

AKT isoforms have discrete expression in Triple Negative breast cancers and roles in cisplatin sensitivity.

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Additional file 1; Supplementary Figure

Figure S1. Western blot images correlating the expression of AKT1, AKT2 and AKT3 isoforms with malignant breast cancer

Figure S2. AKT1 promotes cell proliferation in MCF-10A and BT-549 cell lines

Figure S3. AKT2 isoform is correlated with aggressiveness of cells and contributes to metastasis

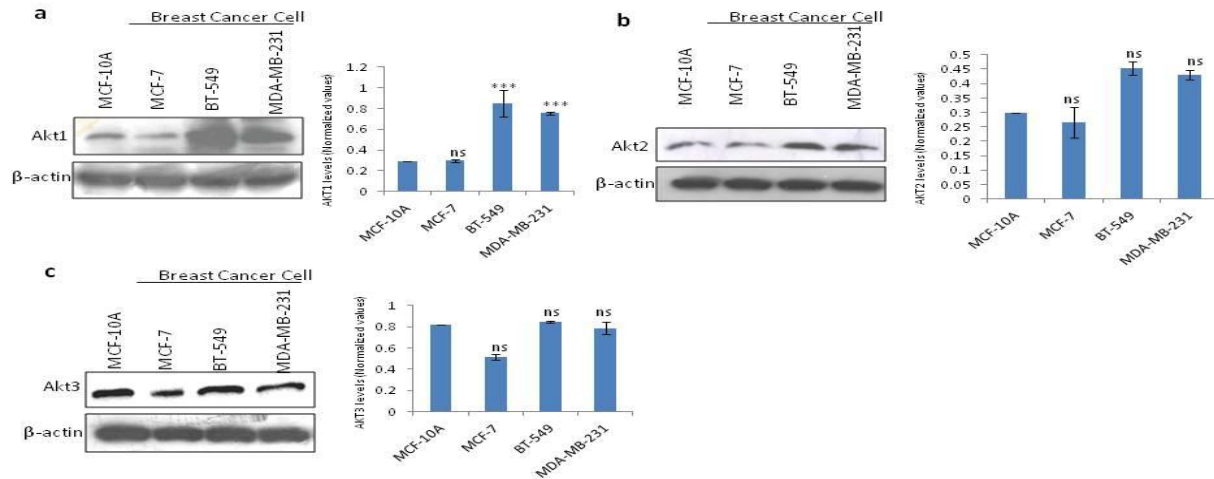
Figure S4. AKT1 knockdown leads to prolonged G2 arrest

Figure S5. AKT1 knockdown abrogated cisplatin sensitivity

Figure S6. Zosuquidar, a p-gp pump drug efflux inhibitor promoted the apoptotic effect of cisplatin in knockdown of AKT1 (shAKT1)

