

# Protein Encapsulation of Experimental Anticancer Agents 5F 203 and Phortress: Towards Precision Drug Delivery

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**Introduction:** Advancement of novel anticancer drugs into clinical use is frequently halted by their lack of solubility, reduced stability under physiological conditions, and non-specific uptake by normal tissues, causing systemic toxicity. Their progress to use in the clinic could be accelerated by the development of new formulations employing suitable and complementary drug delivery vehicles.

**Methods:** A robust method for apoferritin (Aft)-encapsulation of antitumour benzothiazoles has been developed for enhanced activity against and drug delivery to benzothiazole-sensitive cancers.

**Results:** More than 70 molecules of benzothiazole 5F 203 were encapsulated per Aft cage. Post-encapsulation, the size and integrity of the protein cages were retained as evidenced by dynamic light scattering. ToF-SIMS depth profiling using an argon cluster beam confirmed 5F 203 exclusively within the Aft cavity. Improved encapsulation of benzothiazole lysyl-amide prodrugs was achieved (~130 molecules of Phortress per Aft cage). Transferrin receptor 1, TfR1, was detected in lysates prepared from most cancer cell lines studied, contributing to enhanced anticancer potency of the Aft-encapsulated benzothiazoles (5F 203, Phortress, GW 610, GW 608-Lys). Nanomolar activity was demonstrated by Aft-formulations in breast, ovarian, renal and gastric carcinoma cell lines, whereas  $GI_{50} > 50 \mu M$  was observed in non-tumourigenic MRC-5 fibroblasts. Intracellular 5F 203, a potent aryl hydrocarbon receptor (AhR) ligand, and inducible expression of cytochrome P<sub>450</sub> (CYP) 1A1 were detected following exposure of sensitive cells to Aft-5F 203, confirming that the activity of benzothiazoles was not compromised following encapsulation.

**Conclusion:** Our results show enhanced potency and selectivity of Aft-encapsulated 5F 203 against carcinomas derived from breast, ovarian, renal, colorectal as well as gastric cancer models, and offer realistic prospects for potential refinement of tumour-targeting and treatment, and merit further in vivo investigations.

**Keywords:** benzothiazole, apoferritin, transferrin receptor, anticancer activity, drug delivery

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## Introduction

Development of drug delivery vehicles is of great importance for advancement of novel drugs into the clinic, and could be used to address their limited solubility, stability in physiologically relevant solvents, provide targeted delivery of the drug, hence limiting systemic toxicities.<sup>1</sup> These disadvantages are often only apparent at later phases in drug development. Nanomedicine has already enabled significant progress in the development of advanced therapeutic agents and further investigations are ongoing to achieve successful translation into the clinic.<sup>2-4</sup> Several drug delivery

nano-vehicles have been identified, including ones derived from lipids,<sup>5</sup> synthetic polymers<sup>6,7</sup> and proteins.<sup>8</sup> Of particular interest are protein capsules as they are biocompatible, biodegradable and species-specific, hence they are not expected to accumulate in the body or induce an immune response if species matched.<sup>9</sup> Of additional benefit is the possibility to genetically modify the protein surface for attachment of targeting molecules.<sup>10</sup> Proteins from the ferritin family, such as apoferritin (Aft) and Dps have emerged as promising candidates for drug delivery and have been successfully used to encapsulate small molecules and imaging agents.<sup>11–13</sup> Encapsulation of anticancer therapeutic agents into Aft has attracted considerable attention due to internalisation of Aft-encapsulated cargo via transferrin receptor 1 (TfR1)-mediated endocytosis<sup>14,15</sup> and cellular recognition of modified Aft by receptors, such as EGFR, which are overexpressed in specific cancers.<sup>16</sup> Despite promising applications, Aft encapsulation of drugs remains challenging and the procedure requires optimization.

The experimental antitumour agents 2-(4-amino-3-methylphenyl)-5-fluorobenzothiazole (5F 203, [Figure 1a](#)) and 2-(3,4-dimethoxyphenyl)-5-fluorobenzothiazole (GW610; [Supplementary Information Scheme S1](#)) elicit exquisitely potent and selective anticancer activity in certain cancer models via a mechanism distinct from current clinical anticancer therapies.<sup>17,18</sup> The compounds are potent aryl hydrocarbon receptor (AhR) ligands, and induce their own cytochrome P450 (CYP) 1A1 and 2W1-catalyzed metabolism to generate electrophilic species which then form DNA adducts leading to lethal DNA double-strand breaks; this occurs exclusively in sensitive phenotypes.<sup>19</sup> However, systemic exposure and adverse toxicity in potentially susceptible healthy tissues (e.g. lung, liver)<sup>20</sup> and the poor water-solubility of 5F 203 limit its advance into the clinic.

Here we develop a new formulation of 5F 203 addressing its toxicity and solubility. The agent has been encapsulated within horse spleen Aft protein cages. Aft-encapsulation protocols have been systematically investigated to optimize encapsulation efficiency; complementary characterization techniques have been employed to study Aft integrity and confirm 5F 203 encapsulation. Direct determination of encapsulation of 5F-203 inside the Aft cage was achieved by TOF-SIMS. The antitumour activity of the Aft-encapsulated agents was interrogated in breast, ovarian, renal, colorectal, lung and gastric cancers; concurrently, studies were undertaken to verify that the cellular mechanism of

action of 5F 203 was not compromised following its encapsulation in Aft nanocages.

