Copper Physiology in Ruminants: Trafficking of systemic copper, adaptations to
 variation in nutritional supply and thiomolybdate challenge
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8 ABSTRACT

9 Ruminants are recognised to suffer from copper responsive disorders. Present understanding of copper transport and metabolism is limited and inconsistent across vets and veterinary 10 professionals. There has been much progress from the studies of the 1980s and early 90s in 11 cellular copper transport and liver metabolism which has not been translated into agricultural 12 practice. Copper metabolism operates in regulated pathways of copper trafficking rather in than 13 14 pools of copper lability. Copper in the cell is chaperoned to enzyme production, retention within metallothionein or excretion via the Golgi into the blood. The hepatocyte differs in that 15 16 copper-containing caeruloplasmin can be synthesized to provide systemic copper supply and excess copper is excreted via bile. The aim of this review is to improve understanding and 17 18 highlight the relevant progress in relation to ruminants through the translation of newer findings from medicine and non-ruminant animal models into ruminants. 19

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21 **KEYWORDS:** Ruminant, copper transport, liver metabolism, thiomolybdate

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23 INTRODUCTION

Copper metabolism in ruminants remains poorly understood in practice ⁽¹⁻⁵⁾. Developments in the fundamental understanding of copper physiology have been insufficiently translated into livestock nutrition. While there is some awareness among industry professionals of the effects of 'copper deficiency' and of the potential nutritional effects by antagonists it is inconsistently understood ⁽⁶⁾. Vets vary in their response to copper-related problems some may discourage supplementation in fear of toxicity problems, while others may continue to supplement ^(3,5-7). There is considerable marketing pressure from mineral suppliers for their products and an inclination from producers to seek a 'quick fix' for trace element supplementation ⁽⁸⁾.

Recent surveys have found UK sheep and cattle are commonly affected by different forms of 32 copper imbalance, including toxicity and deficiency ^(9,10). Kendall et al. ⁽¹⁰⁾ reported as many 33 as 40% of British dairy cattle may be accumulating excessive liver copper, with up to 52% of 34 them above the Animal Health Veterinary Laboratories Agency (AHVLA) reference range of 35 300-8,000 µmol/kg DM (10). Copper imbalance was the most common mineral problem 36 reported between 2004 and 2014; with ~300 fatal occurrences each year reported for cattle and 37 sheep combined for both toxicity and deficiency (11-13). Indications from academic studies, 38 government reports and industry suggest that copper imbalance is still highly prevalent ^(3,5,14,15). 39 Highlighting that copper supplementation remains a problem in ruminant production. 40

This review focusses on post-absorptive trafficking and systemic regulation of copper and describes the interference of thiomolybdates on these mechanisms. A review of the role of the rumen in thiomolybdate formation has been previously published ⁽¹⁶⁾.

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45 COPPER METABOLISM AT CELLULAR LEVEL

Most recent fundamental knowledge generated on copper biology has been produced with models such as cell culture, *c.elegans*, laboratory animals and humans ⁽¹⁷⁾. These selected species concentrate on a medical or nutritional perspective. The lack of emphasis on ruminants, and the limited overlap with human focused sciences, has prevented dissemination of this new understanding; resulting in a lack of progress from the classic ideas on copper in ruminants.

The copper chaperones and enzymes which exist in ruminants are the same as those studied in other mammalian species ⁽¹⁷⁾. At cellular level, basic copper metabolism appears to be consistent throughout eukaryotic life and can be traced from laboratory animals to humans through their shared evolution ⁽¹⁸⁾; demonstrating that copper in the systemic circulation is trafficked in the same manner in mammalian cells thus providing opportunities to expand our understanding of copper metabolism in ruminants ⁽¹⁷⁾.