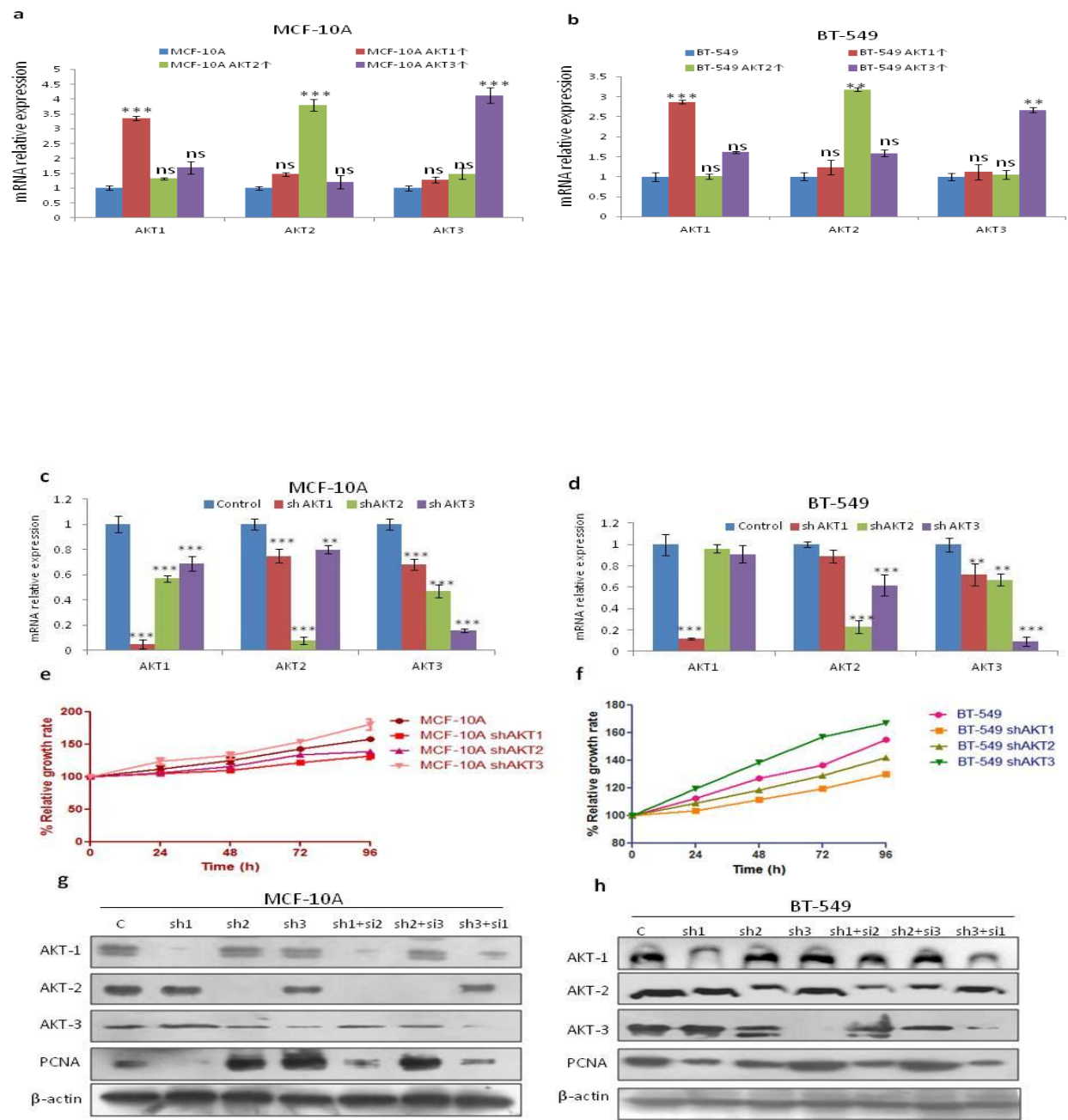
Figure S7. Western blot images of the expression of anti-apoptosis