## Materials and Methods

### Encapsulation Methods

Encapsulation of 5F 203 was performed using both the nanoreactor and reassembly routes ([Figure S1](#)). In the nanoreactor route, molecules are encapsulated by passive diffusion through 3 and 4 Angstrom channels at the protein junctions in the Aft shell. In the reassembly route, the ability of the Aft cage to disassemble into its subunits and reassemble is used to entrap the cargo. Two triggers were used for reassembly: pH-dependent and urea-induced reassembly. Horse spleen ferritin (Sigma Aldrich) was used to prepare Aft and was used for encapsulation.

The optimized nanoreactor method was used for subsequent encapsulation of Phortress, GW610 and GW 608-Lys. The chemical structure of all test agents is shown in [Scheme S1](#).

Characterisation methods: UV-vis spectroscopy: The benzothiazole concentrations were determined by measurement of absorbance at 360, 334, 325 and 336 nm for 5F 203, GW 610, Phortress and GW 608-Lys, respectively, on a Varian Cary50 UV-Vis spectrometer, using a calibration curve ( $R^2 > 0.999$ ) obtained from samples of known benzothiazole concentration. Transmission electron microscopy: For transmission electron microscopy (TEM), the samples were deposited onto graphene oxide-coated copper grid and images were acquired on a JEM-2100F, JOEL operated at 200 kV. Dynamic Light Scattering (DLS): DLS analyses were carried out at 25°C using a Zetasizer Nano spectrometer (Malvern Instruments Ltd) equipped with a 633 nm laser at a fixed angle of 173°. ToF-SIMS: Time of Flight Secondary Ion Mass Spectrometry (ToF SIMS) analysis was carried out on a ToF-SIMS IV instrument (ION-TOF GmbH, Munster, Germany) equipped with a bismuth liquid metal ion gun and a single-stage reflectron analyser. Bradford assay: The Aft concentration was determined by Bradford assay. Briefly, 10  $\mu$ L of sample was diluted in 90  $\mu$ L of d.H<sub>2</sub>O in a 96-well microliter plate and 140  $\mu$ L of Bradford reagent was added. After 5-min incubation, the plate was read at 595 nm using a Perkin Elmer plate reader. A calibration curve was constructed, using bovine serum albumin (BSA) from 0.1 to 1.0 mg mL<sup>-1</sup> and used to convert absorbance readings to protein concentrations.

## In vitro Cell Culture Studies

All cell lines were purchased from the American Type Culture Collection (ATCC) and cultured in RPMI 1640 medium supplemented with 10% v/v foetal bovine serum (FBS) at 37°C in an atmosphere of 5% CO<sub>2</sub>. Cells were passaged twice weekly to maintain continuous logarithmic growth. In this work, we have used the following human-derived cell lines: non-tumourigenic foetal fibroblast MRC-5; breast cancer cell lines MCF-7, MDA-MB-468, renal cancer cell TK-10, ovarian cancer cell IGROV-1, colorectal cancer cells HCT 116, HCC 2998 and KM12; gastric cancer cells MKN-45, NCI-N87, KATO-III, BGC-823 and SGC-7901.

For MTT assays, cells were seeded at a density of  $5 \times 10^3$  cells per well into 96-well plates and allowed 24 hrs to adhere. Serial dilutions were prepared in medium and added to cells ( $n = 4$  per concentration). Viable cells at the time of test agent addition (T<sub>0</sub>), and following 72 hrs of drug exposure were determined by cell-mediated 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) reduction. MTT was added to each well (final concentration 400  $\mu\text{g mL}^{-1}$ ) and incubated for 3 hrs at 37°C. Well supernatants were aspirated and formazan solubilized with 150  $\mu\text{L}$  DMSO. Absorbance was read at 550 nm using Perkin Elmer Envision plate reader. The GI<sub>50</sub> values were determined by interpolation.

For clonogenic assays, 250 cells per well were seeded into 6-well plates and allowed 24 hrs to attach. Test compounds were added at GI<sub>50</sub> concentrations. After 24 hrs exposure, the medium was aspirated and cells washed with PBS ( $2 \times 1 \text{ mL}$ ) before incubation in medium (2 mL) for 7–9 days. Experiments were terminated when colonies  $\geq 50$  cells were observed in control wells. Colonies were washed (PBS,  $2 \times 1 \text{ mL}$ ), fixed (methanol, 15 mins), stained (0.5% methylene blue, 15 mins) and counted.

## Confocal Microscopy

Following required treatments, cells were lysed and proteins separated by SDS-PAGE then electroblotted onto PVDF membranes, Membranes were blocked in Tris-buffered saline (TBS) containing 5% milk and 0.1% Tween-20 (1 hr), then incubated with 1° Abs (CYP 1A1, TfR1; Cell Signaling; O/N, 4°C). Membrane incubation with 2° Abs preceded ECL detection. Images were examined on a C-DiGit blot scanner (LI-Cor-Biosciences). Cells were seeded at a density of  $2 \times 10^5$  in wells with coverslips and allowed 24 hrs to attach. Following 24 hrs exposure to the test compound, live images of the cells were taken using

a Zeiss LSM 510 fluorescence microscope equipped with a UV laser.

