

## **Drug-induced injury due to flucloxacillin: relevance of multiple HLA alleles**

\*Paola Nicoletti,<sup>1,2</sup> \*Guruprasad P. Aithal,<sup>3</sup> Thomas C. Chamberlain,<sup>4§</sup> Sally Coulthard,<sup>4</sup> Mohammad Alshabeeb,<sup>4,5</sup> Jane I. Grove,<sup>3,6</sup> Raul J. Andrade,<sup>7</sup> Einar Bjornsson,<sup>8</sup> John F. Dillon,<sup>9</sup> Par Hallberg,<sup>10</sup> M. Isabel Lucena,<sup>7</sup> Anke H. Maitland-van der Zee,<sup>11</sup> Jennifer H. Martin,<sup>12</sup> Mariam Molokhia,<sup>13</sup> Munir Pirmohamed,<sup>14</sup> Mia Wadelius,<sup>10</sup> Yufeng Shen,<sup>15,16</sup> Matthew R. Nelson,<sup>17</sup> and Ann K. Daly<sup>4</sup> for the International Drug-induced Liver Injury consortium (iDILIC)\*\*

\*Joint first authors

<sup>1</sup>Department of Genetics and Genomic Sciences, Icahn School of Medicine at Mount Sinai, New York, USA

<sup>2</sup>Sema4, a Mount Sinai venture, Stamford, Connecticut, USA

<sup>3</sup>National Institute for Health Research (NIHR) Nottingham Biomedical Research Centre, Nottingham University Hospital NHS Trust and University of Nottingham, Nottingham, UK

<sup>4</sup>Institute of Cellular Medicine, Newcastle University, Newcastle upon Tyne, UK

<sup>5</sup>Developmental Medicine Department, King Abdullah International Medical Research Center, Riyadh, Kingdom of Saudi Arabia

<sup>6</sup>Nottingham Digestive Diseases Centre, School of Medicine, University of Nottingham, Nottingham, UK

<sup>7</sup>UGC Digestivo y Servicio de Farmacología Clínica, Instituto de Investigación Biomédica de Málaga (IBIMA), Hospital Universitario Virgen de la Victoria, Universidad de Málaga, Málaga, Spain and Centro de Investigación Biomédica en Red de Enfermedades Hepáticas y Digestivas (CIBERehd), Madrid, Spain

<sup>8</sup>Division of Gastroenterology and Hepatology, Department of Internal Medicine, The National University Hospital of Iceland, Reykjavik, Iceland

<sup>9</sup>Medical Research Institute, University of Dundee, Ninewells Hospital, Dundee, UK

<sup>10</sup>Department of Medical Sciences and Science for Life Laboratory, Uppsala University, Uppsala, Sweden

<sup>11</sup>Department of Respiratory Medicine, Academic Medical Center (AMC), University of Amsterdam, Amsterdam, Netherlands

<sup>12</sup>School of Medicine and Public Health, University of Newcastle, New South Wales, Australia

<sup>13</sup>School of Population Health and Environmental Sciences, Faculty of Life Sciences and Medicine, King's College, London, UK

<sup>14</sup>Department of Molecular and Clinical Pharmacology, University of Liverpool, Liverpool, UK

<sup>15</sup>The Herbert Irving Comprehensive Cancer Center, Columbia University, New York, USA

<sup>16</sup>Department of Biomedical Informatics, Columbia University, New York, USA

<sup>17</sup>Target Sciences, GSK, King of Prussia, PA, USA

\*\*A complete list of iDILIC investigators is provided in the Supplementary Material

§Current address: University of British Columbia, Vancouver, Canada

Conflict of interest: MRN is an employee of GSK. The other authors declare no conflicts of interest.

Correspondence to: Professor Ann K. Daly, Institute of Cellular Medicine, Newcastle University, Framlington Place, Newcastle upon Tyne NE2 4HH, UK, email a.k.daly@ncl.ac.uk, Tel: +44 191 208 7031

Keywords: drug-induced liver injury, HLA genes, adverse drug reactions, genetic polymorphisms

Electronic word count: main text 3918 words

2 figures and 5 tables

Funding information: The genome-wide association study and iDILIC case enrolment and sample collection was funded by the International Serious Adverse Events Consortium with (Phase 2) membership support from Abbott, Amgen, Daiichi-Sankyo, GlaxoSmithKline, Merck, Novartis, Pfizer, Roche, Sanofi-Aventis, Takeda, and the Wellcome Trust. TCC was funded by a BBSRC-Industrial CASE studentship with AstraZeneca (PI AKD). This is a summary of independent research partly (the DILIGEN and iDILIC sample collection) funded by the National Institute for Health Research (NIHR) Nottingham Digestive Diseases Biomedical Research Unit at the Nottingham University Hospitals NHS Trust and University of Nottingham. The views expressed are those of the authors and not necessarily those of the NHS, the NIHR or the Department of Health. GPA is the gastrointestinal and liver disorder theme lead for the NIHR Nottingham BRC (Reference no: BRC-1215-20003). The EUDRAGENE collaboration (MM) received support from the EC 5th Framework program (QLRI-CT-2002-02757). The Spanish DILI Registry (RJA, MIL) is partly funded by the Spanish Medicine Agency, Fondo Europeo de Desarrollo Regional - FEDER (P10-CTS-6470, FIS PI12/00378, PI16/01748). CIBERehd is funded by Instituto de Salud Carlos

III. The Swedish case collection (SWEDEGENE) (PH, MW) has received support from the Swedish Medical Products Agency, the Swedish Society of Medicine (2008-21619), Swedish Research Council (Medicine 521-2011-2440 and 521-2014-3370), and Swedish Heart and Lung Foundation (20120557). The Swedish Twin Registry which provided control data is managed by Karolinska Institutet and receives funding through the Swedish Research Council under the grant no 2017-00641. MM was supported by the National Institute for Health Research (NIHR) Biomedical Research Centre at Guy's and St Thomas' NHS Foundation Trust and King's College London. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

## Abstract

Some patients prescribed flucloxacillin (~0.01%) develop drug-induced liver injury (DILI). *HLA-B\*57:01* is an established genetic risk factor for flucloxacillin DILI. To consolidate this finding, identify additional genetic factors and assess relevance of risk factors for flucloxacillin DILI in relation to DILI due to other penicillins, we performed a genome-wide association study involving 197 flucloxacillin DILI cases and 6835 controls. We imputed SNP and HLA genotypes. *HLA-B\*57:01* was the major risk factor (allelic OR=36.62,  $P=2.67 \times 10^{-97}$ ). *HLA-B\*57:03* also showed an association (OR=79.21,  $P=1.2 \times 10^{-6}$ ). Within the HLA-B protein sequence, imputation showed valine<sup>97</sup>, common to *HLA-B\*57:01* and *HLA-B\*57:03*, had the largest effect (OR=38.1,  $P=9.7 \times 10^{-97}$ ). We found no *HLA-B\*57* association with DILI due to other isoxazolyl penicillins (n=6) or amoxicillin (n=15) and no significant non-HLA signals for any penicillin-related DILI. (125 words)

Flucloxacillin is a very common cause of drug-induced liver injury (DILI) in a number of countries worldwide, especially in Northern Europe and Australasia.(1) The incidence of DILI due to this drug is 8.5 per 100,000 people prescribed flucloxacillin increasing to 35 per 100,000 in those receiving one consecutive prescription and 110.5 per 100,000 in those aged >70 years who have received two or more prescriptions.(2) Most patients recover completely but some may develop chronic disease with effects such as vanishing bile duct syndrome.(1) The toxicity is occasionally fatal.(3) Though flucloxacillin DILI was originally thought to be due to impaired drug metabolism (4), features such as rash and fever more typical of hypersensitivity reactions are often seen.(1, 5) The relevance of the immune system to this form of adverse drug reaction was indicated by the finding that 85% of a case cohort (n=51) were positive for *HLA-B\*57:01*.(6) Further studies showed that CD8-positive T cells from *HLA-B\*57:01*-positive donors could be activated by flucloxacillin in a mechanism that appeared to involve the formation of drug-protein conjugates,(7) though a non-hapten mechanism has also been proposed.(8) Although carriage of *HLA-B\*57:01* is also a strong risk factor for abacavir hypersensitivity, the predictive power of *B\*57:01* for this adverse drug reaction is higher than for flucloxacillin-related DILI and the underlying mechanism, which involves a change in the peptide repertoire in the peptide binding groove of *B\*57:01* appears different.(9)

Despite the strong association of flucloxacillin DILI with *HLA-B\*57:01*, overall sensitivity and specificity of genotyping to predict susceptible individuals are both too low to justify using this test routinely prior to prescription since only approx. 1 in every 500 *HLA-B\*57:01* carriers, who comprise approx. 5% of Europeans, are likely to develop DILI if treated with flucloxacillin.(6) However, this form of DILI is more common than estimated previously with individuals over 70 years particularly at risk, with the number needed to test for *B\*57:01* in this group to prevent one case now estimated at 2512.(2) Finding additional genetic risk factors might lower this number further. To investigate this possibility both in those positive for *B\*57:01* and those who don't carry the genotype, we have extended the size of our case group and performed a further genome-wide association study (GWAS)

with improved imputation of genotypes together with additional HLA typing. We have also investigated whether *B\*57:01* is relevant to the risk of DILI due to the other related isoxazolyl penicillins including cloxacillin and dicloxacillin, which are often used as alternatives to flucloxacillin and are generally considered to be less important causes of DILI. We assess possible overlap in genetic risk factors for DILI due to other beta lactam antibiotics by studies on amoxicillin-related DILI.(5)

## **Results**

### **Characteristics of the study subjects**

The flucloxacillin DILI cases in the study were from two separate recruitment phases. Phase I consisted of cases included in two previous studies (all from UK DILIGEN study) (n=75)(6, 10) and phase II more recently recruited cases for the iDILIC study (n=122) from UK, Sweden, Netherlands and Australia. The majority of the phase I cases (n=51) were included in an earlier GWAS on flucloxacillin DILI.(6) The combined 197 cases were from the UK (n=156), Sweden (n=37), the Netherlands (n=3) and Australia (n=1). Clinical inclusion criteria for all cases were those described by Aithal et al.(11) Causality assessment and inclusion criteria were as described previously.(12) Clinical data for the flucloxacillin DILI cases (phase I and phase II) included in the GWAS are summarized in Table 1. Thirty percent of the cases (n=59) were aged 70 years or over and 68% (n=134) were female.

