

1 **Circulating leukocyte cell-derived chemotaxin 2 and fibroblast growth factor 21 are**
2 **negatively associated with cardiorespiratory fitness in healthy volunteers**

3 **Running Title:** cardiorespiratory fitness and hepatokines

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33 Hepatokines, LECT2, FGF21, sedentary time, exercise, physical activity

34 **Competing Interests**

35 The authors declare there are no competing interests.

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38 **Abstract**

39 Leukocyte cell-derived chemotaxin-2 (LECT2) and fibroblast growth factor 21 (FGF21) are
40 hepatokines which are regulated by energy balance and mediate insulin sensitivity and
41 glycaemic control. This cross-sectional study examined the independent associations of
42 cardiorespiratory fitness (CRF), moderate-to-vigorous intensity physical activity (MVPA), and
43 sedentary time, with circulating LECT2 and FGF21. Data were combined from two previous
44 experimental studies in healthy volunteers (n=141, male=60%, mean \pm SD age=37 \pm 19 years,
45 body mass index (BMI)=26.1 \pm 6.3 kg·m⁻²). Sedentary time and MVPA were measured via an
46 ActiGraph GT3X+ accelerometer while magnetic resonance imaging quantified liver fat. CRF
47 was assessed using incremental treadmill tests. Generalized-linear models examined the
48 association of CRF, sedentary time and MVPA with LECT2 and FGF21 whilst controlling for
49 key demographic and anthropometric variables. Interaction terms explored the moderating
50 influence of age, sex, BMI, and CRF. In the fully adjusted models, each SD increase in CRF
51 was independently associated with a 24% (95% CI: -37% to -9%, *P* = 0.003) lower plasma
52 LECT2 concentration and 53% lower FGF21 concentration (95% CI: -73% to -22%, *P* =
53 0.004). Each SD increase in MVPA was independently associated with 55% higher FGF21
54 (95% CI: 12% to 114%, *P* = 0.006) and this relationship was stronger in those with lower BMI
55 and higher levels of CRF. These findings demonstrate that CRF and wider activity behaviours
56 may independently modulate the circulating concentrations of hepatokines and thereby
57 influence inter-organ cross-talk.

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62 **Introduction**

63 The liver plays a central role in system-wide metabolic homeostasis through inter-organ
64 crosstalk with other tissues (Watt et al., 2019). Many ‘hepatokines’ have been characterised as
65 vehicles of systemic communication, with important roles in regulating energy/substrate
66 metabolism, insulin sensitivity, and glycaemic control (Jensen-Cody & Potthoff, 2021). The
67 regulation of hepatokines has received scientific interest in recent years, with recognition that
68 anthropometric variables, ectopic lipids, and glycaemic control are associated with many
69 hepatokines (Jensen-Cody & Potthoff, 2021). Most prominently, experimental studies have
70 shown that liver fat directly modulates the hepatic proteome (Kirpich et al., 2011; Meex et al.,
71 2015). Emerging evidence indicates that physical activity and cardiorespiratory fitness (CRF)
72 also influence hepatokine metabolism. However, knowledge in this area is rudimentary
73 (Ennequin et al., 2019; Weigert et al., 2019).

74 Leukocyte cell-derived chemotaxin 2 (LECT2) is a hepatokine known to promote insulin
75 resistance in skeletal muscle and adipose tissue (Jung et al., 2018; Lan et al., 2014).
76 Observational studies have shown that circulating LECT2 is positively associated with body
77 mass index (BMI), circulating lipids, and insulin resistance (Okumura et al., 2013). Moreover,
78 in one analysis, visceral adipose tissue was cited as the strongest predictor of circulating
79 LECT2 concentrations in humans (Tanisawa et al., 2017). Mechanistically, LECT2 is
80 negatively regulated by AMP-activated protein kinase (AMPK), therefore energy balance may
81 also influence LECT2 metabolism (Garcia et al., 2019; Lan et al., 2014). Acute exercise
82 suppressed circulating LECT2 in rodents (Lan et al., 2014); however, this finding has not been
83 replicated in humans (Sargeant et al., 2018; Willis et al., 2019). Additional research is needed
84 to better understand the interaction between physical activity, metabolic status, and LECT2.

85 A second hepatokine which has received significant attention is fibroblast growth factor 21
86 (FGF21). Although the FGF21 protein is produced in several tissues, circulating levels are
87 predominantly liver-derived (Markan et al., 2014; van Baak et al., 2020). Preclinical studies
88 have shown that the augmentation of FGF21 action elicits favourable effects on substrate
89 (glucose and lipid) metabolism, insulin sensitivity, ectopic fat, and energy expenditure (Coskun
90 et al., 2008; Xu et al., 2009). In clinical trials, FGF21 analogues have conferred positive
91 metabolic effects in humans (e.g. glycaemic control and lipid metabolism) (Cui et al., 2020).
92 Counter-intuitively, circulating FGF21 levels are positively related to BMI and the metabolic
93 syndrome (Chow et al., 2013), which may be a compensatory response to the metabolic stress
94 of overnutrition (Giralt et al., 2015). Furthermore, FGF21 is acutely regulated by circulating
95 free fatty acids and the glucagon-to-insulin ratio; factors which are sensitive to physical activity
96 (Hansen et al., 2015; Mai et al., 2009). Previous research has identified associations between
97 circulating FGF21, CRF, and habitual physical activity, however, these studies have failed to
98 separate these associations from the confounding influence of liver fat (Cuevas-Ramos et al.,
99 2010, 2012; Matsui et al., 2019; Taniguchi et al., 2014). Given that liver fat is a key determinant
100 of circulating FGF21 levels (Okumura et al., 2013), further studies are warranted to disentangle
101 the relationship between these variables.