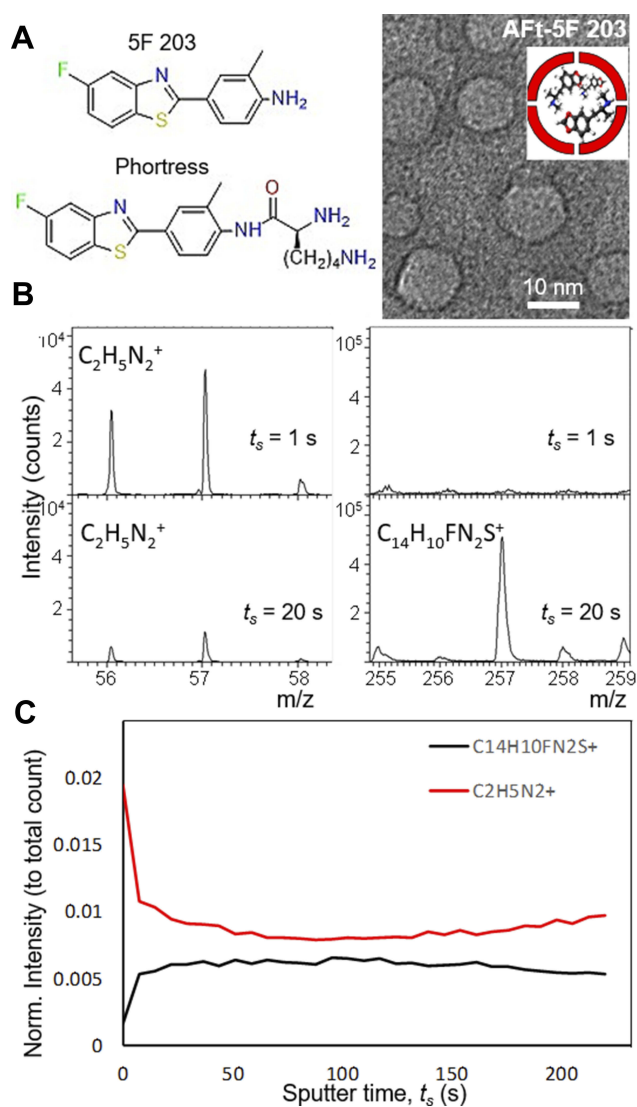
## Statistical Analysis

All experiments were repeated at least three times and results are reported as mean  $\pm$  standard deviation (SD). Statistical significance was assessed using ANOVA.

## Results and Discussion

### Development and Optimization of AfT Encapsulation Method

Encapsulation of 5F 203 was performed using both nanoreactor and reassembly routes (Figure 1), following the



**Figure 1** (A, left) Chemical structure of test agents 5F 203 and Phortress and (right) a cartoon and a representative dark field TEM image of Apoferritin-encapsulated 5F 203. (B) TOF SIMs Mass spectra of AfT-5F 203. (C) A summary of intensity change with increasing sputter time.

modified method developed for encapsulation of nanocrystals.<sup>21,22</sup> Initially, a ratio of five molecules of 5F 203 per AFt cage (24 protein subunits per cage, horse spleen AFt) were mixed together under dialysis (see Experimental section), however significant amounts of precipitate formed. The precipitate was removed and the supernatant was used to assess the concentrations of 5F 203 and AFt by UV-vis spectroscopy and Bradford assays, respectively. For both the nanoreactor and reassembly routes, only ~0.5–1.5% 5F 203 and <1% AFt was detected.

In the nanoreactor route, the number of 5F 203 molecules was increased to 100 per AFt cage, improving the encapsulation efficiency to 47%; however, the loss of the protein during encapsulation was still significant. The choice of buffer is of great importance for AFt stability and solubility. Hence, sodium acetate buffer (pH 5.5) was replaced by TRIS-buffer (pH 7.4) which had previously been reported for encapsulation of doxorubicin [10], but this failed to decrease protein precipitation. The use of 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid (HEPES) buffer and a reduced concentration of AFt (halved to ~1.5 mg mL<sup>-1</sup>) enabled encapsulation without significant loss of protein, with up to 72% of AFt remaining in solution. Hence, HEPES buffer was used to further increase the number of test agent molecules per AFt cage. Since the aqueous solubility of benzothiazoles is very low, 5F 203 was added in aliquots (equivalent of 100 molecules per AFt cage was added at 30 mins intervals). Using this method, we achieved encapsulation of 71 molecules per AFt cage. Dynamic light scattering (DLS) studies of the resulting solution revealed an average size of 13.1 ± 1.6 nm, as expected for single AFt cages, confirming that the size and the integrity of the AFt had not been affected by encapsulation of 5F 203. Transmission electron microscopy (TEM) studies confirmed that integrity of the AFt capsule is not affected by the encapsulation. We observed the AFt capsules with an average diameter of 11.6 ± 1.9 nm (Figure 1A).