DILI cases relating to dicloxacillin (n=2), cloxacillin (n=2), and oxacillin (n=2) were recruited in USA and Iceland from the DILIN and iDILIC studies.(10, 12) Cases of DILI relating to amoxicillin alone were also recruited from a range of European centres (n=13) and from USA (n=2) as described previously.(12) In addition, two DILI cases from Spain relating to cloxacillin were analyzed. Data for the cases due to other isoxazolyl penicillins (n=6) and the amoxicillin only cases (n=15) are summarized in Table S1.

Since DILI has a very low prevalence, we used general population samples as study controls. Flucloxacillin cases were then compared with a total of 6835 northern European ancestry controls from multiple available sources: Wellcome Trust Case Control Consortium,(13) the population reference sample (POPRES)(14) and PGX40001.(15) In order to increase the case/control ratio for Swedish cases, we added samples from the Swedish Twin Registry.(16) For other penicillins, we compared drug-specific cases to a total of 10588 European controls reported previously which included the 6835 northern European ancestry controls described above.(12)

### **GWAS and HLA analysis on flucloxacillin cases**

Following the initial GWAS, imputation to assign SNP and HLA genotypes was performed. Imputation methods were described in detail previously.(12) For HLA, four digit HLA alleles and amino acid sequences were inferred using SNP2HLA.(17)

Principal component analysis of the imputed SNP data confirmed the self-reported ethnicity for the cases and divided the controls into two major clusters which represent mainly UK and Swedish cases respectively (Figure 1). The GWAS results for flucloxacillin DILI are summarized in Figures 2 and S1. The only genome-wide significant peak was on chromosome 6 in the MHC region with the top SNP rs2395029 (Odds ratio (OR) = 35.48; 95% CI 25.38-49.58; P =  $5.7 \times 10^{-97}$ ). Considering only newly recruited phase II cases (n=146), the rs2395029 association showed equivalent OR and AF (OR=31, 95%CI 22.28-45.86, P= $6.9 \times 10^{-79}$ , allele frequency (AF) of 40% in cases) to the OR and AF from our smaller original phase I study.(6) A number of other SNPs in the MHC region were also genome-wide significant. The genotype data for all cases was used to impute HLA alleles. In line with the GWAS findings, the strongest imputed HLA association was with *HLA-B\*57:01* (OR = 36.62; 95% CI 26.14-51.29; P =  $2.67 \times 10^{-97}$ ) followed by *HLA-C\*06:02*, *HLA-DQB1\*03:03*, *HLA-DRB1\*07:01*, *HLA-DQA1\*02:01* and *HLA-A\*01:01* which together form part of the most common B57 haplotype (allelefreqencies.net). A second group of alleles, *HLA-C\*07:02*, *HLA-B\*07:02* and

*HLA-DQB1\*03:01* were protective and showed p values  $<10^{-5}$  (Table 2). Haplotype analysis showed that *HLA-B\*57:01*-containing haplotypes confer risk while *B\*57:01*-negative haplotypes seem to be protective, suggesting that *B\*57:01* and no other alleles within the haplotype is the main risk factor (Table S2).

Reciprocal conditional analyses on rs2395029 and *HLA-B\*57:01* demonstrated that *HLA-B\*57:03* was a MHC-significant independent risk allele (OR = 79.21; 95% CI 13.57-462.4; P =  $1.2 \times 10^{-6}$ ). *HLA-B\*57:03* is rare in Caucasians (AF = 0.0003) (Table 2). In our case group, two samples were predicted to be positive for this allele from the imputation; this was confirmed for one of the two cases by direct HLA typing. The positive *HLA-B\*57:03* cases were in both the ancestry clusters. There was no difference in AF between two ancestry clusters (UK and Sweden) for *HLA-B\*57:01* alleles (AF<sub>sweden</sub> = 0.03 vs AF<sub>uk</sub> = 0.04, p-value for direct comparison between the clusters = 0.4) and *HLA-B\*57:03* (AF<sub>sweden</sub> = 0.0002 vs AF<sub>uk</sub> = 0.0003, p-value for direct comparison between the clusters = 0.9).

The GWAS was repeated to include the *B\*57*-carriers only (n=163) in the case group but with the same control group as used in the main analysis. We detected no additional genome-wide significant signals in this further analysis (data not shown).

We also analyzed polymorphic amino acid residues in the HLA proteins to assess their individual contribution to flucloxacillin DILI susceptibility. Valine at position 97 (V<sup>97</sup>) in the HLA-B protein (OR = 38.1, 95% CI 27.07-53.62, P =  $9.7 \times 10^{-97}$ ) had a stronger association compared to the two significant single HLA *B\*57* alleles (Table 3) because V<sup>97</sup> is shared by *HLA-B\*57:01* and *HLA-B\*57:03* (Figure S2). This amino acid is also shared by other *B\*57* alleles (16 in total) and non-*B\*57* alleles (6 alleles). (18) Other than for *B\*57*, the V<sup>97</sup>-associated alleles are extremely rare or not present in Caucasian populations. A complete list of the alleles and their frequency in Caucasians is shown in Table S3. Except for *HLA-B\*57:03*, the cases did not carry any rare alleles positive for V<sup>97</sup>. Six different amino acid variants at position 97 of HLA-B are known. Except for valine, all had a



protective effect against flucloxacillin DILI, but only arginine (R)<sup>97</sup> and serine (S)<sup>97</sup> showed significant protection (OR= 0.43, P= 5.13x10<sup>-14</sup> and OR= 0.53, P=9.82x10<sup>-7</sup>) (Table 3).

### **Relationship between genetic and clinical risk factors for flucloxacillin DILI**

We analyzed the contribution of age and gender to DILI risk in the cases and a subset of controls with the necessary data on age and sex (380 controls (POPRES) (14)). Although, sex, age and HLA-B\*57 were significant in an univariate model, in a multivariate analysis model only age above 70 years and HLA-B\*57 remained significantly associated with the DILI (Table S4). These data suggest that for those older than 70 years there was an 7 fold increase in DILI risk.

We also compared selected clinical characteristics of the cases positive for any B\*57 allele with those entirely negative but there appeared to be no significant difference for any of the characteristics examined (Table S5). The proportion of patients aged 70 or over in the *HLA-B\*57:01*-negative cases was compared with the positive group. Among the *B\*57:01*-negative cases, 22% were in this older age group compared with 32% of those positive for *B\*57:01* but this difference was not statistically significant (p=0.32).

### **HLA genotypes in B\*57-negative flucloxacillin DILI cases**

To determine whether additional HLA alleles were risk factors for DILI in this set of 34 cases (17.25% of all cases) that were negative for rs2395029, *HLA-B\*57:01* and *HLA-B\*57:03*, the imputed HLA genotypes for the group were analyzed separately against a negative *HLA-B\*57* alleles control set (n= 6321). While no strong risk factors emerged, it was found that the rare allele *HLA-A\*02:02* (0.001 in controls) was enriched in the negative cases (OR = 15.24, 95% CI 1.89-123.1, P = 0.01) but absent in the group of HLA-B\*57-carriers. Two other class I alleles, *HLA-A\*30:01* and *HLA-B\*13:02*, showed nominally significant associations (Table 4). *HLA-B\*07:02* also showed a protective effect in this group, similar to the *B\*57:01*-positive group. When amino acid analysis on HLA proteins was performed on these B\*57-negative cases, we found that the most associated

variants were deletions of the first 30 and last 58 amino acids of the HLA-A protein (Table S6). The deletions showed a higher effect size and significance compared to the single top HLA-A alleles (OR = 19.67, 95% CI = 2.35-164.4, P = 0.006). This suggests that the short *A\*02:02* alleles (including *A\*02:02:02*, *A\*02:02:04* and *A\*02:02:05*) as well as some short *A\*30:01* alleles (such as *A\*30:01:03*, *A\*30:01:04*, *A\*30:01:05*, *A\*30:01:06*, *A\*30:01:07*, *A\*30:01:10*) might be enriched in *HLA-B\*57:01* negative cases. No samples were positive for the B\*57-related V<sup>97</sup>.

### **GWAS and HLA analysis on cases due to other penicillins**

A total of 21 additional European penicillin DILI cases were available to us: 15 samples due to amoxicillin only (not amoxicillin-clavulanate) and 6 due to dicloxacillin, cloxacillin or oxacillin (Table S1). The majority of these cases (62%) had a cholestatic or mixed phenotype but the percentage was slightly lower than the 81% seen in the flucloxacillin DILI cases. As for the flucloxacillin DILI cases, GWAS analysis (Figures S3 to S5) and HLA imputation were performed. In both groups, the effect of *HLA-B\*57:01* was not significant though there was a trend in the direction of a positive association (Table S7). Considering non-flucloxacillin isoxazolyl penicillin cases, the uncommon *HLA-C\*07:04* and the corresponding amino acid (phenylalanine (F)<sup>95</sup>) showed associations just above the threshold for MHC significance (both showing OR = 12.97, P= 0.001, Table 5 and Table S8). The amoxicillin DILI cases showed a significant association with *HLA-B\*58:01* (OR= 20.29, 95%CI 4.25-96.94, P= 0.0002) and borderline associations with *HLA-DPBI\*01:01*, *HLA-A\*01:01* and *HLA-C\*03:02* (Table 5). The top amino acid association was with methionine (M)<sup>67</sup> or F<sup>67</sup> as B pocket residues of the HLA-B gene product but this was not significant (OR=3.55, 95%CI 1.69-7.40, P=0.0008). No other genetic variants were found to be genome-wide or MHC significant when the penicillin cases were analyzed, either together conditioning for *HLA-B\*57* alleles or by individual drug.

Two additional cloxacillin DILI cases from Spain were not included in the main GWAS analysis which was confined to Northern Europeans, but these samples were typed directly for HLA-B alleles.

Neither was positive for *B\*57:01*; the HLA-B genotypes were *B\*07:02/B\*44:02* and *B\*08:01/B\*35:02*.