Since 1966 radiolabelled copper, cell fractionation and isolation of intracellular membrane
components have been used to develop mathematical models to describe copper movement in
rat liver ^(19,20). This led to the concept that separate pools, of varying availability existed ⁽²¹⁾.
Initially, the pools were designated as 'storage', 'synthetic' and 'excretory' ⁽¹⁹⁾. The

61 relationship between the pools appeared complex, with no evidence of reversible movement between them. It was suggested the copper pools were able to become saturated, and the 62 regulation or exchange between the pools was not determined ^(21,22). The number and function 63 of the pools was not easily apparent. Most studies agreed hepatocyte copper could be divided 64 into at least two pools, one a readily available, extractable copper pool accounting for the 65 majority of copper. The second, a less readily available pool containing the remainder of 66 soluble copper and potentially a third, non-extractable, insoluble pool which could be 67 considered a potential subset of the second pool ^(20,22). By 1987 it was proposed that three 68 separate pools existed within the liver representing bile, caeruloplasmin and 'storage' which 69 was not further defined ⁽²¹⁾. 70

Subsequent research has mapped the intracellular movement of copper and improved our 71 understanding of copper distribution in cells ^(23–27). Fundamentally, this new knowledge does 72 not contradict the description of copper as cellular pools, but it illustrates copper physiology in 73 terms of copper trafficking. Free copper ions rarely exist within cells, thus copper is kept 74 complexed to prevent intracellular damage ⁽²⁸⁾. Distinct intracellular pathways exist where 75 copper is bound to chaperones and channelled across membranes rather than a series of storage 76 compartments as the older model suggests. However, the persistence of the term 'pool', even 77 78 in current literature, conjures images of discrete areas. It is perhaps better to update our terminology, and start discussing the 'pathways' of copper trafficking, rather than its 'pools' 79 80 of availability to better reflect the process and improve understanding of the process as a continuous regulation instead of discrete compartments of varying lability. 81

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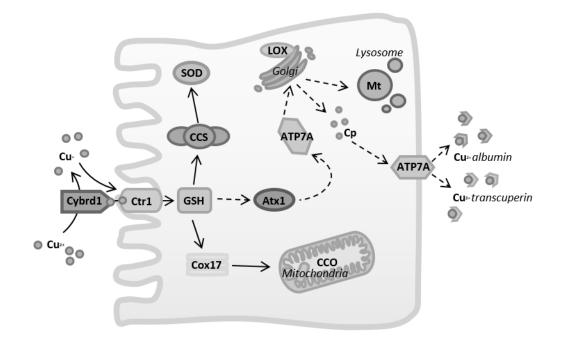
83 OVERVIEW OF COPPER TRAFFICKING IN ENTEROCYTES

The one aspect of copper metabolism that differentiates ruminants from other species is their 84 unique digestive system. Copper availability in the ruminant gastrointestinal tract presents 85 peculiarities that are extensively reviewed elsewhere (16,29,30). However, the process of 86 absorption is well-preserved across the animal kingdom (31-33). In order for copper to be 87 absorbed, it must be reduced into its most reactive state (Cu⁺). At the intestinal brush border a 88 copper specific transporter (Ctr1) is responsible for ~70% of copper uptake into the enterocyte, 89 the remainder is taken up by the non-specific transporter Divalent Metal Transporter 1 (DMT1) 90 91 ⁽³⁴⁾. Where copper is trafficked through the DMT1 route direct competition for the transporter with dietary elements such as iron and zinc may be more biologically relevant ⁽³⁵⁾. Once inside 92

93 the cell, copper chaperone proteins bind copper and transport it to other specific proteins or 94 incorporate it into enzymes. The pathway via the Golgi is known as the secretory pathway. 95 Copper in excess of cellular requirements enters the secretory pathway to be bound to 96 metallothionein by the Golgi and is stored in the lysosome, which acts as a buffer restricting 97 free cellular copper. Once the metallothionein reaches its saturation capacity copper continues 98 through the secretory pathway from the Golgi via its chaperone to the basolateral membrane 99 for efflux from the cell.

100 The process in detail

101 *Figure 1 below illustrates the process described.*



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Figure 1: Copper trafficking pathways using the copper chaperones from the intestinal lumen.
Atx1, antioxidant 1; CCO, cytochrome c oxidase; CCS, copper chaperone protein; Cox17,
cyclo-oxygenase 17; Cp, caeruloplasmin; Ctr1, copper transporter 1; Cybrd1, cytochrome B
reductase; GSH, glutathione; LOX, lysyl oxidase; Mt, metallothionein; SOD, superoxide
dismutase.