## Examining Encapsulation Efficiency of the Agent

In order to investigate whether the encapsulation efficiency could be improved using the reassembly route, two methods were trialed: pH-induced and urea-induced reassembly. For pH-induced reassembly, 5F 203 was added to AFt subunits (disassembly at pH = 2) and the pH was raised to pH 4.0 (using 1M, NaOH) and then to 7.4 (by addition of 1M, tris

buffer), to gradually reform the AFt cages. The AFt remained in solution, but significant loss of test agent was observed (only 1.3% of test agent encapsulated). To address obstacles posed by agent solubility, we used a glycine/HEPES buffer and reduced the concentration of AFt (1.5 mg mL<sup>-1</sup>). The reassembly of AFt capsules was initiated by direct addition of base; this was necessary to limit the time that the solution was kept in the pH 4.5–6.5 range, which is close to the isoelectric point of the AFt subunits and could lead to precipitation.<sup>12</sup> This method resulted in >55% of AFt remaining in solution and encapsulation of 25 molecules of 5F 203 per AFt cage. However, DLS studies indicated that the reassembly of the AFt capsule under these conditions was altered leading to the formation of larger protein cages (average diameter 20.9 ± 2.8 nm). Following filtration, only a small fraction of the samples demonstrated the expected diameter of 12.7 ± 1.8 nm. These results could be associated with the formation of nanocages larger than the expected 24-mer, or possible damage to AFt leading to the formation of holes in the AFt surface.<sup>23</sup> It was hypothesised that disassembly of AFt using the urea method may provide more efficient protein reassembly;<sup>24</sup> However, the results of encapsulation were similar to those obtained using the pH method. Since the reassembly routes failed to increase the encapsulation efficiency, the nanoreactor method with gradual addition of the test agent as multiple aliquotes was adopted for all subsequent studies. Encapsulation using the nanoreactor route was performed at pH 5.5. We anticipate no significant leakage of the encapsulated agent during storage (pH 7 and T = 4°C) as AFt pore size is reduced under these conditions.

To confirm that 5F 203 was encapsulated within the AFt cavity and not bound to the surface, time of flight secondary ion mass spectrometry (ToF-SIMS) was performed (Figure 1B and C). We observed a series of distinctive peaks corresponding to AFt at 56–58 m/z range and to 5F 203 at 255–295 m/z range. With increasing sputter time, the intensity of the peaks at 56–58 m/z is reduced and accompanied by increased signal at the 5F 203 range. The peaks of the drug are not visible in the AFt, until the protein shell is removed. We note that the signals corresponding to common contaminants such as Na<sup>+</sup>, K<sup>+</sup>, and PDMS did not reduce with increasing sputter time. Hence, depth profiling using an Argon cluster beam to remove a layer of the sample was applied and revealed 5F 203 peaks exclusively below the AFt surface, within the cavity. These ToF-SIMS results confirm that the drug is encapsulated inside the AFt



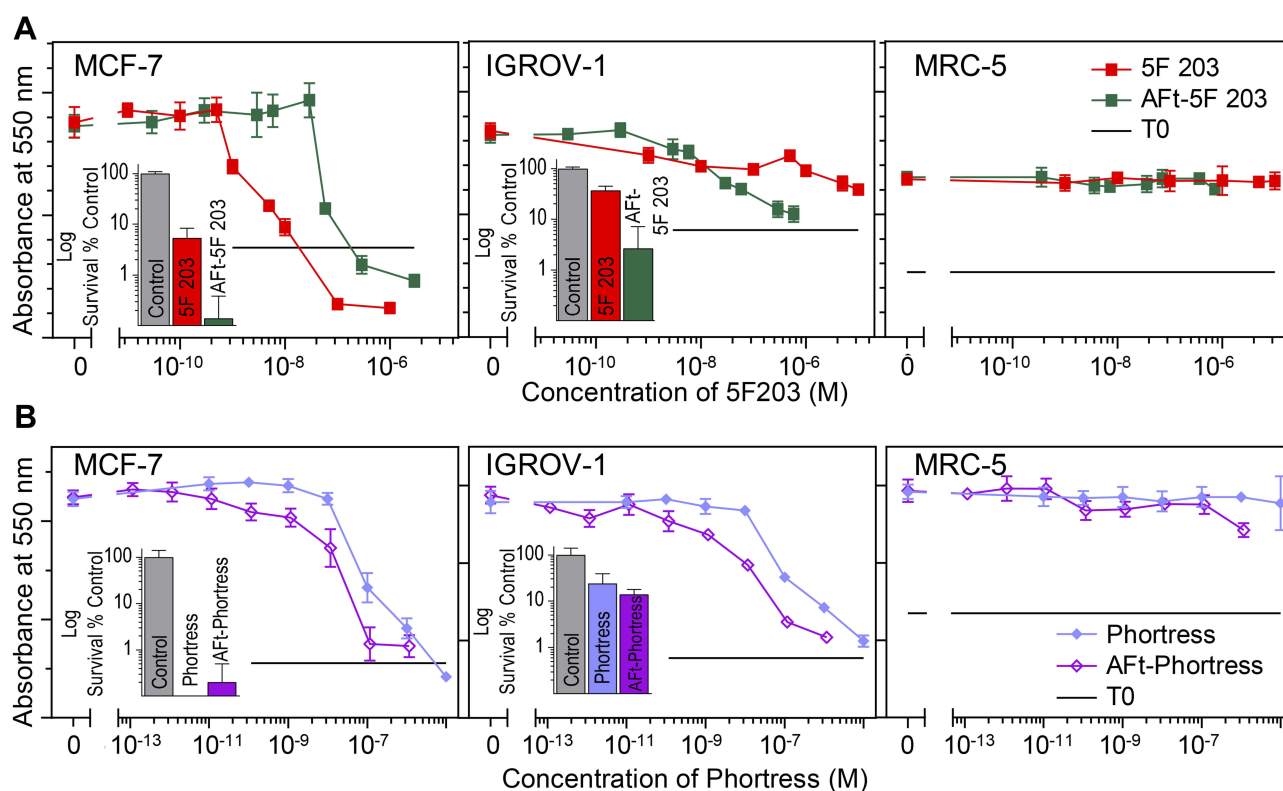
cavity and provide the first direct examination of the morphology of the protein encapsulated structures.

