



**Periconceptional Environment and the Developmental
Origins of Disease**

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Periconceptional Environment and the Developmental Origins of Disease

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19 **Abstract**

20 The concept emerging from Professor David Barker's seminal research on the developmental
21 origins of later life disease has progressed in many directions since first published. One critical
22 question is *when* during gestation might environment alter the developmental programme with such
23 enduring consequences. Here, we review the growing consensus from clinical and animal research
24 that the period around conception, embracing gamete maturation and early embryogenesis, might be
25 the most vulnerable period. We focus on four types of environmental exposure shown to modify
26 periconceptual reproduction and offspring development and health: maternal overnutrition and
27 obesity; maternal undernutrition; paternal diet and health; and assisted reproductive **technology**.
28 These conditions may act through diverse epigenetic, cellular and physiological mechanisms to alter
29 gene expression and cellular signalling and function in the conceptus affecting offspring growth and
30 metabolism and leading to increased risk for cardiometabolic and neurological disease in later life.

31
32 **Key words**

33 Embryo; sperm; parental nutrition; assisted reproductive **technology** (ART); epigenetics;
34 cardiometabolic disease.

35

36 Introduction

37 The concept of the early origins of disease associated with *in utero* environmental factors has been
38 advanced in both clinical and biological directions since the pioneering and ground-breaking
39 epidemiological discoveries by Professor David Barker and his colleagues. Developmental
40 programming of disease has been tested experimentally across global populations providing
41 confirmation of its veracity. In addition, numerous animal models have been generated for insight
42 on mechanisms across physiological, cellular, molecular and epigenetic levels. Much progress on
43 the understanding of the hypothesis, now known as the Developmental Origins of Health and Adult
44 Disease (DOHaD) concept, has been achieved as evidenced by the varied reviews in this special
45 issue of *J Endocrinology* dedicated to Professor Barker's seminal work. One critical issue and the
46 subject of our review is the question of **when** environment may interact with reproduction to initiate
47 a change in the developmental programme leading to DOHaD-related responses and later disease
48 risk.

49
50 A growing consensus has emerged that the period around conception is critical in DOHaD. This
51 consensus has come from both animal and human studies, ranging across different environmental
52 exposures from the quality of maternal and paternal nutrition to assisted reproductive technology
53 (ART) (**Figure 1**). The stages of gamete maturation, fertilisation and early embryo development are
54 collectively known as the **periconceptual** period. These are characterised by the parental
55 genomes being superseded by the new embryonic genome and the establishment and differentiation
56 of early cell lineages from a pluripotent cellular stock, required for development of the new
57 organism (Graham and Zernicka-Goetz 2016; Li, et al. 2013). Such processes involve significant
58 epigenetic, cellular and metabolic activity (Gardner and Harvey 2015; Lim, et al. 2016; White, et al.
59 2016) and, from fertilisation, occur within the confines of the maternal oviduct and uterine lumens,
60 long recognised to facilitate the stepwise progression in gamete and embryo maturation culminating
61 in implantation (Coy, et al. 2012; Ghersevich, et al. 2015; Matsumoto 2017).

62

63 It has become apparent that these periconceptual stages in reproduction are vulnerable to
64 environmental factors that may cause changes, either through perturbation or via adaptive
65 compensatory responses, which may persist beyond the periconceptual period affecting phenotype
66 across the lifespan. We have recently reviewed the vulnerability of periconception in the context of
67 adverse developmental programming with a focus on the consequences of maternal and paternal
68 over- and under-nutrition and of ART in human and animal models (Fleming, et al. 2018). Maternal
69 or paternal lifestyle factors such as nutritional quality will influence parental physiology in many
70 ways and there is evidence that diet can modify oviduct and uterine transport activities and thereby
71 alter the nutrient composition of luminal compartments and the direct environment experienced by
72 early embryos (Eckert, et al. 2012; Jordaens, et al. 2017). A similar disturbance to the seminal
73 tubule and sperm microenvironment by paternal diet has also been reported (Fan, et al. 2015).
74 Given the clinical implications raised for next generation health from a time when many women
75 may not know they are pregnant, these discoveries of environmental susceptibility of
76 periconceptual stages have contributed to the call for preconception health of both partners to be
77 considered before pregnancy (Barker, et al. 2018; Stephenson, et al. 2018).

78

79 Here, we summarise the key processes, mechanisms and DOHaD-induced outcomes during the
80 periconceptual window with respect to maternal and paternal nutrition and ART. We focus in
81 particular on new understanding of themes previously presented in our earlier review (Fleming et al.
82 2018), reflecting the dynamic nature of this subject.

83

84 **Maternal Overnutrition and Obesity**

85 High maternal body mass index (BMI) and obesity has long been associated with reduced fertility
86 and the occurrence of obesity in children, mediated by raised maternal metabolites such as glucose
87 and insulin promoting increased placental transport of macronutrients and subsequent increased

fetal growth in late gestation (Godfrey, et al. 2017; Musial, et al. 2017; Nam, et al. 2017; Nicholas, et al. 2016). The risk of metabolic syndrome in offspring from obese mothers has been substantiated mechanistically in animal models (Nicholas et al. 2016; Samuelsson, et al. 2008).