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109 Upon arrival at the intestinal brush border the membrane reductase Cybrd1 (Cytochrome B 110 Reductase 1) and ascorbate (Vitamin C) reduce any dietary copper which is present as Cu^{2+} 111 into Cu^{+} (^{36–38)}. Reduced copper is carried across the membrane by high-affinity Copper 112 transporter 1 (Ctr1) (^{34,39–41)}. Once inside the cell it is immediately incorporated onto its specific

chaperones (CCS, Atx1 and Cox17) within the cytosol ^(42,43). Copper chaperone protein (CCS) 113 transports copper within the cytosol where the metalloenzyme Superoxide dismutase (SOD) is 114 synthesised ⁽¹⁷⁾. Cyclo-oxygenase 17 (Cox17) transports copper to proteins in the mitochondria 115 where the metalloenzyme Cytochrome c oxidase (CCO) is synthesised ^(44,45). Anti-oxidant 1 116 (Atx1) and ATP7A transport copper to the Golgi lumen where dopamine β -hydroxylase, 117 peptidylglycine α -amidating monooxygenase, lysyl oxidase (LOX), SOD, tyrosinase, 118 caeruloplasmin (Cp) and hephaestin vital for nerve and connective tissue function and for 119 copper and iron transport are synthesised ^(18,46). Surplus copper is bound to Metallothionein 120 (Mt) and held in the lysosome after processing by the Golgi (18,44,47,48). Upon reaching the 121 metallothionein carrying capacity in the lysosome, surplus copper from the Golgi is transported 122 using the ATP7A secretory pathway and effluxed from the enterocyte into circulation ^(17,18,45). 123 At the point of release from the cell membrane the oxygen tension of the interstitial fluid is 124 sufficient to elicit spontaneous oxidation of the Cu⁺ to oxidised Cu²⁺ without the need for an 125 oxidase in the membrane $^{(49)}$. 126

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128 COPPER MOVEMENT IN THE BLOOD

Following efflux from the enterocytes copper is bound to albumin; an abundant plasma protein 129 accounting for 15-20% of total copper transport, and transcuprein; a small protein which in 130 contrast to albumin, is a specific copper carrier in plasma carrying 10-30% of total transported 131 copper ^(34,50–53). The concentration of albumin in blood plasma exceeds that of transcuprein, but 132 transcuprein has a higher affinity for copper. Around a third of the copper entering the blood 133 from the small intestine is bound to transcuprein ⁽⁵³⁾. These two proteins transport copper from 134 the intestines through the systemic circulation to the liver. Metabolic studies have demonstrated 135 that absorbed dietary copper from the portal circulation is cleared by the liver and appears in 136 newly synthesised caeruloplasmin ⁽⁵⁴⁾. Caeruloplasmin is the predominant copper transporter 137 in the systemic blood and is responsible for distribution of copper to the tissues after its 138 synthesis in the liver ^(55,56). In ruminants around 88% (range 86-90%) of total plasma copper is 139 present bound to caeruloplasmin⁽⁵⁷⁾. 140

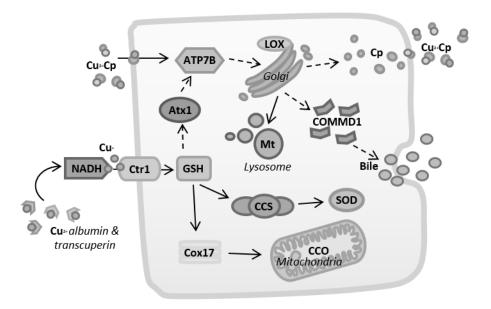
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142 OVERVIEW OF HEPATIC COPPER TRAFFICKING

143 The liver has a major role in the regulation of copper ⁽²⁸⁾. This homeostatic control acts 144 primarily through regulating the secretion of copper into bile ^(36,43,50,58). Copper reaching the 145 liver is transported in a similar mechanism to the enterocytes. At the membrane the arriving copper is reduced and trafficked into the cell by the same copper transporter (Ctr1). Once inside 146 the hepatocyte the chaperones fulfil their respective roles with one notable difference. The 147 secretory pathway for efflux via the Golgi has a unique chaperone (ATP7B) which directs the 148 majority of copper to be incorporated into caeruloplasmin which is then effluxed into 149 circulation for distribution to other tissues. However, when caeruloplasmin bound copper from 150 the peripheral tissues re-enters the circulation and returns to the liver the whole molecule of 151 caeruloplasmin is absorbed for destruction and excretion through the biliary route. 152

153 The process in detail

154 *Figure 2 illustrates the process described below.*



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Figure 2: Copper trafficking pathways using the copper chaperones into hepatocytes and out
into systemic circulation. Atx1, antioxidant 1; CCO, cytochrome c oxidase; CCS, copper
chaperone protein; COMMD1, copper metabolism MURR1 domain; Cox17, cyclo-oxygenase
17; Cp, caeruloplasmin; Ctr1, copper transporter 1; Cybrd1, cytochrome B reductase; GSH,
glutathione; LOX, lysyl oxidase; Mt, metallothionein; SOD, superoxide dismutase.

