nutriepigenomics in environmental-associated oxidative stress. *Antioxidants* 12(3):771
- Zheng C, Yu G, Su Q, Wu L, Tang J, et al. 2023. The deficiency of N6-methyladenosine demethylase ALKBH5 enhances the neurodegenerative damage induced by cobalt. Sci. Total Environ. 881:163429
- Herst PM, Dalvai M, Lessard M, Charest PL, Navarro P, et al. 2019. Folic acid supplementation reduces
  multigenerational sperm miRNA perturbation induced by in utero environmental contaminant exposure.

  Environ. Epigenetics 5(4):dvz024

- Santini L, Halbritter F, Titz-Teixeira F, Suzuki T, Asami M, et al. 2021. Genomic imprinting in mouse blastocysts is predominantly associated with H3K27me3. Nat. Commun. 12:3804
- Kobayashi EH, Shibata S, Oike A, Kobayashi N, Hamada H, et al. 2022. Genomic imprinting in human placentation. Reprod. Med. Biol. 21(1):e12490
- 222. Eggermann T. 2024. Human reproduction and disturbed genomic imprinting. Genes 15(2):163
- 223. Young LE, Fernandes K, McEvoy TG, Butterwith SC, Gutierrez CG, et al. 2001. Epigenetic change in *IGF2R* is associated with fetal overgrowth after sheep embryo culture. *Nat. Genet.* 27(2):153–54
- 224. Chen Z, Robbins KM, Wells KD, Rivera RM. 2013. Large offspring syndrome: a bovine model for the human loss-of-imprinting overgrowth syndrome Beckwith–Wiedemann. *Epigenetics* 8(6):591–601
- 225. Li Y, Xiao P, Boadu F, Goldkamp AK, Nirgude S, et al. 2023. The counterpart congenital overgrowth syndromes Beckwith-Wiedemann Syndrome in human and large offspring syndrome in bovine involve alterations in DNA methylation, transcription, and chromatin configuration. medRxiv. https://doi.org/10.1101/2023.12.14.23299981
- Shirane K, Miura F, Ito T, Lorincz MC. 2020. NSD1-deposited H3K36me2 directs de novo methylation in the mouse male germline and counteracts Polycomb-associated silencing. Nat. Genet. 52(10):1088–98
- Hanna CW, Kelsey G. 2021. Features and mechanisms of canonical and noncanonical genomic imprinting. Genes Dev. 35(11–12):821–34
- Fang S, Chang KW, Lefebvre L. 2024. Roles of endogenous retroviral elements in the establishment and maintenance of imprinted gene expression. Front. Cell Dev. Biol. 12:1369751
- Liao J, Szabo PE. 2024. Role of transcription in imprint establishment in the male and female germ lines. Epigenomics 16(2):127–36
- 230. Watanabe T, Tomizawa S, Mitsuya K, Totoki Y, Yamamoto Y, et al. 2011. Role for piRNAs and noncoding RNA in de novo DNA methylation of the imprinted mouse *Rasgrf1* locus. *Science* 332(6031):848–52
- Anvar Z, Chakchouk I, Demond H, Sharif M, Kelsey G, et al. 2021. DNA methylation dynamics in the female germline and maternal-effect mutations that disrupt genomic imprinting. Genes 12(8):1214
- 232. Monk D, Mackay DJG, Eggermann T, Maher ER, Riccio A. 2019. Genomic imprinting disorders: lessons on how genome, epigenome and environment interact. *Nat. Rev. Genet.* 20(4):235–48
- 233. Shi H, Strogantsev R, Takahashi N, Kazachenka A, Lorincz MC, et al. 2019. ZFP57 regulation of transposable elements and gene expression within and beyond imprinted domains. *Epigenetics Chromatin* 12:49
- 234. Yan R, Cheng X, Gu C, Xu Y, Long X, et al. 2023. Dynamics of DNA hydroxymethylation and methylation during mouse embryonic and germline development. Nat. Genet. 55(1):130–43
- Robles-Matos N, Artis T, Simmons RA, Bartolomei MS. 2021. Environmental exposure to endocrine disrupting chemicals influences genomic imprinting, growth, and metabolism. *Genes* 12(8):1153
- Pietryk EW, Clement K, Elnagheeb M, Kuster R, Kilpatrick K, et al. 2018. Intergenerational response
  to the endocrine disruptor vinclozolin is influenced by maternal genotype and crossing scheme. *Reprod. Toxicol.* 78:9–19
- Tindula G, Murphy SK, Grenier C, Huang Z, Huen K, et al. 2018. DNA methylation of imprinted genes in Mexican-American newborn children with prenatal phthalate exposure. *Epigenomics* 10(7):1011–26
- Zhao Y, Chen J, Wang X, Song Q, Xu HH, Zhang YH. 2016. Third trimester phthalate exposure is associated with DNA methylation of growth-related genes in human placenta. Sci. Rep. 6:33449
- Bowman A, Peterson KE, Dolinoy DC, Meeker JD, Sanchez BN, et al. 2019. Phthalate exposures, DNA
  methylation and adiposity in Mexican children through adolescence. Front. Public Health 7:162
- 240. Choi YJ, Lee YA, Hong YC, Cho J, Lee KS, et al. 2020. Effect of prenatal bisphenol A exposure on early childhood body mass index through epigenetic influence on the insulin-like growth factor 2 receptor (*IGF2R*) gene. *Environ. Int.* 143:105929
- 241. Drake AJ, O'Shaughnessy PJ, Bhattacharya S, Monteiro A, Kerrigan D, et al. 2015. In utero exposure to cigarette chemicals induces sex-specific disruption of one-carbon metabolism and DNA methylation in the human fetal liver. BMC Med. 13:18
- Ow MC, Hall SE. 2023. Inheritance of stress responses via small non-coding RNAs in invertebrates and mammals. *Epigenomes* 8(1):1

