1	Prevalence of feline lungworm Aelurostrongylus abstrusus in England
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#### 26 Abstract

27 Infection of cats with lungworm Aelurostrongylus abstrusus has recently been documented in 28 the UK. Here, we aimed to study the prevalence of A. abstrusus in fecal samples from cats across England. A total of 950 fecal samples were collected from cats together with information 29 30 on their age, breed, gender, geographic region, lifestyle, and treatment history. A total of 17 31 (1.7%) cats were positive for A. abstrusus based on species-specific morphological features of 32 the larvae isolated by Baermann's technique. There was no statistically significant difference 33 in the proportion of positive samples between females (506; 53.2%) and males (444; 46.7%). 34 Multiple regression analysis showed that prevalence of feline lungworm was significantly 35 different across geographic regions: in comparison with East Midlands, some regions had 36 shown significantly increased odds of A. abstrusus-positive samples (South East [odds ratio 37 [OR] = 7.68; 95% confidence interval [CI] = 1.70 to 32.76; p = 0.01]; West Midlands [OR =38 6.20; 95% CI = 1.21 to 26.84; p = 0.02]), while other regions had also increased odds although 39 not statistically significant (Greater London [OR = 9.63; 95% CI = 0.43 to 84.05; p = 0.07]; 40 North West [OR = 4.25; 95% CI = 0.59 to 20.89; p = 0.09]; South West [OR = 2.48; 95% CI = 0.12 to 17.64; p = 0.43]; and North East [OR = 1.88; 95% CI = 0.10 to 12.24; p = 0.57]). 41 42 Keeping cats inside was protective against the risk of infection compared with those having 43 outdoor access (OR = 0.09; 95% CI = 0.01 to 0.48; p = 0.02). On the other hand, age, breed, 44 gender and deworming history did not have any significant effect on the likelihood of infection. 45 Our data indicate that A. abstrusus is a parasite of potential significance in cats, in particular 46 those from certain geographic regions in England. To reduce the spread of this parasite, an 47 integrated feline lungworm control program needs to be implemented.

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*Keywords*: Cats; lungworm; *Aelurostrongylus abstrusus*; prevalence; survey; risk factors

### 51 **1. Introduction**

52 The gastropod-borne nematode Aelurostrongylus abstrusus (Railliet, 1898) is the most 53 common lungworm of domestic and wild felids, and is found in many parts of the world, 54 including Europe, USA, South America and Australia (Scott, 1973; Elsheikha et al., 2016; 55 Giannelli et al., 2017; Penagos-Tabares et al., 2018). This parasite has a considerable impact 56 on the health and welfare of cats. Also, it has shown both regional endemicity and geographic 57 expansion across Europe. Infected cats exhibit chronic wasting, cough, dyspnea, pulmonary 58 wheezes and other signs of lower airway disease, although asymptomatic cases, shedding high 59 number of larvae in feces, may also occur (Genchi et al., 2014; Elsheikha et al., 2016; Hansen 60 et al., 2017). In addition to A. abstrusus, recent studies have detected other metastrongyloids, 61 such as Troglostrongylus brevior (Crenosomatidae) and Oslerus rostratus (Filaroididae) and 62 the trichurid Eucoleus aerophilus (syn. Capillaria aerophila) in the lungs of cats (Pennisi et 63 al., 2015; Giannelli et al., 2017).

Biological and epidemiological drivers (Traversa et al., 2009; Beugnet et al., 2014; Hansen 64 65 et al., 2017), some of them yet unconfirmed, appear to be increasing the risk of infection in 66 cats in certain parts of the world. However, important gaps remain in the available literature 67 surrounding the prevalence of feline lungworm infection and its epidemiological patterns as 68 well as determinants. Lack of understanding of these changing patterns may have serious 69 implications from a clinical standpoint, given that a delay in diagnosis and treatment can lead 70 to severe lesions and even death of the infected cat. Recently, more cases have begun to be 71 observed by clinicians (Gunn-Moore and Elsheikha, 2018). Despite this increasing frequency 72 of A. abstrusus in cats, there is lack of epidemiological studies that assess the prevalence and 73 distribution of this parasite in cats in The UK. A pan-European study involving 12 countries, 74 reported 0% A. abstrusus infection rate in fecal samples collected from Cambridge, UK (Giannelli et al., 2017). However, this study is not representative to the status of A. abstrusus 75

infection in The UK due to its very small sample size. Given the paucity of data on *A. abstrusus*in The UK, a larger survey involving more samples collected from diverse geographic areas is
needed in order to provide important insight into the transmission potential of *A. abstrusus* in
cats.