## Discussion

This study has confirmed that *HLA-B\*57:01* is an important risk factor for DILI relating to flucloxacillin. By repeating the original analysis in a larger cohort, the p value for this risk factor was lowered but also no evidence for signals outside the chromosome 6 region was obtained. The previous GWAS had found a contribution by a SNP in *ST6GAL1* which was of borderline significance when analysis was performed using cases carrying *HLA-B\*57:01* only,(6) but this was not confirmed in the current study with a substantially larger group of patients, even when B\*57-carrier only analysis was performed. Improved ability to impute HLA genotypes as well as an increase in numbers of cases and some additional direct HLA typing indicated that in addition to *HLA-B\*57:01*, *HLA-B\*57:03* is also a risk factor for flucloxacillin DILI. *B\*57:03* is very rare among Northern Europeans (for example estimated allele frequencies of 0 (Northern Ireland), 0.002 (Irish Republic) and 0.0007 (Germany) ([www.allelefrequencies.net](http://www.allelefrequencies.net))). This low allele frequency makes meaningful comparisons challenging but suggests that the *HLA-B\*57:01* association for flucloxacillin DILI is less specific than the association between *HLA-B\*57:01* and abacavir hypersensitivity. Based mainly on studies *in vitro*, alleles related closely to *HLA-B\*57:01* such as both *HLA-B\*57:02* and *HLA-B\*57:03* are not risk factors for abacavir hypersensitivity.(9) In view of a recent report involving an African population which reports that *HLA-B\*57:02* and *HLA-B\*57:03* are risk factors for an unusual form of DILI seen when both antiretroviral and anti-TB drugs are prescribed(19) as well as the strong homology in the amino acid sequence for the gene product, it is also possible that *HLA-B\*57:02* and other rare B\*57 alleles may be risk factors for flucloxacillin DILI. *HLA-B\*57:02*, like *HLA-B\*57:03*, is very rare in Northern Europeans and was not detected at all in our population but does code for V<sup>97</sup> in the HLA-B gene product similar to both *HLA-B\*57:01* and *HLA-B\*57:03*. While R<sup>97</sup> is the most common residue, encoded by a number of different common *HLA-B* alleles,(20) S<sup>97</sup> is a characteristic

residue for the protective *HLA-B\*07:02*, which is the main flucloxacillin DILI-associated protective allele (see Table 2). The residues methionine, isoleucine and tryptophan which are of similar hydrophobicity to valine, are also found in position 97 in other HLA class I proteins. When HLA-A proteins sharing these hydrophobic residues at position 97 are considered, the alleles *HLA-A\*31:01*, *A\*33:01* and *A\*33\*03*, which are strong predictors of susceptibility to skin rash or DILI due to various drugs (12, 21), each encode M<sup>97</sup>. This suggests this position and the nature of the amino acid present may be of importance in presentation of modified peptides for both HLA-A and HLA-B.

In addition to V<sup>97</sup>, D<sup>114</sup> and S<sup>116</sup> are considered to be important in the interaction between abacavir and the B\*57:01 gene product.(9) As summarised in Figure S2, amino acids 114 and 116 are not conserved in the B\*57:02 and B\*57:03 proteins. This finding of an apparent difference in HLA genotype selectivity for flucloxacillin DILI compared with abacavir hypersensitivity is in line with findings suggesting that direct interaction of flucloxacillin with the HLA-B\*57:01 gene product is unlikely.(22) There is also *in vitro* data indicating covalent binding of flucloxacillin to cellular proteins during the T cell activation process, in contrast to what is believed to occur during activation with abacavir.(7, 22) These studies also suggested that a T cell response involving *HLA-B\*58:01* might occur in flucloxacillin DILI, though the effect was less convincing than for B\*57:01.(7) The B\*58:01 protein sequence does not include V<sup>97</sup> with Arg present instead at this position. No *HLA-B\*58:01* positive carriers were found in our flucloxacillin cases. Since this allele is approx. 10 times more common in Europeans than B\*57:03, we believe our study was powered adequately to detect this if present and we did detect it in our controls (frequency 0.03%). B\*58:01 did show an increased frequency compared with controls among the amoxicillin DILI cases so it appears to be a possible separate risk factor for DILI with this drug. HLA-B\*57:01 carriage also appears to be a risk factor for DILI due to pazopanib, which does not appear to have any structural homology to flucloxacillin or abacavir, though the effect size is much smaller than that observed for flucloxacillin.(23)

We also detected significant associations with additional HLA alleles and haplotypes. The association with the B57 haplotype demonstrated the clear specific association with *B\*57:01*. Novel protective associations involving various alleles including *HLA-DQA1\*03:01*, *HLA-C\*07:02*, *HLA-B\*07:02* and *HLA-DRB1\*04:01* were seen. This protective effect for *HLA-B\*07:02* is also detectable at the amino acid level with S<sup>97</sup> in HLA-B associated with significant protection together with R<sup>97</sup> which is the most common amino acid in this position in Europeans. Protective HLA associations have generally not been reported previously for DILI, except for one report of a protective HLA class II genotype against amoxicillin-clavulanate DILI(24) (25). *DRB1\*04:01* is a well-established risk factor for autoimmune hepatitis,(26) so the protective effect observed here against flucloxacillin DILI is interesting.

The previous study on flucloxacillin DILI was relatively small and the number of cases that were *HLA-B\*57:01*-negative was low.(6) In this larger study population, the frequency of *HLA-B\*57:01* carriage of 82% is comparable to that in the original study and it seems likely that there is a genuine subgroup of cases (a total of 34 when two cases positive for the related *B\*57:03* are excluded) that are not positive either for *HLA-B\*57:01* or a closely related allele. This is supported by both the similarity in phenotype between the two groups and the fact that rigorous adjudication of cases was performed. We therefore examined genetic risk factors for this group in more detail, especially the possibility that the DILI reaction in these individuals might relate to penicillins more generally, since widely used penicillins such as amoxicillin occasionally give rise to DILI reactions (27-29) and we had positively adjudicated cases relating to both amoxicillin and other isoxazolyl penicillins available for study. GWAS analysis involving either the *B\*57:01*-negative cases only or the combined group of *HLA-B\*57:01* cases and any other penicillin (excluding amoxicillin-clavulanate) did not yield any genome-wide significant signals though this could reflect the small patient group. In view of the strong biological plausibility for a HLA association, we examined imputed HLA genotypes in detail with some additional direct typing. The *B\*57*-negative flucloxacillin cases show significantly

increased frequencies for several HLA class I alleles, but, these are seen at low frequencies in Northern Europeans (e.g. *HLA-A\*02:02* has a frequency of 0.0008 in Germans and *A\*30:01* is seen at 0.016). The pattern of HLA alleles seen in the amoxicillin DILI cases was not similar to that seen in either the *B\*57*-negative flucloxacillin cases or the other isoxazolyl penicillin cases, suggesting that there is not a general risk factor which relates T cell responses to common structural features of these penicillins, at least *in vivo*. In line with a number of previous GWAS showing HLA associations for DILI(30, 31) and other adverse drug reactions(32) where small numbers of cases comparable in number to those in the current amoxicillin DILI group were studied, we believe *B\*57:01* is not a risk factor for this form of DILI. However, interestingly, one of the HLA associations seen for the amoxicillin DILI group is with *B\*58:01* which is closely related to *B\*57:01* and strongly associated with cutaneous hypersensitivity reactions induced by allopurinol.(33) As mentioned previously, we believe that sufficient isoxazolyl penicillin cases including flucloxacillin cases were available to us to demonstrate no flucloxacillin DILI association with *B\*58:01*, again suggesting the absence of a common "penicillin DILI" risk factor. In line with this, the structure of the side chain on a penicillin is generally considered the major determinant of T cell response(34) though cross-reactivity between some T cell clones in response to flucloxacillin and other isoxazolyl penicillins as well as amoxicillin has been demonstrated.(7) Therefore, the question of overall specificity is still not completely clear. The finding that *B\*57:01* was not common among the other isoxazolyl penicillin DILI group is interesting, especially since it is considered that, despite a strong structural homology to flucloxacillin, these beta lactamase-resistant penicillins are associated with a lower rate of DILI.(1, 5) Since patient numbers were small, this finding still needs to be treated with caution and should be investigated further. Estimates for rates of DILI due to non-flucloxacillin isoxazolyl penicillins in Iceland (where flucloxacillin is not licensed) are 1 in 26000 approx. compared with 1 in 12000 reported for first time users of flucloxacillin in the UK,(2, 35) though as with the genetic study this conclusion is based on small numbers of patients and prescriptions. *In vitro* studies also generally

failed to show differences between the isoxazolyl penicillins in terms of T-cell interaction.(7) It is possible that the lower rate of DILI with these drug such as dicloxacillin and cloxacillin could partly reflect pharmacokinetic differences,(36) but, the failure to see any significant increased B\*57-carriage in the cases that were available to us indicates that replacing the fluoride in flucloxacillin with a chloride or other group may change the structure of the putative peptide-drug complex sufficiently to eliminate initial presentation by B\*57.

The association between *HLA-B\*57:01* and flucloxacillin DILI remains the most significant report of a genetic association for DILI. General implementation of *B\*57:01* testing in the clinic prior to flucloxacillin prescription appears impractical at present, but in the era of stratified medicine it seems possible that data on patient genotypes may become more generally available to prescribers. Using such data with other risk factors such as age and gender, especially in view of recent pharmacovigilance data showing increased risk above age 70,(2) could be a useful means of decreasing the incidence of flucloxacillin DILI in the future. Our current findings, which are also consistent with these age and gender effects,(2) suggest that additional genetic risk factors for flucloxacillin DILI are unlikely to be discovered in Europeans and that, despite the strong *B\*57:01/03* association, approx. 20% of DILI cases relating to flucloxacillin would not be reliably predictable on the basis of carriage of this allele or any other common HLA allele.

A limitation of the current study is that we have predicted and not directly sequenced HLA alleles. The cost of sequencing and sample availability are barriers to direct HLA typing on all cases. HLA imputation is standard practice for research grade HLA analysis and the accuracy is consistently high, especially for HLA class I alleles, as shown in detail previously.(37) However, accuracy for rare allele assignment remains limited, so we cannot exclude the possibility that the flucloxacillin DILI cases might include additional extremely rare alleles positive for V<sup>97</sup>.