Fig. S1



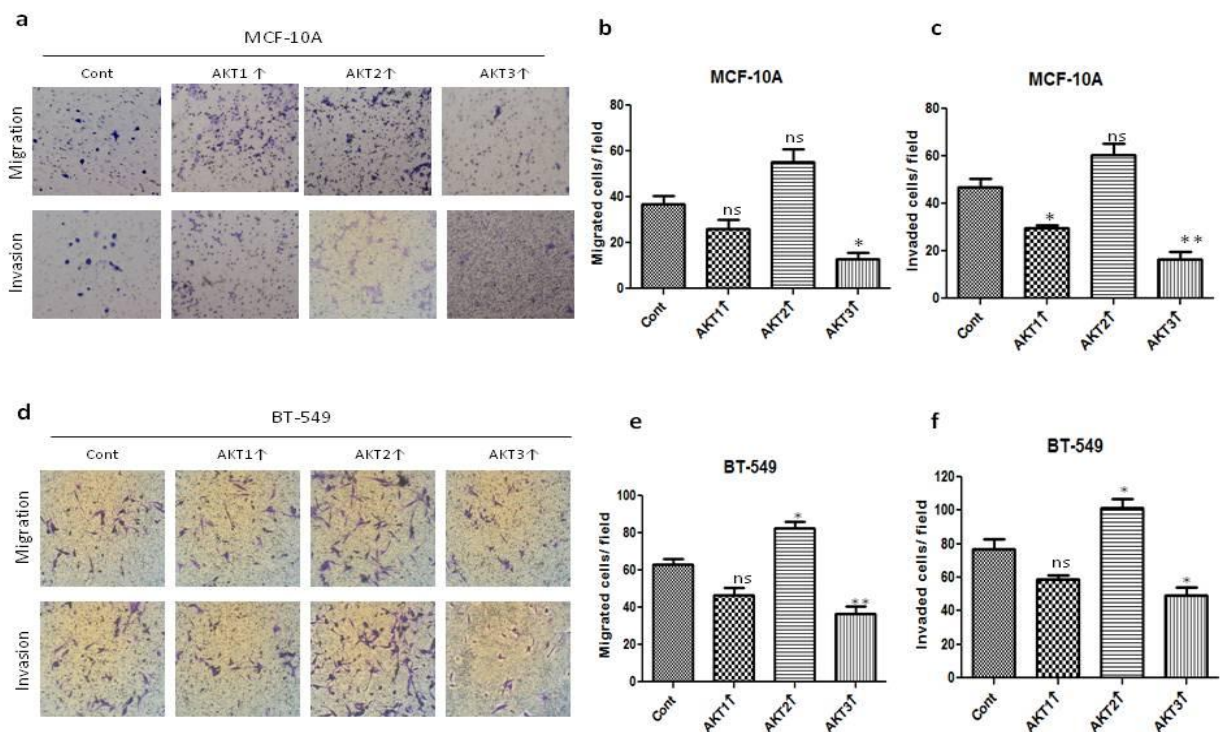
Supplementary Figure S1. Western blot images correlating the expression of AKT1, AKT2 and AKT3 isoforms with malignant breast cancer. (a-c) AKT isoform differential expression between breast normal and cancer cell lines. Data are presented as the mean $\pm$ standard deviation of three independent experiments.