- Chen Q, Yan W, Duan E. 2016. Epigenetic inheritance of acquired traits through sperm RNAs and sperm RNA modifications. *Nat. Rev. Genet.* 17(12):733–43
- Fitz-James MH, Cavalli G. 2022. Molecular mechanisms of transgenerational epigenetic inheritance. Nat. Rev. Genet. 23(6):325–41
- 245. Hammond SS, Matin A. 2009. Tools for the genetic analysis of germ cells. Genesis 47(9):617-27
- Tang F, Kaneda M, O'Carroll D, Hajkova P, Barton SC, et al. 2007. Maternal microRNAs are essential for mouse zygotic development. *Genes Dev.* 21(6):644

  –48
- Conine CC, Sun F, Song L, Rivera-Pérez JA, Rando OJ. 2018. Small RNAs gained during epididymal transit of sperm are essential for embryonic development in mice. *Dev. Cell* 46(4):470–80.e3
- 248. Beck D, Ben Maamar M, Skinner MK. 2021. Integration of sperm ncRNA-directed DNA methylation and DNA methylation-directed histone retention in epigenetic transgenerational inheritance. *Epigenetics Chromatin* 14(1):6
- Nilsson EE, Ben Maamar M, Skinner MK. 2022. Role of epigenetic transgenerational inheritance in generational toxicology. *Environ. Epigenetics* 8(1):dvac001
- Stouder C, Paoloni-Giacobino A. 2011. Specific transgenerational imprinting effects of the endocrine disruptor methoxychlor on male gametes. Reproduction 141(2):207–16
- Brevik A, Lindeman B, Rusnakova V, Olsen AK, Brunborg G, Duale N. 2012. Paternal benzo[a]pyrene exposure affects gene expression in the early developing mouse embryo. *Toxicol. Sci.* 129(1):157–65
- Li G, Chang H, Xia W, Mao Z, Li Y, Xu S. 2014. F0 maternal BPA exposure induced glucose intolerance of F2 generation through DNA methylation change in Gck. *Toxicol. Lett.* 228(3):192–99
- 253. Chen J, Wu S, Wen S, Shen L, Peng J, et al. 2015. The mechanism of environmental endocrine disruptors (DEHP) induces epigenetic transgenerational inheritance of cryptorchidism. PLOS ONE 10(6):e0126403
- Thompson RP, Nilsson E, Skinner MK. 2020. Environmental epigenetics and epigenetic inheritance in domestic farm animals. Anim. Reprod. Sci. 220:106316
- Stenz L, Rahban R, Prados J, Nef S, Paoloni-Giacobino A. 2019. Genetic resistance to DEHP-induced transgenerational endocrine disruption. PLOS ONE 14(6):e0208371
- Rajapakse N, Silva E, Kortenkamp A. 2002. Combining xenoestrogens at levels below individual noobserved-effect concentrations dramatically enhances steroid hormone action. *Environ. Health Perspect*. 110(9):917–21
- Vandenberg LN, Colborn T, Hayes TB, Heindel JJ, Jacobs DR Jr., et al. 2012. Hormones and endocrinedisrupting chemicals: low-dose effects and nonmonotonic dose responses. *Endocr. Rev.* 33(3):378–455