In conclusion, this study has detected a novel association between *HLA-B\*57:03* and flucloxacillin DILI in addition to the *HLA-B\*57:01* association detected previously and confirmed that

approximately 20% of the flucloxacillin DILI cases studied do not show any *B\*57* association. The apparently lower risk for development of DILI with isoxazolyl penicillins other than flucloxacillin reported previously by others (1, 5) is consistent with our failure to see an association with *B\*57:01* and other *B\*57* alleles for DILI due to these drugs. Other alleles which code for a HLA protein positive for *V<sup>97</sup>* could be risk factors for flucloxacillin DILI in non-Europeans. It also remains possible that rare variants in HLA and other genes not covered by the GWAS chip or imputation may contribute to the risk of DILI with flucloxacillin and the other penicillins, but, population effect sizes from these seem unlikely to be large though for individuals the impact of rare variants may be higher.

## **Materials and methods**

### **Enrollment details**

All participants provided written informed consent and each study had been approved by the appropriate national or institutional ethical review boards. In the United Kingdom, ethical approval was via the Leeds East Research Ethics committee (approval reference 04/Q1206/91).

### **Causality assessment**

The iDILIC cases were evaluated by application of the Council for International Organizations of Medical Science (CIOMS) scale, also called the Roussel Uclaf Causality Assessment Method (RUCAM)(11) and by expert review by a panel of three hepatologists. The pattern of liver injury was classified according to the International Consensus Meeting Criteria.(38) Only cases having at least a possible causality (score  $\geq 3$ ) were included in the study. For DILIN cases, causality assessment was by expert consensus as previously described.(12)

### **DNA preparation and Genome-wide genotyping of Phase II cases**

For iDILIC cases, DNA was prepared as described previously.(6) DILIN DNA was extracted from lymphocytes and stored at the NIDDK biosample repository at Rutgers University, Piscataway, NJ.



Genome-wide genotyping of the phase II cases was performed by the Broad Institute, Boston on the Illumina Infinium HumanCoreExome BeadChip. iDILIC and DILIN cases were genotyped separately. Full details of the genotyping and quality control processes have been described previously.(12)

### **SNP and HLA Imputation**

SNP imputation was performed as described recently.(12) Four digit HLA alleles and amino acid changes were also inferred using SNP2HLA using reference data collected by the Type 1 Diabetes Genetics Consortium (T1DGC).(17)

### **Direct HLA genotyping**

Samples from phase I were genotyped for *HLA-B\*57* using *HLA-B\*57:01* genotyping with the Dynal AllSet Gold SSP B17 high resolution kit (Invitrogen). Selected *HLA-B\*57:01* negative flucloxacillin DILI cases (n=33) were genotyped for all HLA-B alleles using an AllSet<sup>TM</sup> Gold sequence-specific primer (SSP) HLA-B Locus High Res Kit (Invitrogen) according to the manufacturer's instructions. Following PCR, products were applied to 2% agarose gels containing ethidium bromide (0.5 µg/ml) and electrophoresis was performed in 1 X TBE buffer. Positive lane amplifications were identified. HLA-B alleles were assigned by analysis with UniMatch® PLUS 6.0 SSP software (Invitrogen).

### **Statistical analysis**

All genetic analyses were performed as described previously.(12) For HLA analysis, we tested for association of carriage of each HLA allele/AA/HLA haplotype. MHC significance was defined using the Bonferroni correction threshold of  $P < 0.00025$  (0.05/200 accounting for 200 observed HLA alleles). Conditional analyses in the MHC region were undertaken and the genotypes at the conditioning SNP(s) were included as covariates under an additive model. For clinical variable comparisons, Fisher's exact test was used for categorical covariates such as gender and pattern of

liver damage. Univariate and multivariate analysis for studies combining genetic and clinical risk factors was performed by STATA15.

## **Study Highlights**

### **WHAT IS THE CURRENT KNOWLEDGE ON THE TOPIC?**

Flucloxacillin is known to be a common cause of drug-induced liver injury. Previously, carriage of *HLA-B\*57:01* has been demonstrated to be a strong genetic risk factor for this adverse drug reaction in a small genome-wide association study.

### **WHAT QUESTION DID THIS STUDY ADDRESS?**

To extend the previously reported *HLA-B\*57:01* association in an enlarged cohort, identify additional genetic factors and assess the relevance of risk factors for flucloxacillin-related liver injury to liver injury with other penicillins, both other isoxazolyl penicillins such as dicloxacillin and also amoxicillin.

### **WHAT DOES THIS STUDY ADD TO OUR KNOWLEDGE?**

We found that a related allele *HLA-B\*57:03* was also a risk factor and that the amino acid valine at position 97 which is common to both B\*57:01 and B\*57:03 HLA proteins was the key risk factor at the amino acid level. We detected no *HLA-B\*57* association with liver injury due to other isoxazolyl penicillins or amoxicillin and no significant non-HLA signals for any penicillin-related liver injury.

### **HOW THIS MIGHT CHANGE CLINICAL PHARMACOLOGY OR TRANSLATIONAL SCIENCE?**

Prior knowledge of *HLA-B\*57* genotype may affect the decision to prescribe flucloxacillin especially in patients aged 70 years and older who have an increased risk of liver injury with this drug.

**Acknowledgements** We are extremely grateful to Arthur Holden of iSAEC for his continuing support. Collaboration and helpful discussions with Paul Watkins and colleagues in the DILIN network (see <http://dilin.org/publications/>) are acknowledged with thanks. We are grateful to Julia Patch, Julian Leathart and Julian Arbuckle for technical help, study management and assistance with recruitment on the iDILIC study, to Rob Delahay for assisting with the illustration of HLA-B\*57:01 and to Daniele Cusi (Hypergenes), Patrik K. Magnusson (Swedish Twin Registry) and Javier Martin (Spanish DNA bank) for provision of control data. Other contributors to case recruitment-UK: J. Henderson, R. Wake (Newcastle University); C Davies, S. Henry (Nottingham Digestive Diseases Centre); K. Hawkins, A. Hanson, J. Evely (University of Liverpool), S. Cleary (Dundee). H. Hussaini (Truro), W. Griffiths (Addenbrooks Hospital, Cambridge), J. Collier (John Radcliffe Infirmary, Oxford), A. Brind (North Staffordshire), N. Fisher (Dudley), J. Shearman (South Warwick), A. Grant (Leicester Royal Infirmary), A. Austin (Derby), F. Gordon (Bristol), M. Cramp (Plymouth), S. Saksena (North Durham), H J McMurtry (Chorley), S. Verma (Brighton), H. Mitchison (Sunderland), A. M. Elsharkawy (Birmingham), H. Dallal (Middlesbrough), C. McDonald (Carlisle), J. Metcalf (Hartlepool); Sweden: Eric Eliasson (Karolinska Institutet, Stockholm); Ulrica Ramqvist, Elisabet Stjernberg, Sofie Collin, Eva Prado, Agnes Wadelius, Martha Wadelius, Agnes Kataja Knight and Hugo Kohnke at SWEDEGENE (Department of Medical Sciences, Uppsala University); Netherlands: Renate Udo and Marie L. De Bruin (Utrecht Institute for Pharmaceutical Sciences, Utrecht University), L.C. Baak (Onze Lieve Vrouwe Gasthuis, Amsterdam), J.T. Brouwer (Reinier de Graaf Gasthuis, Delft), J.P.H. Drenth (Radboud University Medical Center, Nijmegen), M. Klemt-Kropp (Medisch Centrum Alkmaar), T.C.M.A. Schreuder (Slingeland Ziekenhuis, Doetichem), L.A. van der Waaij (Martini Ziekenhuis, Groningen), F.H.J. Wolfhagen (Tweesteden Ziekenhuis, Tilburg). Author contributions: Study concept and design: GPA, MRN and AKD; case recruitment and data acquisition: GPA, EB, PH, MW, AHM-Z, JHM, JFD, MIL, MM, MP and AKD; case adjudication:

GPA, EB, RJA; sample preparation and laboratory analysis: SAC, TC and MA; data analysis and interpretation: PN, YS, JIG, GPA, MRN and AKD; writing the manuscript: PN, GPA and AKD

## References

1. Devereaux, B.M., Crawford, D.H., Purcell, P., Powell, L.W. & Roeser, H.P. Flucloxacillin associated cholestatic hepatitis. An Australian and Swedish epidemic? *Eur J Clin Pharmacol* **49**, 81-5 (1995).
2. Wing, K. *et al.* Quantification of the risk of liver injury associated with flucloxacillin: a UK population-based cohort study. *J Antimicrob Chemother* **72**, 2636-46 (2017).
3. Bjornsson, E., Jerlstad, P., Bergqvist, A. & Olsson, R. Fulminant drug-induced hepatic failure leading to death or liver transplantation in Sweden. *Scand J Gastroenterol* **40**, 1095-101 (2005).
4. Lakehal, F. *et al.* Indirect cytotoxicity of flucloxacillin toward human biliary epithelium via metabolite formation in hepatocytes. *Chem Res Toxicol* **14**, 694-701 (2001).
5. Olsson, R., Wiholm, B.-E., Sand, C., Zettergren, L., Hultcrantz, R. & Myrhed, M. Liver damage from flucloxacillin, cloxacillin and dicloxacillin. *J Hepatol* **15**, 154-61 (1992).
6. Daly, A.K. *et al.* HLA-B\*5701 genotype is a major determinant of drug-induced liver injury due to flucloxacillin. *Nat Genet* **41**, 816-9 (2009).
7. Monshi, M.M. *et al.* Human leukocyte antigen (HLA)-B\*57:01-restricted activation of drug-specific T cells provides the immunological basis for flucloxacillin-induced liver injury. *Hepatology* **57**, 727-39 (2013).
8. Wuillemin, N., Adam, J., Fontana, S., Krahenbuhl, S., Pichler, W.J. & Yerly, D. HLA haplotype determines hapten or p-i T cell reactivity to flucloxacillin. *J Immunol* **190**, 4956-64 (2013).
9. Illing, P.T. *et al.* Immune self-reactivity triggered by drug-modified HLA-peptide repertoire. *Nature* **486**, 554-8 (2012).
10. Urban, T.J. *et al.* Limited contribution of common genetic variants to risk for liver injury due to a variety of drugs. *Pharmacogenet Genomics* **22**, 784-95 (2012).
11. Aithal, G.P. *et al.* Case Definition and Phenotype Standardization in Drug-Induced Liver Injury. *Clin Pharmacol Ther* **89**, 806-15 (2011).
12. Nicoletti, P. *et al.* Association of Liver Injury From Specific Drugs, or Groups of Drugs, With Polymorphisms in HLA and Other Genes in a Genome-Wide Association Study. *Gastroenterology* **152**, 1078-89 (2017).
13. *The Wellcome Trust Case Control Consortium*. <<https://www.wtccc.org.uk/>>. Accessed November 1st 2018.
14. Novembre, J. *et al.* Genes mirror geography within Europe. *Nature* **456**, 98-101 (2008).
15. Shen, Y. *et al.* Genome-wide association study of serious blistering skin rash caused by drugs. *Pharmacogenomics J* **12**, 96-104 (2012).
16. *The Swedish Twin Registry*. <[https://ki.se/en/research/the-swedish-twin-registry?\\_ga=2.140851515.829713908.1541074484-1049565503.1541074484](https://ki.se/en/research/the-swedish-twin-registry?_ga=2.140851515.829713908.1541074484-1049565503.1541074484)>. Accessed November 1st 2018.
17. Jia, X. *et al.* Imputing amino acid polymorphisms in human leukocyte antigens. *PLoS One* **8**, e64683 (2013).
18. *HLA Epitope Registry*. <<http://www.epregistry.com.br/>>. Accessed 1st November 2018.
19. Petros, Z., Kishikawa, J., Makonnen, E., Yimer, G., Habtewold, A. & Aklillu, E. HLA-B\*57 Allele Is Associated with Concomitant Anti-tuberculosis and Antiretroviral Drugs Induced Liver Toxicity in Ethiopians. *Front Pharmacol* **8**, 90 (2017).
20. *Immuno Polymorphism Database*. Hinxton, Cambridgeshire, UK: *The European Bioinformatics Institute (EBI) (Release 2.0.0 June 2018)*. <<https://www.ebi.ac.uk/ipd/imgt/hla/>>. Accessed 1st November 2018.
21. McCormack, M. *et al.* HLA-A\*3101 and carbamazepine-induced hypersensitivity reactions in Europeans. *N Engl J Med* **364**, 1134-43 (2011).