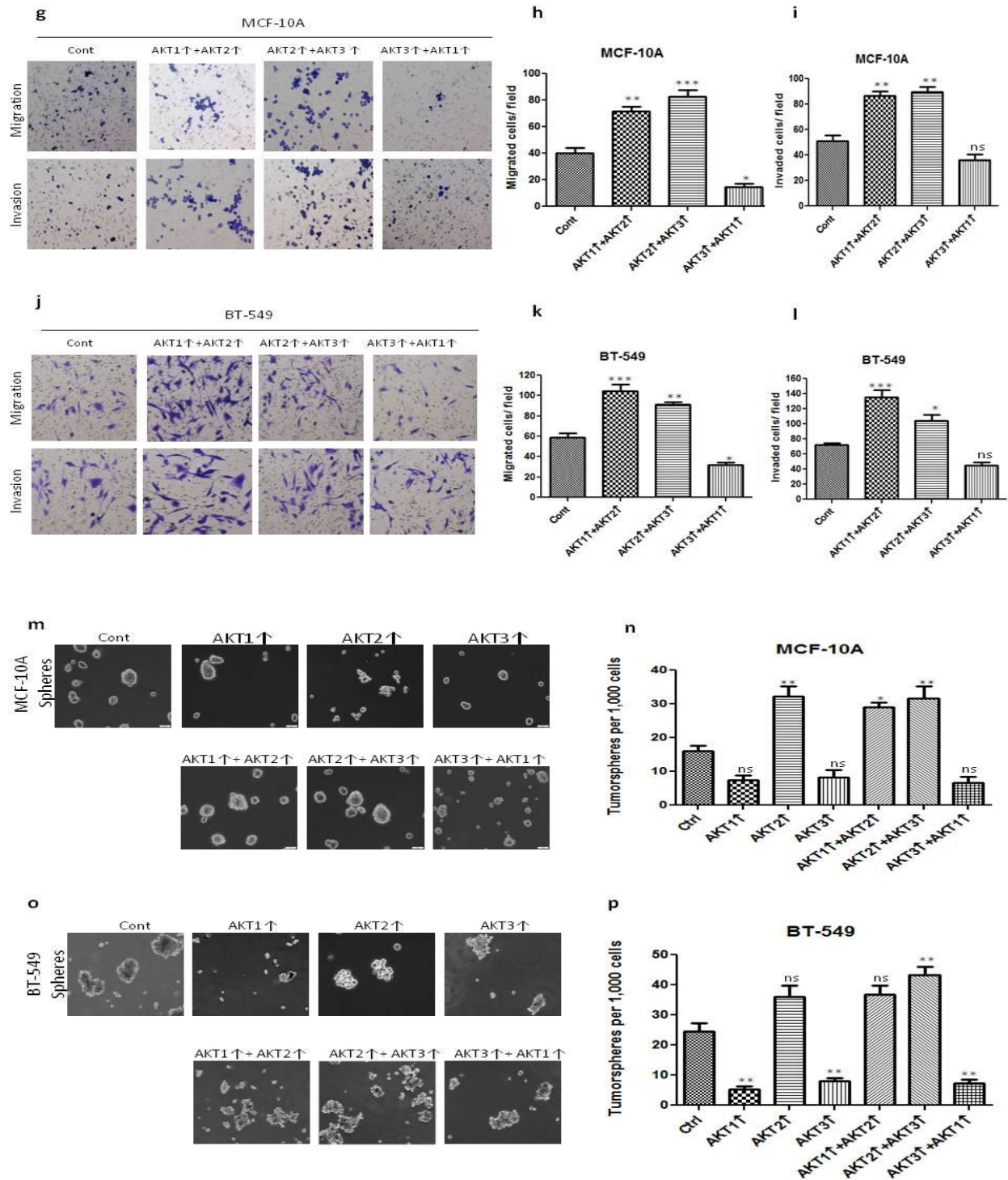
Fig. S2



Supplementary Figure S2. AKT1 promotes cell proliferation in MCF-10A and BT-549 cell lines. qPCR analysis confirming the overexpression of AKT1, AKT2 and AKT3 isoforms expression in (a) MCF-10A and (b) BT-549 cells. qPCR analysis confirming the shRNA-mediated downregulation of AKT1, AKT2 and AKT3 expression in (c) MCF-10A and (d) BT-549 cells. (e) Cell viability was analyzed by MTT at different time intervals in MCF-10A and in (f) BT-549. (g) Western blot analysis of PCNA in individual and dual AKT isoform silencing in MCF-10A and (h) BT-549. All the results show the means of the three independent experiments. Error bars indicate SD. Columns, mean; bars, SD with *** $p < 0.001$, ** $p < 0.01$, * $p < 0.05$ versus control.