- 258. Chauvigne F, Menuet A, Lesne L, Chagnon MC, Chevrier C, et al. 2009. Time- and dose-related effects of di-(2-ethylhexyl) phthalate and its main metabolites on the function of the rat fetal testis *in vitro*. *Environ. Health Perspect.* 117(4):515–21
- 259. Welsh M, Saunders PT, Fisken M, Scott HM, Hutchison GR, et al. 2008. Identification in rats of a programming window for reproductive tract masculinization, disruption of which leads to hypospadias and cryptorchidism. *7. Clin. Investig.* 118(4):1479–90
- Auharek SA, de Franca LR, McKinnell C, Jobling MS, Scott HM, Sharpe RM. 2010. Prenatal plus
  postnatal exposure to di(n-butyl) phthalate and/or flutamide markedly reduces final Sertoli cell number
  in the rat. Endocrinology 151(6):2868–75
- vom Saal FS, Hughes C. 2005. An extensive new literature concerning low-dose effects of bisphenol A shows the need for a new risk assessment. Environ. Health Perspect. 113(8):926–33
- Murray TJ, Maffini MV, Ucci AA, Sonnenschein C, Soto AM. 2007. Induction of mammary gland ductal hyperplasias and carcinoma in situ following fetal bisphenol A exposure. Reprod. Toxicol. 23(3):383–90
- 263. Kobayashi K, Miyagawa M, Wang RS, Sekiguchi S, Suda M, Honma T. 2002. Effects of in utero and lactational exposure to bisphenol A on somatic growth and anogenital distance in F1 rat offspring. *Ind. Health.* 40(4):375–81
- Lv Z, Cheng J, Huang S, Zhang Y, Wu S, et al. 2016. DEHP induces obesity and hypothyroidism through both central and peripheral pathways in C3H/He mice. Obesity 24(2):368–78
- 265. Sun D, Zhou L, Wang S, Liu T, Zhu J, et al. 2022. Effect of di-(2-ethylhexyl) phthalate on the hypothalamus-pituitary-thyroid axis in adolescent rat. *Endocr.* 7. 69(2):217–24

- 266. Birru RL, Liang HW, Farooq F, Bedi M, Feghali M, et al. 2021. A pathway level analysis of PFAS exposure and risk of gestational diabetes mellitus. *Environ. Health* 20:63
- Klenke U, Constantin S, Wray S. 2016. BPA directly decreases GnRH neuronal activity via noncanonical pathway. *Endocrinology* 157(5):1980–90
- 268. Lama A, Del Piano F, Annunziata C, Comella F, Opallo N, et al. 2023. Bisphenol A exacerbates anxiety-like behavior and neuroinflammation in prefrontal cortex of adult obese mice. Life Sci. 313:121301
- Li LA. 2007. Polychlorinated biphenyl exposure and CYP19 gene regulation in testicular and adrenocortical cell lines. Toxicol. in Vitro 21(6):1087–94
- 270. Peshdary V, Styles G, Gagné R, Yauk CL, Sorisky A, Atlas E. 2020. Depot-specific analysis of human adipose cells and their responses to bisphenol S. *Endocrinology* 161(6):bqaa044
- 271. Sharma U, Rando OJ. 2017. Metabolic inputs into the epigenome. Cell Metab. 25(3):544-58
- Wiese M, Bannister AJ. 2020. Two genomes, one cell: mitochondrial-nuclear coordination via epigenetic pathways. Mol. Metab. 38:100942