The periconceptual period is critical in the transmission of disease risk from maternal obesity to offspring. Women with high BMI transfer excess metabolites and hormones such as insulin, triglycerides, leptin and lactate from the circulation into ovarian tissue and especially the follicular fluid of maturing follicles (Robker, et al. 2009). These metabolites subsequently accumulate within oocytes, affecting their metabolic function and leading to diminished embryo developmental potential after fertilisation (Yang, et al. 2012). Interestingly, increased lipid accumulation within human follicular fluid coincides with increased inflammatory mediators that may contribute to the reduced potential of embryos from obese mothers (Gonzalez, et al. 2018). Notably, the size of human oocytes is reduced by high maternal BMI and lead to poorer quality embryos with excess triglycerides and diminished glucose consumption (Leary, et al. 2015).

Animal models have been used to identify the metabolic defects in oocytes and early embryos caused by maternal overnutrition. Mitochondria become severely affected in their structure and organisation of cristae, in their cellular distribution and rate of biogenesis, and critically in their capacity for generating energy in response to maternal overnutrition (Igosheva, et al. 2010; Luzzo, et al. 2012). These defective mitochondria are more likely to be preserved in embryos since obesity further reduced mitophagy (Boudoures, et al. 2017). Moreover, accumulating lipids in oocytes induces endoplasmic reticulum and oxidative stress, impairing developmental potential and increasing aneuploidy (Hou, et al. 2016; Igosheva et al. 2010; Luzzo et al. 2012). Maternal diabetes may similarly modulate embryo metabolism, recently investigated in a rabbit model of developmental programming. Here, significant remodelling of several metabolic pathways occurred with a critical role identified for adiponectin in generating lipid accumulation leading to oxidative

114 metabolic stress (Fischer, et al. 2017). Further evidence of periconceptional metabolic induction of
115 programming from maternal overnutrition has come from supplementing the diet of obese mice
116 with coenzyme Q10 injection which restored mitochondrial functioning (Boots, et al. 2016). Animal
117 *in vitro* studies have also confirmed that increased levels of fatty acids **impairs** follicular maturation
118 and oocyte potential leading to blastocysts with altered transcription and epigenome profiles
119 (Desmet, et al. 2016; Van Hoeck, et al. 2013). **Such studies also demonstrate fatty acid modulation**
120 **of oviductal barrier function to influence embryo exposure to nutrient levels (Jordaens et al. 2017).**

121 Epigenetic effects have also been demonstrated in the oocytes from obese mouse dams with altered
122 levels of DNA and histone methylation regulators (Hou et al. 2016). Epigenetic change associated
123 with genes regulating metabolic health in offspring has also been shown in an ovine model of
124 maternal overnutrition (Nicholas, et al. 2013).

125
126 Recent mouse studies have identified a role for PGC7/Stella protein in mediating maternal obesity
127 effects on adverse programming of embryos (Han, et al. 2018). Stella is known to regulate the
128 asymmetry in global DNA demethylation between paternal and maternal genomes and protect
129 imprinted genes from demethylation (Nakamura, et al. 2007) and **becomes** depleted in oocytes from
130 obese mothers coinciding with global hypomethylation of the embryonic genome (Han et al. 2018).
131 Notably, restoring Stella expression **reverses** both the epigenetic status of embryos from obese dams
132 and their developmental defects (Han et al. 2018). A further study has identified reduced expression
133 of TIGAR (TP53-induced glycolysis and apoptosis regulator) in oocytes from obese mothers which
134 may contribute to the increased oxidative stress and meiotic spindle defects in such oocytes (Wang,
135 et al. 2018).

136
137 These metabolic perturbations induced in oocytes and embryos by maternal overnutrition persist
138 during later development. Mouse fetuses from obese mothers **exhibit** an altered growth trajectory
139 and **give** rise to offspring with increased adiposity and metabolic dysfunction such as glucose

140 intolerance (Jungheim, et al. 2010). Such physiological responses also coincides with underlying
141 transcriptional and epigenetic changes both in the fetus and placenta (Mahany, et al. 2018).
142 Moreover, metabolic dysfunction in offspring from maternal obesity has been shown to persist over
143 three mouse generations, likely reflecting the inheritance of defective maternally-derived
144 mitochondria (Saben, et al. 2016).
145
146 The importance of the periconceptual origin of adverse programming from maternal obesity has
147 been demonstrated using embryo transfer to healthy recipients in mouse and sheep models with the
148 persistence of fetal and postnatal metabolic dysfunction despite a normal uterine environment
149 (Luzzo et al. 2012; Nicholas et al. 2013). A similar periconceptual origin of adverse programming
150 in response to maternal diabetes has been shown by mouse transfer of zygotes to healthy recipients
151 (Wyman, et al. 2008). Lastly, consistent with the above, in assisted conception practice, there is
152 some evidence that the maternal BMI of oocyte donors negatively influences reproductive outcomes
153 despite not carrying the pregnancy (Cardozo, et al. 2016).

155 **Maternal Undernutrition**

156 The original datasets revealing adverse adult health outcomes derived from *in utero* experience by
157 David Barker and colleagues implicated maternal undernutrition during pregnancy followed by
158 accelerated ‘catch-up’ growth postnatally as causative (Barker and Thornburg 2013). Supporting
159 human evidence linking maternal undernutrition and subsequent adult health risks linked to
160 cardiometabolic and neurological dysfunction have come from well-researched historical famines,
161 particularly the Dutch Hunger Winter of 1944-45 and the Chinese Great Famine over 1959-61 (Liu,
162 et al. 2018; Roseboom, et al. 2011; van den Broek and Fleischmann 2017). Whilst such human
163 epidemiological studies are complex and wide-ranging, it has been possible to identify early
164 gestation and the periconceptual period as a vulnerable window for adverse programming. Thus,
165 those individuals conceived during the 5-month Dutch famine exhibit poorer cardiometabolic and

166 neurological outcomes in adulthood, including accelerated aging that those where the famine
167 experience occurred later in their gestation (Franke, et al. 2018; Roseboom et al. 2011; Tobi, et al.
168 2014). A similar increased risk of first trimester exposure has also been shown in the Chinese
169 famine (Wang, et al. 2012; Zimmet, et al. 2018). In addition, the Dutch famine research has shown
170 that periconceptional exposure **leads** to epigenetic dysregulation of genes involved in growth and
171 metabolism such as conserved hypomethylation of the imprinted *IGF2* gene into adulthood (Tobi et
172 al. 2014).