102 Using a sample of precisely phenotyped volunteers, this cross-sectional study examined
103 independent associations between objectively-measured CRF, physical activity and sedentary
104 time with circulating LECT2 and FGF21. A secondary aim was to explore whether sex, age
105 and BMI moderated the associations between exposure and outcome variables.

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109 **Methods**

110 *Ethical approval*

111 This cross-sectional study pooled data from two experiments which used identical procedures
112 for outcomes included in the present analysis (Goltz et al., 2019; Roberts et al., 2022). Both
113 studies obtained institutional ethical approval and written informed consent from all
114 participants.

115 *Participants*

116 Data were available for 141 individuals (85 men, 56 women) who were white European or
117 South Asian. Participants did not smoke, were weight stable, were not taking medications
118 known to affect study outcomes, and were free of established cardiometabolic disease (e.g.,
119 type 2 diabetes and cardiovascular disease). Pre-menopausal women reported not being
120 pregnant and their tests were completed during the follicular phase of the menstrual cycle.
121 Although there was a wide-range, the majority of participants were physically active.

122 *Study procedures*

123 Data collection took place at Loughborough University within laboratory visits that occurred
124 between November 2016 and September 2019. Although the data included in this manuscript
125 were pooled from two separate studies, each study was undertaken in the same laboratory using
126 identical techniques and standard operating procedures. In all cases, participants abstained
127 from alcohol, caffeine, and structured exercise in the 24 h before data collection.

128 *Anthropometry*

129 Height and body mass were measured using an integrated stadiometer and scale (Seca Ltd,
130 Hamburg, Germany). Body mass index was calculated as weight (kg) divided by height (m)
131 squared.

132 *Measurement of liver fat*

133 Participants underwent an MRI scan to quantify liver fat. Scans used a dual-echo Dixon fat and
134 water sequence on a 3T MRI scanner (MR750w, GE Healthcare, Chicago, USA). The IDEAL-
135 IQ sequence was used to assess proton density fat fraction (West et al., 2016). After collection,
136 anonymised scans were analysed by AMRA medical using the AMRATM profiler (AMRA
137 Medical AB, Linköping, Sweden) (Borga et al., 2015).

138 *Measurement of physical activity and sedentary time*

139 Habitual physical activity and sedentary time were assessed over 7-days using a waist-worn
140 ActiGraph GT3X+ accelerometer (ActiGraph, Pensacola, USA). For inclusion, participants
141 were required to submit at least four days of valid wear time (≥ 600 minutes). Data were
142 analysed over 15 second epochs (ActiLife, Actigraph corporation, Florida, USA) and classified
143 as: sedentary time < 25 counts per 15 seconds and MVPA ≥ 488 counts per 15 seconds (Byrom
144 et al., 2016; Freedson et al., 1998). Sixty minutes of continuous zero counts were classified as
145 non-wear time.

146 *Measurement of cardiorespiratory fitness*

147 Cardiorespiratory fitness was assessed as participants' peak oxygen uptake ($\dot{V}O_2$ peak) and was
148 measured directly via indirect calorimetry for 111 individuals. Within these tests, participants
149 completed an incremental protocol on a treadmill until volitional exhaustion, with heart rate
150 (Polar T31; Polar Electro, Kempele, Finland) and perceived exertion (Borg, 1973) measured

151 throughout. Oxygen uptake was measured continuously during the test via a breath-by-breath
152 analyser (Metalyzer 3B, Cortex, Leipzig, Germany), with $\dot{V}O_2$ peak determined as the highest
153 oxygen consumption value averaged over 20 seconds. In the remaining 30 participants, $\dot{V}O_2$
154 peak was measured indirectly using the Bruce test (Bruce et al., 1973) given their higher cardio-
155 metabolic risk (central obesity). This indirect measure of $\dot{V}O_2$ peak correlates strongly ($r =$
156 0.97) with that measured directly (Bruce et al., 1973; Foster et al., 1984).

157 *Biochemical analysis*

158 Fasted venous blood samples were drawn from an antecubital or forearm vein after participants
159 had rested in a semi-supine position for at least 5 minutes. Samples were collected into pre-
160 chilled EDTA monovettes (Sarstedt, Leicester, United Kingdom) and spun immediately in a
161 refrigerated centrifuge for 10 minutes (4°C, 1500 x g) (Labofuge 400R, ThermoScientific,
162 Langenselbold, Germany). The plasma supernatant was collected and stored at -80°C before
163 analysis. Commercially available enzyme-linked immunosorbent assays (ELISAs) were used
164 to measure plasma concentrations of LECT2 (BioVendor, Czech Republic) and FGF21 (R&D
165 Systems, Minneapolis, United States). The coefficient of variation for LECT2 and FGF21 were
166 4.8% and 5.0%, respectively.