80 We previously conducted a cross-sectional survey in England and in the initial phase we 81 detected A. abstrusus larvae in the feces of 2.2% (14 out of 629) of cats (Elsheikha et al., 2017). 82 Herein, we report a more up-to-date A. abstrusus prevalence rate, after the completion of the 83 survey, based on the analysis of 950 fecal samples from cats across seven main geographic 84 regions of England. Our study established a new background prevalence of A. abstrusus in cats 85 in England and identified outdoor access as a potential risk factor for A. abstrusus infection. 86 This new knowledge may lead to more insight into the real burden and risk of feline lungworm 87 infection in the UK, which will ultimately lead to improved sustainable management strategies 88 for feline aelurostrongylosis.

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- 91 **2.** Materials and methods
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93 2.1. Fecal sample	es and data collection
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From January 2016 to January 2018, fecal samples (n = 950) were collected from cats, 506 females and 444 males, across seven administrative regions of England. The study was designed to include feral and street cats in addition to domestic cats. Fecal samples were collected from cats from shelters, catteries and privately owned cats, and were examined using Baermann's technique in order to isolate the first stage larvae (L1s). Morphological identification of the isolated *A. abstrusus* L1s and its differentiation from L1s of other metastrongyloids was achieved via microscopic examination using previously described morphometric features of *A. abstrusus* larvae (Gerichter, 1949; Brianti et al., 2014; Giannelli
et al., 2014; Giannelli et al., 20117).

103 Data on age (kitten [0 - 6 months]; junior [7 months - 2 years]; prime [3 years - 6 years]; 104 mature [7 years – 10 years]; and senior [11 years – 14 years]), breed, gender (male vs female), 105 main geographic regions in England (North East, North west, South East, South West, East 106 Midlands, West Midlands, and Greater London), lifestyle (indoor, outdoor access, feral, and 107 stray) and deworming history (recently treated using anthelmintics, such as emodepside [Profender<sup>®</sup>] or macrocyclic lactones, which have reported efficacy against A. *abstrusus* vs left 108 109 un-treated) were collected. Ethical approval was granted by the Research Ethics Committee of 110 School of Veterinary Medicine and Science, University of Nottingham.

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## 112 2.2. Prevalence and risk factor analyses

113 Statistical relationships were assessed between fecal shedding of A. abstrusus larvae and 114 defined risk factors, such as age, breed, gender, geographic location, cat lifestyle, and animal 115 treatment status at the time of fecal sampling. The overall parasite prevalence was determined 116 by dividing the number of parasite-positive fecal samples by the total number of samples collected within each risk factor category. Test of independence for contingency tables was 117 118 used to evaluate associations between each risk factor (e.g., animal age, breed, gender etc.) and 119 presence of A. abstrusus larvae. Multiple logistic regression was used to investigate the 120 associations between host-specific, demographic, and environmental risk factors with respect 121 to test outcome (e.g., parasite present or absent). Risk factors that were significant at a p level of < 0.1 were then incorporated in a forward-stepping manner into multiple logistic regression 122 123 models. These multivariable models yielded adjusted odds ratios (OR) that simultaneously 124 measured the strength of associations between multiple risk factors and the presence of parasite larvae in cat's feces. 125

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#### 127 **2.** Results and discussion

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## 129 2.1. Characteristics of the cat population

A total of 950 cats were examined with ages ranging from 2 to 240 months (mean age 53.1  $\pm$  37.7 months). Approximately, 53.2% (506) of the cats were females and 46.7% (444) were males. Breed distribution included 910 (95.78%) domestic short hair, 27 (2.84%) domestic longhair and 13 (1.36%) belonged to British Semi longhair (*n* = 6), British longhair (*n* =3), Bengal (*n*=1), Burmese (*n*=1), Cornish Rex (*n*=1), and Maine Coon (*n*=1).