173

174 A further critical human dataset linking maternal periconceptional undernutrition with later adult
175 disease has come from studies on populations in The Gambia. Here, nutritional quality is seasonal
176 and associated with later life mortality and health risk. The quality of maternal nutrition at
177 conception has been shown to alter the pre-gastrulation epigenome at metastable epialleles, domains
178 characterised by inter-individual variation in DNA methylation, in a manner that **persists** into
179 childhood and adolescence (Waterland, et al. 2010). Such alterations in epigenetic signatures further
180 **associate** with genomic regions predictive of immune status, obesity risk and tumorigenesis
181 (Kuhnen, et al. 2016; Silver, et al. 2015). Indeed, metastable epialleles are present in human early
182 embryos and may provide a suitable epigenetic basis for environment to induce persistent
183 phenotypic change during developmental programming (Kessler, et al. 2018).

184

185 Animal DOHaD studies involving rodents, sheep and cattle have further demonstrated the close
186 association between maternal undernutrition and later-life risk of poor health and again underscore
187 the criticality of the periconceptional period (Fleming et al. 2018; Hansen, et al. 2016; Sinclair and
188 Watkins 2013). From our own work, a maternal low protein diet, effectively 50% of normal protein
189 recommendation, targeted exclusively to the mouse and rat preimplantation period of embryo
190 development (Emb-LPD) has been shown sufficient to cause adult offspring cardiovascular,
191 metabolic and behavioural dysfunction, especially in female progeny (Gould, et al. 2018; Kwong, et

192 al. 2000; Watkins, et al. 2008). The stepwise mechanistic pathway responsible for Emb-LPD
193 adverse programming has been closely examined. The diet results in reduced concentrations of
194 circulating insulin and amino acids (especially the branched-chain amino acids (BCAAs), leucine,
195 isoleucine and valine) within dams that, through analysis of uterine luminal fluids, also changed the
196 metabolite milieu of the immediate environment of embryos (Eckert et al. 2012). Insulin and
197 BCAAs are potent activators of the mTOR signalling pathway regulating cellular growth (Wang
198 and Proud 2009) and, as a consequence of dietary-induced reduction in these metabolites, blastocyst
199 mTOR activity **is** reduced by Emb-LPD (Eckert et al. 2012). This early maternal-embryo interaction
200 is critical since it **activates** later adverse programming as shown both by an *in vitro* culture model in
201 medium reduced in insulin and BCAAs (Velazquez, et al. 2018) and by embryo transfer of Emb-
202 LPD blastocysts into control, normal-fed, recipients (Watkins et al. 2008).

203

204 The subsequent development of the Emb-LPD blastocyst after maternal dietary induction **is** altered
205 and in distinct ways for extra-embryonic (trophectoderm, TE; primitive endoderm, PrE) and
206 embryonic (epiblast) cell lineages. These phenotypic modulations **impact** on the growth trajectory
207 of the fetus which in turn positively **correlates** with later adult disease risk (Watkins et al. 2008).
208 Both TE and PrE cell lineages, in response to maternal Emb-LPD, **undergo** cellular changes that
209 collectively **are** compensatory, likely to augment nutrient delivery to the developing embryo and
210 fetus. These include increased proliferation of the lineages and their capacity for endocytosis of
211 extracellular fluids, thought to increase nutrient supply (Eckert et al. 2012; Sun, et al. 2014). The
212 TE also **adopts** a more invasive migratory phenotype likely to enhance endometrial implantation
213 (Eckert et al. 2012; Watkins, et al. 2015). Extra-embryonic adaptations induced by maternal protein
214 restriction **persist** through pregnancy with evidence of improved nutrient delivery via the
215 chorioallantoic placenta (Coan, et al. 2011) and visceral yolk sac (Watkins et al. 2008), the latter
216 coinciding with altered epigenetic regulation of the *Gata6* transcription factor that has a central role
217 in PrE differentiation (Sun, et al. 2015).

218

219 In contrast to extra-embryonic lineages, the somatic tissues of the fetus derived from the epiblast,
220 such as liver and kidney, **alter** their growth trajectory to match prevailing maternal nutrient
221 availability. This **is** achieved via the rate of ribosome biogenesis, the fundamental unit of
222 biosynthesis, and specifically ribosomal RNA (rRNA) transcription, which **is** reduced if the
223 maternal dietary restriction **is** maintained, but increased beyond control levels, if the dietary
224 challenge **is** lifted as in Emb-LPD. The manipulation of ribosome biogenesis **is** regulated
225 epigenetically through the level of DNA methylation at the rDNA gene promotor and **coincides**
226 with altered expression of the ribosome factor Rrn3, known to link ribosome biogenesis with
227 mTOR nutrient signalling (Denisenko, et al. 2016). Thus, the combination of extra-embryonic and
228 embryonic lineage adaptations to maternal Emb-LPD from implantation, comprising increased
229 extra-embryonic nutrient delivery and increased capacity for fetal biosynthesis, in addition to
230 improved maternal protein diet, all act to promote late fetal overgrowth as a basis for postnatal
231 disease derived from periconceptual environment (Fleming et al. 2018; Watkins et al. 2008).