167 *Statistical analyses*

168 A formal sample size calculation was not performed for this exploratory study. Statistical
169 analyses were carried out using SPSS version 26 (SPSS Inc., Chicago, Illinois). To examine
170 the distribution of the data, histograms and Kolmogorov-Smirnov tests were used. Data are
171 presented as the mean \pm standard deviation (SD) for normally distributed data, median
172 (interquartile range) for non-normally distributed data, or number (percentage) for categorical
173 groups. Correlations between physical activity variables, CRF, and liver fat with LECT2 and

174 FGF21 were explored using Pearson's correlation coefficient (r) for normally distributed data,
175 or Spearman's correlation coefficient (ρ) for non-normally distributed data. LECT2 was
176 normally distributed, whereas FGF21 was not normally distributed. Generalized linear
177 modelling was used to examine the independent associations of CRF, sedentary time, and
178 MVPA with circulating LECT2 and FGF21. All exposure variables were standardised for this
179 analysis. A normal distribution with a log link was used for LECT2 while a gamma distribution
180 with a log link was used for FGF21 (due to the right-skewed distribution of the variable). The
181 consistent use of log links across models and the standardisation (per SD) of exposure variables
182 allowed for the strength of the association to be reported as a fold change per SD for each
183 model, allowing direct comparison between models. In model 1, adjustment was made for
184 demographic variables including study, age (continuous), sex, ethnicity (white European/South
185 Asian), BMI, plus device wear time (continuous) where accelerometer-measured variables
186 (sedentary time and MVPA) were included in the model as exposures. Model 2 was adjusted
187 for the same variables as model 1, as well as for CRF or accelerometer-measured variables
188 including sedentary time and MVPA (all continuous). Model 3 was adjusted for the same
189 variables as model 2 plus liver fat (continuous). Multicollinearity between covariates was
190 assessed using a correlation matrix. Significant associations in model 3 were then explored
191 further by simultaneously adding interactions terms into the models to assess whether these
192 associations were modified by sex, age, and BMI. In addition, because interventions to reduce
193 sedentary behaviour have been shown to be more effective at improving metabolic health in
194 those with lower fitness (McCarthy et al., 2017), we further assessed interactions between CRF
195 and physical activity variables. Only significant interactions are presented. To facilitate
196 interpretation, interactions between continuous variables were also stratified using the median
197 split. Coefficients were back-transformed to generate fold-change. All data for the regression

198 analyses are presented as fold change (95% confidence intervals). Statistical significance was
199 set at $P < 0.05$ for main effects and interactions.

200 **Results**

201 The characteristics of included participants are shown in Table 1. Due to technical issues with
202 the accelerometer, sedentary time and MVPA are presented for $n = 130$. Additionally, liver fat
203 could not be determined in 15 participants due to motion artefacts; therefore, these data are
204 presented for $n = 126$. Moreover, circulating FGF21 data for one participant were removed
205 from the analysis due to being an outlier (z -score = 7); therefore, these values are presented for
206 $n = 140$.

207

208 *Insert Table 1*

209

210 *Simple correlations between LECT2, FGF21 and key study variables*

211 Figure 1 shows the simple correlations of LECT2 and FGF21 with CRF, sedentary time,
212 MVPA, and liver fat. Pearson's correlation coefficients revealed that plasma LECT2 was
213 negatively associated with CRF ($r = -0.38$, $P < 0.001$) and positively associated with liver fat
214 ($r = 0.35$, $P < 0.001$). Plasma LECT2 was not associated with sedentary time or MVPA ($P >$
215 0.05). Furthermore, Spearman's correlation coefficients revealed that plasma FGF21 was
216 negatively associated with CRF ($\rho = -0.42$, $P < 0.001$) and positively associated with both
217 sedentary time ($\rho = 0.23$, $P = 0.007$) and liver fat ($\rho = 0.47$, $P < 0.001$). Plasma FGF21 was
218 not associated with MVPA ($P > 0.05$).

219 *Independent associations of LECT2 with CRF, sedentary time and MVPA*

220 Adjusted associations of plasma LECT2 with CRF, sedentary time, and MVPA are shown in
221 Table 2 and Figure 2. After adjustment for demographic variables in model 1, only CRF was
222 associated with plasma LECT2 ($P = 0.001$). Each SD increase in CRF was associated with a
223 26% (95% CI: -40% to -11%) lower plasma LECT2 concentration. After additional adjustment
224 for sedentary time, MVPA and liver fat in model 3, the association remained statistically
225 significant ($P = 0.003$) such that each SD increase in CRF was associated with a 24% (95% CI:
226 -37% to -9%) lower plasma LECT2 concentration. There were no associations between LECT2
227 and sedentary time, or MVPA. To study the relationship further, interaction analyses with sex,
228 age, and BMI were performed; however, no significant interactions were observed.