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## 136 2.2. Prevalence of and risk factors associated with infection

137 In our study, 1.7% (17/950) of the fecal samples tested were positive. According to a recent 138 epidemiological survey conducted across 12 European countries, feline lungworms were the 139 second most frequent group of nematodes diagnosed in cats, and A abstrusus was the most 140 frequently detected lungworm species across Europe, but none of the samples tested from cats 141 in Cambridge (UK) was positive for A. abstrusus (Giannelli et al., 2017). However, the 0% 142 prevalence reported previously from Cambridge may not be representative of the cat population 143 in The UK due to the very small sample size examined. In fact, prevalence obtained in our 144 study seems to fall within the prevalence range reported in Europe, which varied greatly from 145 0.38% in Croatia (Grabarević et al., 1999), 1% in Germany (Mundhenke and Daugschies, 146 1999) 2.08% in Ireland (Garcia-Campos et al., 2018), 2.3% in Switzerland (Zottler et al., 2019), 147 2.6% in the Netherlands (Robben et al., 2004), 8.3% in the Denmark (Hansen et al., 2017), 148 26.5% in in Italy (Genchi et al., 2014), to 43.1% in Albania (Knaus et al., 2011). 149 The prevalence rate of A. abstrusus can also vary significantly within the same country, for example in Denmark local prevalence rates varied from 0% [95% CI: 0.0-8.8] to 31.4% 150

151 [95% CI: 16.9–49.3] (Hansen et al., 2017). Difference in prevalence rates was also detected 152 among three regions in Italy (Giannelli et al., 2017). A similar trend was detected in our study 153 where significant differences were observed in the geographic regions in regard to their association with the increase in the odds of A. abstrusus infection, in comparison with East 154 155 Midlands region (Table 1). The disparity among the prevalences of feline lungworm A. 156 abstrusus across geographic regions may reflect the level of transmission or availability of 157 intermediate and paratenic reservoir hosts that are able to maintain A. abstrusus life cycle in 158 certain areas. The broad geographic distribution of A. abstrusus in our study indicates that A. 159 *abstrusus* is circulating in cat population and not restricted to a certain locality in England. 160 Further epidemiological studies are required to determine the factors that drive the transmission 161 of A. abstrusus in the areas where this parasite is highly prevalent.

162 In this study, the larvae per gram of feces (LPG) were determined using Baermann's 163 technique and ranged from 8 to 22 (11.6  $\pm$ 3.3). This was surprisingly low compared to a 164 previous study that detected a mean of 508.7 LPG (Giannelli et al., 2017). The larval survival 165 tend to decline, due to dehydration, depending on the cat litter type and the duration of time 166 faecal samples remain in the litter; a reduction in the viability of 80% of larvae occurred after 167 3 h and reached almost 100% after 24 h (Abbate et al., 2018). In our study, the length of time 168 samples remained in the litter before collection ranged from 1 to 4 h. Thus, we must be 169 cognizant of the potential influence of dehydration on the larval viability, given the low-170 parasitic load and the time elapsed while the samples are present in the cat litter, which may 171 have underestimated the isolation rate of larvae in our study. It is also worth mentioning that 172 although Baermann's technique is specifically used for direct isolation of lungworm larvae 173 from feces, its diagnostic performance and sensitivity can be compromised by various factors. 174 These include the inability to isolate larvae in the pre-patent period, inconsistent shedding of the larvae especially in cases with low parasite burdens, or cessation of shedding larvae by 175

some cats, despite being infected, which in turn lead to false negative results (Hamilton, 1968; Elsheikha et al., 2016). To increase the accuracy of detection of *A. abstrusus* in future surveys, Baermann's technique should be performed on freshly voided fecal samples collected on three consecutive days. Serological detection of antibodies has dramatically improved the sensitivity of detection of lungworms (Zottler et al., 2017), adding more value for the diagnosis of feline aelurostrongylosis. Thus, a greater emphasis on the use of a serological assay in conjunction with fecal analysis may be warranted to achieve more accurate laboratory diagnosis.

We examined the association between age, breed, gender, lifestyle, deworming history, and the risk of infection with *A. abstrusus*. Our risk assessment analysis did not detect any effect of the age or breed on the frequency of infection. However, the risk of *A. abstrusus* infection in Denmark was lower in kittens younger than 11 weeks compared to older cats (Hansen et al., 2017). Another study reported significantly higher prevalence in cats younger than 2 years and in cats co-infected with other gastrointestinal parasites (Giannelli et al., 2017). In the present and previous studies, gender was not a risk factor for infection with *A. abstrusus* 

190 (Traversa et al., 2008; Barutzki and Schaper, 2013; Olsen et al., 2015; Hansen et al., 2017).

