

1 SOS: Speed of Stomata opening and closing is influenced by 2 vapour pressure deficit

3
4 Alexandra J. Burgess^{1,2*} and José Manuel Ugalde^{1,3*}

5 ¹Assistant Features Editor, Plant Physiology, American Society of Plant Biologists, USA

6 ²Agriculture and Environmental Sciences, School of Biosciences, University of Nottingham, Sutton
7 Bonington Campus, Loughborough, LE12 5RD, UK

8 ³INRES-Chemical Signalling, University of Bonn, Friedrich-Ebert-Allee 144, 53113 Bonn, Germany

9
10 Authors for correspondence: alexandra.burgess@nottingham.ac.uk (A.J.B), jugaldev@uni-bonn.de
11 (J.M.U.)

12
13 Stomata, small pores on the surface of above ground plant tissues, are critical for all aspects of plant
14 physiology and metabolism (Lawson and Blatt, 2014). The stomata structure consists of two
15 specialised guard cells, surrounding a central pore. The appearance of these guard cells differs, from
16 kidney shaped within dicots to dumbbell shaped in the monocot grasses. However, both perform the
17 same function; to control the exchange of gases between the plant and the atmosphere. When
18 stomata are open, carbon dioxide (CO₂) is able to enter the intercellular space for use in
19 photosynthesis, and simultaneously, water and oxygen (O₂) are lost to the environment. Therefore,
20 stomata are critical for determining both photosynthesis and water use efficiency (WUE), the
21 balance between carbon gained and water lost. As such, stomata represent key targets for crop
22 improvement and water-use efficiency under climate change (Condon, 2020).

23 The aperture (i.e. size) of the stomatal pore is a function of guard cell morphology and cell turgor
24 pressure. Internal and external signals control this pressure, with environmental cues including high
25 light intensity, low vapour pressure deficit (VPD), and low internal CO₂ concentration triggering
26 stomatal opening (Raschke, 1975). The speed of stomata opening (SOS) is one of the most important
27 determinants of carbon uptake and WUE in fluctuating environments (Lawson and Blatt, 2014; Long
28 et al., 2022), as evidenced in recent studies which genetically manipulate opening speed (e.g.
29 Horaruang et al., 2022). Therefore, understanding the mechanistic control of stomatal opening
30 under changing conditions is of critical importance.

31 Within the angiosperms and members of the fern family Marsiliaeaceae, there is mechanical linkage
32 between the stomata and the surrounding epidermal cells. In these plants, guard cells achieve larger
33 stomatal apertures by displacing the neighbouring epidermal cells. Therefore, the overall aperture of
34 the stomatal pore is a result of opposing pressures between the two sets of cell types. Previous
35 studies indicate that within these species, the epidermal turgor pressure has a greater influence on
36 overall aperture size (Buckley et al., 2003; Franks and Farquhar, 2007). As such, when water status
37 decreases, there is a transient opening of stomata, resulting in a temporary increase in water loss,
38 prior to closure (Buckley, 2005). Modelling studies indicate that when epidermal turgor pressure is
39 low, stomatal opening in the light should be faster in species with mechanical linkage (Mott et al.,
40 1999). Evidence for this is complex due to the wide-range of different guard cell morphologies, with

41 dumbbell shaped guard cells predicted to achieve higher opening speeds. Nevertheless, the link
42 between stomatal opening speeds and the presence of mechanical linkage has not previously been
43 explored.