229

230 *Insert Table 2*

231

232 *Independent associations of FGF21 with CRF, sedentary time and MVPA*

233 Associations of plasma FGF21 with CRF, sedentary time, and MVPA are shown in Table 2
234 and Figure 2. After adjustment for demographic variables (model 1), the association with CRF
235 was statistically significant for plasma FGF21 ($P = 0.007$). Each SD increase in CRF was
236 associated with a 51% (95% CI: -71% to -19%) lower plasma FGF21 concentration. This
237 association remained in the fully adjusted model (model 3) ($P = 0.004$), where each SD increase
238 in CRF was associated with a 53% (95% CI: -73% to -22%) lower plasma FGF21
239 concentration. Additionally, there was a statistically significant association between plasma

240 FGF21 and MVPA ($P = 0.006$), whereby each SD increase in MVPA was associated with a
241 55% (95% CI: 12% to 114%) higher plasma FGF21 concentration.

242 Interaction analyses showed that the relationship between plasma FGF21 and MVPA was
243 moderated by BMI ($P = 0.052$) (Table 3). Specifically, each SD increase in MVPA (per 30
244 minute) was associated with an 86% (95% CI: -48% to 576%) higher plasma FGF21
245 concentration in those with a lower BMI ($< 26.1 \text{ kg}\cdot\text{m}^{-2}$), whereas no relationship was evident
246 in those with a higher BMI ($\geq 26.1 \text{ kg}\cdot\text{m}^{-2}$) (Table 3). Additionally, CRF also moderated the
247 relationship between plasma FGF21 and MVPA ($P = 0.088$), such that each SD increase in
248 MVPA (per 30 minute) was associated with a 104% (95% CI: 35% to 202%) higher plasma
249 FGF21 concentration in those with a higher CRF ($\geq 40.1 \text{ mL}\cdot\text{kg}^{-1}\cdot\text{min}^{-1}$) (Table 3).

250

251 *Insert Table 3*

252

253 **Discussion**

254 This study examined the associations of circulating LECT2 and FGF21 with CRF, sedentary
255 time, and MVPA. The primary findings are that circulating concentrations of LECT2 and
256 FGF21 are inversely associated with CRF. Importantly, these relationships were apparent after
257 controlling for liver fat and other anthropometric, demographic, and accelerometer-measured
258 variables, each of which are key mediators of circulating hepatokine concentrations. Whilst
259 observational in nature, these data suggest that an individual's CRF may be an additional
260 mediator of hepatokine metabolism.

261 LECT2 is a hepatokine that has several detrimental effects on metabolic homeostasis (Slowik
262 & Apte, 2017). Specifically, LECT2 has been shown to promote skeletal muscle and adipose
263 tissue insulin resistance in preclinical models (Jung et al., 2018; Lan et al., 2014) and has most
264 recently been implicated in the development of hepatic inflammation (Takata et al., 2021). In
265 humans, circulating LECT2 concentrations are elevated in individuals with obesity (Okumura
266 et al., 2013), type 2 diabetes (Zhang et al., 2018) and NAFLD (Yoo et al., 2017), and are
267 positively associated with adiposity, dyslipidaemia, and markers of insulin resistance (Lan et
268 al., 2014; Okumura et al., 2013; Zhang et al., 2018). Liver fat is thought to be a key mediator
269 of LECT2 production given that LECT2 mRNA is almost exclusively expressed in the liver
270 and circulating concentrations are positively associated with the presence of NAFLD
271 (Yamagoe et al., 1998; Yoo et al., 2017). Our data support and extend these findings as we
272 observed novel positive associations between circulating LECT2 and MRI-derived liver fat in
273 both simple correlations and fully adjusted generalised linear models (data not shown in results
274 tables).

275 In the present study, we report for the first time that circulating concentrations of LECT2 are
276 inversely and independently associated with CRF. Only one previous study has examined the
277 relationship between circulating LECT2 and CRF, and the authors reported no statistically
278 significant associations after adjusting for age and visceral adipose tissue (Tanisawa et al.,
279 2017). We opted to adjust our models for liver fat given the importance of liver fat in the
280 regulation of LECT2. Furthermore, discrepancies between the studies may be related to the
281 more homogenous population of middle-aged and elderly men in the study by Tanisawa et al.
282 (2017), and cultural/lifestyle differences between their Japanese population and our European
283 population. Importantly, our findings were independent of liver fat, suggesting that CRF may
284 be an additional mediator, and raises the possibility that improving CRF through exercise

285 training could potentially reduce circulating LECT2 concentrations. Previous research has
286 demonstrated that an acute, exhaustive bout of exercise in mice can reduce hepatic LECT2
287 expression and secretion via increasing phosphorylation of hepatic AMPK (Lan et al., 2014);
288 however, human studies are yet to replicate these findings (Sargeant et al., 2018; Willis et al.,
289 2019). Experimental studies are required to determine whether improvements in CRF reduce
290 circulating LECT2 concentrations in humans.