22. Norcross, M.A. *et al.* Abacavir induces loading of novel self-peptides into HLA-B\*57: 01: an autoimmune model for HLA-associated drug hypersensitivity. *AIDS* **26**, F21-9 (2012).
23. Xu, C.F. *et al.* HLA-B\*57:01 Confers Susceptibility to Pazopanib-Associated Liver Injury in Patients with Cancer. *Clin Cancer Res* **22**, 1371-7 (2016).
24. Donaldson, P.T. *et al.* Human leucocyte antigen class II genotype in susceptibility and resistance to co-amoxiclav-induced liver injury. *J Hepatol* **53**, 1049-53 (2010).
25. Andrade, R.J. *et al.* HLA class II genotype influences the type of liver injury in drug-induced idiosyncratic liver disease. *Hepatology* **39**, 1603-12 (2004).
26. de Boer, Y.S. *et al.* Genome-wide association study identifies variants associated with autoimmune hepatitis type 1. *Gastroenterology* **147**, 443-52.e5 (2014).
27. Davies, M.H., Harrison, R.F., Elias, E. & Hubscher, S.G. Antibiotic-associated acute vanishing bile duct syndrome: a pattern associated with severe, prolonged, intrahepatic cholestasis. *J Hepatol* **20**, 112-6 (1994).
28. Schwarze, C., Schmitz, V., Fischer, H.P., Sauerbruch, T. & Spengler, U. Vanishing bile duct syndrome associated with elevated pancreatic enzymes after short-term administration of amoxicillin. *Eur J Gastroenterol Hepatol* **14**, 1275-7 (2002).
29. Bjornsson, E.S. & Hoofnagle, J.H. Categorization of drugs implicated in causing liver injury: Critical assessment based on published case reports. *Hepatology* **63**, 590-603 (2016).
30. Nicoletti, P. *et al.* HLA-DRB1\*16: 01-DQB1\*05: 02 is a novel genetic risk factor for flupirtine-induced liver injury. *Pharmacogenet Genomics* **26**, 218-24 (2016).
31. Kowalec, K. *et al.* Common variation near IRF6 is associated with IFN-beta-induced liver injury in multiple sclerosis. *Nat Genet* **50**, 1081-5 (2018).
32. Cheung, C.L. *et al.* HLA-B\*38:02:01 predicts carbimazole/methimazole-induced agranulocytosis. *Clin Pharmacol Ther* **99**, 555-61 (2016).
33. Hung, S.I. *et al.* HLA-B\*5801 allele as a genetic marker for severe cutaneous adverse reactions caused by allopurinol. *Proc Natl Acad Sci U S A* **102**, 4134-9 (2005).
34. Meng, X. *et al.* Definition of the Nature and Hapten Threshold of the beta-Lactam Antigen Required for T Cell Activation In Vitro and in Patients. *J Immunol* **198**, 4217-27 (2017).
35. Bjornsson, E.S. Drug-induced liver injury due to antibiotics. *Scand J Gastroenterol* **52**, 617-23 (2017).
36. Paton, D.M. Comparative bioavailability and half-lives of cloxacillin and flucloxacillin. *Int J Clin Pharmacol Res* **6**, 347-9 (1986).
37. Zheng, X. *et al.* HIBAG--HLA genotype imputation with attribute bagging. *Pharmacogenomics J* **14**, 192-200 (2014).
38. Benichou, C., Danan, G. & Flahault, A. Causality assessment of adverse reactions to drugs--II. An original model for validation of drug causality assessment methods: case reports with positive rechallenge. *J Clin Epidemiol* **46**, 1331-6 (1993).

**Table 1 Summary of clinical data for the flucloxacillin DILI cases**

Number of cases	197
Gender (F/M)	133/64 (68% female)
Mean Age at onset (years) (with standard deviation)	62 ± 13
Mean Time to onset (days) (with standard deviation)	24 ± 18
Total days on drug (with standard deviation)	10 ± 6
<u>Pattern of liver injury</u>	
Cholestatic	74 (38%)
Hepatocellular	39 (20%)
Mixed	84 (43%)
<u>Scoring (CIOMS/RUCAM)</u>	
3-5 (possible)	22 (11%)
6-8 (probable)	90 (46%)
>8 (highly probable)	85 (43%)



**Table 2 The most significant HLA alleles for flucloxacillin DILI**

<b>ALLELE</b>	<b>OR</b>	<b>95% CI</b>	<b>P</b>	<b>Cond OR</b>	<b>Cond P</b>	<b>AF cases</b>	<b>AF controls</b>
<i>B*57:01</i>	36.62	26.14-51.29	2.6x10 <sup>-97</sup>	-	-	0.42	0.04
<i>C*06:02</i>	10.11	7.88-12.97	4.3x10 <sup>-74</sup>	1.32	0.23	0.45	0.09
<i>DQB1*03:03</i>	10.18	7.77-13.34	1.1x10 <sup>-63</sup>	0.96	0.84	0.31	0.05
<i>DRB1*07:01</i>	4.02	3.23-5.02	3.8x10 <sup>-35</sup>	1.01	0.94	0.38	0.13
<i>DQAI*02:01</i>	4.02	3.22-5.01	4.5x10 <sup>-35</sup>	1.01	0.95	0.38	0.13
<i>A*01:01</i>	1.86	1.5-2.31	1.8x10 <sup>-8</sup>	0.95	0.69	0.30	0.18
<i>DQAI*03:01</i>	0.42	0.3-0.58	3.0x10 <sup>-7</sup>	0.61	0.009	0.10	0.21
<i>C*07:02</i>	0.33	0.22-0.51	4.2x10 <sup>-7</sup>	0.63	0.04	0.06	0.16
<i>B*07:02</i>	0.32	0.2-0.5	5.7x10 <sup>-7</sup>	0.60	0.04	0.05	0.15
<i>DQB1*03:01</i>	0.51	0.36-0.71	7.3x10 <sup>-5</sup>	0.76	0.14	0.10	0.17
<i>DQB1*06:02</i>	0.46	0.31-0.69	0.0001	0.69	0.09	0.07	0.14
<i>DRB1*04:01</i>	0.46	0.3-0.7	0.0003	0.67	0.09	0.06	0.12
<i>DQB1*03:02</i>	0.47	0.3-0.73	0.0007	0.68	0.11	0.05	0.11
<i>B*57:03</i>	19.77	3.37-116.1	0.001	79.21	0.000001	0.005	0.0003

OR = Odds Ratio; 95% CI = 95% Confidence Interval of the odds ratio; P = logistic p-value; Cond OR = Odds Ratio from the reciprocal conditional analyses on rs2395029 and *HLA-B\*57:01*; Cond P = logistic p-value from the conditional analysis; AF cases = allele frequency in cases; AF controls = allele frequency in controls. The p value required for significance was P<0.00025.

**Table 3 Effect size of the imputed amino acid residues located at position 97 on HLA-B gene product**

<b>Residues</b>	<b>OR</b>	<b>95% CI</b>	<b>P</b>	<b>AF cases</b>	<b>AF controls</b>
Valine (V)	38.10	27.07-53.62	$9.7 \times 10^{-97}$	0.43	0.04
Arginine (R)	0.43	0.34-0.53	$5.13 \times 10^{-14}$	0.28	0.48
Serine (S)	0.53	0.41-0.68	$9.82 \times 10^{-7}$	0.18	0.30
Asparagine (N)	0.31	0.14-0.69	0.004	0.02	0.05
Threonine (T)	0.64	0.42-0.96	0.03	0.06	0.10
Tryptophan (W)	0.87	0.5-1.54	0.64	0.03	0.04

Residue = name of the associated amino acid at position 97; OR = Odds Ratio; 95% CI = 95% Confidence Interval of the odds ratio; P = logistic p-value; AF cases = allele frequency in cases ; AF controls = allele frequency in controls. The significance threshold was  $p < 0.00025$ .