Fig. S3

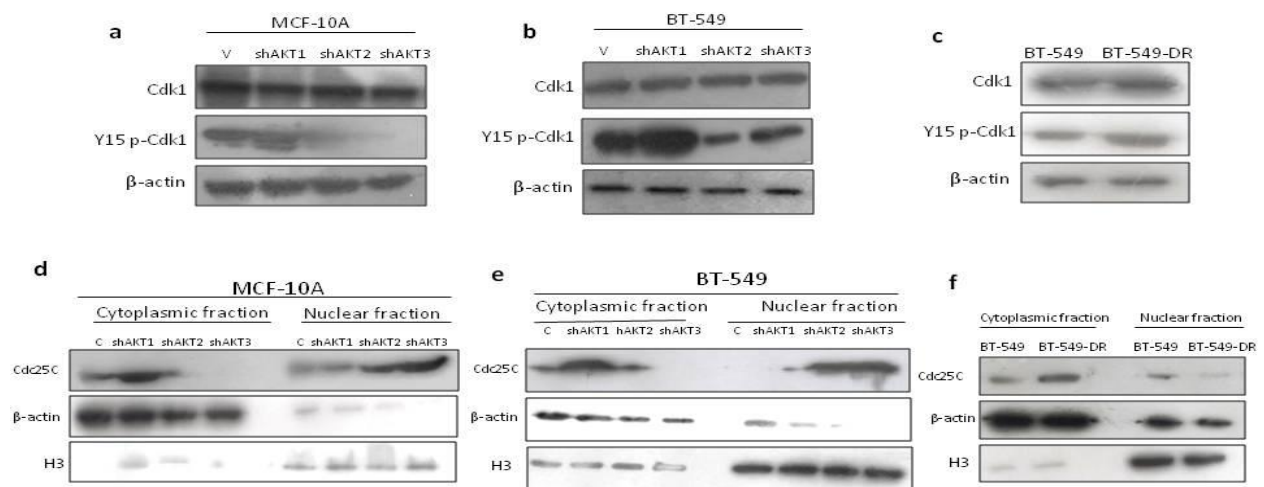




Supplementary Figure S3. AKT2 isoform is correlated with aggressiveness of cells and contributes to metastasis. (a-f) Effect of migration and invasion in MCF-10A and BT-549 cells with the overexpressed AKT isoforms. (g-l) Representative images of migration and invasion analysis in dual AKT isoform overexpression in MCF-10A and BT-549 cells. (m-n)

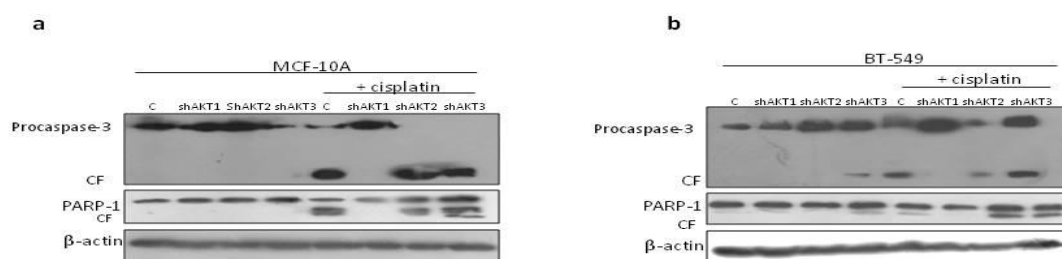
Representative images of the mammospheres formed by overexpressed variants of AKT isoforms in MCF-10A. (o-p) Representative images of the mammospheres formed by overexpressed variants of AKT isoforms in BT-549. The results are shown as mean $\pm$ SD of one representative experiment (from three independent experiments) performed in triplicate. Statistically significant differences (***) $p < 0.001$, (**) $p < 0.01$, (*) $p < 0.05$ are indicated.