## In vitro Assessment of Anticancer Activity

We recently reported that conjugation to a lysine moiety significantly enhances the encapsulation of benzothiazole GW 610 in AFt, resulting in a more potent formulation.<sup>25</sup> Moreover, the lysylamide prodrug of 5F 203 (Phortress) has advanced to clinical evaluation because of its selective potent antitumour activity in vitro and in vivo against breast and ovarian cancer models, unique mode of action and favourable pharmacokinetic properties.<sup>20,26</sup> Therefore, Phortress, was encapsulated within AFt. Promising encapsulation results were achieved with 130 molecules of Phortress per AFt cage, with a corresponding encapsulation efficiency (EE) of 8% and drug loading of 10%. Phortress encapsulation efficiency was slightly higher compared to 5F 203, likely a consequence of the difference in polarity and lipophilicity of the test agents ([Supplementary Information S11](#), [Scheme S1](#) and [Figure S2](#)).

The rate at which test agents are released from a delivery capsule could significantly impact its therapeutic properties. Hence, release rates were studied at physiologically relevant pH values of 7.4 and 6.5. It has been reported that a lower pH of ~6.5 is normally associated with tumour microenvironments.<sup>27</sup> [Figure 2](#) shows that the release of 5F 203 occurs steadily over 24 hrs with 100% release observed <24 hrs at both pH 6.5 and 7.4 at 37°C. Phortress was readily released from the AFt core with 100% release observed within 9 hrs. This observation is in contrast to the reported release of doxorubicin from AFt,<sup>24</sup> where no significant release was observed at pH 7.4 after 60 hrs. This difference may be attributed to the combination of the size and polarity of the test agents, i.e. doxorubicin is a larger (544 g mol<sup>-1</sup>), more polar molecule than 5F 203 (258 g mol<sup>-1</sup>) and Phortress (386 g mol<sup>-1</sup>), hence its diffusion out from the lumen/core through the AFt pores (3–4 Å diameter) is unlikely at near neutral pH.

To understand the effects of encapsulation on the in vitro antitumour activity of the benzothiazoles, both non-encapsulated (naked) and encapsulated forms of 5F 203 and Phortress were tested against a range of cell lines



**Figure 2** MTT assay (72 hrs exposure) dose–response curves of **(A)** 5F 203 and AFt-5F 203 and **(B)** Phortress and AFt-Phortress in MCF-7, IGROV-1 and MRC-5 cell lines. Data points are mean  $\pm$  SD, n = 4 in one representative trial, number of independent trials = 3. Insets: Clonogenic assay displaying survival fractions following 24 exposure treated at GI<sub>50</sub> values in corresponding cell lines. Data points are mean  $\pm$  SD, taken from 3 independent trials where n=4 per trial.

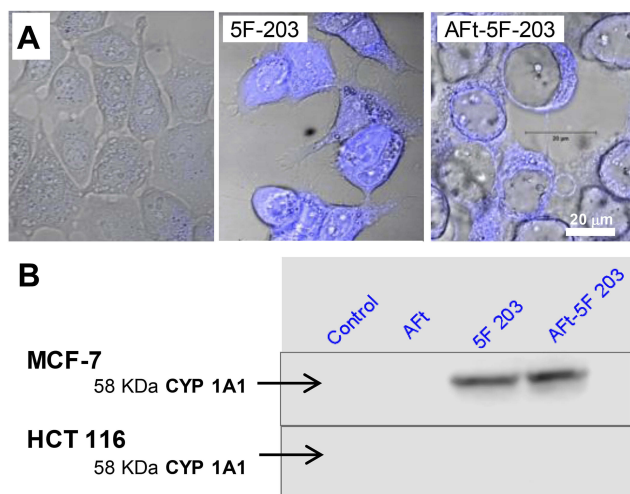
**Table 1** GI<sub>50</sub> Values Calculated from MTT Assay (72 Hrs) of Naked and Encapsulated Forms of 5F 203 and Phortress. GI<sub>50</sub> Is Mean  $\pm$  SD from Three Independent Trials, Where N = 4