232

233 Recent work has shown that Emb-LPD and sustained LPD treatment throughout pregnancy have a
234 negative influence on neurogenesis. Both treatments **lead** to a decline in neural stem cells (NSCs)
235 during fetal development through reduced proliferation and increased apoptosis. The loss of NSCs
236 **coincides** with an altered rate of neural differentiation and a postnatal phenotype of altered cortex
237 thickness and short-term memory loss in both males and females (Gould et al. 2018). These
238 findings **extend** earlier behavioural outcomes from the mouse Emb-LPD model (Watkins et al.
239 2008) and **confirm** periconceptual maternal undernutrition as critical in DOHaD for postnatal
240 health across diverse systems.

241

242 **Assisted reproductive technologies**

243 Assisted reproductive technology (ART) refers to any technique that interferes with the normal
244 biological pathways of reproductive-related events and/or structures in order to contribute to the
245 establishment of pregnancy with the final goal of producing healthy offspring. In general, ART
246 manipulates events and/or structures related to ovulation, fertilization and embryo development
247 (Velazquez 2008). Current estimates from the International Committee Monitoring for Assisted
248 Reproductive Technologies (ICMART) indicate that since the first ART-derived baby in 1978 over
249 8 million babies have been born through ART worldwide (De Geyter 2018). It should be
250 emphasised that most ART-derived babies appear healthy. But giving the adverse effects associated
251 to ART reported in some human and animal studies (see below), there is an active effort to ensure
252 an efficient and safe application of human ART, including monitoring of the health status of the
253 resultant offspring.

254

255 Data from Finland indicated that children up to 4 years of age whose mothers were subjected to
256 ovulation induction with or without intrauterine insemination (IUI) showed an increased risk of
257 cerebral palsy, allergy and asthma, along with longer periods of hospitalization (Klemetti, et al.
258 2010). A Danish study found that the risk of developing type 1 diabetes during childhood was
259 increased in children conceived through the use of FSH in ovulation induction protocols or in
260 combination with IUI (Kettner, et al. 2016). Analysis of UK data revealed that babies derived from
261 ARTs such as *in vitro* fertilization (IVF), intracytoplasmic sperm injection (ICSI), IUI, gamete
262 intra-fallopian transfer (GIFT) and ovulation induction had an increased risk of developing
263 respiratory distress and infection during the first week of life when compared to naturally conceived
264 counterparts (Waynforth 2018). Similarly, a meta-analysis of 45 studies suggested that the risk of
265 developing birth defects can be increased by IVF and ICSI (Hansen, et al. 2013), something that has
266 been confirmed in a more recent meta-analysis (Zhao, et al. 2018).

267

Another recent meta-analysis indicated that children conceived by IVF and ICSI showed a lower weight during the first 4 years of age, with the difference disappearing afterwards (Bay, et al. 2018), indicating an enhanced growth velocity during early development. Rapid growth during early childhood can increase the risk of developing obesity and hypertension later in life (Lei, et al. 2015; Mahrshahi, et al. 2011). Indeed, IVF children with rapid growth during early childhood (1-3 years of age) showed higher blood pressure levels compared to spontaneously conceived counterparts at 8-18 years of age (Ceelen, et al. 2009). Increases in blood pressure in IVF/ICSI-derived children has been detected in several studies (Meister, et al. 2018; Sakka, et al. 2010; Scherrer, et al. 2012; Valenzuela-Alcaraz, et al. 2013; Valenzuela-Alcaraz, et al. 2018). Reproductive potential seems to be affected as well, especially in males. Young adults conceived through ICSI showed low sperm concentration and motile sperm count compared to men born after spontaneous conception (Belva, et al. 2016). Interestingly, the impaired sperm production was not associated with significant changes in reproductive hormones (Belva, et al. 2017).

Current evidence seems to indicate that the incidence of certain diseases and some developmental features might not be strongly affected by ART. For instance, the available data indicate that the overall cancer risk does not seem to be increased in ART-derived children, although some studies found a small increased risk for specific types of cancer (Chen and Heilbronn 2017; Wainstock, et al. 2017; Williams, et al. 2018). Studies in the Netherlands reported that behavioural and cognitive performance was not affected in ICSI-derived children at 5 years of age when compared to the general Dutch population (Meijerink, et al. 2016) and that subfertility rather than ART *per se* seems to be the underlying cause of impaired cognitive and behavioural development during childhood observed in some ART-derived children (Schendelaar, et al. 2016). A recent study from the UK also found that IVF and ICSI do not seem to impair children's early cognitive outcomes up to age 11 years (Barbuscia and Mills 2017). Similarly, a recent systematic review revealed that ART treatments such as preimplantation genetic diagnosis/screening do not seem to affect cognitive and

behavioural development, but they can mildly affect psychomotor development (e.g. dysregulation in posture, muscle tone) of children in their first two years of life. However this subtle psychomotor dysfunction was not detected in follow up studies in children up to 9 years of age (Natsuaki and Dimler 2018).