291 No previous studies have explored the associations of circulating LECT2 with objectively
292 measured physical activity and sedentary time. In our fully adjusted model, we found no
293 statistically significant associations between circulating LECT2 and sedentary time, or MVPA.
294 Whilst it may be expected that MVPA would be negatively associated with circulating LECT2
295 given our observed association with CRF, it is important to note that CRF is determined by
296 both genetic factors and habitual physical activity, in which vigorous intensity is a key
297 determinant (Bouchard et al., 2015). Notably, our objective measurement of MVPA does not
298 enable us to differentiate between these two intensities; thus, it is possible that the inclusion of
299 moderate intensity physical activity dampened our ability to detect differences. Therefore,
300 further research is warranted to specifically examine the associations of circulating LECT2
301 with more purposeful physical activity of vigorous intensity.

302 FGF21 is another hepatokine that has gained extensive attention due to its favorable effects on
303 glucose and lipid metabolism (BonDurant & Potthoff, 2018). Synthetic FGF21 analogues have
304 shown promise as novel medicinal therapies for metabolic disease (Cui et al., 2020).
305 Administration of recombinant FGF21 reduces body weight, liver fat content and circulating
306 glucose and lipid concentrations, and improves insulin sensitivity in mice with obesity and type
307 2 diabetes (Berglund et al., 2009; Coskun et al., 2008; Kharitonov et al., 2005). FGF21 is
308 considered a marker of physiological stress since its production may be induced by several

309 acute metabolic stress signals such as fasting (Nygaard et al., 2018), (Sargeant, et al., 2018;
310 Willis et al., 2019), and overfeeding (Lundsgaard et al., 2017; Willis et al., 2020). Notably,
311 however, circulating FGF21 concentrations are chronically elevated in obesity, the metabolic
312 syndrome (Zhang et al., 2008), type 2 diabetes (Chavez et al., 2009), and NAFLD (Dushay et
313 al., 2010), potentially as a compensatory mechanism to alleviate the obesity-related metabolic
314 dysfunction. Similar to LECT2, liver fat may also be an important determinant of circulating
315 FGF21 concentrations (Okumura et al., 2013). Our data corroborate this notion as circulating
316 FGF21 was positively associated with liver fat in our sample of volunteers.

317 Furthermore, we found circulating FGF21 to be negatively associated with CRF independent
318 of anthropometric, demographic, physical activity variables, and liver fat. This finding is in
319 agreement with two previous studies reporting inverse associations between circulating FGF21
320 and $\dot{V}O_2$ peak in middle-aged and elderly men and women (Matsui et al., 2019; Taniguchi et
321 al., 2014). These data are also consistent with experimental research by Taniguchi et al. (2016)
322 who showed that five weeks of exercise training reduces circulating FGF21 concentrations
323 alongside improvements in CRF and reductions in liver fat content (Taniguchi et al., 2016). In
324 a subsequent regression analysis, the authors concluded that the liver fat reduction may be
325 mediating the exercise-induced decrease in circulating FGF21. Importantly, our regression
326 analysis demonstrated that the negative association between FGF21 and CRF was independent
327 of liver fat. Henceforth, this raises the possibility that interventions aimed at improving CRF
328 may be able to reduce FGF21 independent of changes in liver fat. Due to the observational
329 nature of the present study, future studies are needed to confirm this in experimental trials.

330 Additionally, we found that circulating FGF21 concentrations were independently positively
331 associated with greater objectively measured MVPA. Given our inverse association between
332 circulating FGF21 and CRF, the positive association observed with MVPA may appear

333 unexpected. However, our data are consistent with the work of others (Cuevas-Ramos et al.,
334 2010, 2012) who have previously observed positive associations between circulating FGF21
335 and MVPA when measured using self-report questionnaires. However, the study by Cuevas-
336 Ramos et al. (2012) demonstrated that two weeks of daily supervised physical activity was
337 sufficient to increase circulating FGF21 concentrations. Consequently, our observed positive
338 association with MVPA could represent a more transient acute response to recent physical
339 activity, whereas CRF is a global marker of longer-term trends in higher intensity physical
340 activity and healthy lifestyle practices. In contrast to our findings, Matsui et al. (2020) recently
341 reported an inverse association between circulating FGF21 concentrations and objectively
342 measured MVPA after adjustment for potential confounders (Matsui et al., 2020). It must be
343 noted, however, that this association was only evident in their older cohort (mean age = 70
344 years), whilst the participants in the present study ranged from 18 to 59 years; thus, older age
345 may be an important factor mediating this relationship.

346 Interestingly, although represented as statistical tendencies, our interaction analyses showed
347 that the relationship between circulating FGF21 and MVPA may be modified by both BMI and
348 CRF. Specifically, the positive association between circulating FGF21 and MVPA was
349 stronger in those with lower BMI, and higher levels of CRF. This finding is in agreement with
350 Slusher et al. (2015) who observed that the circulating FGF21 response to an acute bout of
351 exercise was blunted in individuals with overweight or obesity, potentially due to the greater
352 FGF21 resistance in these individuals (Slusher et al., 2015). Therefore, when split based on
353 median CRF and BMI, the fitter and leaner individuals in our study cohort may possess a
354 greater FGF21 sensitivity and are thus more responsive to regular bouts of physical activity.
355 This supports the idea that chronic exercise training may act as an FGF21 sensitizer (Fletcher
356 et al., 2012), potentially through increasing CRF and reducing body weight, which in turn could

357 increase the responsiveness of FGF21 to regular physical activity. Appropriately designed
358 rodent and human studies are required to test this hypothesis in an experimental setting.