44 In this issue of *Plant Physiology*, Pichaco et al. (2024) tested if the mechanical interaction between
45 guard cells and epidermal cells modulates the light-dependent speed of stomata opening under
46 variable VPD conditions. The authors compared the stomatal opening speed, measured as stomatal
47 conductance over time, between plant species that have the mechanical linkage advantage between
48 guard cells and epidermal cells (using the angiosperms: *Erythrina sandwicensis*, *Senecio minimus*,
49 and *Umbellularia californica*, and exceptionally the fern, *Marsilea minuta*), against plants that does
50 not have this mechanical linkage advantage (the gymnosperms *Callitris tuberculata*, *Xanthocyparis*
51 *vietnamensis*, and the fern *Pteris vittate*; Fig. 1A). Their results show that a higher VPD increases the
52 stomatal opening speed in the light only in plants that have a mechanical linkage between the guard
53 cells and epidermal cells. On the contrary, plants that do not have this mechanical interaction did
54 not show any difference in their stomata opening speed, regardless of local VPD (Fig. 1B). Moreover,
55 after analyzing the variety in stomata anatomy among the species with mechanical interaction, the
56 authors found a direct correlation between the ratio of guard- and epidermal cell size and the light-
57 dependent stomata opening induced by VPD. Plants with a smaller guard cell/epithelial cell ratio,
58 such as *M. minuta*, have the greatest increase in stomata opening induced by VPD. In comparison,
59 plants with a larger guard cell/ epithelia cell ratio, such as *U. californica*, had a slower stomata
60 opening following an increase in VPD (Fig. 1C). The latter is an important observation since there are
61 only a few studies that have linked physiological functions to the noticeable range of diversity in
62 epidermal anatomy.

63 The results of Pichaco et al. (2024) supports the hypothesis that decreased turgor pressure in
64 epidermal cells increases stomata opening in a VPD-induced process. They established that the VPD
65 dependency of light-induced stomata opening is limited to plants that have mechanical interaction
66 between guard cells and epidermal cells, such as angiosperms and, exceptionally, the Marsiliaeaceae
67 ferns (Fig. 1). Furthermore, the anatomy of the epidermis might be a determinant factor of the
68 effect that epidermal turgor has on stomatal responses. Since a comparison of the stomata opening
69 speed between genotypes is traditionally measured under constant VPD, the findings of this work
70 highlight the possible underestimation of stomata response, when not measured under variable
71 VPDs. These characteristics will be highly important to consider when searching for plants able to
72 respond rapidly to an ever-changing environment.

73

74 **Figure Legends**

75

76 **Figure 1: Stomata opening speed dependency on vapor pressure difference (VPD).** **A)** Examples of
77 plant species that have a mechanical interaction between guard cells and epidermal cells:
78 angiosperms *Erythrina sandwicensis*, *Senecio minimus*, *Umbellularia californica*, and the fern,
79 *Marsilea minuta*, and plants that do not have a mechanical linkage advantage: gymnosperms *Callitris*
80 *tuberculata*, *Xanthocyparis vietnamensis*, and the fern *Pteris vittata*. Most plant silhouettes were
81 obtained from <https://www.phylopic.org/> **B)** The dynamics of light-induced stomata opening was
82 measured in the indicated plants as stomatal conductance over time under different VPD conditions.
83 Plants with mechanical interaction displayed an increased speed of stomata (SOS) opening at higher
84 VPD, while plants that did not have such interaction did not change their SOS opening upon
85 exposure to different VPD conditions. **C)** Among the plants that respond to VPD changes, there is a
86 direct correlation between the ratio of the guard cells size over the epidermal size. Plants with a
87 lower ratio have a higher VPD influence (e.g., *M. minuta*), while plants with a lower ratio are less
88 sensitive to changes in VPD (e.g., *U. californica*). Results in B and C are adapted from Pichaco et al.
89 (2024). The figure was made by J.M.U using Affinity Designer version 2.3.1.

90

91 **Acknowledgements**

92 A.J.B. is partly supported by the BBSRC International partnership in AI for Biosciences [grant number
93 BB/Y513866/1]- H2YOLO: plant water status from cellular to whole plant level. As part of this,
94 www.stomatahub.com has been set up to encourage collaboration and sharing of image datasets.
95 J.M.U is supported by the Deutsche Forschungsgemeinschaft (DFG, German Research Foundation)
96 [grant number UG 74/1-1].