191 Also, we did not detect any differences between neutered and intact cats

192 Out of the 17 infected cats, 13 had outdoor access, three were stray cats and one was an 193 indoor cat (Table 2). Keeping cats indoor was associated with significant protection against 194 infection (odds ratio [OR] = 0.09; 95% confidence interval [CI] = 0.01 to 0.48; p = 0.02). In 195 contrast, no significant difference was detected between stray cats and cats with outdoor access 196 (OR = 1.01; 95% CI = 0.22 to 3.49; p = 0.99); probably due to the small number of cats in 197 these categories. These findings lend further support to previously reported findings where 198 rural origin, feral lifestyle and outdoor access have been shown to correlate with an increased 199 risk of infection (Traversa et al., 2009; Beugnet et al., 2014; Hansen et al., 2017).

200 Out of the 17 infected cats, four cats were treated with various anthelmintics on the day of 201 sample collection and 13 cats were non-treated. Our analysis has shown a lack of correlation 202 between deworming history and risk of infection. Although deworming was not associated with a significant reduction in the infection risk, treatment is still a key factor that influences the 203 204 frequency of infection with lungworm in cats. This is because many cats with outdoor access 205 have more opportunity to acquire infection by preying on intermediate and transport hosts. 206 Also, the frequency and timing of deworming are likely to affect the likelihood of infection. 207 Additionally, these results should be interpreted with caution given the small number of 208 positive samples in the stratified categories, which may not have been sufficient to identify any 209 protective effect of deworming.

210 In conclusion, the present study addressed a significant aspect of the epidemiology of A. 211 abstrusus, a potential serious health problem in feline medicine. Our findings demonstrate that 212 A. abstrusus is present in 1.7% of cats in England, and infection frequency seems to vary 213 according to the geographic region and lifestyle. These findings suggest that A. abstrusus 214 should be considered a potential cause of respiratory tract disease in cats presenting with 215 pulmonary manifestations including cats with mild respiratory signs. However, it is possible 216 that cats can be infected and shed high number of larvae in feces without presenting clear 217 clinical signs. Therefore, integrated strategies for management of A. abstrusus as well as other 218 feline metastrongyloid lungworms (Troglostrongylus spp., Oslerus rostratus, Capillaria aerophila), should be implemented and can be achieved through using preventative 219 220 anthelmintics, enhanced diagnostics and increased awareness of feline lungworms. To this end, 221 awareness and education campaigns, launched by pharma and professional organizations, such 222 as European Scientific Counsel Companion Animal Parasites (ESCCAP) and Companion 223 Animal Parasite Council (CAPC), which promote adherence to lungworm prophylaxis should 224 be tailored to at-risk cat populations.

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232	References
233	
234	Abbate, J.M., Arfuso, F., Gaglio, G., Napoli, E., Cavalera, M.A., Giannetto, S., Otranto, D.,
235	Brianti, E., 2018. Larval survival of Aelurostrongylus abstrusus lungworm in cat litters. J
236	Feline Med Surg. Nov 12:1098612X18811168. doi: 10.1177/1098612X18811168. [Epub
237	ahead of print]
238	Barutzki, D., Schaper, R., 2013. Occurrence and regional distribution of Aelurostrongylus
239	abstrusus in cats in Germany. Parasitol. Res. 112, 855-861.
240	Beugnet, F., Chalvet-Monfray, K., Cozma, V., Farkas, R., Guillot, J., Halos, L., Joachim, A.,
241	Losson, B., Miro, G., Otranto, D., Renaud, M., Rinaldi, L., 2014. Parasites of domestic
242	owned cats in Europe: co-infestations and risk factors. Parasit. Vectors 7,: 291.
243	Brianti, E., Giannetto, S., Dantas-Torres, F., Otranto, D., 2014. Lungworms of the genus
244	Troglostrongylus (Strongylida: Crenosomatidae): neglected parasites for domestic cats.
245	Vet. Parasitol. 202, 104-112.
246	Elsheikha, H.M., Schnyder, M., Traversa, D., Di Cesare, A., Wright, I., Lacher, D.W., 2016.
247	Updates on feline aelurostrongylosis and research priorities for the next decade. Parasit.
248	Vectors. 9(1), 389.