Fig. S4



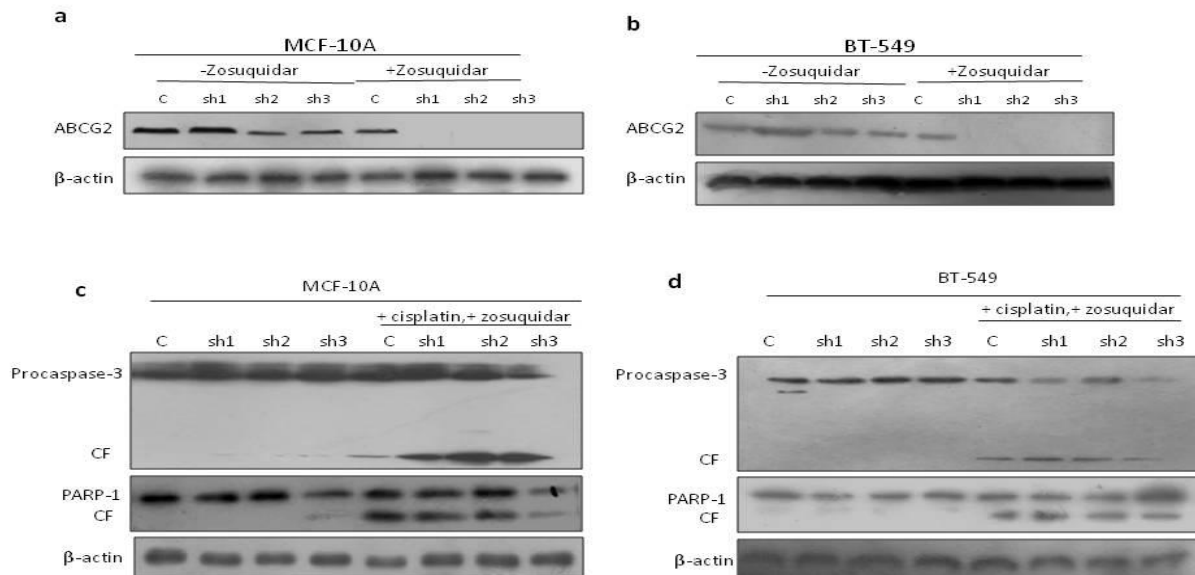
Supplementary Figure S4. AKT1 knockdown leads to prolonged G2 arrest. (a-c) Western blotting images of Cdk1 and Y15p-Cdk1 in (a) MCF-10A AKT isoforms knockdown variants, (b) BT-549 AKT isoforms knockdown variants, (c) BT-549 and BT-549-DR cells. (d-f) Western blot analysis of Cdc25C in subcellular fractions of (d) AKT isoforms knockdown variants in MCF-10A, (e) Akt isoforms knockdown variants in BT-549, (f) BT-549 and BT-549-DR cells.

Fig. S5



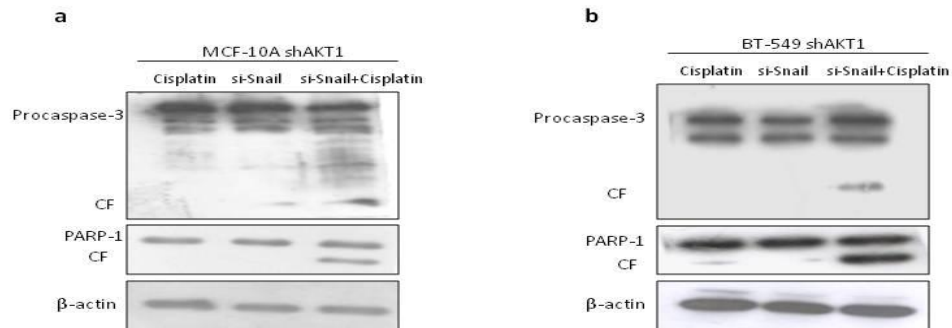
Supplementary Figure S5. AKT1 knockdown abrogated cisplatin sensitivity. (a-b) Western blot analysis of caspase-3 and PARP-1 in presence and absence of cisplatin in (a) MCF-10A and (b) BT-549.

Fig. S6



Supplementary Figure S6. Zosuquidar, a p-gp pump drug efflux inhibitor promoted the apoptotic effect of cisplatin in knockdown of AKT1 (shAKT1). (a-b) Western blot analysis of ABCG2 expression by P-gp inhibitor, zosuquidar. (c-d) Western blot images of apoptotic protein, Procaspase-3 and PARP-1 in AKT isoforms knockdown in (c) MCF-10A and (d) BT-549 respectively. Following 24 h pre-treatment with 2.5 μ M zosuquidar, cells were exposed to cisplatin.

Fig. S7



Supplementary Figure S7. Western blot images of the expression of anti-apoptosis. (a) Knockdown of Snail in shAKT1 MCF-10A and (b) in BT-549 altered the expression of apoptotic-related proteins, Procaspase-3 and PARP-1.