97

98 A.B. and J.M.U. wish to thank Plant Physiology for the opportunity to act as Assistant Feature
99 Editors.

100

101 **Data availability**

102 No new data was collected as part of this manuscript.

103

104 **Conflict of Interest**

105 The authors declare no conflict of interest.

106

107 **References**

108 Buckley, T., 2005. The control of stomata by water balance. *New Phytologist*.
109 <https://doi.org/10.1111/j.1469-8137.2005.01543.x>

- 110 Buckley, T., Mott, K., Farquhar, G., 2003. A hydromechanical and biochemical model of stomatal
111 conductance. *Plant Cell Environ* 26, 1767–1785.
112 <https://doi.org/https://doi.org/10.1046/j.1365-3040.2003.01094.x>
- 113 Condon, A., 2020. Drying times: plant traits to improve crop water use efficiency and yield. *J Exp Bot*
114 71, 2239–2252. <https://doi.org/10.1093/jxb/eraa002>
- 115 Franks, P., Farquhar, G., 2007. The mechanical diversity of stomata and its significance in gas-
116 exchange control. *Plant Physiol* 143, 78–87. <https://doi.org/10.1104/pp.106.089367>
- 117 Horaruang, W., Klejchová, M., Carroll, W., Silva-Alvim, F., Waghmare, S., Papanatsiou, M., Amtmann,
118 A., Hills, A., Alvim, J., Blatt, M., Zhang, B., 2022. Engineering a K⁺ channel ‘sensory antenna’
119 enhances stomatal kinetics, water use efficiency and photosynthesis. *Nat Plants* 8, 1262–1274.
120 <https://doi.org/10.1038/s41477-022-01255-2>
- 121 Lawson, T., Blatt, M., 2014. Stomatal Size, Speed, and Responsiveness Impact on Photosynthesis and
122 Water Use Efficiency. *Plant Physiol* 164, 1556–1570. <https://doi.org/10.1104/pp.114.237107>
- 123 Long, S., Taylor, S., Burgess, S., Carmo-Silva, E., Lawson, T., De Souza, A., Leonelli, L., Wang, Y., 2022.
124 Into the Shadows and Back into Sunlight: Photosynthesis in Fluctuating Light. *Annu Rev Plant*
125 *Biol* 73, 617–648. <https://doi.org/10.1146/annurev-arplant-070221-024745>
- 126 Mott, K., Shope, J., Buckley, T., 1999. Effects of humidity on light-induced stomatal opening:
127 evidence for hydraulic coupling among stomata. *J Exp Bot* 50, 1207–1213.
- 128 Pichaco, J., Manandhar, A., Mcadam, S., 2024. Mechanical advantage makes stomatal opening speed
129 a function of evaporative demand. *Plant Physiol*.
- 130 Raschke, K., 1975. Stomatal Action. *Annu Rev Plant Physiol* 26, 309–340.
131 <https://doi.org/10.1146/annurev.pp.26.060175.001521>
- 132

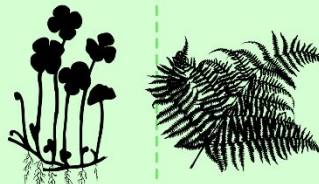
A

Angiosperms



E. sandwicensis *S. minimus* *U. californica*

Ferns



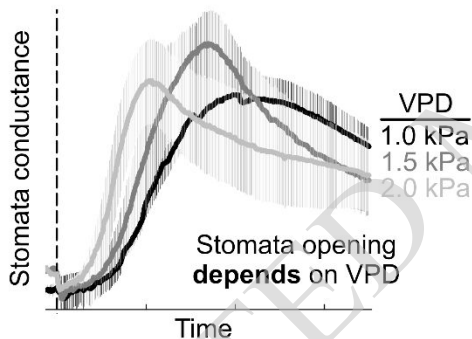
M. minuta *P. vittate*

Gymnosperms

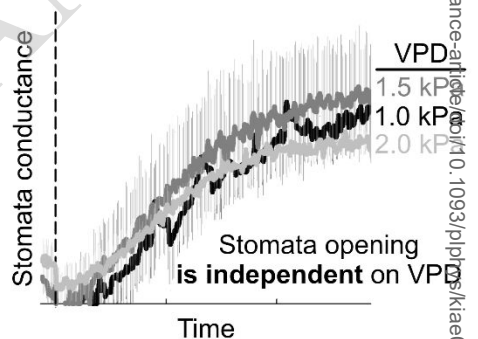


X. vietnamensis *C. tuberculata*

B Species **with** mechanical interaction
between **guard cells** and **epidermal cells**