**Table 4 HLA allele frequency data for the *HLA-B\*57:01* and *HLA-B\*57:03* negative flucloxacillin DILI cases**

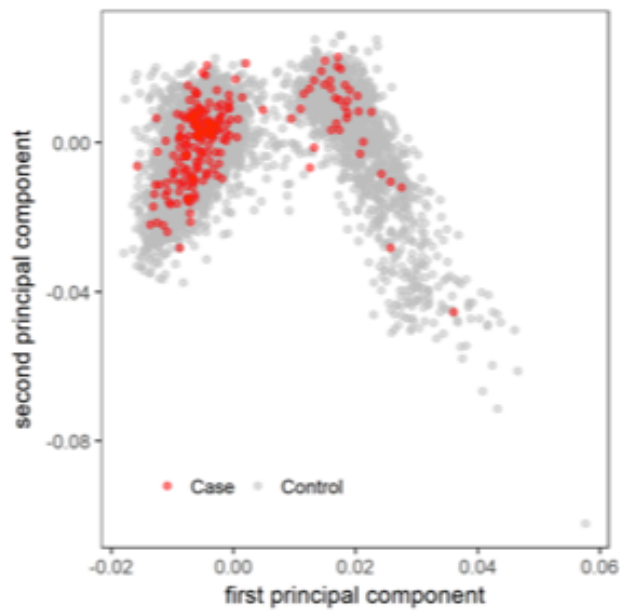
<b>ALLELE</b>	<b>OR</b>	<b>95% CI</b>	<b>P</b>	<b>AF cases</b>	<b>AF controls</b>
<i>HLA-A*02:02</i>	15.24	1.89-123.1	0.01	0.01	0.001
<i>HLA-A*30:01</i>	4.447	1.38-14.34	0.01	0.04	0.01
<i>HLA-B*13:02</i>	3.496	1.22-10.06	0.02	0.06	0.02
<i>HLA-DQB1*03:02</i>	0.1228	0.02-0.89	0.04	0.01	0.11
<i>HLA-B*07:02</i>	0.347	0.13-0.95	0.04	0.06	0.15

OR = Odds Ratio; 95% CI = 95% Confidence Interval of the odds ratio; P = logistic p-value; AF cases = allele frequency in cases; AF controls = allele frequency in controls. The p value required for significance was P<0.00025.

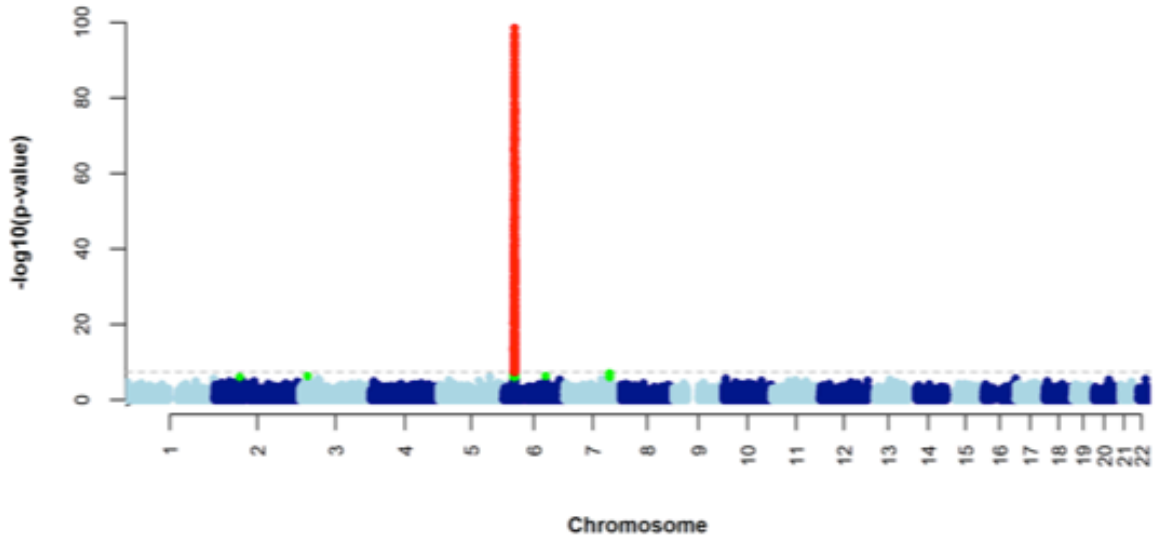
**Table 5 HLA associations across different penicillin classes**

<b>Group</b>	<b>HLA Allele</b>	<b>OR</b>	<b>95% CI</b>	<b>P</b>	<b>AFcases</b>	<b>AF controls</b>
Other isoxazolyl penicillins (n=6)	<i>C*07:04</i>	12.97	1.58-134.6	0.001	0.17	0.02
	<i>DQB1*06:09</i>	14.57	2.18-12.16	0.02	0.08	0.01
Amoxicillin (n=15)	<i>B*58:01</i>	20.68	4.31-99.12	0.0002	0.07	0.01
	<i>DPB1*01:01</i>	4.84	2.07-11.32	0.0003	0.233	0.054
	<i>A*01:01</i>	3.25	1.59-6.62	0.001	0.433	0.159
	<i>C*03:02</i>	30.09	3.55-255.2	0.002	0.033	0.002

OR = Odds Ratio; 95% CI = 95% Confidence Interval of the odds ratio; P = logistic p-value; AF cases = allele frequency in cases ; AF controls = allele frequency in controls. The p value required for significance was P<0.00025.



**Figure 1 Scatterplot representing the case control distribution of study cohort.** The axes represent the first two principal components where the red dots are the cases and the gray dots the controls. The controls cluster in two groups representing the UK and Swedish major control populations.



**Figure 2 Manhattan plot from the GWAS analysis of 197 flucloxacillin cases and 6835 controls.** Manhattan plot displays the negative log of the significance for each tested variant ranked by chromosomes and position; SNPs in green have a significance level less than  $5 \times 10^{-6}$  and red have a significance level less than  $5 \times 10^{-8}$  which was taken as the threshold for genome-wide significance.

### **Supplementary Information Titles**

Supplied as single PDF file with the following content:

iDILIC investigators

Supplementary Tables 1-8

Supplementary Figures 1-5



**Supplementary Material for "Drug-induced injury due to flucloxacillin: relevance of multiple HLA alleles"**

\*Paola Nicoletti, \*Guruprasad P. Aithal, Mohammad Alshabeeb, Thomas C. Chamberlain, Sally Coulthard, Jane I. Grove, Raul Andrade, Einar Bjornsson, John F. Dillon, Par Hallberg, M. Isabel Lucena, Anke H. Maitland-van der Zee, Jennifer H. Martin, Mariam Molokhia, Munir Pirmohamed, Mia Wadelius, Aris Floratos, Yufeng Shen, Matthew R. Nelson and Ann K. Daly for the International Drug-induced Liver Injury consortium (iDILIC)

**Table of contents**

iDILIC investigators.....3

Supplementary Tables 1-8.....4

Supplementary Figures 1-5.....13

### **iDILIC investigators (in alphabetical order)**

Guruprasad P. Aithal, National Institute for Health Research (NIHR) Nottingham Digestive Diseases Biomedical Research Unit, Nottingham University Hospital NHS Trust and University of Nottingham, Nottingham, UK; Raul J. Andrade, IBIMA Hospital Universitario Virgen de la Victoria, Universidad de Málaga, Málaga, Spain and CIBERehd, Madrid, Spain; Fernando Bessone, Universidad Nacional de Rosario, Rosario, Argentina; Einar Bjornsson, Division of Gastroenterology and Hepatology, Department of Internal Medicine, The National University Hospital of Iceland, Reykjavik, Iceland; Ingolf Cascorbi, Institute for Experimental and Clinical Pharmacology, University Hospital Schleswig-Holstein, Kiel, Germany; Ann K. Daly, Institute of Cellular Medicine, Newcastle University, Newcastle upon Tyne, UK; John F. Dillon, Ninewells Hospital and Medical School, Dundee, UK; Christopher P. Day, Institute of Cellular Medicine, Newcastle University, Newcastle upon Tyne, UK; Par Hallberg, Uppsala University, Uppsala, Sweden; Nelia Hernández, Universidad de la Republica, Montevideo, Uruguay; Luisa Ibanez, Hospital Universitari Vall d'Hebron, Barcelona, Spain; Gerd A. Kullak-Ublick, University of Zurich, Zurich, Switzerland; Tarja Laitinen, Helsinki University Central Hospital, Helsinki, Finland; Dominique Larrey, Hôpital Saint Eloi, Montpellier, France; M. Isabel Lucena, IBIMA Hospital Universitario Virgen de la Victoria, Universidad de Málaga, Málaga, Spain and CIBERehd, Madrid, Spain; Anke Maitland-van der Zee, AMC, Amsterdam, Netherlands; Jennifer H. Martin, University of Newcastle, Newcastle, NSW, Australia; Dick Menzies, MUHC and McGill University, Montreal Chest Institute, Montreal, Canada; Mariam Molokhia, King's College, London, UK; Munir Pirmohamed, Institute of Translational Medicine, University of Liverpool, Liverpool, UK; Shengying Qin, Shanghai Jiao Tong University, Shanghai, China; Mia Wadelius, Uppsala University, Uppsala, Sweden

**Table S1****Clinical information for other non-flucloxacillin DILI cases**

<b>Clinical information</b>	<b>Amoxicillin</b>	<b>Isoxazolyl penicillins</b>
Total number of cases	15	6
Gender (F/M)	9/6 (57%)	3/3 (50%)
Mean Age at onset (years) (with standard deviation)	60 ±19	60 ±25
Mean Time to onset (days) (with standard deviation)	25 ±22	18 ±10
Total days on drug (with standard deviation)	13 ±14	17± 12
<b>Pattern of liver injury</b>		
Cholestatic	9	2
Hepatocellular	5	2
Mixed	1	1
Unknown	-	1
<b>CIOMS/RUCAM scoring</b>		
3-5 (possible)	2	-
6-8 (probable)	10	4
>8 (highly probable)	3	2