	Cells Line	Test Agent			
		5F 203	AFt-5F 203	Phortress	AFt- Phortress
GI <sub>50</sub> $\pm$ SD ( $\mu$ M)	MCF-7	0.005 $\pm$ 0.0006	0.033 $\pm$ 0.009	0.085 $\pm$ 0.017	0.063 $\pm$ 0.030
	MDA-468	0.014 $\pm$ 0.007	0.008 $\pm$ 0.008	0.11 $\pm$ 0.013	0.11 $\pm$ 0.035
	TK-10	0.11 $\pm$ 0.09	0.028 $\pm$ 0.004	6.3 $\pm$ 2.9	0.098 $\pm$ 0.008
	IGROV-1	12.8 $\pm$ 10.2	0.039 $\pm$ 0.011	0.17 $\pm$ 0.070	0.058 $\pm$ 0.022
	HCT-116	>50	>0.61	>50	0.14 $\pm$ 0.059
	MRC-5	>50	>0.61	>50	>2.5

via MTT assays following 72 hrs exposure of cells to test agent. The cell lines selected for study included: MCF-7, MDA-MB-468 breast, TK10 renal, IGROV-1 ovarian, HCT 116 colorectal (CRC) cancer cell lines and non-tumorigenic MRC-5 fibroblasts. The results of the MTT tests are summarised in Table 1 and representative dose-response curves are illustrated in Figure 2 and SI Figures S3 and S4. In the majority of sensitive cell lines, the encapsulated benzothiazole is more potent than the naked agent. The increase in potency for example in TK10 renal and IGROV-1 ovarian cell lines, respectively, is  $\sim$ 4 $\times$  and  $>$ 300 $\times$  for AFt-5F 203, and  $>$ 60 $\times$  and 3 $\times$  for AFt-Phortress. In the benzothiazole-insensitive cell line HCT-116 and in non-tumorigenic fibroblasts (MRC-5), both 5F 203 and Phortress were inactive (GI<sub>50</sub> values  $>$ 50  $\mu$ M). No growth inhibition was encountered following AFt-

encapsulation of either agent in MRC-5 cells (Table 1) confirming that cancer cell-selectivity is preserved following encapsulations. Surprisingly, HCT 116 CRC cells (which do not express inducible CYP1A1; SI Figure 3) did demonstrate sensitivity to AFt-Phortress (Table 1 and Supplementary Information SI2 and Figures S3 and S4). Similar to the naked agent, AFt-5F 203 was more active than AFt-Phortress in all sensitive cell lines tested, exemplified by increased potency (with correspondingly reduced GI<sub>50</sub> values) by factors of  $\sim$ 1.5 $\times$  in IGROV-1, and 3.5 $\times$  in TK-10. However, if GI<sub>50</sub> values in relation to AFt concentration are considered, AFt-Phortress is more potent (than AFt-5F 203) in the sensitive cell lines tested, increasing potency by a factor ranging from  $\sim$ 3 $\times$  for MCF-7 and up to  $\sim$ 22 $\times$  for TK-10. The in vitro antitumour activity observed in the MTT assays was corroborated by clonogenic assays (Figure 2, SI S3 and S4). Clonogenic survival was inhibited in all sensitive cell lines tested and AFt-encapsulated benzothiazoles were either equally, or more potent than free drug. Thus, not only is the enhanced potency of encapsulated benzothiazoles confirmed, but importantly, clonal survival of benzothiazole-sensitive cancer cells is inhibited following brief exposure of cells to AFt-5F 203 and AFt-Phortress. AFt alone did not cause growth inhibition or toxicity in either the MTT or clonogenic assays (up to 1 mg mL<sup>-1</sup>), confirming the biocompatibility of this protein and its potential for drug delivery applications.

We envisage that the enhanced toxicity of the encapsulated agents is a consequence of cellular recognition of AFt by the transferrin receptor (TfR1) and receptor-mediated endocytosis.<sup>28</sup> Membrane TfR1 expression is markedly upregulated in many cancers including colon and breast.<sup>14,16</sup> To examine this hypothesis, we assessed TfR1 expression in the studied cell lines. The results confirm TfR1 expression in lysates of most cancer cells,



**Figure 3** (A) Confocal microscopy images of MCF-7 cells and following 24 hrs exposure of cells to AFt, 5F 203 alone and AFt-5F-203 prepared via nanoreactor route. (B) Selective induction of CYP1A1 protein in lysates of sensitive MCF-7 cells only following exposure to naked and encapsulated agent (24 hrs; 1  $\mu$ M 5F 203); lysates prepared from HCT 116 cells express neither inducible nor constitutive CYP1A1.