359 A crucial strength of the present study is our robust measurement of physical activity variables
360 and sedentary time using accelerometers, and the use of MRI to quantify liver fat percentage.
361 Furthermore, our sample is a diverse group of community volunteers spanning a wide range of
362 demographic and physical variables. Some limitations of this study must also be recognized,
363 however. The cross-sectional nature of the present study means that causality cannot be
364 inferred. Notably, CRF is a global marker of overall health status that reflects genetic,
365 environmental, and behavioural factors. Therefore, the associations reported here, could be
366 confounded by unmeasured determinants of CRF. Additionally, the study participants were
367 free from chronic disease; thus, future studies are needed to test our identified associations in
368 clinical populations such as type 2 diabetes and NAFLD. Finally, whilst this study examined
369 whether activity behaviours were independently associated with hepatokines, future studies
370 should determine the interactive effects of sedentary time and physical activity (Julian et al.,
371 2022).

372 In conclusion, the present study found that in a sample of community volunteers, CRF is
373 negatively associated with both circulating LECT2 and FGF21 concentrations. Furthermore,
374 circulating FGF21 is positively associated with MVPA, and this relationship may be stronger
375 in those with a lower BMI and higher CRF. These findings suggest that independent of key
376 demographics, sedentary time, physical activity, and liver fat, CRF is an important determinant
377 of circulating concentrations of LECT2 and FGF21. Additional studies are now required to
378 determine if reported association are in causal nature by undertaking interventions aimed at
379 increasing CRF through chronic structured exercise training in both community volunteers and
380 clinical populations.

381

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387

388 **Authors Contribution**

389 SM, JAK and SAW developed the initial idea for this secondary data analysis, which was
390 further refined with DJS and SM. MJR, FRG, AET and DB collected the primary data which
391 this secondary analysis is based on. SM, SAW and JAK led the analysis of this paper with
392 support from JH, DHB and all other authors. All authors approved the final version of this
393 manuscript and hold accountability for all aspects of the work.

394

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396 **Data Availability Statement**

397 The datasets analysed during the current study are available from the corresponding author on
398 reasonable request.

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643 **Table 1. Participant characteristics.**

Demographic variables	Combined		Female		Male	
	(n = 141)		(n = 56)		(n = 85)	
Ethnicity (white European)	119	[84]	51	[91]	68	[80]
Age (years)	37.0	(19.0)	34.5	(14.7)	38.0	(17.0)
Height (cm)	172.8	± 8.9	165.2	± 6.1	177.8	± 6.6
Body mass (kg)	80.9	± 19.7	66.5	± 11.1	90.3	± 18.3
Anthropometric variables						
BMI (kg·m ⁻²)	26.1	(6.3)	24.1	(4.9)	27.4	(6.4)
MRI-derived variables						
Liver fat (%) ^a	1.8	(2.1)	1.3	(0.9)	2.3	(5.8)
Cardiorespiratory fitness, sedentary time, and physical activity						
CRF (mL·kg ⁻¹ ·min ⁻¹)	40.8	± 9.8	38.9	± 6.0	42.1	± 11.5
Sedentary time (mins·d ⁻¹) ^a	580	± 95	557	± 82	595	± 100
MVPA (mins·d ⁻¹) ^a	50	(41)	46	(41)	50	(41)
Device wear time (mins·d ⁻¹) ^a	925	(73)	917	(63)	926	(83)
Hepatokines						
LECT2 (ng·mL ⁻¹)	25	± 6	25	± 5	25	± 7
FGF21 (pg·mL ⁻¹) ^a	116	(162)	88	(107)	145	(211)

644 Data are presented as mean ± SD, median (interquartile range) or number [column percentage]. BMI,
645 body mass index; CRF, cardiorespiratory fitness; MRI, magnetic resonance imaging; MVPA, moderate-
646 vigorous intensity physical activity; LECT2, leukocyte cell-derived chemotaxin 2; FGF21, fibroblast
647 growth factor 21. ^aPlease note n = 130 for physical activity data, n = 126 for liver fat data and n = 140
648 for FGF21 data

649 **Table 2. Associations of cardiorespiratory fitness and objectively measured sedentary time and physical activity with circulating**
 650 **hepatokines**