Copper reaches the liver bound to either transcuprein or albumin which are reduced on arrival
 by NADH oxidase ⁽⁵²⁾. Uptake of the reduced copper into the hepatocyte is mediated by Ctr1
 ⁽⁵⁹⁾. Once inside, CCS and Cox 17 traffic their copper payload to the cytosol and mitochondria

respectively and Atx1 delivers copper to the Golgi body via ATP7B⁽⁶⁰⁾. ATP7A is not 165 expressed in the liver, instead hepatocytes express a unique version ATP7B (44). ATP7B directs 166 the majority of copper to be incorporated into caeruloplasmin to be subsequently returned to 167 the circulation for distribution to other tissues ^(17,28,40,44,60). When caeruloplasmin returns from 168 systemic circulation to the hepatocytes the whole molecule is absorbed. The endothelial 169 hepatocytes must first remove sialic acid residues from the caeruloplasmin to allow the 170 underlying hepatocytes to absorb the caeruloplasmin molecule for proteolysis and destruction 171 through the biliary route ⁽⁵⁸⁾. The excess hepatic copper is exported into the bile using the 172 chaperones COMMD1 (copper metabolism MURR1 domain) and potentially also XIAP (X-173 linked inhibitor of apoptosis protein) ^(36,40,60). COMMD1 binds to the N-terminal region of 174 ATP7B but not to ATP7A, explaining the difference in ATPase channel expression between 175 hepatocytes and other cells ^(60,61). 176

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178 ADAPTATIONS TO CHANGING DIETARY COPPER SUPPLY

Under copper-limiting conditions the movement of copper into the secretory pathway (Atx1-179 ATP7A) is diminished in all tissues ^(25,50). Copper bound to metallothionein is mobilised using 180 the acidic pH of the lysosome to partially degrade the metallothionein held within the lysosome 181 and release its copper into the cytosol ^(18,62,63). The released copper is delivered, likely by 182 glutathione (GSH), to the copper chaperones (cytosolic CCS and mitochondria targeting 183 Cox17) equally, but not into the secretory pathway $(Atx1)^{(25,63,64)}$. This redirection diminishes 184 copper supply to the secretory pathway resulting in the production and secretion into the 185 bloodstream of the copper-empty apo-caeruloplasmin, rather than its copper-containing holo 186 187 form ⁽⁶³⁾. This process inhibits excretion and retains copper for intracellular use ⁽⁶⁵⁾.

Under copper replete conditions in the tissues each of the copper transporters and proteins are 188 down-regulated ^(25,48). The down-regulation of copper transporter (Ctr1) in the membrane 189 prevents any further copper uptake into the cell ^(66–68). ATP7A (a chaperone in the secretory 190 pathway) moves out of the trans-Golgi network into vesicles that move towards the membrane. 191 These vesicles accumulate copper and intermittently fuse with the membrane to efflux the 192 remaining excess copper from the cell into the blood before returning to the cytoplasm ⁽⁶⁹⁾. 193 Increased metallothionein expression (regulated by Metal transcription factor MTF1) exerts 194 intracellular homeostatic control through binding excess copper and acting as storage buffer 195 protecting the cell ^(18,65). 196

When hepatocytes are exposed to increasing copper concentrations they behave similarly to other cells with one exception; ATP7B (from the hepatocyte secretory pathway) leaves the trans-Golgi network but instead of moving towards the membrane it moves towards the lysosome at the canalicular membrane ^(50,65). Here, the ATP7B imports copper into the lysosomal lumen for temporary storage. Increasing intracellular copper concentrations induce exocytosis of the lysosome releasing the excess copper into the biliary canal (mediated by the secretory chaperones ATP7B and COMMD1) ^(25,36,60,70,71).