249	Elsheikha, H.M., Schunack, B., Schaper, R., 2017. Prevalence of feline lungworm
250	Aelurostrongylus abstrusus in England. Abstract Book , 407, The 26th International
251	Conference of the World Association for the Advancement of Veterinary Parasitology,
252	WAAVP, Kuala Lumpur, Malaysia.
253	Garcia-Campos, A., Power, C., O'Shaughnessy, J., Browne, C., Lawlor, A., McCarthy, G.,
254	O'Neill, E.J., de Waal, T., 2018. One-year parasitological screening of stray dogs and cats
255	in County Dublin, Ireland. Parasitology. Dec 18:1-7. doi: 10.1017/S0031182018002020.
256	[Epub ahead of print]
257	Genchi, M., Ferrari, N., Fonti, P., De Francesco, I., Piazza, C., Viglietti, A., 2014. Relation
258	between Aelurostrongylus abstrusus larvae excretion, respiratory and radiographic signs
259	in naturally infected cats. Vet. Parasitol. 206 (3/4), 182-187.
260	Gerichter, C.B., 1949. Studies on the nematodes parasitic in the lungs of Felidae in Palestine.
261	Parasitology 39, 251-262.
262	Giannelli, A., Capelli, G., Joachim, A., Hinney, B., Losson, B., Kirkova, Z., René-Martellet,
263	M., Papadopoulos, E., Farkas, R., Napoli, E., Brianti, E., Tamponi, C., Varcasia, A.,
264	Margarida, Alho A., Madeira de Carvalho, L., Cardoso, L., Maia, C., Mircean, V.,
265	Mihalca, A.D., Miró, G., Schnyder, M., Cantacessi, C., Colella, V., Cavalera, M.A.,

- Latrofa, M.S., Annoscia, G., Knaus, M., Halos, L., Beugnet, F., Otranto, D., 2017.
- 267 Lungworms and gastrointestinal parasites of domestic cats: a European perspective. Int.
- 268 J. Parasitol. 47, 517-528.
- 269 Giannelli, A., Ramos, R.A., Annoscia, G., Di Cesare, A., Colella, V., Brianti, E., Dantas-
- 270 Torres, F., Mutafchiev, Y., Otranto, D., 2014. Development of the feline lungworms
- 271 *Aelurostrongylus abstrusus* and *Troglostrongylus brevior* in *Helix aspersa* snails.
- 272 Parasitology 141, 563-569.

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- 273 Grabarević Ž., Ćurić S., Tustonja A., Artuković B., Šimec Z., Ramadan K., Živičnjak T.
- 274 1999. Incidence and regional distribution of the lungworm *Aelurostrongylus abstrusus* in
  275 cats in Croatia. Veterinarski Arhiv 69, 279-287.
- Gunn-Moore, D., Elsheikha, H.M., 2018. Current status of feline lungworm in the UK. Vet.
  Rec. 182(4), 113-114.
- Hamilton, J.M., 1968. Studies on re-infestation of the cat with *Aelurostrongylus abstrusus*. J.
  Comp. Pathol. 78(1), 69-72.
- 280 Hansen, A.P., Skarbye, L.K., Vinther, L.M., Willesen, J.L., Pipper, C.B., Olsen, C.S., Mejer,
- H., 2017. Occurrence and clinical significance of *Aelurostrongylus abstrusus* and other
  endoparasites in Danish cats. Vet. Parasitol. 234, 31-39.
- Knaus, M., Kusi, I. Rapti, D., Xhaxhiu, D., Winter, R., Visser, M., Rehbein, S., 2011.
  Endoparasites of cats from the Tirana area and the first report on *Aelurostrongylus abstrusus* (Railliet, 1898) in Albania. Wien. Klin. Wochenschr. 123, 31-35.
- Mundhenke, H., Daugschies, A., 1999. Studies on the prevalence of endoparasites in cats in
  Hannover and surroundings. Wien. Tierarztl. Monatsschr. 86 (2), 43-48.
- 288 Olsen, C.S., Willesen, J.L., Pipper, C.B., Mejer, H., 2015. Occurrence of Aelurostrongylus
- *abstrusus* (Railliet, 1898) in Danish cats: A modified lung digestion method for isolating
  adult worms. Vet. Parasitol. 210, 32-39.
- 291 Penagos-Tabares, F., Lange, M.K., Chaparro-Gutiérrez, J.J., Taubert, A., Hermosilla, C., 2018.
- 292 Angiostrongylus vasorum and Aelurostrongylus abstrusus: Neglected and underestimated
- 293 parasites in South America. Parasit. Vectors 11, 208.
- 294 Pennisi, M.G., Hartmann, K., Addie, D.D., Boucraut-Baralon, C., Egberink, H., Frymus, T.,
- 295 Gruffydd-Jones, T., Horzinek, M.C., Hosie, M.J., Lloret, A., Lutz, H., Marsilio, F.,
- 296 Radford, A.D., Thiry, E., Truyen, U., Möstl, K.; European Advisory Board on Cat