298

Although these results have been taken as reassuring for ART outcomes affecting offspring mental health (Meijerink et al. 2016), these studies were carried out during early childhood and the truly long-term consequences (i.e. in adulthood) for mental health remain to be determined. Furthermore, there is more uncertainty with some neurodevelopmental disorders. For instance, the occurrence of autism and cerebral palsy in IVF/ICSI-derived children was found to be increased in some (Goldsmith, et al. 2018; Kamowski-Shakibai, et al. 2015; Lehti, et al. 2013; Sandin, et al. 2013; Schieve, et al. 2017; Stromberg, et al. 2002) but not all studies (Fountain, et al. 2015; Kallen, et al. 2010; Kissin, et al. 2015; Reid, et al. 2010). Both autism (Fountain et al. 2015) and cerebral palsy (Goldsmith et al. 2018) has been strongly associated with multiple births in ART pregnancies highlighting the need to reduce multiple pregnancies in women undergoing ART (Pinborg 2018).

309

Most of the above-discussed studies used as comparison group children naturally conceived by fertile couples, which has been suggested not to be the best control group. Instead, naturally conceived children from sub-fertile parents that managed to achieve pregnancy while waiting for ART treatment will be a more appropriate comparison group (Zhao et al. 2018). Although studies using this control group are available, a substantial proportion of human ART studies still have methodological limitations that hamper the ability to provide reliable conclusions (Guo, et al. 2017; Liu, et al. 2017; Rumbold, et al. 2017), to the point that some authors believe their findings (e.g. increased risk of type diabetes due to ovulation induction protocols) are a statistical artefact (Kettner et al. 2016).

319

320 Nevertheless, animal models have provided experimental evidence supporting the notion that
321 cardiovascular (Rexhaj, et al. 2013; Watkins, et al. 2007), metabolic (Cerny, et al. 2017; Chen, et al.
322 2014; Feuer, et al. 2014), immunological (Karimi, et al. 2017), reproductive (Calle, et al. 2012), and
323 behavioural (Lopez-Cardona, et al. 2015) activity during postnatal development can be affected by
324 ART. These postnatal alterations can be induced by the microenvironment to which embryos are
325 exposed to during *in vitro* procedures. For example, mice and bovine models have demonstrated
326 that *in vitro* exposure during the preimplantation period to specific constituents of culture media
327 such as metabolic hormones (e.g. insulin), amino acids, pyruvate, lactate, and growth factors can
328 induced alterations in birth weight, body growth rate, and cardiovascular function (Banrezes, et al.
329 2011; Kannampuzha-Francis, et al. 2015; Velazquez et al. 2018). A similar situation has been
330 found in humans, where the culture medium composition induced changes in birth weight
331 (Kleijkers, et al. 2016) and body weight and body mass index examined at 9 years of age (Zandstra,
332 et al. 2018). Importantly, animal models have revealed that culture media modification (e.g.
333 melatonin supplementation) can reverse some of these altered phenotypes (e.g. cardiovascular
334 dysfunction) (Rexhaj et al, 2015).

335
336 The current consensus is that the effects of ART on offspring health may have an epigenetic origin
337 (Huntriss, et al. 2018). Indeed, a meta-analysis revealed that the incidence of rare imprinting
338 disorders in IVF/ICSI-derived children is higher than in spontaneously conceived children, although
339 the exact underlying epigenetic mechanism is unknown (Lazaraviciute, et al. 2014). Nevertheless,
340 compared to methylation levels in somatic and embryonic stem cells, a perturbed methylation of
341 imprinted genes such as SNRPN, KCNQ1OT1 and H19 was found in ART-derived human
342 preimplantation embryos (White, et al. 2015). Similarly, changes in DNA methylation were
343 observed in the placenta (Choufani, et al. 2018; Katari, et al. 2009; Melamed, et al. 2015) and cord
344 blood (Katari et al. 2009; Melamed et al. 2015) from ART-derived babies when compared to
345 naturally conceived counterparts. A study comparing natural conception with oocyte donation (i.e.

346 young fertile oocyte donors/no male infertility) also found differences in placental DNA
347 methylation levels between the groups, suggesting a strong effect of ART and not infertility (Song,
348 et al. 2015). Several regulatory regions, metastable epialleles and imprinted genes, including IGF2,
349 were hypomethylated in blood spots from ART-conceived newborns relative to those conceived
350 naturally (Estill, et al. 2016). The methylation levels of SNRPN, a paternal imprinted gene, were
351 increased in the buccal cells of 2 year old children conceived by ICSI, but not by IVF. This
352 hypermethylation is believed to be associated with the greater degree of *in vitro* manipulation taking
353 place during ICSI (Whitelaw, et al. 2014).

354
355 These epigenetic changes are partially attributed to the microenvironment in which embryos are
356 cultured in, as animal models have revealed that media culture composition can alter DNA
357 methylation profiles in preimplantation embryos (Canovas, et al. 2017; Market-Velker, et al. 2010).
358 Furthermore, oxygen tension (5% vs 20%) during culture and type of embryo transferred (fresh vs
359 frozen) have the capacity to alter placental methylation levels from ART-conceived babies when
360 compare to natural conception. Importantly, data from pigs indicate than modification of culture
361 media to resemble *in vivo* composition can induced methylation levels in preimplantation embryos
362 more similar to those of produced *in vivo* (Canovas et al. 2017).

363
364 In contrast, DNA methylation was not affected in blood from prepubertal children conceived
365 through IVF (Oliver, et al. 2012). This suggest that ART-induced changes in DNA methylation
366 could be gene- and/or tissue-specific or that postnatal environment masked any subtle changes in
367 DNA methylation induced by ART. The latter emphasises the complexity of epigenetic studies in
368 humans and the need to consider several methodological issues to produce useful epigenetic data
369 (Lazaraviciute et al. 2014). Also, a critical step in elucidating the long-term effects of ART in
370 human populations is the development of databases for ART surveillance (i.e. health monitoring of
371 ART-derived offspring), something that has been implemented just in a few countries (Pinborg

2018). The first ART-derived baby turned 40 years just recently, hence the long-term repercussions (or lack of) of ART for healthy aging are far from being elucidated. This highlights the current need for more research throughout the lifespan of ART-derived offspring.