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205 RUMINANT COPPER SENSITIVITY

206 When discussing the unique characteristics of ruminant copper handling it is important to first note that metallothionein knock-out animals, even from monogastric species, are 207 hypersensitive to copper ⁽⁷²⁾. Sheep have a limited ability to synthesise metallothionein in 208 response to rising copper concentration and they appear to have a restricted capacity to 209 accumulate copper bound to metallothionein in the liver ^(56,73). In comparison to rats, sheep 210 reach a point where metallothionein synthesis is unable to keep up with rising copper at a much 211 lower dietary inclusion resulting in less copper sequestering by the lysosome ⁽⁷³⁾. Additionally, 212 sheep have a limited ability to increase biliary copper excretion in response to copper intake 213 ⁽⁷⁴⁾. Cattle also have a lower capacity to store copper bound to metallothionein in comparison 214 to monogastric species and a limited capacity to induce metallothionein in response to copper 215 intake ^(56,75). Furthermore, in cattle and sheep the copper-buffering capacity decreases as 216 hepatic copper loading increases alongside the Cu:Zn ratio ⁽⁷⁶⁾. If the influx of copper exceeds 217 the capacity of the metallothionein and lysosomal uptake, unbound copper will occur in the 218 cytosol and begin to enter the nucleus, causing severe cell damage ^(76,77). While, pigs and dogs 219 have around 500-600 mg/kg, sheep and cattle have only ~200 mg/kg metallothionein in their 220 livers ⁽⁷⁷⁾. Additionally, the metallothionein transcription in the lysosome of cattle and sheep 221 does not effectively respond to rapid increases in copper ^(75,78), seemingly reaching a plateau of 222 total copper concentration ~1,607 mg/kg DM (25,347 µmol/kg DM) in cattle and ~571-643 223 mg/kg DM (9,006- 10,142 µmol/kg DM) in sheep ^(74,75,77,78). Potentially this plateau is linked 224 225 to the limited production of metallothionein and an inhibited biliary copper excretion ⁽⁷⁴⁾, theoretically explaining why cattle appear to be more copper tolerant than sheep and why both 226 species appear sensitive in comparison to monogastric species such as pigs. 227

Further to species differences, breed differences among ruminants have also been documented. 228 Texel sheep are more sensitive to copper than Landrace breeds ^(79,80). In cattle, Holstein and 229 Angus breeds are more copper tolerant than Jersey, Charolais and Simmental^(81–83). In cattle, 230 the more copper tolerant breeds exhibit a greater expression of duodenal Ctr1 and ATP7A, and 231 232 a higher hepatic expression of; Ctr1, Cox17, ATP7B, CCS and SOD where copper supply is inadequate ^(84,85). These suggest the ability to increase expression of copper transporters and 233 chaperones allows more effective uptake and utilisation where copper supply is insufficient; 234 reducing the susceptibility of these breeds to deficiency in comparison to their counterparts 235 ^(84,85). This research highlights a potential mechanism for the observed breed differences, but 236 further studies in a wider range of breeds and in sheep, under elevated and copper replete 237 conditions would further clarify the role of transporter expression in copper sensitivity. 238

239 THIOMOLYBDATE DISRUPTION

Thiomolybdate is known to interact with copper. It naturally forms in the reducing environment 240 of the rumen between dietary sulphur and molybdenum. Thiomolybdate poses a problem for 241 copper availability and post-absorptive utilisation ^(29,86–88). Thiomolybdates interact with 242 available copper in the digestive tract forming an insoluble precipitate greatly reducing copper 243 availability ^(29,86–89). If there is insufficient copper where thiomolybdates form to 'de-toxify' 244 them they can be absorbed into systemic circulation, where they exert their affinity for copper 245 by complexing with copper contained in biological compounds rendering them biologically 246 inactive ^(16,90). Thiomolybdates are able to cross cell membranes but the mechanism by which 247 248 this takes place is unknown. However, once inside the cell they have the potential to disrupt copper transport through binding to copper located on the copper chaperones, transporters and 249 enzymes⁽¹⁷⁾. 250