Species **without** mechanical interaction
between **guard cells** and **epidermal cells**

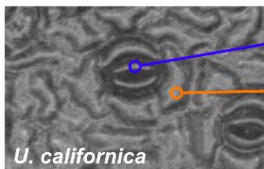
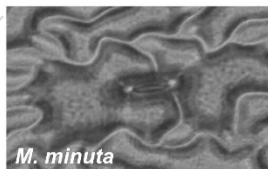


C

Low

guard cells:epidermal cells size ratio

High



High

VPD influence on stomata opening

Low

Parsed Citations

- Buckley, T., 2005. The control of stomata by water balance. *New Phytologist*. <https://doi.org/10.1111/j.1469-8137.2005.01543.x>
Google Scholar: [Author Only](#) [Title Only](#) [Author and Title](#)
- Buckley, T., Mott, K., Farquhar, G., 2003. A hydromechanical and biochemical model of stomatal conductance. *Plant Cell Environ* 26, 1767–1785. <https://doi.org/https://doi.org/10.1046/j.1365-3040.2003.01094.x>
Google Scholar: [Author Only](#) [Title Only](#) [Author and Title](#)
- Condon, A., 2020. Drying times: plant traits to improve crop water use efficiency and yield. *J Exp Bot* 71, 2239–2252. <https://doi.org/10.1093/jxb/eraa002>
Google Scholar: [Author Only](#) [Title Only](#) [Author and Title](#)
- Franks, P., Farquhar, G., 2007. The mechanical diversity of stomata and its significance in gas-exchange control. *Plant Physiol* 143, 78–87. <https://doi.org/10.1104/pp.106.089367>
Google Scholar: [Author Only](#) [Title Only](#) [Author and Title](#)
- Horaruang, W., Klejchová, M., Carroll, W., Silva-Alvim, F., Waghmare, S., Papanatsiou, M., Amtmann, A., Hills, A., Alvim, J., Blatt, M., Zhang, B., 2022. Engineering a K⁺ channel 'sensory antenna' enhances stomatal kinetics, water use efficiency and photosynthesis. *Nat Plants* 8, 1262–1274. <https://doi.org/10.1038/s41477-022-01255-2>
Google Scholar: [Author Only](#) [Title Only](#) [Author and Title](#)
- Lawson, T., Blatt, M., 2014. Stomatal Size, Speed, and Responsiveness Impact on Photosynthesis and Water Use Efficiency. *Plant Physiol* 164, 1556–1570. <https://doi.org/10.1104/pp.114.237107>
Google Scholar: [Author Only](#) [Title Only](#) [Author and Title](#)
- Long, S., Taylor, S., Burgess, S., Carmo-Silva, E., Lawson, T., De Souza, A., Leonelli, L., Wang, Y., 2022. Into the Shadows and Back into Sunlight: Photosynthesis in Fluctuating Light. *Annu Rev Plant Biol* 73, 617–648. <https://doi.org/10.1146/annurev-arplant-070221-024745>
Google Scholar: [Author Only](#) [Title Only](#) [Author and Title](#)
- Mott, K., Shope, J., Buckley, T., 1999. Effects of humidity on light-induced stomatal opening: evidence for hydraulic coupling among stomata. *J Exp Bot* 50, 1207–1213.
Google Scholar: [Author Only](#) [Title Only](#) [Author and Title](#)
- Pichaco, J., Manandhar, A., Mcadam, S., 2024. Mechanical advantage makes stomatal opening speed a function of evaporative demand. *Plant Physiol*.
Google Scholar: [Author Only](#) [Title Only](#) [Author and Title](#)
- Raschke, K., 1975. Stomatal Action. *Annu Rev Plant Physiol* 26, 309–340. <https://doi.org/10.1146/annurev.pp.26.060175.001521>
Google Scholar: [Author Only](#) [Title Only](#) [Author and Title](#)