**Table S2**

**Haplotype association results for flucloxacillin DILI using the six HLA alleles belonging to B57 haplotype**

	<b>FREQ in cases</b>	<b>FREQ in Controls</b>	<b>OR</b>	<b>P</b>
<b>Positive B*57:01 haplotypes</b>				
PPPPA	0.15	0.013	18.7	6.19E-59
PPPPP	0.14	0.013	18.6	9.12E-53
PPAAAA	0.06	0.005	15.4	7.45E-24
PPAAAP	0.04	0.004	16.1	5.55E-19
PAPPPP	0.01	0.0003	61.5	7.27E-08
PPAPPA	0.009	0.0007	36.8	4.66E-06
PAPPPA	0.004	0.0006	7.65	0.03
PPAPPP	0.002	0.0003	17.4	0.07
PAAAAP	0.0019	0.0006	4.34	0.3
PAAAAA	0.0009	0.0006	1.53	0.8
PAAPPA	0	0.0003	4.37E-190	1.0
PAAPPP	0	0.0001	3.17E-129	1
PPAPAP	0.002	0	3.45E+11	1
PPPAAP	0	3.19E-05	2.07E-52	1
PPPAAA	0	3.62E-06	1.94E-157	1
PAPPAA	0	0	0	0
PAAPAA	0	0	0	0
<b>Negative B*57:01 haplotypes</b>				
AAAAAA	0.40	0.6739	0.278	2.39E-27
AAAAAP	0.09	0.1401	0.578	0.003
AAAPPA	0.04	0.06	0.482	0.01
AAAPPP	0.001	0.006	0.00224	0.06
AAPAAA	0.003	0.01326	0.147	0.1
AAPPPA	0.0030	0.0083	0.267	0.2
APPPPP	0.0013	0.0005	5.15	0.4
AAPPPP	0.0002	0.0016	0.003	0.4
AAPAAP	0.0001	0.0011	4.92E-06	0.5
APAPPP	0.0018	0.0031	0.296	0.5
APAAAA	0.0150	0.0196	0.745	0.5
APAAAP	0.0073	0.0088	0.776	0.7

AAAPAA	0.0001	0.0003	0.24	0.8
APPPA	0.0003	0.0005	0.335	0.8
AAAPAP	4.95E-05	0.0002	0.237	0.9
APAPPA	0.02	0.02	0.96	0.9
AAAAPA	0	0.0001		1.0
APPAPP	0	5.96E-05		1
APPAAP	0	2.97E-05		1
AAPAPA	0	1.20E-05		1
APAPAA	0	2.91E-05		1
APPAAA	0	2.72E-05		1
AAPPAA	0	0	0	0

Odds ratios (OR) and p-values (P) are presented after correcting for population stratification for each combination of alleles belonging to B57 haplotype. The six letters in the first column (haplotype) reflects in order of HLA-B\*57:01|HLA-C\*06:02|HLA-DQB1\*03:03|HLA-DRB1\*07:01|HLA-DQA1\*02:01|HLA-A\*01:01 status. The risk alleles status is represented by P = present or A = absent.

**Table S3****HLA alleles which share valine at position 97**

<b>ALL</b>	<b>MAF</b>	<b>Dataset</b>
B*57:01	0.04	USA NMDP European Caucasian
B*57:03	0.0007	USA NMDP European Caucasian
B*57:10	0.0003	Poland DKMS
B*57:02	0.0002	USA NMDP European Caucasian
B*57:07	0.00001	USA NMDP European Caucasian
B*57:04	0.000003	USA NMDP European Caucasian
B*40:30	0.0000008	USA NMDP European Caucasian
B*40:34	0.0000008	USA NMDP European Caucasian
B*57:14	0.0000008	USA NMDP European Caucasian
B*57:16	0.0000008	
B*57:08	0.0000004	USA NMDP European Caucasian
B*57:15	0.0000004	USA NMDP European Caucasian
A*26:32	0	
A*30:28	0	
B*55:14	0	
B*57:06	0	USA NMDP European Caucasian
B*57:09	0	
B*57:12	0	
B*57:13	0	
B*57:17	0	
B*57:18	0	
B*57:19	0	
B*58:14	0	

**Table S4. Case-control analysis on effect of age, gender and B\*57 genotype on risk of flucloxacillin DILI**

<b>Univariate analysis</b>			
<b>FACTOR</b>	<b>OR</b>	<b>95% CI</b>	<b>P</b>
age	1.09	1.06-1.10	$7.1 \times 10^{-20}$
>70 years	6.27	3.75-10.50	$2.7 \times 10^{-12}$
Gender (Female)	1.69	1.14-2.36	$4.2 \times 10^{-3}$
HLA B*57 allele	43.09	24.19-76.74	$2.1 \times 10^{-37}$
<b>Multivariate analysis</b>			
<b>FACTOR</b>	<b>OR</b>	<b>95% CI</b>	<b>P</b>
Gender (Female)	1.66	0.92-3.00	$9.1 \times 10^{-2}$
>70 years	6.70	3.07-15.02	$2.2 \times 10^{-6}$
HLA B*57 allele	42.45	23.04-78.14	$2.5 \times 10^{-33}$

**Age is analyzed as a continuous variable in the univariate analysis**



**Table S5. Comparison of selected parameters between HLA-B\*57-positive and negative cases relating to flucloxacillin**

	B*57 positive (n=163)	B*57 negative (n=34)
Gender (F/M)	114F/49M (70% female)	19F/15M (56% female)
Age at onset (years) (mean $\pm$ SD)	63 $\pm$ 13	61 $\pm$ 10
Time to onset (days) (mean $\pm$ SD)	23 $\pm$ 13	30 $\pm$ 30
Total days on drug (mean $\pm$ SD)	10 $\pm$ 5	9 $\pm$ 7
<u>Pattern of liver injury</u>		
Cholestatic	63	11
Hepatocellular	31	8
Mixed	69	17
<u>Scoring (CIOMS/RUCAM)</u>		
3-5 (possible)	18	4
6-8 (probable)	73	17
>8 (highly probable)	72	13

There were no statistically significant differences between the two groups for the parameters listed

**Table S6. The most associated amino acid deletions in the HLA-B\*57:01-negative flucloxacillin case analysis**

<b>POSITION OF DELETED RESIDUE IN HLA-A</b>	<b>OR</b>	<b>95%CI</b>	<b>P</b>
276	17.85	2.16-147.4	0.007
282	17.85	2.16-147.4	0.007
283	17.85	2.16-147.4	0.007
288	17.85	2.16-147.4	0.007
294	17.85	2.16-147.4	0.007
297	17.85	2.16-147.4	0.007
298	17.85	2.16-147.4	0.007
299	17.85	2.16-147.4	0.007
307	17.85	2.16-147.4	0.007
310	17.85	2.16-147.4	0.007
311	17.85	2.16-147.4	0.007
321	17.14	2.07-141.6	0.008
334	17.14	2.07-141.6	0.008
-11	16.55	2.02-135.4	0.009
-15	16.55	2.02-135.4	0.009
-20	16.55	2.02-135.4	0.009
-22	16.55	2.02-135.4	0.009

Residue in HLA-A= Amino acid position along the HLA-A protein product; OR = Odds Ratio; 95% CI = 95% Confidence Interval of the odds ratio; P = logistic p-value

**Table S7 Effect size of the HLA-B\*57:01 association across different drug groups**

<b>DRUG GROUP</b>	<b>OR</b>	<b>95%CI</b>	<b>PV</b>	<b>AF<sub>cases</sub></b>
Flucloxacillin	36.62	26.14-51.29	$2.6 \times 10^{-97}$	0.48
Amoxicillin	2.80	0.84-9.42	0.09	0.1
Other isoxazolyl penicillins	3.09	0.37-25.78	0.28	0.1

OR = Odds Ratio; 95% CI = 95% Confidence Interval of the odds ratio; P = logistic p-value; AF cases = allele frequency in cases.

**Table S8**

**The most associated amino acids in the analysis on DILI due to penicillins other than flucloxacillin**

<b>DRUG GROUP</b>	<b>LOCUS &amp; POSITION</b>	<b>AMINO ACID</b>	<b>OR</b>	<b>95% CI</b>	<b>P</b>
Isoxazolyl penicillins (n=6)	HLA-C (95)	Phenylalanine	12.97	2.69-62.64	0.001
	HLA-C (156)	Aspartic acid	12.97	2.69-62.64	0.001
Amoxicillin (n=15)	HLA-DPB1 (194)	Glutamine	4.86	2.08-11.35	0.0003
		Phenylalanine /Methionine	3.55	1.7-7.41	0.0008
	HLA-B(67)	Lysine	3.23	1.59-6.57	0.0012
	HLA-A(67)	Methionine	3.23	1.59-6.57	0.0012

OR = Odds Ratio; 95% CI = 95% Confidence Interval of the odds ratio; P = logistic p-value

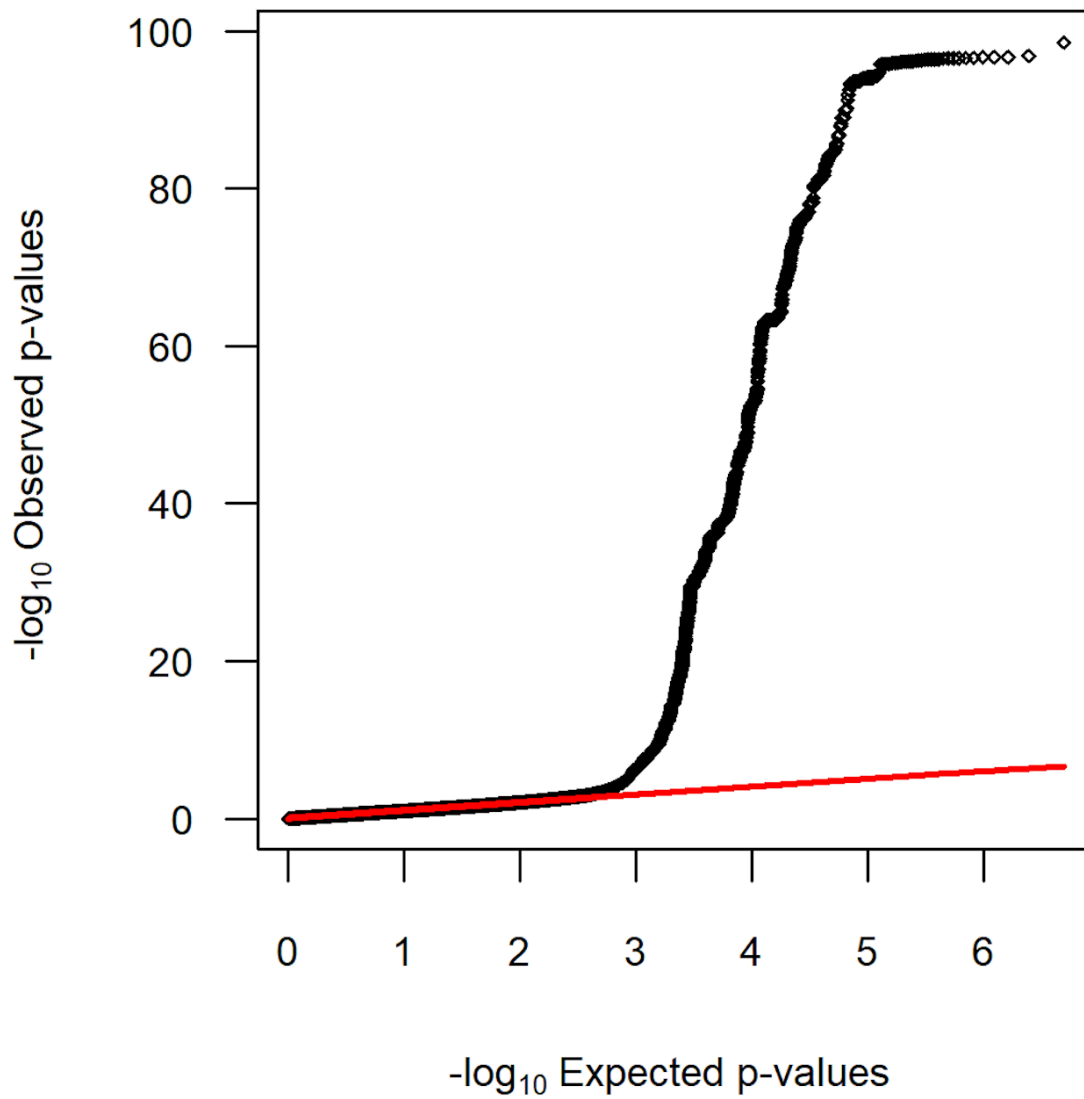
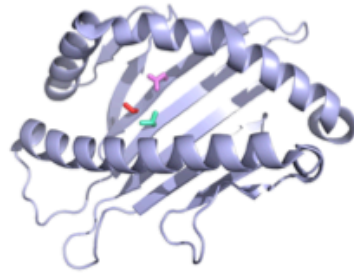