Thiomolybdates can bind to copper in cuproenzymes including; caeruloplasmin, 251 metallothionein, CCO, SOD (90-93), and Atx1 (94). Binding does not remove the copper 252 component but renders it unable to perform redox reactions (vital to its biological function) 253 through the formation of a stable complex ^(16,29,95,96). Superoxide dismutase has been shown to 254 differ and copper may be partially stripped from this enzyme ^(97,98). In the case of the chaperone 255 Atx1, thiomolybdate supresses the incorporation of copper into the products of the secretory 256 pathway disrupting the activity of the Atx1⁽⁹⁴⁾. Thiomolybdates have a high affinity for copper 257 and they have no effect on other trace metals with similar properties such as iron, zinc or 258 cadmium (99,100). 259

261 PRACTICAL IMPLICATIONS

Copper provision in ruminants requires a careful balance between intake and availability. The 262 263 inhibited capacity of these species to adapt to copper influx explains their sensitivity to overloading. Routine calculation of copper intake at farm level is not routinely undertaken 264 which can lead to over-supply ^(11,101). Calculation of copper supply in combination with 265 monitoring of biological parameters as part of routine management allows a more accurate 266 assessment of copper status across the entire flock or herd to be made ⁽¹⁰²⁾. At present, liver 267 sampling is an under-utilised as a measure of herd or flock copper status, especially where there 268 269 is a history of oversupply. Annual monitoring of a representative sample, from cull animals or from biopsy, allows more effective long-term decisions to be made for copper provision. It has 270 271 been recently demonstrated that a significant linear relationship exists between increasing hepatic copper concentrations and the abundance of rhodamine stained granules in hepatic 272 tissue histology ⁽¹⁵⁾. This staining technique detects the copper-filled lysosomes which occur 273 as the cellular mechanism for copper storage becomes overwhelmed ⁽¹⁵⁾. In effect, their 274 presence has the potential to be used as an indicator that copper concentrations are in excess; 275 although this technique is not yet used in practice. Little correlation exists between hepatic 276 copper concentrations and copper concentrations in blood parameters (30,103). It is useful to bear 277 this in mind and employ both techniques in conjunction with each other to establish animal 278 status (30,103). 279

280 The potential danger posed through absorption of thiomolybdate causing disruption to systemic copper chaperones and cuproenzymes should also not be neglected. The use of blood assay is 281 282 of importance to help monitor changes in shorter-term copper status. Decreases in caeruloplasmin activity can be a useful indicator of systemic thiomolybdate presence or copper 283 deficiency over and above the use of caeruloplasmin concentration ⁽⁹¹⁾. Since the apo-protein 284 will continue to be synthesised in the absence of adequate, available hepatic copper while its 285 activity can be reduced to nil ⁽¹⁰⁴⁾. This measure is not without flaws, as caeruloplasmin is an 286 acute phase protein and can be elevated by infection or stress leading to falsely elevated 287 measures of copper status ^(30,105,106). Unfortunately, a single, reliable measure for copper status 288 does not yet exist. Therefore, it is important to use both blood and hepatic measures in 289 monitoring ruminant copper status in addition to monitoring nutritional input ^(11,101). 290 Furthermore, it is important in practice to provide an appropriate copper source, or combination 291

of sources, which will be sufficient to 'de-toxify' thiomolybdate before it is absorbed and retain
a sufficient supply of labile copper for absorption which does not provide an excess or exceed
legal restriction ⁽¹⁰¹⁾.

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296 CONCLUSION

297 Advances in understanding of the physiology of intracellular copper transport from fundamental biology have not effectively penetrated the field of ruminant nutrition leading to 298 299 widespread misunderstanding and consequently widespread copper imbalance in practice. The pathways of copper transport are synonymous with other mammalian species and much 300 301 information is available to underpin nutritional theory for ruminants. Greater understanding of the trafficking pathways and their response to over and under copper supply allows decisions 302 for copper supply to be more informed. In ruminants and in particular sheep, these pathways 303 have a limited ability to respond to changes in dietary copper supply which explains this species 304 sensitivity to copper oversupply. Thiomolybdates formed under ruminal conditions have been 305 shown to be able to interfere with the copper chaperone pathways leading to cellular disruption 306 of their function, if they are not effectively 'de-toxified' preventing their entry into systemic 307 circulation. Considering the cellular pathways for copper and their potential disruption through 308 thiomolybdate absorption can help to better inform supplemental actions to remedy copper-309 related disorders in practice. 310

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316 CONFLICT OF INTEREST

317 None

318 AUTHORSHIP

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322 Proofing of concept and article content and wording- NR Kendall, J Martin-Tereso, S Paine

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