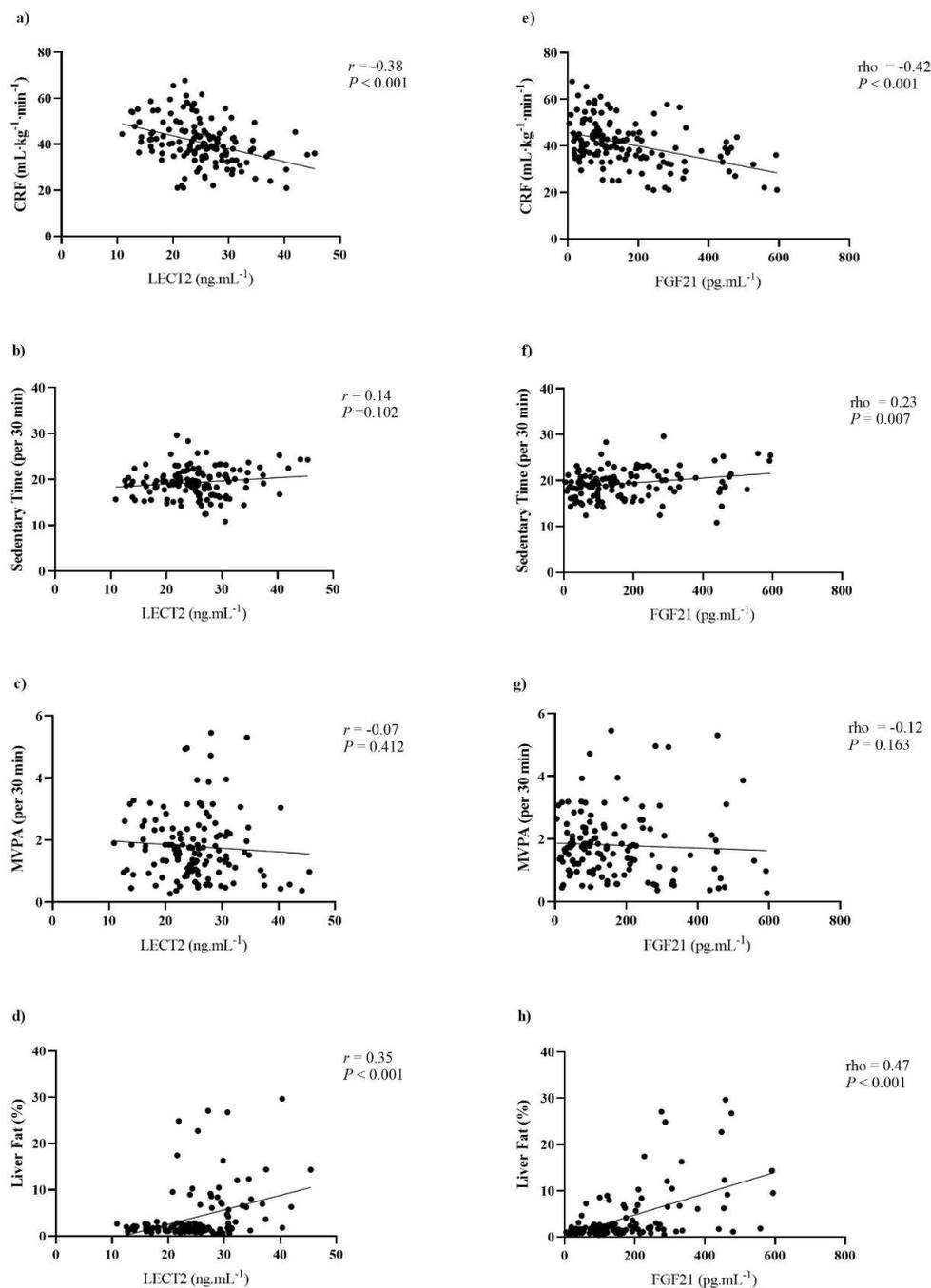
	<i>CRF</i> (mL·kg ⁻¹ ·min ⁻¹) ^{bc}		Sedentary time (per 30 mins) ^{ac}		MVPA (per 30 mins) ^{ab}	
	Fold-change (95% CI)	<i>P</i> -value	Fold-change (95% CI)	<i>P</i> -value	Fold-change (95% CI)	<i>P</i> -value
Model 1						
LECT2 (ng.mL ⁻¹)	0.74 (0.60 to 0.89)	0.001	1.10 (0.95 to 1.23)	0.233	1.00 (0.89 to 1.12)	0.971
FGF21 (pg.mL ⁻¹)	0.49 (0.29 to 0.81)	0.007	1.20 (0.85 to 1.70)	0.306	1.20 (0.91 to 1.62)	0.202
Model 2						
LECT2 (ngm.L ⁻¹)	0.72 (0.59 to 0.87)	0.001	1.10 (0.95 to 1.26)	0.178	1.07 (0.95 to 1.20)	0.234
FGF21 (pgm.L ⁻¹)	0.42 (0.25 to 0.71)	0.001	1.35 (0.93 to 1.91)	0.109	1.12 (1.12 to 2.09)	0.009
Model 3						
LECT2 (ngm.L ⁻¹)	0.76 (0.63 to 0.91)	0.003	1.07 (0.93 to 1.23)	0.276	1.10 (0.98 to 1.23)	0.130
FGF21 (pgm.L ⁻¹)	0.47 (0.27 to 0.78)	0.004	1.29 (0.91 to 1.78)	0.150	1.55 (1.12 to 2.14)	0.006

651 Data were back-transformed to show fold-change (95% CI). Model 1 adjusted for study, sex, ethnicity, age, BMI, and device wear time. Model 2
 652 adjusted for all of the previous plus ^a*CRF*, ^b*sedentary time*, or ^c*MVPA*. Model 3 adjusted for all of the previous covariates plus liver fat. *CRF*,
 653 cardiorespiratory fitness; *MVPA*, moderate-vigorous intensity physical activity; *LECT2*, leukocyte cell-derived chemotaxin 2; *FGF21*, fibroblast
 654 growth factor 21.