Figure S1. QQ plot for the main Flucloxacillin GWAS.



	100	110	120
B*07:02	GSHTLQSMYG	CDVGPDGRL	RGHDQYAYDG
B*57:01	GSHIIQVMYG	CDVGPDGRL	RGHDQ SAYDG
B*57:02	GSHIIQVMYG	CDVGPDGRL	RGHNQYAYDG
B*57:03	GSHIIQVMYG	CDVGPDGRL	RGHNQYAYDG

Figure S2. Molecular structure of the HLA-B\*57:01 antigen binding cleft with key amino acids contributing to abacavir binding marked. The structure shown is an illustration of HLA-B\*57:01 visualised in PyMOL (Version 1.2b2; <http://www.pymol.org>) based on the Illing structure (3VRI)(Illing, P.T. *et al.* Immune self-reactivity triggered by drug-modified HLA-peptide repertoire. *Nature* **486**, 554-8 (2012)) available from Protein Data Bank. Residue 97 (valine) is highlighted in green, 114 (aspartic acid) in pink and 116 (serine) in red. Alignment of amino acids in the region 90 to 120 is shown for B\*57:01, B\*57:02 and B\*57:03 with B\*07:01, the most common B allele in European populations as reference. Key amino acid differences are highlighted.

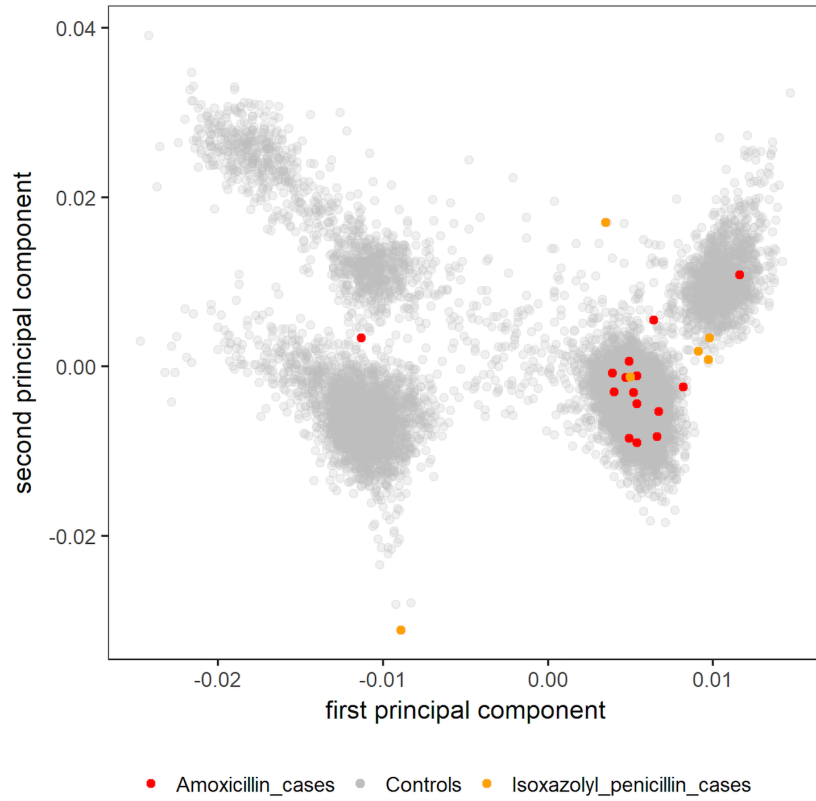


Figure S3. Scatterplots representing the first two principal components of the European study cohort. Amoxicillin cases are highlighted in red and the isoxazoly penicillin cases in orange while controls are highlighted in gray.

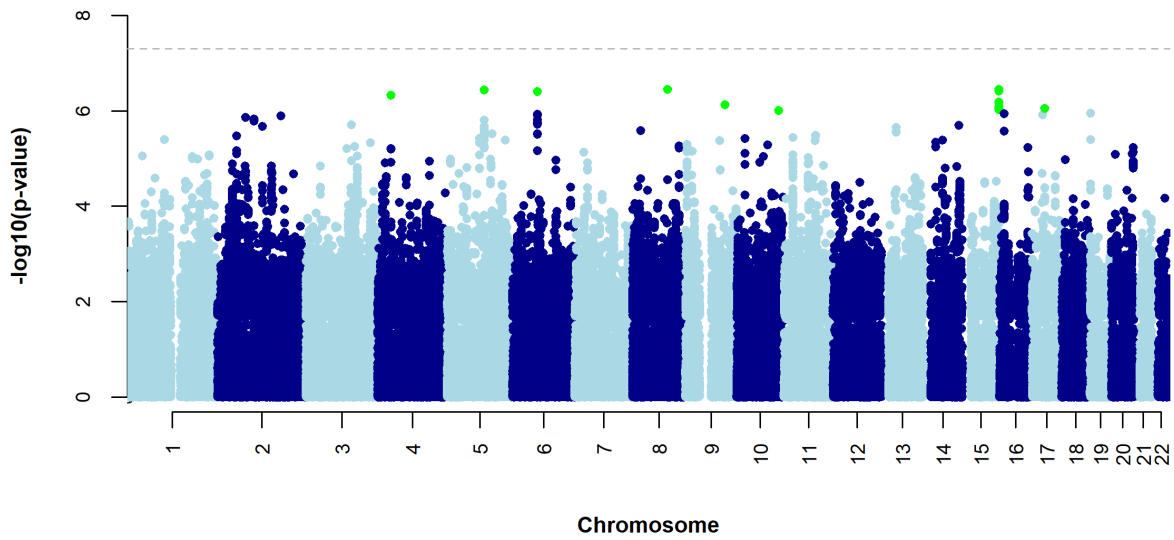


Figure S4. Manhattan plot from the GWAS analysis of 6 isoxazolyl penicillin cases and 10588 controls. Manhattan plot displays the negative log of the significance for each tested variant ranked by chromosomes and position; SNPs in green have a significance level less than  $5 \times 10^{-6}$  and red have a significance level less than  $5 \times 10^{-8}$ .



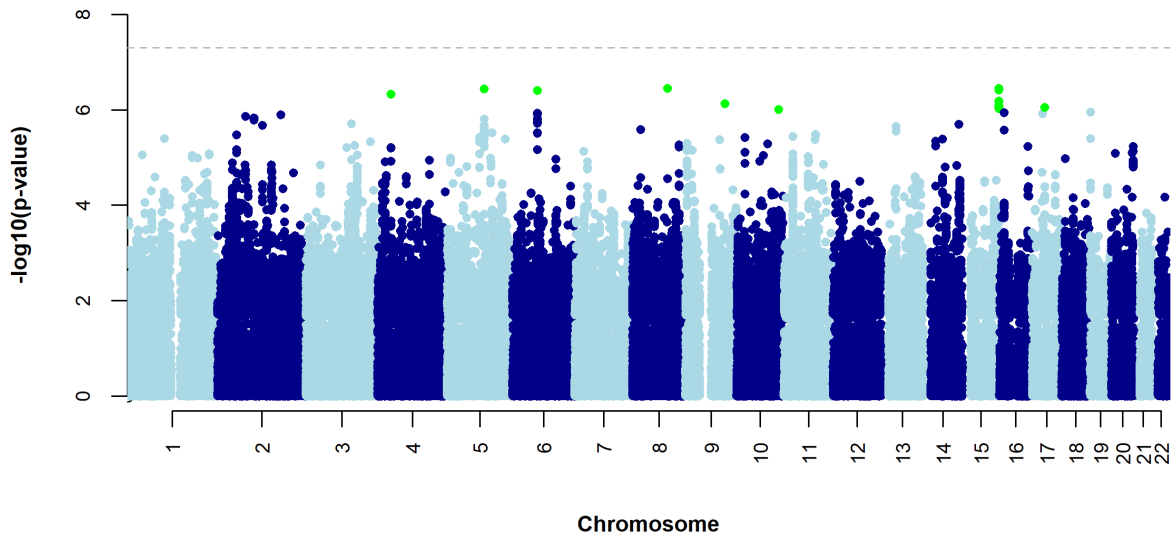


Figure S5. Manhattan plot from the GWAS analysis of 15 amoxicillin cases and 10588 controls. Manhattan plot displays the negative log of the significance for each tested variant ranked by chromosomes and position; SNPs in green have a significance level less than  $5 \times 10^{-6}$  and red have a significance level less than  $5 \times 10^{-8}$ .