Paternal origin of periconceptional programming

In contrast to the substantial epidemiological and animal model research linking maternal well-being with offspring programming, our understanding of how a father influences the development and cardiometabolic health of his offspring has been largely overlooked. However, there is now a significant body of data indicating paternal physiological status, lifestyle and environmental exposure to a range of factors not only impact on sperm quality, but also affect the long-term health of his offspring (Fleming et al. 2018). In line with maternal programming studies, animal models have become critical tools for not only defining the underlying paternal mechanisms involved but also identifying central biomarkers of paternal programming ahead of studies using human samples. Studies from humans and animal models have revealed the complexity of both sperm and the seminal plasma, identifying novel processes by which perturbed paternal health at the time of conception affect a dynamic range of reproductive and developmental processes and ultimately, long-term offspring health.

Paternal reproductive health and sperm quality are impaired in response to paternal physiological and lifestyle factors. Mirroring changes in oocyte quality in response to maternal obesity, elevated paternal BMI has been associated with reduced semen volume, sperm number and sperm motility (Chavarro, et al. 2010; Ma, et al. 2018). Furthermore, sperm from overweight or obese men show higher levels of DNA damage when compared to sperm from normal weight males (Campbell, et al. 2015; Kort, et al. 2006). As obesity is associated with multiple disturbances in metabolic profile including elevated levels of inflammatory markers and metabolic intermediates, the detrimental effects of increasing male BMI on sperm quality is believed to be mediated through increased

oxidative damage. Indeed, in both men and rodents, obesity has been shown to result in increased reactive oxygen species generation (Palmer, et al. 2011; Tunc, et al. 2011) and sperm DNA damage (Duale, et al. 2014; Zhao, et al. 2014). Furthermore, consumption of high energy diets have also been associated with reduced sperm morphology, motility and DNA integrity (Agbaje, et al. 2007), perturbed testicular metabolism (Rato, et al. 2013) and reduced fertility (Bener, et al. 2009) in both mice and men. Similar to the effects of paternal overnutrition, deficiency of specific nutrients, or even nutritional imbalance also affect sperm quality. Many macronutrients such as zinc, vitamins and glutathione act as antioxidants to prevent excessive damage from reactive oxygen species. Sperm from infertile men show higher rates of DNA damage which can be reduced following treatment with supplement of selenium and vitamin E (Moslemi and Tavanbakhsh 2011). In mice, the negative effects of paternal undernutrition on sperm DNA damage can be prevented through dietary supplementation with vitamins and minerals (McPherson, et al. 2016).

Poor paternal health not only impacts on sperm quality, but can also affect post-fertilisation development and offspring well-being. In men, some studies have identified associations between obesity and reduced rates of blastocyst development and live birth following IVF (Bakos, et al. 2011). Such observations are supported by a recent, large meta-analysis in which the link between paternal obesity and live birth rates after ART cycles was examined in 115,158 patients (Campbell et al. 2015). Here, the authors reported a significant negative impact of increased male BMI on non-viable pregnancy outcomes. In mice, paternal obesity has been reported to increase rates of one-cell block, decrease blastocyst cell number and perturb embryo carbohydrate metabolism (Binder, et al. 2012; Mitchell, et al. 2011). Our own studies have revealed that a paternal low protein diet (LPD) decreased blastocyst expression of multiple genes involved in the 5' AMP-activated protein kinase (AMPK) pathway including genes for metabolism, regulation of transcription and protein synthesis (Watkins, et al. 2017). Interestingly, similar decreases in several of these AMPK pathway genes were still evident in late gestation fetal liver tissues and associated with increased rates of fetal

424 growth (Watkins et al. 2017). As in studies of poor maternal diet during pregnancy, we observed
425 that the enhanced fetal growth programmed by paternal LPD **was** associated subsequently with
426 increased adiposity, impaired glucose metabolism, hypotension and vascular dysfunction in adult
427 offspring (Watkins and Sinclair 2014). Separately, other studies have shown significant changes in
428 fetal (Carone, et al. 2010; Lambrot, et al. 2013) and postnatal offspring development and metabolic
429 health (Anderson, et al. 2006; McPherson et al. 2016; Ryan, et al. 2018) in response to paternal diet
430 or food intake in mice. Interestingly, recent studies have demonstrated robust transgenerational
431 effects of chronic paternal stress on offspring well-being and hypothalamic pituitary adrenal axis
432 function (Gapp, et al. 2014; Rodgers, et al. 2015).