655 **Table 3. Statistically significant interaction analyses with body mass index, cardiorespiratory fitness and objectively measured physical**
 656 **activity.**

Outcome	Variable	n	<i>P</i>-value for interaction	Category 1 Fold-change (95% CI)	Category 2 Fold-change (95% CI)
BMI				< 26.1 kg·m⁻²	≥ 26.1 kg·m⁻²
FGF21 (pg·mL ⁻¹)	MVPA (per 30 mins) ^{ab}	129	0.052	1.86 (0.52 to 6.76)	0.83 (0.46 to 1.51)
Cardiorespiratory fitness				< 40.1 mL·kg⁻¹·min⁻¹	≥ 40.1 mL·kg⁻¹·min⁻¹
FGF21 (pg·mL ⁻¹)	MVPA (per 30 mins) ^{ab}	129	0.088	1.07 (0.91 to 2.82)	2.04 (1.35 to 3.02)

657 Models adjusted for study, sex, ethnicity, age, device wear time, BMI, interaction term and all previous plus ^aCRF and ^bsedentary time. Data are
 658 presented as *P*-values for the interaction term and as fold-changes (95% confidence intervals) for categorical variables and variables stratified
 659 using the median split. BMI, body mass index; CRF, cardiorespiratory fitness; FGF21, fibroblast growth factor 21; LECT2, leukocyte cell-derived
 660 chemotaxin 2; MVPA, moderate-vigorous intensity physical activity.



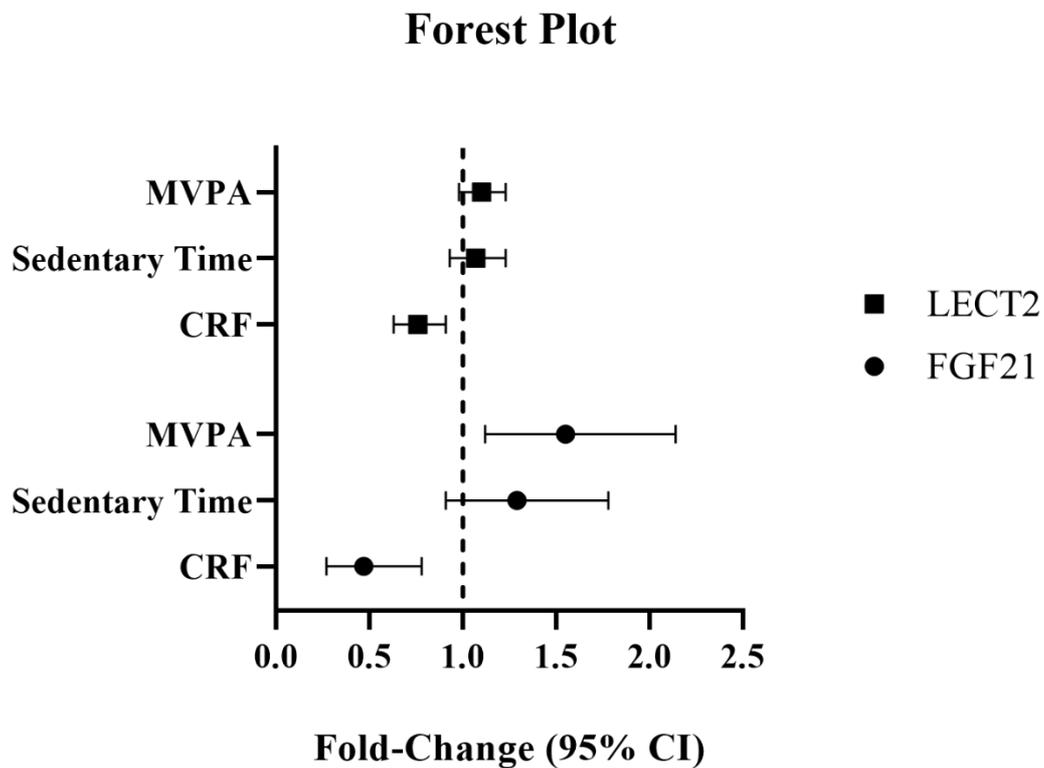
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663 **Figure 1.** Correlations of plasma LECT2 and FGF21 with CRF (a, e), sedentary time (b, f),
 664 MVPA (c, g), and liver fat (d, h). CRF, cardiorespiratory fitness; FGF21, fibroblast growth
 665 factor 21; LECT2, leukocyte cell-derived chemotaxin 2; MVPA, moderate-vigorous intensity
 666 physical activity.

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671 **Figure 2.** Forest plot showing the associations of cardiorespiratory fitness, sedentary time and
672 objectively measured moderate-vigorous physical activity with plasma LECT2 and FGF21.
673 Values represent fold-change and 95% CI for each SD change in CRF and physical activity
674 metrics (model 3). CRF, cardiorespiratory fitness; FGF21, fibroblast growth factor 21; LECT2,
675 leukocyte cell-derived chemotaxin 2; MVPA, moderate-vigorous intensity physical activity.

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