revealing an order of TfR1 expression in benzothiazole-sensitive carcinoma cell lines as follows: KM-12 > MCF-7 > TK-10 > HCC-2998 > IGROV-1 > MDA-MB-468 (Supplementary Information S12 and Figures S5 and S6). Insensitive MRC-5 fibroblasts showed undetectable levels of TfR1 expression, whereas TfR1 expression was evident in HCT 116 lysates. These results are consistent with observed differences in GI<sub>50</sub> values following benzothiazole encapsulation. For example, the highest expression of TfR1 was seen in KM-12 cells; as reported, enhanced potency of Aft-encapsulated benzothiazoles GW 610 and GW608-Lys, by ~50- and >750-fold, respectively, was seen in this cell line.<sup>25</sup> Moderate expression of TfR1 in TK-10 (and HCC-2998) cells corresponded to ~4-fold and ~64-fold enhanced activity of Aft-encapsulated 5F 203 and Phortress in TK10 cells. No TfR1 expression was detected in MDA-MB-468 cells, and no difference was observed in GI<sub>50</sub> values of naked or encapsulated Phortress (0.11 μM).

In the benzothiazole-insensitive HCT 116 cell line, growth inhibition was observed following exposure of cells to Aft-Phortress, facilitated (we speculate) by TfR1-mediated endocytosis of the Aft formulation. However, direct correlation between TfR1 expression and GI<sub>50</sub> values was not observed in MCF7 and IGROV-1 cell lines. There is strong TfR1 expression in MCF-7 cells; however, there was no enhanced potency of the encapsulated benzothiazoles; in IGROV-1 cells, there is minimal TfR1 expression, however dramatically increased potency following encapsulation of 5F 203 was observed. Abundant cytosolic AhR expression in MCF-7, IGROV-1 (and MDA-468) cells promotes rapid depletion of 5F 203 from nutrient medium, sequestration and ligand-receptor binding.<sup>29</sup> Un-encapsulated (naked) benzothiazoles rely on passive diffusion and their hydrophobic nature to cross the cell membrane and enter cells. Cytosolic AhR expression in cancer cells correlates with rapid sequestration of anti-tumour benzothiazoles such as 5F 203. These observations correlate with CYP1A1 induction and generation of DNA adducts over the initial 24 hrs exposure of MCF-7 and IGROV-1 cells to 5F 203, or Phortress.<sup>30,31</sup>

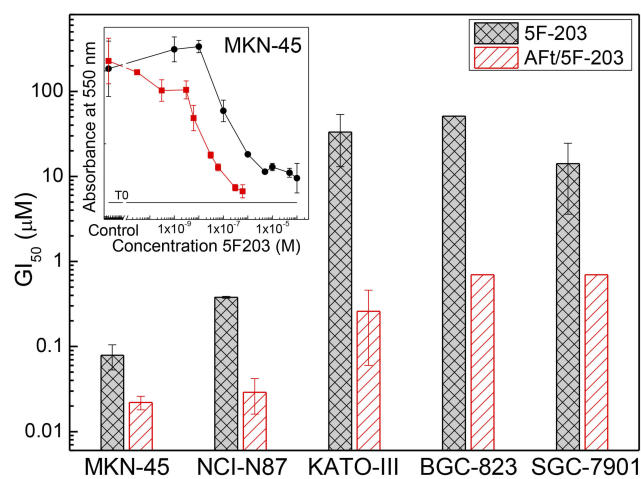
## Cellular Uptake of Aft-Encapsulated Agents

Confocal microscopy was performed following exposure of MCF-7 cells to naked and Aft-encapsulated 5F 203 (Figure 3). Additionally, lysates were prepared from MCF-

7 and HCT 116 cells exposed to naked and Aft-encapsulated 5F 203 and induction of CYP1A1 protein expression was examined to investigate AhR-signal transduction activation by 5F 203 (a potent AhR ligand).<sup>32</sup> Intracellular 5F 203 was clearly observed in MCF-7 cells exposed to both naked and Aft-encapsulated 5F 203 (Figure 3A). Moreover, clear expression of CYP1A1 protein exclusively in benzothiazole-sensitive MCF-7 cells exposed to naked and Aft-encapsulated 5F 203 provided evidence of AhR signal transduction. Aft alone did not induce CYP1A1, and no CYP1A1 protein was detected in lysates of insensitive HCT 116 cells exposed to 5F 203 or Aft-5F 203 (Figure 3B). As inferred, the surprising activity observed following exposure of HCT 116 cells to Aft-Phortress maybe a consequence of its TfR1-mediated cellular internalization. We conclude that CYP1A1-inducibility by anti-tumour benzothiazoles is not compromised following encapsulation in Aft protein nanocages.

## Studies in Gastric Cell Lines

Encapsulation of benzothiazoles into the Aft cavity offers a number of advantages compared to other delivery vehicles, such as enhanced solubility, bioavailability and selective uptake. Passive tumour-targeting to the tumour microenvironment can be achieved via EPR, and active tumour targeting via TfR1-mediated internalization (and cytosolic AhR-mediated sequestration). Only a subset of cancers demonstrates sensitivity to anti-tumour benzothiazoles with ≥4 log-fold selectivity between sensitive and



**Figure 4** Histograms of GI<sub>50</sub> values for all gastric cell lines treated with 5F 203 and Aft-5F 203; means ± SEM from 3 independent trials; n=4 per trial. (Inset) Representative dose-response profiles following exposure of gastric cell line MKN-45 to 5F 203 and Aft-5F 203. MTT assays were performed after 72 hrs treatments.