433
434 The fact that many paternal **programming** studies identify consistent transgenerational programming
435 effects (Fullston, et al. 2013; Gapp et al. 2014) indicates changes in sperm epigenetic status as one
436 potential mechanism linking paternal well-being with offspring development. Over recent years the
437 epigenetic complexity of mammalian sperm has been revealed. In contrast to the oocyte, sperm
438 contain almost no cytoplasm and the DNA is packaged using protamines rather than histones.
439 Inappropriate protamine packaging of the sperm DNA, or perturbed histone to protamine transition
440 can be indicative of impairments in the fundamental process of spermatogenesis (Sakkas, et al.
441 2002) or damage due to excessive exposure to reactive oxygen species (Sakka et al. 2010).
442 Furthermore, atypical chromosome packaging and localisation within the sperm or perturbed
443 telomere-centromere interactions has been associated with infertility in some men (Zalensky and
444 Zalenskaya 2007), while sperm chromatin maturation level has been link with pregnancy
445 establishment rates (de Lamirande, et al. 2012). While the majority of the sperm DNA is re-
446 packaged with protamines, specific genomic sequences retain their histone marks. What is
447 interesting is that the location of these retained histones is not random, but specific to important
448 developmental genes (Hammoud, et al. 2009) and retrotransposable long and short interspersed
449 nuclear elements in both men and mice (Samans, et al. 2014). Furthermore, some of these sperm-

specific histones have been shown to be retained within the oocyte and contribute to the zygotic genome (van der Heijden, et al. 2008).

452

In addition to sperm chromatin structure, differential profiles of DNA methylation have also been linked to sperm quality in infertile men (Hammoud, et al. 2010). In studies looking at success rates of women undergoing IVF, the genome-wide methylation profile of their partner's sperm correlated with embryo quality (Aston, et al. 2015) and was indicative of pregnancy failure (Benchaiib, et al. 2005). In mice, significant changes in sperm DNA methylation profiles have also been identified in response to paternal obesity (Fullston et al. 2013), low protein (Carone et al. 2010) or low folate (Lambrot et al. 2013) diets. Our own studies have showed that feeding male mice a LPD results in global sperm hypomethylation associated with reduced testicular expression of central regulators of DNA methylation and 1-carbon metabolism (Watkins, et al. 2018). Interestingly, analysis of the sperm DNA hypomethylation revealed significant reductions at multiple genes involved in calcium signalling which correlated with our earlier reported impairments in cardiovascular function and cardiac calcium signalling gene expression in adult offspring of LPD fed males (Watkins and Sinclair 2014). In addition to histone and DNA modifications, sperm have been shown to contain a range of RNA species including mRNA, micro-RNA, short and long non-coding RNA and small interfering RNAs (Colaco and Sakkas 2018). The significance of sperm-derived RNAs for post-fertilisation development has been demonstrated in animal models where the depletion of specific sperm micro RNAs results in developmental delay of the zygote (Liu, et al. 2012). In addition, injection of tRNA-derived small RNAs from sperm of high fat diet fed male mice into control zygotes resulted in impaired glucose metabolism and insulin secretion in the resultant offspring (Chen, et al. 2016).

473

Separate to the epigenetic status of the sperm, fathers may also influence the development of their offspring via seminal plasma-specific modulations of the maternal reproductive tract environment

(Robertson and Sharkey 2016). In both mice and women, deposition of seminal plasma within the reproductive tract initiates a significant inflammatory and immunological response culminating in uterine vascular remodelling, the recruitment of leukocytes and the priming of regulatory T cells (T-regs) and the production of a myriad of cell signalling molecules such as colony stimulating factor-2 (CSF2), leukemia inhibitory factor (LIF) and interleukin 6 (IL-6) (Schjenken and Robertson 2014). Interestingly, studies have demonstrated positive associations between a woman's unprotected exposure to her partner's seminal plasma and a reduced risk for her developing preeclampsia during pregnancy (Robillard, et al. 1994). In mice, lack of seminal plasma at the time of conception has been shown to impair embryo development, fetal growth and adult offspring cardiometabolic health (Bromfield, et al. 2014). Our own studies have shown that offspring growth and metabolic health appear equally compromised in response to either sperm or seminal plasma from male mice fed a LPD (Watkins et al. 2018).

Conclusions

It is clear from the above four types of exposure during periconceptional reproduction that altered developmental programming may emerge from diverse environments (summarised in **Table 1**). Whilst here we focus on parental nutrition *in vivo* and embryo manipulations *in vitro*, the spectrum of exposures with enduring consequences is undoubtedly broader. For example, periconceptional maternal alcohol consumption prior to embryo implantation in a rat model resulted in abnormal trophoblast placental function, altered expression of epigenetic regulators for DNA methylation in the fetal liver, culminating in postnatal glucose and insulin intolerance and increased risk of offspring obesity (Gardebjer, et al. 2015; Gardebjer, et al. 2018; Kalisch-Smith, et al. 2016). In another example, maternal sickness and systemic inflammation at the time of conception has been shown in a mouse model to alter blastocyst morphogenesis with long-term consequences for adult offspring immune function (Williams, et al. 2011). Here, reproductive function and embryo implantation are in part regulated by the activity of maternal immune cells and the balance of pro-

502 and anti-inflammatory cytokines can have significant influence not only on embryo survival but
503 long-term health of offspring (Robertson, et al. 2015).

504

505 The extent to which periconceptional exposure can associate with adult DOHaD consequences is
506 also influenced by intrinsic processes such as maternal ageing. Whilst it is well established that
507 fertility declines with age, the developmental potential of oocytes with advancing age is also
508 affected. In a recent mouse study, preimplantation embryos from aged versus young mothers, both
509 sired by young males and transferred to young recipients to carry the pregnancy, gave rise to
510 offspring with altered growth and increased cardiometabolic dysfunction (Velazquez, et al. 2016).
511 Oocytes from older mothers exhibit mitochondrial dysfunction and perturbed energy homeostasis
512 (Dumesic, et al. 2015) which may indicate adverse programming derives from similar processes as
513 occurs following maternal overnutrition, although mechanisms are underexplored.