- Diseases., 2015. Lungworm disease in cats: ABCD guidelines on prevention and
  management. J. Feline Med. Surg. 17, 626-636.
- 299 Robben, S.R., le Nobel, W.E., Döpfer, D., Hendrikx, W.M., Boersema, J.H., Fransen, F.,
- 300 Evsker, M.E., Infections with helminths and/or protozoa in cats in animal shelters in the
- 301 Netherlands.2004. Tijdschr Diergeneeskd. 129(1), 2-6.
- Scott, D.W., 1973. Current knowledge of aelurostrongylosis in the cat. Literature review and
   case reports. Cornell Vet. 63, 483-500.
- 304 Traversa, D., Lia, R.P., Boari, A., Di Cesare, A., Capelli, G., Milillo, P., Otranto, D.,
- 305 2009. Feline aelurostrongylosis: epidemiological survey in central and southern Italy.
- 306 Veterinaria 23, 41-45.
- 307 Traversa, D., Lia, R.P., Iorio, R., Boari, A., Paradies, P., Capelli, G., Avolio, S., Otranto, D.,
- 308 2008. Diagnosis and risk factors of *Aelurostrongylus abstrusus* (Nematoda, Strongylida)
  309 infection in cats from Italy. Vet. Parasitol. 153, 182-186.
- 310 Zottler, E.M., Strube, C., Schnyder, M., 2017. Detection of specific antibodies in cats
- 311 infected with the lung nematode *Aelurostrongylus abstrusus*. Vet Parasitol. 235, 75-82.
- 312 Zottler, E.M., Bieri, M., Basso, W., Schnyder, M., 2019. Intestinal parasites and lungworms
- in stray, shelter and privately owned cats of Switzerland. Parasitol. Int. 69, 75-81.
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# **Table 1.**

	Geographic region	Prevalence*	Odd ratios	95% CI	<i>p</i> -value
	Greater London	1/17 (5.8)	9.632	0.43 to 84.05	0.07
	South East	4/72 (5.5)	7.68	1.70 to 32.76	0.01
	West Midlands	3/69 (4.3)	6.2	1.21 to 26.84	0.02
	North West	2/57 (3.5)	4.25	0.59 to 20.89	0.09
	South West	1/46 (2.1)	2.48	0.12 to 17.64	0.43
	North East	1/49 (2.0)	1.88	0.10 to 12.24	0.57
324	* Using East Midlands	as a reference [5	infected out of 6	523 tested (80.2%)]	, Greater London,
325	North West, South	East, West Midl	ands showed sign	nificant differences	. Prevalence is
326	shown as number	of positive sample	es/total number to	ested (%).	
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323 The prevalence of *A. abstrusus* across seven administrative regions in England.

## Table 2.

- 341 Lifestyle distribution and positivity rates of *A. abstrusus* in cats examined in this study.
- 342 Correlation was established only between cats living indoor and *A. abstrusus* infection.

	No. of	No. of	Larvae per	
Lifestyle category	uninfected cats	infected cats	gram of feces	
Indoor	375	1	12.0	
Outdoor Access	455	13	11.5	
Stray	81	3	12.0	
Feral	22	0	0.0	
Total ( $n = 950$ )	933	17	11.6	