**Table 2** Summary of GI<sub>50</sub> Values Calculated from MTT Assay (72 Hrs) in Gastric Cell Lines for All Test Agents. GI<sub>50</sub> Is Mean ± SD from Three Independent Trials, Where N = 4

Test Agent	GI <sub>50</sub> ± SD (µM)				
	Cell Line				
	MKN-45	NCI-N87	KATO-III	BGC-823	SGC-7901
5F 203	0.079±0.026	0.38	33.3±20.9	>51	14.1±18.5
AFt/5F 203	0.011±0.004	0.029±0.013	0.26±0.2	>0.7	>0.7
GW610	0.43±0.37	0.41±0.34	7.7±1.1	18.6±14.1	35.1±8.6
AFt-GW610	0.036±0.032	0.48±0.34	0.42±0.33	1.4±0.5	>1.8
GW608Lys	22.1±16.8	38.1	41.7±21.4	>50	25.9±22.5
AFt/GW608Lys	0.21±0.13	0.59	>3.5	>3.5	>3.5

inherently resistant phenotypes.<sup>33–36</sup> Sensitive tumour types can be identified through ex vivo examination of activity and pharmacodynamic (PD) biomarkers of sensitivity (AhR, DNA adducts).<sup>37</sup>

An additional important benefit of AFt encapsulation, as alluded to in ref. [27], is the sensitivity of the protein capsule to pH, which could offer drug delivery and treatment advantage for gastric cancer. The AFt cage would disassemble in this highly acidic environment releasing its cargo, in this case a potent AhR ligand, locally, at the tumour site. AhR is expressed by gastric cancer cells and plays a role in gastric carcinogenesis.<sup>38,39</sup> Moreover, CYP1A1 mRNA and protein expression are inducible in gastric cancer, and CYP 2W1 protein, embryonically expressed in gastrointestinal tissues is highly expressed in gastric cancer.<sup>40</sup>

Our studies in gastric cancer models further validated the selective activity in vitro and efficacy in vivo of 5F 203, revealing correlation in this cancer phenotype between benzothiazole activity and CYP 1A1 induction/CYP2W1 expression.<sup>41</sup> In our preliminary studies, ~10-fold enhanced potency was observed for AFt-encapsulated compared to naked 5F-203 in NCI-N87 cancer cells (GI<sub>50</sub> values of 0.04 µM and 0.38 µM, respectively, were obtained); ~7-fold increased activity was seen in MKN-45 cancer cells following 5F 203 AFt-encapsulation (GI<sub>50</sub> values: 5F 203, 0.011 µM; AFt-5F 203, 0.079 µM). In KATO-III cells, GI<sub>50</sub> values of 33.3 µM (5F 203) and 0.26 µM (AFt-5F 203) reflect >100-fold enhanced potency following AFt-encapsulation (Figure 4A and Table 2). Further enhancement of the activity of observed following AFt-encapsulation of benzothiazoles GW 610 and GW608-Lys (~180 and ~380 molecules/AFt cage, respectively),<sup>25</sup> where up to 100-fold reduction in GI<sub>50</sub> values was observed for encapsulated agents in the MKN-45 gastric cell line (Figure 4B and C and Table 2).

We envisage that potency may be further enhanced in vivo where agents will be exposed to pH ~2. These results demonstrate a significant advantage of the AFt delivery capsule and merit further assessment in vivo.

## Conclusions

In conclusion, we have developed and optimized a robust biocompatible, formulation for antitumour benzothiazoles that involves encapsulation within AFt protein cages for selective treatment of benzothiazole-sensitive cancer phenotypes. Improved encapsulation has been demonstrated for lysylamide prodrugs Phortress and GW 608-Lys compared to parent 5F 203 and GW 610, respectively; enhanced in vitro antitumour activity of AFt-encapsulated formulations compared to naked 5F 203, Phortress, GW 610 and GW 608-Lys has been encountered; finally, potency against carcinomas derived from breast, ovarian, renal, colorectal as well as gastric cancer models, for which cargo-release at low pH is pertinent, has been demonstrated. Our results herald exciting prospects for potential refinement of tumour-targeting, imaging (e.g. fluorescence of 5F 203 itself or <sup>18</sup>F-labelled 5F 203) and treatment, and merit further in vivo investigations.

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## Author Contributions

All authors have made substantial contributions to conception and design of the study, to data acquisition, analysis and interpretation, and towards drafting and critically revising the paper, gave final approval of the version to be published, and agreed to be accountable for all aspects of the work. Major individual contributions are listed as follows. AFB and MLC performed encapsulation experiments, AFB optimized encapsulation procedure, characterized the material and has done in vitro studies. DS provided expertise and performed ToF SIMS measurements. JZ contributed to studies in gastric cell lines. AFB, GW, NRT, LT and TDB conceptualized and designed the project with contribution from all co-authors. AFB, LT and TDB co-wrote the manuscript.

## Disclosure

The authors report no conflicts of interest in this work.

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