514

515 A consistent feature across the research field of periconceptional programming has been the
516 involvement of epigenetic dysregulation as a means by which effects on gene expression and
517 cellular phenotype may persist through gestation and later life (Steegers-Theunissen, et al. 2013).
518 Manipulation of periconception maternal diet composition to reduce the availability of methyl
519 donors for DNA and histone methylation via one-carbon metabolism has been shown to alter the
520 offspring epigenome with accompanying cardiometabolic disease outcomes (Sinclair, et al. 2007).
521 Provision of methyl donors can also reverse adverse programming mediated through the rat
522 maternal LPD model (Lillycrop, et al. 2005). Animal oocytes and early embryos are known to
523 express key enzymes in the methionine/folate cycles (Kwong, et al. 2010) and a role for mTOR
524 signalling has been identified for sensing the levels of folate available for placental development
525 and fetal growth (Gupta and Jansson 2018; Rosario, et al. 2017). Variability across individuals and
526 ethnic groups in regulatory genes involved in one-carbon metabolism may contribute to the relative
527 susceptibility to adverse programming (Clare, et al. 2018). What is clear is that health of both

528 parents in terms of diet and physiological condition is an important factor to establish before
529 conception rather than later in pregnancy to protect the health of the next generation.

530

531 **Declaration of interests**

532 The authors declare that there is no conflict of interest that could be perceived as prejudicing the
533 impartiality of the research reported.

534

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539

540

541 **Figure legends**

542

543 **Figure 1.**

544 Summary diagram of the periconceptional period covering gamete maturation and early
 545 embryogenesis with key developmental stages and events identified, shown both in vivo and during
 546 ART, and with long-term risks for offspring health from adverse exposures listed.

547

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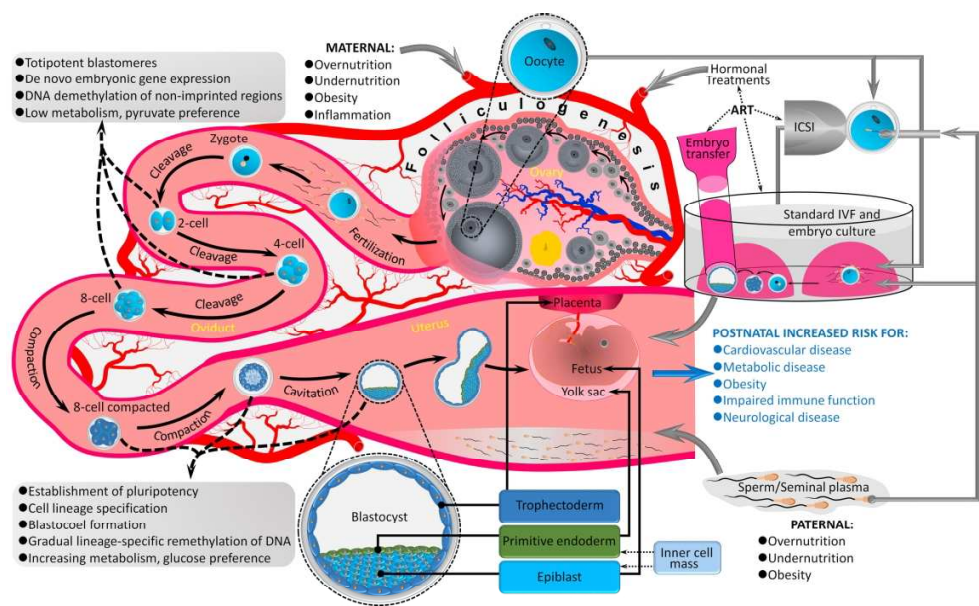
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Insult

	<u>Maternal overnutrition</u>	<u>Maternal undernutrition</u>	<u>Paternal nutrition</u>	<u>Assisted Reproductive Technologies (ART)</u>
Impact on gamete quality and parental environment	<ul style="list-style-type: none"> • Excess follicular metabolite concentration • Reduction in oocyte size and embryo quality • Increased oocyte lipid accumulation, ER stress and mitochondrial dysfunction • Perturbed expression of epigenetic regulators 	<ul style="list-style-type: none"> • Altered uterine metabolite concentrations 	<ul style="list-style-type: none"> • Elevated sperm DNA damage • Altered sperm epigenome • Altered sperm RNA content • Altered seminal plasma composition 	
Impact on embryo development	<ul style="list-style-type: none"> • Increased oxidative metabolic stress • Altered profiles of transcription 	<ul style="list-style-type: none"> • Reduced blastocyst mTOR signalling • Extra-embryonic cellular adaptations to enhance nutrient retrieval 	<ul style="list-style-type: none"> • Reduced APMK gene expression • Altered maternal uterine immunological environment 	<ul style="list-style-type: none"> • Altered epigenetic status
Impact on offspring phenotype and health	<ul style="list-style-type: none"> • Increased fetal growth • Altered placental epigenetic status • Increased offspring adiposity • Cardio-metabolic dysfunction 	<ul style="list-style-type: none"> • Altered epigenetic status • Altered ribosome biogenesis • Increased fetal growth • Increased adiposity • Cardio-metabolic dysfunction • Neurodevelopmental dysfunction • Perturbed imprinted gene epigenetic status 	<ul style="list-style-type: none"> • Perturbed fetal growth • Increased offspring adiposity • Cardio-metabolic dysfunction 	<ul style="list-style-type: none"> • Altered birthweight • Increased early life growth • Poorer cardio-metabolic health • Reduced sperm counts • Increased rates of imprinting disorders

Table 1. Summary of main environmental exposures discussed in the review and their impact during development and health outcomes in later life.

For Review Only